Clinical Microbiology: Open Access

Kim et al., Clin Microbiol 2015, 4:5
http://dx.doi.org/10.4172/2327-5073.1000220

Research Article Open Access

Differential Diagnosis of *Brucella abortus* by Real-Time PCR based on a Single-Nucleotide Polymorphisms

Ji-Yeon Kim[1#], Sung-Il Kang[1#], Jin Ju Lee[1], Kichan Lee[1], So-Ra Sung[1], Suk Chan Jung[1], Yong Ho Park[2], Han-Sang Yoo[2] and Moon Her[1*]

[1]*Animal and Plant Quarantine and Inspection Agency, 175 Anyang-ro, Anyang-city, Gyeonggi-do, 430-757, Republic of Korea*

[2]*Seoul National University, Gwanak-ro 1, Gwanak-gu, Seoul, 151-742, Republic of Korea*

[*]***Corresponding author:*** *Moon Her, Animal and Plant Quarantine and Inspection Agency, 175 Anyang-ro, Anyang-city, Gyeonggi-do, 430-757, Republic of Korea, Tel: +82-31-467-1776; Fax: +82-31-467-1778; E-mail:* herm@korea.kr

[#]Authors contribute equally

Received date: August 12, 2015; **Accepted date:** September 17, 2015; **Published date:** September 24, 2015

Copyright: © 2015 Kim JY, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

To diagnose brucellosis effectively, many genus-and species-specific detection methods based on PCR have been developed. With conventional PCR assays, real-time PCR techniques have been developed as rapid diagnostic tools. Among them, real-time PCR using hybridization probe (hybprobe) has been recommended for bacteria with high DNA homology among species, with which it is possible to make an accurate diagnosis by means of an amplification curve and melting peak analysis. A hybprobe for *B. abortus* was designed from a specific single-nucleotide polymorphism (SNP) on the fbaA gene. This probe only showed specific amplification of *B. abortus* from approximately the 14th cycle, given a melting peak at 69°C. The sensitivity of real-time PCR was revealed to be 20 fg/µl by 10-fold DNA dilution, and the detection limit was 4 CFU in clinical samples. This real-time PCR showed greater sensitivity than that of conventional PCR and previous real-time PCR based on Taqman probe. Therefore, this new real-time PCR assay could be helpful for differentiating *B. abortus* infection with rapidity and accuracy.

Keywords: Real-time PCR; Hybprobe; SNP; fbaA gene; *Brucella abortus*

Introduction

Brucellosis is known as a major zoonotic disease that can cause reproductive problems, such as abortion, stillbirth or infertility in livestock and wild animals [1,2]. The genus *Brucella* consists of ten species; six species (*Brucella abortus*, *B. melitensis*, *B. suis*, *B. canis*, *B. ovis* and *B. neotomae*) considered classic members and four species (*B. ceti*, *B. pinnipedialis*, *B. microti*, and *B. inopinata*) considered atypical types of *Brucella*. So far, classification of *Brucella* species has been mainly based on host preferences and classical phenotypic biotyping [2,3]. Moreover, the genus expansion is still being processed, with the recent addition of *B. papionis* from baboons [3].

In terms of the diagnosis of brucellosis, serological assays and bacterial cultivation have mainly been used. Serologic methods are very sensitive and rapid methods to perform, but sometimes false-positive reactions occur with cross-reactive bacteria, such as *Yersinia enterocolitica* O:9, due to the similar structure of the O-chain in the smooth lipopolysaccharide portion [4,5]. In contrast, bacterial culture is considered a 'gold standard' with high specificity, but it is time-consuming and also requires a highly trained workforce and a well-equipped laboratory due to the biohazard risks with *Brucella* [6].

To overcome these disadvantages, molecular detection methods have been introduced as an alternative for diagnosing brucellosis. Many genus- or species-specific PCR assays, using 16S rRNA and the bcsp31, IS711, and omp2 genes, have been developed [5,7-9]. Additionally, various multiplex PCRs that can differentiate at the species level have been established [10-12]. All of these molecular detection methods are very effective for detecting *Brucella* strains [13]. Since the development of conventional PCR assays, real-time PCR and loop-mediated isothermal amplification assay (LAMP)-PCR have been introduced as rapid diagnostic tools. Recently, the application of SNPs (single nucleotide polymorphisms) in the microbiological field has shown some merits for diagnosing bacteria with high homology of their DNA [14]. SNP-based PCR assays have been introduced for differentiating bacterial strains and species [15].

Here, we developed a new real-time PCR assay with a hybprobe from a specific SNP to distinguish *B. abortus* from other *Brucella* species. Real-time PCR assay using this hybprobe could diagnose rapidly, using an amplification curve with real-time monitoring, and exactly, using melting peak analysis, so it is expected to provide more sensitive, rapid and accurate diagnostic efficiency in detecting *B. abortus* infection.

Materials and Methods

Bacterial strains and DNA samples

A total of 296 *Brucella* strains were included: 22 *Brucella* reference strains; 110 Mongolian isolates (16 *B. abortus* and 94 *B. melitensis*); 156 Korean isolates (84 *B. abortus* and 72 *B. canis*); and 8 non-*Brucella* strains reported to be serologically cross-reactive or phylogenetically similar bacteria (Table 1). *B. abortus* and *B. melitensis* from Mongolia were provided by institute of veterinary medicine (IVM), through a collaborative project conducted from 2012 to 2014. The isolates from Mongolia were *B. abortus* biovar (bv.) 3 (nine strains) and untype (seven), and *B. melitensis* bv. 1 (67), bv. 3 (10), and Rev. 1 (17). In addition, all of the Korean *B. abortus* bv. 1 was obtained from slaughtered cattle with brucellosis beginning in 2008, and the *B. canis* were from dog-breeding farms during 2002-2011. All of the *Brucella*

isolates were identified by the classical biotyping assay including colony morphology, lysis by phages, oxidase, urease activity, growth on dyes and agglutination with monospecific sera (anti-A, anti-M, and anti-R) [16] and also confirmed specific bands for *Brucella* species by the differential multiplex PCR [12]. Genomic DNA for real-time PCR was extracted using a blood and tissue kit (Qiagen Korea, Ltd.) per the manufacturer's instructions.

Species	Strains	PCR results		
		16S rRNA	BaSS	Realtime PCR
Brucella **species**				
B. abortus bv. 1 (544)	ATCC 23448	+	+	+
B. abortus bv. 2 (86/8/59)	ATCC 23449	+	+	+
B. abortus bv. 3 (Tulya)	ATCC 23450	+	±[a]	+
B. abortus bv. 4 (292)	ATCC 23451	+	+	+
B. abortus bv. 5 (B3196)	ATCC 23452	+	±[a]	+
B. abortus bv. 6 (870)	ATCC 23453	+	±[a]	+
B. abortus bv. 9 (C68)	ATCC 23455	+	±[a]	+
B. canis (RM6/66)	ATCC 23365	+	±[a]	-
B. suis bv. 1 (1330)	ATCC 23444	+	±[a]	-
B. suis bv. 2 (Thomsen)	ATCC 23445	+	±[a]	-
B. suis bv. 3 (686)	ATCC 23446	+	±[a]	-
B. suis bv. 4 (40)	ATCC 23447	+	±[a]	-
B. suis bv. 5 (513)	NCTC 11996	+	±[a]	-
B. ovis (63/290)	ATCC 25840	+	±[a]	-
B. neotomae (5K33)	ATCC 23459	+	±[a]	-
B. melitensis bv. 1 (16M)	ATCC 23456	+	±[a]	-
B. melitensis bv. 2 (63/9)	ATCC 23457	+	±[a]	-
B. melitensis bv. 3 (Ether)	ATCC 23458	+	±[a]	-
B. ceti (B1/94)	NCTC 12891	+	±[a]	-
B. pinnipedialis (B2/94)	NCTC 12890	+	±[a]	-
B. microti (CCM 4915)	BCCN 07-01	+	±[a]	-
B. inopinata (B01)	BCCN 09-01	+	±[a]	-
Mongolian *B. abortus*	16 isolates	+	±[a]	+
Mongolian *B. melitensis*	94 isolates	+	±[a]	-
Korean *B. abortus*	84 isolates	+	+	+
Korean *B. canis*	72 isolates	+	±[a]	-
Non-*Brucella* organisms				
Ochrobactrum anthropi	Field strain	+	-	-
Escherichia coli O157:H7	Field strain	-	-[b]	-
Pasteurella multocida	ATCC 43017	-	-	-
Salmonella typhimurium	ATCC 14028	-	-	-

Campylobacter jejuni	ATCC 33560	-	-	-
Yersinia enterocolitica O:9	NCTC 11174	-	-	-
Staphylococcus aureus	Field strain	+	-	-
Clostridium perfringens type A	ATCC 13124	-	-	-
±[a]: They were identified with *Brucella* spp. and showed two amplified products (180 bp and 800 bp).				

Table 1: Bacterial strains used in the present study and comparison of the two conventional PCR methods.

Hybprobe design and real-time PCR conditions

To develop *B. abortus*-specific real-time PCR assays, comparative sequence analysis, using fbaA gene region in whole genome sequences and/or partial sequences of 22 *Brucella* reference strains, was performed with the CLC Main Workbench software program version 6.0 (Insillicogen Inc., Aarhus N, Denmark). Based on the new *B. abortus*-specific SNP sites, the primer and probe sets were designed and developed using BEACON designer (Sigma-Aldrich, St. Louis, MO, USA).

Real-time PCR with hybprobe was performed using 4.0 µl of 5x genotyping master mix, 0.5 µl of each primer, 0.3 µl of each hybprobe, 13.4 µl of DW, and 1.0 µl of DNA in a 20 µl total volume. After centrifugation for the removal of bubbles from the PCR plate, amplification and melting curve analysis were conducted using a LightCycler® 480 II (Roche Diagnostic, Mannheim, Germany). The real-time PCR amplification was performed with an initial denaturation step of 95°C for 5 min, followed by 45 cycles of 95°C for 10 s, 65°C for 15 s, and 68°C for 15 s. After amplification, melting analysis was performed at between 40°C and 80°C at a rate of increase of 0.1°C.

Specificity and sensitivity of real-time PCR assay

The specificity of real-time PCR assay using 22 *Brucella* reference strains, *Brucella* isolates and non-*Brucella* bacteria was assessed (Table 1). Its sensitivity was determined from a DNA concentration of 1 ng/µl to 1 fg/µl by serial 10-fold dilution of the *B. abortus* 544 reference strain. DNA concentration was measured using a Nanodrop ND-1000 UV/UVS spectrophotometer (Nanodrop Tech., USA). These results were compared with those of a 16S rRNA [13] and BaSS-PCR assays [9], which were used to identify *Brucella* species and *B. abortus* biovars 1, 2 and 4 conventionally.

Detection limits of real-time PCR assay

To compare the analytical sensitivity of real-time PCR assay in the clinical specimens, artificial inoculation using a *B. abortus* strain in the clinical samples was conducted. Briefly, ten-fold serial dilutions of the *B. abortus* strain with 0.85% saline were processed into the macerated lymphoid tissue, and then each spiked sample was cultivated on three tryptic soy agars and was calculated in colony-forming units (CFU). The DNA of the spiked samples was extracted using a commercial blood and tissue kit (Qiagen Ltd., South Korea) according to the manufacturer's protocols.

Evaluation of real-time PCR assay

To apply real-time PCR assay to the clinical specimens, twelve samples (supramammary, submandibular, inguinal, and parotid lymph node, testicle and buffy coat) were acquired from seropositive Korean native cattle on a breeding farm. These specimens were ground in 1 ml of PBS buffer and spread onto tryptic soy agar supplemented with 5% bovine serum (GIBCO, Grand Island, NY, USA) and 5% sheep blood agar for 3 days-4 days at 37°C and 5% CO_2. Genomic DNA from 200 µl of ground sample was extracted using a Blood & Tissue kit (Qiagen Ltd., South Korea) per the manufacturer's instructions and was submitted to real-time PCR assay.

Results

B. abortus-specific SNPs were detected at the fbaA gene of *B. abortus* chromosome II (Genbank accession No. AE 017224), with cytosine changed to thymine at 360432 on *B. abortus* chromosome II. Based on the sequence of the fbaA gene, a primer set 176 bp in size and a hybprobe with a specific SNP were designed (Table 2).

		Sequence (5'-3')	Comment
Primer	Forward	GATGCGCCGGTTATCCTG	176 bp size
	Reverse	GTGAAGCCCGCCTGGATG	
Probe	Anchor	ACGGAGCATGATGTCATTGGCATAGGAACG	*SNP site
	Sensor	ATAGATTTCCGC*CACGGCATCCATC	

Table 2: Primers and probes for detection of *B. abortus* using specific SNPs.

Real-time PCR assay showed a positive reaction only to *B. abortus* reference strains (biovars 1-6, and 9) and *B. abortus* organisms from clinical specimens, whereas it yielded negative reactions to other *Brucella* species and non-*Brucella* bacterial strains (Table 1 and Figure 1). Only *B. abortus* species showed specific amplification from approximately the 14th cycle (Figure 1a). Additionally, the specific amplification was also confirmed by melting curve analysis. The Tm calling value of *B. abortus* reference strains and isolates was generated in 69°C. In contrast with this finding, other *Brucella* species and non-*Brucella* strains showed less than a low melting peak at a temperature of 62°C (Figure 1b). In the 16S rRNA PCR, some bacterial strains, such as *Ochrabactrum anthropi* and *Staphlyococcus aureus*, were diagnosed as positive reactions with the amplified product of 905 bp. BaSS-PCR showed different results depending on the biovar of *B. abortus*. In case of *B. abortus* bv. 1, 2 and 4, three specific bands (180 bp, 500 bp and 800 bp) appeared, but other bvs. of *B. abortus* and other *Brucella* species showed only two bands (180 bp and 800 bp), and non-*Brucella*

bacteria revealed only an 800 bp-band product as an internal control (Table 1).

Figure 1: ***(a)*** Amplification curves and ***(b)*** melting peak analysis in *B. abortus* 544 reference strain and Korean *B. abortus* isolates.

The sensitivity of real-time PCR was assessed by means of successive 10-fold serial dilution of the genomic DNA of the *B. abortus* 544 reference strain, and it was revealed to be 20 fg/µl (data not shown). In addition, the detection limit for *B. abortus* in the clinical samples was 4 CFU/µl. In contrast, the detection limit of conventional BaSS-PCR showed 80 CFU/ µl. Our new real-time PCR revealed 20 times greater sensitivity than that of BaSS-PCR (Figure 2) but equal sensitivity to 16S rRNA PCR (data not shown).

Figure 2: ***(a)*** Detection limits of the hybridization probe-based real-time PCR and ***(b)*** BaSS-PCR determined by DNA extracted from lymphoid tissue inoculated with 10-fold diluted *B. abortus* strains serially. ***(a)*** There were ranged from 4×10^6 to 4×10^{-2} CFU/µl. ***(b)*** M: 100-bp DNA ladder, lane 1 to 7: 8×10^4 to 8×10^{-2} CFU/µl, lane 8 and 9: internal and negative control (DW).

In addition, the application of the real-time PCR to the clinical specimens was conducted using brucellosis-positive Korean native cattle. Here, *B. abortus* was isolated from tissue samples, such as various lymph nodes and buffy coats. All of the samples were confirmed as positive by generating a fluorescent signal during real-time PCR (Table 3). The range of mean Ct ranged between 28 and 30, except for buffy coat, and the Tm values were almost identical to the reference *B. abortus* 544 strain.

Sample no.	Specimen	Isolation[a]	RBT[b]	Real-time PCR	
				Mean Ct	Tm
1	Buffy coat	+	+	16.89	69.43
2	Buffy coat	+	+	16.88	69.41
3	Calf submandibular LN	+	NT[c]	29.24	69.23
4	Submandibular LN-1	+	+	28.45	69.46
5	Submandibular LN-2	+	+	29.28	69.15
6	Submandibular LN-3	+	+	29.74	69.42
7	Submandibular LN-4	+	+	28.79	69.38
8	Submandibular LN-5	+	+	29.13	69.08
9	Supramammary LN	+	+	28.89	69.40
10	Calf inguinal LN	+	NT[c]	29.29	69.06
11	Parotid LN	+	-	29.27	69.13
12	Testicle	+	+	28.65	69.34

[a]Isolation: The bacterial strains were identified by classical phenotyping and differential multiplex PCR [12]. [b]RBT: Rose-Bengal Test. [c]NT: Not Tested.

Table 3: Direct detection from clinical specimens of brucellosis-positive Korean native cattle by real-time PCR.

Discussion

For decades, PCR-based assays have been developed continually as a form of real-time PCR. It is able to detect target microorganisms more sensitively, specifically and rapidly than conventional PCRs [5,17]. Unlike endpoint detection methods, such as agarose gel electrophoresis, real-time PCR is used for the quantitative measurement of amplified products using fluorescence during each PCR cycle. These reactions can be classified into two main types according to the fluorescent dye and the specificity of the PCR [18]. The former uses double-stranded DNA-intercalating dye (e.g., SYBR Green I), and the latter uses fluoroephores that bind to oligonucleotides. This type can be divided into three subtypes depending on the fluorescent molecules: ***(i)*** primer-probes; ***(ii)*** hydrolysis and hybridization probes; and ***(iii)*** analogs of nucleic acids [18].

First, SYBR Green I, as one of the most commonly used DNA-binding dyes, binds to total amounts of DNA generated during PCR, so it can induce specific and non-specific amplification [18,19]. At the same time, Taqman probe is a representative hydrolysis type, and it is designed to bind to a specific site of the target DNA, so it shows greater specificity than SYBR Green I. However, it has the disadvantage that the primer-dimer can be generated even if the primer design is not appropriate. Contrarily, hybprobe-based real-time PCR offers two main advantages: firstly, it requires two additional probes for binding,

so it can show improved specificity to distinguish between closely related strains; secondly, hybprobe does not rely on the hydrolysis reaction, so melting curve analysis can be applied to differentiate based on the probe Tm [20,21]. Thus, its application has increased in various fields, such as pathogen detection, SNP detection, and so on [18,22].

In particular, the application of SNPs in microbial molecular typing has been increasing in the diagnostic field. SNPs in the conserved region can be very strong markers for detecting and differentiating etiological agents specifically. Therefore, we designed a hybprobe from another *B. abortus*-specific SNP in the conserved fbaA gene, although this gene had been already used in real-time PCR with Taqman probe [23].

With regard to specificity, only *B. abortus* strains revealed specific amplification curves from the 14th cycle, and Tm was 69°C in our new real-time PCR. Not only other *Brucella* species but also highly genetically and serologically related bacteria were not amplified. Especially, it yielded a negative reaction from two cross-reactive bacteria by 16S rRNA PCR-O. anthropi and *S. aureus* [24,25]. In terms of sensitivity using *B. abortus* DNA, our real time PCR assay was equal to or higher than that of 16S rRNA PCR [13]. Application of the spiking test on *B. abortus* in our study showed results that were 20-times higher than those with BaSS-PCR (Figure 2). Moreover, BaSS-PCR has the limitation of detecting *B. abortus* bv. 1, 2 and 4 only, so *B. abortus* infection caused by other biovars could not be differentiated. Therefore, our new real-time PCR could be valuable for diagnosing *B. abortus* infection in terms of its accuracy, specificity and sensitivity.

Besides two conventional PCR assays, our new PCR showed improved analytical sensitivity, compared to other real-time PCR assays. Using serially diluted DNA samples, our assay revealed 20 fg, but two previous studies using 5'-nuclease IS711 yielded 150 fg and 250 fg, respectively [26,27]. Additionally, previous real-time PCR based on the same fba gene reported sensitivity of 50 fg or 15 cells [23], which was lower than in our study (20 fg or 4 CFU). However, Bounaadja et al. [4] reported a 10-times higher detection limit than our assay of 2 fg using three genes (IS711, bcsp31 and per gene), but it was not from clinical samples only from extracted DNA samples. As with clinical specimens, our real-time PCR showed identical results to bacterial isolation with high specificity.

Because the genus *Brucella* is an intracellular bacterium, and the number of bacteria in specimens is usually small [17], a highly sensitive diagnostic technique is required for accurate differential diagnosis. This new real-time PCR could be very useful for directly diagnosing brucellosis caused by *B. abortus* in infected animals due to the high detection limit. In conclusion, our new real-time PCR based on hybprobe could be an efficient diagnostic technique with high sensitivity and rapidity for *B. abortus*-infected animals in the field, and it could also be applicable in public health.

Conflict of Interests

The authors declare that they have no conflict of interest.

Acknowledgements

This study was supported by a grant from the Animal and Plant Quarantine Agency (QIA) of the Ministry of Agriculture, Food and Rural Affairs (MAFRA) of the Republic of Korea during 2012-2014.

References

1. Winchell JM, Wolff BJ, Tiller R, Bowen MD, Hoffmaster AR (2010) Rapid identification and discrimination of *Brucella* isolates by use of real-time PCR and high-resolution melt analysis. J Clin Microbiol 48: 697-702.
2. Gopaul KK, Sells J, Lee R, Beckstrom-Sternberg SM, Foster JT, et al. (2014) Development and assessment of multiplex high resolution melting assay as a tool for rapid single-tube identification of five *Brucella* species. BMC Res Notes 7: 903.
3. Whatmore AM, Davison N, Cloeckaert A, Al Dahouk S, Zygmunt MS, et al. (2014) *Brucella* papionis sp. nov., isolated from baboons (Papio spp.). Int J Syst Evol Microbiol 64: 4120-4128.
4. Bounaadja L, Albert D, Chénais B, Hénault S, Zygmunt MS, et al. (2009) Real-time PCR for identification of *Brucella* spp.: a comparative study of IS71, bcsp31 and per target genes. Vet Microbiol 137: 156-164.
5. Hinić V, Brodard I, Thomann A, Holub M, Miserez R, et al. (2009) IS711-based real-time PCR assay as a tool for detection of *Brucella* spp. in wild boars and comparison with bacterial isolation and serology. BMC Vet Res 5: 22.
6. Wareth G, Melzer F, Elschner MC, Neubauer H, Roesler U (2014) Detection of *Brucella* melitensis in bovine milk and milk products from apparently healthy animals in Egypt by real-time PCR. J Infect Dev Ctries 8: 1339-1343.
7. Ficht TA, Bearden SW, Sowa BA, Marquis H (1990) Genetic variation at the omp2 porin locus of the *Brucella*e: species-specific markers. Mol Microbiol 4: 1135-1142.
8. Cloeckaert A, Verger JM, Grayon M, Grépinet O (1995) Restriction site polymorphism of the genes encoding the major 25 kDa and 36 kDa outer-membrane proteins of *Brucella*. Microbiology 141 : 2111-2121.
9. Bricker BJ, Ewalt DR, Olsen SC, Jensen AE (2003) Evaluation of the *Brucella* abortus species-specific polymerase chain reaction assay, an improved version of the *Brucella* AMOS polymerase chain reaction assay for cattle. J Vet Diagn Invest 15: 374-378.
10. López-Goñi I, García-Yoldi D, Marín CM, de Miguel MJ, Muñoz PM, et al. (2008) Evaluation of a multiplex PCR assay (Bruce-ladder) for molecular typing of all *Brucella* species, including the vaccine strains. J Clin Microbiol 46: 3484-3487.
11. Mayer-Scholl A, Draeger A, Göllner C, Scholz HC, Nöckler K (2010) Advancement of a multiplex PCR for the differentiation of all currently described *Brucella* species. J Microbiol Methods 80: 112-114.
12. Kang SI, Her M, Kim JW, Kim JY, Ko KY, et al. (2011) Advanced multiplex PCR assay for differentiation of *Brucella* species. Appl Environ Microbiol 77: 6726-6728.
13. Romero C, Gamazo C, Pardo M, López-Goñi I (1995) Specific detection of *Brucella* DNA by PCR. J Clin Microbiol 33: 615-617.
14. Scott JC, Koylass MS, Stubberfield MR, Whatmore AM (2007) Multiplex assay based on single-nucleotide polymorphisms for rapid identification of *Brucella* isolates at the species level. Appl Environ Microbiol 73: 7331-7337.
15. McDonald M, Dougall A, Holt D, Huygens F, Oppedisano F, et al. (2006) Use of a single-nucleotide polymorphism genotyping system to demonstrate the unique epidemiology of methicillin-resistant Staphylococcus aureus in remote aboriginal communities. J Clin Microbiol 44: 3720-3727.
16. OIE (2012) Terrestrial Manual:Bovine Brucellosis. World organization for animal health.
17. Surucuoglu S, El S, Ural S, Gazi H, Kurutepe S, et al. (2009) Evaluation of real-time PCR method for rapid diagnosis of brucellosis with different clinical manifestations. Pol J Microbiol 58: 15-19.
18. Navarro E, Serrano-Heras G, Castaño MJ, Solera J (2015) Real-time PCR detection chemistry. Clin Chim Acta 439: 231-250.
19. Smith CJ, Osborn AM (2009) Advantages and limitations of quantitative PCR (Q-PCR)-based approaches in microbial ecology. FEMS Microbiol Ecol 67: 6-20.

20. Newby DT, Hadfield TL, Roberto FF (2003) Real-time PCR detection of *Brucella* abortus: a comparative study of SYBR green I, 5'-exonuclease, and hybridization probe assays. Appl Environ Microbiol 69: 4753-4759.
21. Osińska E, Golke A, Słońska A, Cymerys J, Bańbura MW, et al. (2012) HybProbes-based real-time PCR assay for rapid detection of equine herpesvirus type 2 DNA. Pol J Vet Sci 15: 411-416.
22. Lim SY, Kim BJ, Lee MK, Kim K (2008) Development of a real-time PCR-based method for rapid differential identification of Mycobacterium species. Lett Appl Microbiol 46: 101-106.
23. Gopaul KK, Koylass MS, Smith CJ, Whatmore AM (2008) Rapid identification of *Brucella* isolates to the species level by real time PCR based single nucleotide polymorphism (SNP) analysis. BMC Microbiol 8: 86.
24. Al-Ajlan HH, Ibrahim AS, Al-Salamah AA (2011) Comparison of different PCR methods for detection of *Brucella* spp. in human blood samples. Pol J Microbiol 60: 27-33.
25. Horvat RT, El Atrouni W, Hammoud K, Hawkinson D, Cowden S (2011) Ribosomal RNA sequence analysis of *Brucella* infection misidentified as Ochrobactrum anthropi infection. J Clin Microbiol 49: 1165-1168.
26. Probert WS, Schrader KN, Khuong NY, Bystrom SL, Graves MH (2004) Real-time multiplex PCR assay for detection of *Brucella* spp., B. abortus, and B. melitensis. J Clin Microbiol 42: 1290-1293.
27. Redkar R, Rose S, Bricker B, DelVecchio V (2001) Real-time detection of *Brucella* abortus, *Brucella* melitensis and *Brucella* suis. Mol Cell Probes 15: 43-52.