



Clinical Relevance of High-Resolution Single Nucleotide Polymorphism Array in Patients with Relapsed Acute Lymphoblastic Leukemia with Normal Karyotype: A Report of Three Cases

Sang Hyuk Park, M.D.^{1,2,*}, Seung-Hee Lee, M.D.^{1,*}, Shine Young Kim, M.D.¹, Sun Min Lee, M.D.³, Jongyoun Yi, M.D.¹, In-Suk Kim, M.D.³, Hyung Hoi Kim, M.D.^{1,2}, Chulhun Ludgerus Chang, M.D.³, and Eun Yup Lee, M.D.^{1,2}

Department of Laboratory Medicine¹, Pusan National University School of Medicine, Pusan National University Hospital, Busan; Biomedical Research Institute², Pusan National University Hospital, Busan; Department of Laboratory Medicine³, Pusan National University School of Medicine, Pusan National University Yangsan Hospital, Yangsan, Korea

We report three patients with normal karyotype (NK) ALL, who showed genetic aberrations as determined by high-resolution single nucleotide polymorphism array (SNP-A) analysis at both diagnosis and relapse. We evaluated the clinical relevance of the SNP-A assay for the detection of subtle changes in the size of affected genetic lesions at relapse as well as the prognostic value of the assay. In our patients, application of the SNP-A assay enabled sensitive detection of cryptic changes affecting clinically important genes in NK ALL. Therefore, this assay seems to be more advantageous compared to other conventional methods such as FISH assay, HemaVision (DNA Technology, Denmark), and conventional karyotyping for the detection of an “unstable genotype” at relapse, which may be associated with microscopic clonal evolution and poor prognosis. Further comprehensive studies are required to confirm the issues presented by our case patients in this report.

Key Words: Acute lymphoblastic leukemia, Array, Clonal evolution, Normal karyotype, Prognosis, Single nucleotide polymorphism

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Corresponding author: Eun Yup Lee
Department of Laboratory Medicine and
Biomedical Research Institute, Pusan
National University School of Medicine,
Pusan National University Hospital, 179
Gudeok-ro, Seo-gu, Busan 602-739, Korea
Tel: +82-51-240-7419
Fax: +82-51-247-6560
E-mail: eylee@pusan.ac.kr

*These two authors contributed equally to this work.

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INTRODUCTION

A genome-wide single nucleotide polymorphism array (SNP-A) assay can detect not only the copy number alterations but also the copy-neutral loss of heterozygosity (CN-LOH) on a chromosomal segment with high sensitivity, as demonstrated in MDS/MPN and NK (normal karyotype) AML [1-5]. However, no study has evaluated SNP-A assay in NK ALL, except for one that reported detection of copy number alterations and CN-LOH by

SNP-A in 62% of NK ALL patients as well as smaller deletions compared with those in other leukemia subtypes [6]. In addition, the clinical relevance of unstable genotype (subtle changes in the size of detected alterations at relapse) has not been evaluated. We report three patients with NK ALL who harbored genomic alterations, as detected by a high-resolution SNP-A assay at both diagnosis and relapse. We also assessed the clinical relevance of slight changes in the size of affected genetic lesions at relapse with the maintenance of normal cytogenetics.

CASE REPORT

1. Case 1

A 38-yr-old man was diagnosed as having NK ALL on the basis of the results of negative HemaVision (DNA Technology; Aarhus, Denmark) and metaphase cytogenetics (MC) analysis. He was diagnosed as having B-cell ALL with NK and achieved complete remission (CR) at 10 weeks of treatment. He experienced relapse at 8 weeks after CR and, at relapse, his karyotype was maintained as normal and HemaVision results were also negative.

A genome-wide SNP-A assay was performed by using the Affymetrix CytoGenetics 2.7M Array (Affymetrix; Santa Clara, CA, USA) with bone marrow samples obtained both at diagnosis and relapse. At diagnosis, SNP-A analysis revealed homozygous deletions of 9p21.3 (associated with *CDKN2A*) and 13q14.2 (associated with *RB1*) and heterozygous deletions of 6p22.2, 8q22.1, 9p21.3p21.2, 14q11.2, 18q22.2, and 19p13.2 (Fig. 1A, B). FISH analysis for the detection of *RB1* deletions showed nuc ish (RB1,13q14)×2[200] (Fig. 1C). An identical assay performed at relapse showed loss of heterozygous deletions in 6 affected lesions along with subtle size changes in deleted lesions of 9p21.3 (1,944-1,752 kb) and 13q14.2 (96-89 kb), indicating the persistence of *CDKN2A* and *RB1* deletion at relapse (Fig. 1D, E). FISH analysis for the detection of *RB1* deletions also

showed nuc ish (RB1,13q14)×2[200] (Fig. 1F).

2. Case 2

A 57-yr-old woman was diagnosed as having NK ALL on the basis of the results of HemaVision (negative) and MC analysis. SNP-A analysis at diagnosis revealed heterozygous interstitial deletions of 3q13, 4q23q24, 5q15q21.3, 5q21.3q22.1, 5q22.3q23.1, 5q34, 6q16.3q21, 7q11.23, and 13q14.2q14.3, including *RB1* (Fig. 2A). She achieved CR after 4 weeks of treatment but experienced relapse at 6 months after CR. At relapse, her karyotype was normal and HemaVision results were also negative. Results of SNP-A analysis at relapse showed heterozygous deletions of 3q13, 4q23q24, 5q15q21.3, 5q21.3q22.1, 5q22.3q23.1, 5q34, 6q16.3q21, 7q11.23, and 13q14.2q14.3, including *RB1* (Fig. 2B), which is similar to those at diagnosis. However, the deleted genomic lesions of 3q13, 4q23q24, 5q15q21.3, 5q21.3q22.1, 5q34, and 13q14.2q14.3 at relapse were slightly different from those at diagnosis.

3. Case 3

A 58-yr-old man was diagnosed as having NK ALL as per the results of HemaVision (negative) and MC analysis. He achieved CR after 4 weeks of chemotherapy but experienced relapse at follow-up 32 months after remission. At relapse, the HemaVi-

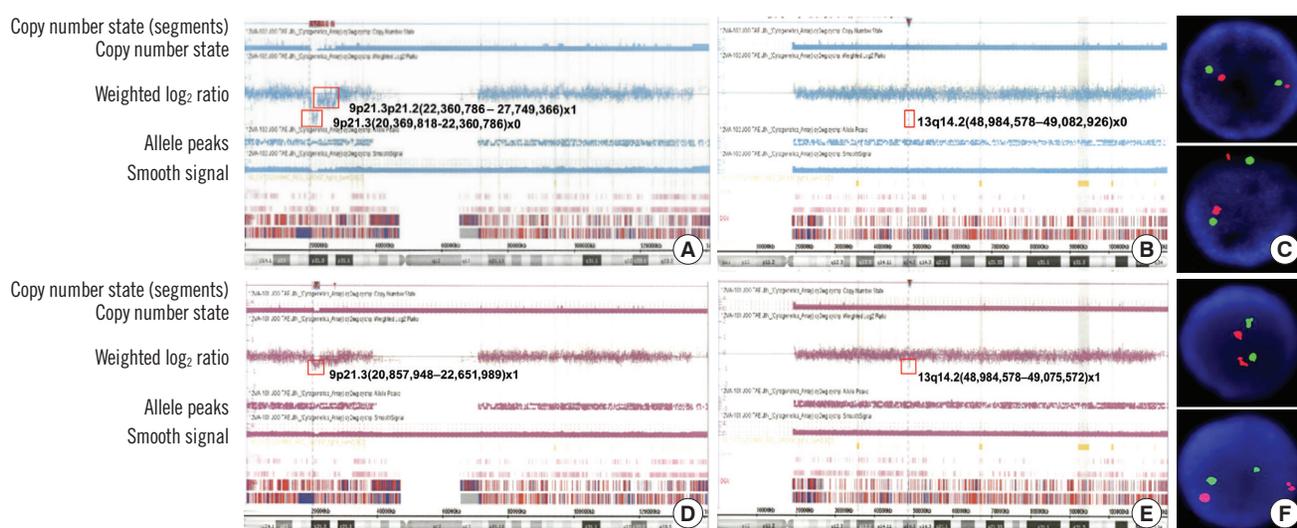


Fig. 1. High-resolution single nucleotide polymorphism array (HR SNP-A) analysis results for chromosome 9 (A) and 13 (B) performed at diagnosis in Case 1. Deleted lesions are indicated with a red box, and detailed array results in each lesion are provided. The results showed a homozygous 1,944 kb deletion of 9p21.3, including *CDKN2A*; a 96 kb deletion of 13q14.2, including *RB1*; and a heterozygous 5,262 kb deletion of 9p21.3p21.2. The FISH analysis for the detection of *RB1* deletion at diagnosis showed nuc ish (RB1,13q34)×2[200], which indicates no deletion of *RB1*(C). Identical HR SNP-A analysis results for chromosome 9 (D) and 13 (E) performed at relapse in Case 1 showed a heterozygous 1,752 kb deletion of 9p21.3, including *CDKN2A* and an 89 kb deletion of 13q14.2, including *RB1*. FISH analysis for the detection of *RB1* deletion at relapse also showed nuc ish (RB1,13q34)×2[200], which indicates no deletion of *RB1*(F).

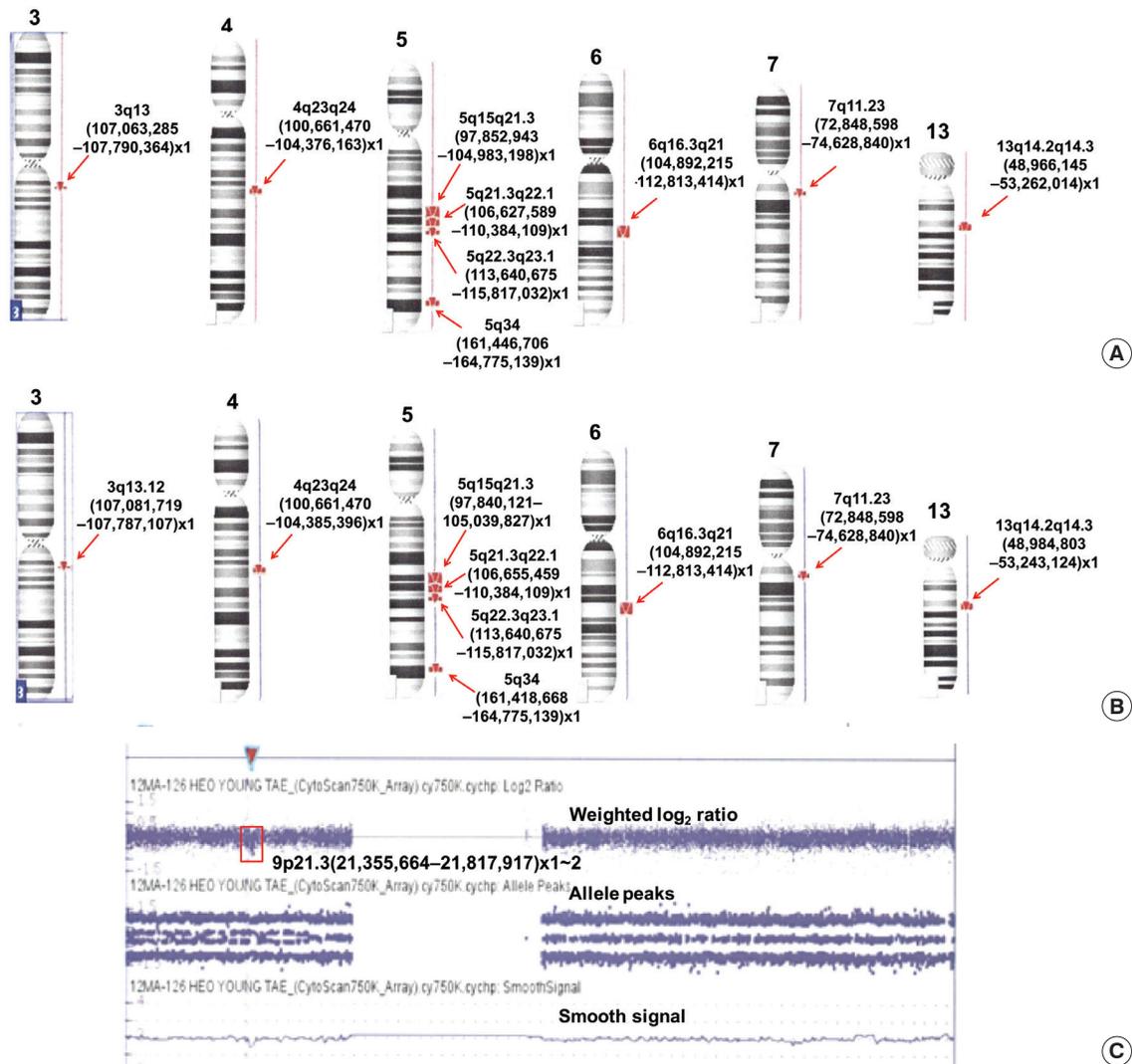


Fig. 2. High-resolution single nucleotide polymorphism array (HR SNP-A) analysis results in whole chromosomes at diagnosis (idiograms of six chromosomes harboring aberrations are provided) in Case 2 (A). The array results showed heterozygous interstitial deletions of 3q13, 4q23q24, 5q15q21.3, 5q21.3q22.1, 5q22.3q23.1, 5q34, 6q16.3q21, 7q11.23, and 13q14.2q14.3, including *RB1* gene. Deleted lesions are indicated separately with red arrows, and the detailed array results for each lesion are provided. Identical array results for whole chromosome at relapse in Case 2 (B) showed similar results, but slight size changes in deleted genomic lesions at 3q13, 4q23q24, 5q15q21.3, 5q21.3q22.1, 5q34, and 13q14.2q14.3 were identified at relapse compared to those at diagnosis. Deleted lesions are also indicated separately with red arrows, and detailed array results for each lesion are provided. In addition, HR SNP-A analysis results of chromosome 9 performed at both diagnosis and relapse in Case 3 (C) are provided. The deleted lesion is indicated with a red box, and detailed array result is provided. The array result showed an interstitial 451 kb deletion of 9p21.3, which includes *IFNA13* and *MIR31* in a mosaic pattern.

sion results were also negative. However, SNP-A analysis demonstrated an interstitial 451 kb deletion of 9p21.3, which includes interferon α -1/13 (*IFNA13*) and micro-RNA 31 (*MIR31*) in a mosaic pattern, at both diagnosis and relapse (Fig. 2C). Demographic findings of the deleted genetic lesions and the affected genes associated with ALL in the three cases are represented in Table 1.

DISCUSSION

In Case 1, SNP-A analysis could detect the presence of interstitial microdeletion including both *RB1* and *CDKN2A*, which is not otherwise detectable by MC or FISH analysis. Because the deletions of *CDKN2A* and *RB1* are associated with poor prognosis in ALL [7, 8], this may suggest that SNP-A assay can provide prognostic information in NK ALL. The SNP-A assay could detect

Table 1. Characteristics of abnormal lesions detected by high-resolution single nucleotide polymorphism array (SNP-A) assay in 3 cases

Case No.	Stage	Chromosome	Affected point (Start)*	Affected point (End)*	Deletion size (kb)	Affected genes associated with ALL†	
1	Diagnosis	6p22.2	26,174,861	26,566,470	382	None	
		8q22.1	95,054,989	95,486,831	422	None	
		9p21.3	20,369,818	22,360,786	1,944	<i>CDKN2A</i>	
		9p21.3p21.2	22,360,786	27,749,366	5,262	None	
		13q14.2	48,984,578	49,082,926	96	<i>RB1</i>	
		14q11.2	22,447,554	23,017,964	557	<i>TRA, TRD</i>	
		18q22.2	66,977,443	67,361,947	375	None	
		19p13.2	12,519,413	12,738,738	214	None	
	Relapse	9p21.3	20,857,948	22,651,989	1,752	<i>CDKN2A</i>	
	13q14.2	48,984,578	49,075,572	89	<i>RB1</i>		
2	Diagnosis	3q13	107,063,285	107,790,364	710	None	
		4q23q24	100,661,470	104,376,163	3,628	None	
		5q15q21.3	97,852,943	104,983,198	6,963	None	
		5q21.3q22.1	106,627,589	110,384,109	3,668	None	
		5q22.3q23.1	113,640,675	115,817,032	2,125	None	
		5q34	161,446,706	164,775,139	3,250	None	
		6q16.3q21	104,892,215	112,813,414	7,736	None	
		7q11.23	72,848,598	74,628,840	1,739	None	
		13q14.2q14.3	48,966,145	53,262,014	4,195	<i>RB1</i>	
		Relapse	3q13.12	107,081,719	107,787,107	689	None
			4q23q24	100,661,470	104,385,396	3,637	None
			5q15q21.3	97,840,121	105,039,827	7,031	None
			5q21.3q22.1	106,655,459	110,384,109	3,641	None
			5q22.3q23.1	113,640,675	115,817,032	2,125	None
			5q34	161,418,668	164,775,139	3,278	None
			6q16.3q21	104,892,215	112,813,414	7,736	None
		7q11.23	72,848,598	74,628,840	1,739	None	
	13q14.2q14.3	48,984,803	53,243,124	4,159	<i>RB1</i>		
3	Diagnosis	9p21.3	21,355,664	21,817,917	451	None	
	Relapse	9p21.3	21,355,664	21,817,917	451	None	

*The locations of affected genetic lesions were aligned using the human genome browser–hg19 assembly ([http:// genome.ucsc.edu/cgi-bin/hgGateway](http://genome.ucsc.edu/cgi-bin/hgGateway)),

†The list of affected genes associated with ALL within each genetic lesion was determined from the review of a web-based database ([http:// atlasgeneticsoncology.org/index.html](http://atlasgeneticsoncology.org/index.html)) and browser (<http:// genome.ucsc.edu/cgi-bin/hgGateway>).

Abbreviations: *CDKN2A*, cyclin-dependent kinase 2a/p16; *RB1*, retinoblastoma; *TRA*, T cell receptor α ; *TRD*, T cell receptor δ .

subtle size changes in the affected lesions (microscopic clonal evolution) and recovery of small-sized interstitial deletions at relapse, which are also not detectable by MC. Thus, the SNP-A assay is more advantageous in detecting microscopic clonal evolution than other methods. In addition, in Case 2, SNP-A assay could detect subtle changes in the size of the affected genetic lesions at relapse (unstable genotype). We may speculate that unstable genotype may indicate microscopic clonal evolu-

tion at relapse. The early relapse in Case 2 patient supports this speculation.

The association between ALL and the loss of *IFNA13* and *MIR31* is unclear. However, as abnormal 9p is an adverse prognostic factor for B-ALL [9], the interstitial deletion of 9p21.3 detected by SNP-A analysis may contribute to poor prognosis, as demonstrated by early relapse in Case 3.

Our report has some limitations. First, the SNP-A assay at re-

lapse in Case 2 did not show any additional abnormalities and slight size changes in the detected genetic lesions as evidence of clonal evolution have not been demonstrated. Therefore, the presence of unstable genotype should be regarded only as a phenomenon suggesting various events, including clonal evolution. Second, we could neither perform SNP-A analysis using samples with CR or fibroblasts, which is important for the discrimination of acquired somatic events from germline aberrations, nor confirm whether all genetic abnormalities detected in our cases were somatic. Third, we could not perform FISH analysis in *RBI* in Case 2 as well as detailed evaluations of the discrepancies between the results of FISH and SNP-A analysis. The discrepancy in the results of FISH and SNP-A analysis regarding the *RBI* gene in Case 1 may be explained from different detection sensitivity of both methods. However, more comprehensive analysis is required for comprehending this discrepancy.

In conclusion, our case report demonstrates that a SNP-A assay can allow sensitive detection of cryptic changes affecting clinically important genes. A SNP-A assay may be more advantageous than other methods in detecting unstable genotype at relapse, which may be associated with microscopic clonal evolution and poor prognosis in ALL.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

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