Selective Small Molecule Induced Degradation of the BET Bromodomain Protein BRD4

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Supporting Information

ABSTRACT: The Bromo- and Extra-Terminal (BET) proteins BRD2, BRD3, and BRD4 play important roles in transcriptional regulation, epigenetics, and cancer and are the targets of pan-BET selective bromodomain inhibitor JQ1. However, the lack of intra-BET selectivity limits the scope of current inhibitors as probes for target validation and could lead to unwanted side effects or toxicity in a therapeutic setting. We designed Proteolysis Targeted Chimeras (PROTACs) that tether JQ1 to a ligand for the E3 ubiquitin ligase VHL, aimed at triggering the intracellular destruction of BET proteins. Compound MZ1 potently and rapidly induces reversible, long-lasting, and unexpectedly selective removal of BRD4 over BRD2 and BRD3. The activity of MZ1 is dependent on binding to VHL but is achieved at a sufficiently low concentration not to induce stabilization of HIF-1α. Gene expression profiles of selected cancer-related genes responsive to JQ1 reveal distinct and more limited transcriptional responses induced by MZ1, consistent with selective suppression of BRD4. Our discovery opens up new opportunities to elucidate the cellular phenotypes and therapeutic implications associated with selective targeting of BRD4.

T he Bromo- and Extra-terminal (BET) family of proteins, including the ubiquitously expressed BRD2, BRD3, and BRD4 and the testis-specific BRDT, recruit transcriptional regulatory complexes to acetylated chromatin thereby controlling specific networks of genes involved in cellular proliferation and cell cycle progression.1 Deregulation of BET protein activity, in particular BRD4, has been strongly linked to cancer and inflammatory diseases, making BET proteins attractive drug targets.2 For example, RNAi screens have identified BRD4 as a therapeutic target in acute myeloid leukemia,3 ovarian carcinoma,4 and siRNA knock down of BRD4, but not of BRD2 or BRD3, induced upregulation of apolipoprotein A1 (ApoA1), aimed at triggering the intracellular destruction of BET proteins. Compound MZ1 potently and rapidly induces reversible, long-lasting, and unexpectedly selective removal of BRD4 over BRD2 and BRD3. The activity of MZ1 is dependent on binding to VHL but is achieved at a sufficiently low concentration not to induce stabilization of HIF-1α. Gene expression profiles of selected cancer-related genes responsive to JQ1 reveal distinct and more limited transcriptional responses induced by MZ1, consistent with selective suppression of BRD4. Our discovery opens up new opportunities to elucidate the cellular phenotypes and therapeutic implications associated with selective targeting of BRD4.
small molecule inhibitor over a prolonged time. This in turn enhances off-target effects and can lead to unwanted side effects or toxicity in a therapeutic setting. To provide an alternative small molecule approach that could address these issues, we hypothesized that it would be possible to design a molecule that can remove BET proteins entirely from the cell as opposed to just inhibiting them, yielding new tools for studying BET bromodomain proteins and validating them as drug targets.

In order to achieve intracellular BET-protein degradation, we applied a small molecule PROTAC (Proteolysis Targeting Chimera) approach. PROTACs are heterobifunctional compounds that contain two ligands connected by a linker unit. One ligand binds an E3 ubiquitin ligase protein, while the other ligand binds to the target protein of interest, thereby bringing the ligase and the target in close proximity. This in turn triggers the polyubiquitination and subsequent proteasome-dependent degradation of the target. Proof-of-concept examples have been described where PROTACs were used to degrade the estrogen20- and androgen-receptor,21 methionine aminopeptidase-2,22 as well as the aryl hydrocarbon receptor.23 However, all first-generation PROTACs included a peptidic moiety as the E3 ligase ligand. For example, a hydroxyproline-containing heptapeptide sequence ALA-Hyp-YIP from the transcription factor Hypoxia-Inducible Factor 1 alpha subunit (HIF-1α) has been widely used,24 as this represents the minimal epitope for HIF-1α binding to the ubiquitously expressed E3 ligase von Hippel-Lindau protein (VHL).25 The high peptidic nature of the first-generation PROTACs resulted in poor physicochemical properties such as low intracellular stability and poor cell permeability, which limited their applicability as chemical probes and their potential therapeutic development. To overcome these limitations here, we develop a nonpeptidic PROTAC approach that exploits our recently discovered and optimized drug-like VHL ligands26 and show that it can be applied to target BET bromodomains and potently induce effective and selective degradation of BRD4.

We began by designing a series of PROTACs that would link together specific VHL ligands and BET bromodomain ligands. Recent work has established compounds VHL-1 and VHL-2 as strong binders with Kd values below 300 nM to VHL (Figure 1a).26 Inspection of the protein−ligand crystal structures show that the methyl group of the terminal acetyl groups in compounds VHL-1 and VHL-2 is solvent exposed, and we therefore reasoned that it could provide a suitable connecting point for a linker (Figure S1). For ΔS and ΔG values, see the Supporting Information.

Figure 1. Design, synthesis, and biophysical and biological evaluation of BET bromodomain PROTACs. (a) Chemical structures of BET-bromodomain inhibitors JQ1 and I-BET762 and binders of von Hippel-Lindau protein VHL-1 and VHL-2. (b) Scheme of the synthesis of PROTAC compounds MZ1−3 and cisMZ1; for detailed synthetic procedures see the Supporting Information. (c) Isothermal titration calorimetry data for titration of MZ1 into the individual members of the BET-bromodomain subfamily. Titrations were performed at 30°C with a protein concentration of 15 μM and ligand concentration of 150 μM (entry 1−6). Titrations of MZ1 and cisMZ1 into VBC at 25°C with identical concentrations (entry 9, 12) and reverse titration of VBC protein (150 μM) into MZ3 (15 μM) at 25°C (entry 10) were conducted. For ΔS and ΔG values, see the Supporting Information. (d) HeLa cells were treated with either siRNA targeting individual BET proteins or negative control siRNA 24 h prior to treatment with the compounds MZ1−3, cisMZ1, and JQ1 or vehicle control (0.01% DMSO) for an additional 24 h. Abundance of individual BET protein was analyzed by Western blotting using corresponding specific antibodies accordingly after SDS-PAGE.
interaction with the BET bromodomains as revealed by cocrystal structures (Figure S1). Linkers with different lengths comprised of polyethylene glycol chains with either three or four ethylene glycol units were chosen to connect JQ1 with the VHL ligand. To achieve the desired ligands, a generally applicable two-step synthetic strategy was devised. First, the linker bearing a carboxylic acid at one end and an azide group at the other end was connected with the terminal free amine of the VHL ligand by a HATU-mediated amide bond formation. In the second step, reduction of the azide group to an amine and subsequent amide bond formation with the carboxylic acid of the ester-hydrolyzed JQ1 analogue afforded the desired PROTAC compounds MZ1, MZ2, MZ3, and cisMZ1 (Figure 1b).

To assess whether PROTAC molecules retained their binding to the target proteins VHL and BET bromodomains in a similar fashion as the parental ligands, isothermal titration calorimetry (ITC) experiments were performed (Figure 1; all ITC titrations are shown in the Supporting Information). MZ1, as a representative of all PROTAC molecules that share the same JQ1 moiety for binding bromodomains, was titrated into individual first and second bromodomains of BRD2, BRD3, and BRD4 (Figure 1c, entries 1–6). The measured binding affinities ($K_d$ of 115–382 nM) and $\Delta H$ ($-6.1$ to $-10.0$ kcal/mol) compared well with those reported for unmodified JQ1 (literature values for BRD4 bromodomains shown in Figure 1c, entries 7, 8), suggesting that JQ1 binding mode is conserved within the context of our PROTACs. Similarly, as binding to...
the VHL protein is crucial for the recruitment of target proteins to the E3 ligase, the binding of MZ1 and MZ3 to the VHL-ElonginB-ElonginC complex (VBC) was also quantified using ITC (Figure 1c, entries 9, 10). The measured affinities ($K_d$ of 150 and 310 nM for MZ1 and MZ3, respectively) and $\Delta H$ ($-6.9$ and $-4.9$ kcal/mol, respectively) compared very closely to those of the parental unmodified ligands VHL-1 ($K_d = 185$ nM, $\Delta H = -5.5$ kcal/mol, entry 11) and VHL-2 ($K_d = 290$ nM, $\Delta H = -5.3$ kcal/mol).26 As the stereochimistry of the hydroxyl group of the central hydroxyproline moiety is crucial for ligand binding to VHL, compound cisMZ1 was synthesized that is structurally identical to MZ1 except for a reversed stereocenter at the C-4 position bearing the hydroxyl group. As expected, cisMZ1 did not exhibit any measurable binding affinity for VHL in the ITC experiment (Figure 1c, entry 12) and thus was elected as a negative control compound in cellular assays.

To demonstrate that PROTACs are able to induce degradation of BET proteins, HeLa cells transfected with control siRNA were treated with 1 $\mu$M of compounds MZ1−3 alongside negative controls JQ1 and cisMZ1 for 24 h (Figure 1d). In parallel, HeLa cells with BRD2, BRD3, and BRD4 individually and separately silenced by transfection with the respective siRNA were treated with vehicle DMSO to compare the protein depletion effect of RNAi knockdown and PROTACs. BET protein abundance was evaluated by SDS-PAGE followed by Western blot using corresponding specific antibodies to probe for BRD2, BRD3 or BRD4, respectively. All three PROTAC compounds demonstrated complete removal of BRD4 with no detectable protein observed after 24 h of treatment. In contrast, removal of BRD2 and BRD3 was not complete after 24 h. MZ1 exhibited the highest efficacy among the three compounds. MZ2, which is structurally analogous to MZ1 except for a longer linker containing four PEG units, showed a weaker removal effect compared to MZ1. MZ3, containing an additional phenylalanine moiety between the linker and the VHL ligand, showed to be the least effective at removing BRD2 and BRD3. Together, the data demonstrate potent and effective degradation of BET proteins and suggested a preferential degradation effect on BRD4 over BRD2 and BRD3. The latter observation was unexpected given the parental compound JQ1 is a pan-BET inhibitor and our PROTACs bind with similar affinities to BET bromodomains. Nevertheless, the attractive opportunity to achieve single target selectivity prompted us to conduct further characterization.

To assess the compound dose- and time-dependent intracellular activities, HeLa cells were first treated with various concentrations of MZ1, MZ2, and MZ3 (Figure 2a). All three compounds showed concentration dependent BET removal activity with higher activity at higher concentrations. As in the initial experiment, MZ1 proved the most active compound, with more than 90% of all BET proteins being removed at compound concentration down to 1 $\mu$M. Remarkably, preferential removal of BRD4 over BRD2 and BRD3 was confirmed with all three compounds. Such preference is more prominent with treatment at lower concentrations, e.g., 0.1−0.5 $\mu$M. To study the activities of our PROTACs over time, HeLa cells were treated with 1 $\mu$M or 0.1 $\mu$M MZ1, and cellular BET protein levels were monitored in a time course experiment (Figure 2b for representative data with MZ1, see Figure S2 for additional data with other compounds and concentrations). Progressive removal of BET proteins over time was observed in all experimental setups, and BRD4 consistently exhibited the strongest and fastest reduction in protein level. Reassuringly, no BET protein degradation was observed in the presence of either DMSO or JQ1 (Figure S3) or cisMZ1 (Figure 3a). To verify whether the observation of preferential removal for BRD4 by
our PROTACs can be observed in another cell line, the same study was carried out in U2OS osteosarcoma cells, and the same activity profile was observed (Figure S4). To visualize the BET protein degradation process, U2OS cells were transfected with a plasmid coding for a green fluorescent protein (GFP)-tagged BRD4 protein, allowing fluorescence readout of BRD4 within the cell nuclei. Cells were induced to express GFP-BRD4 for 24 h and then were treated with either 5 μM MZ1 or 5 μM cisMZ1, and the fluorescence was observed over time. In the presence of the active compound MZ1, a complete depletion of the fluorescence signal was observed after just 3 h, whereas cisMZ1 caused no change in the fluorescence signal over the course of the experiment (Figure 2c, Figure S5 and Supporting Information videos a and b). These data confirmed that BRD4 is removed from the cell nuclei in a time-dependent manner due to the presence of MZ1. Taken together, time and dose—response activity profiles revealed rapid and effective PROTAC-induced preferential degradation of BRD4 over BRD2 and BRD3.

To gain mechanistic insights, the VHL and proteasome dependency of PROTAC-mediated protein degradation was first examined. cisMZ1 was unable to induce degradation of any of the BET proteins over time (Figure 3a), demonstrating that PROTAC efficacy is dependent on productive recruitment of VHL. Next, the reliance of the PROTAC-induced protein degradation on proteasome activity was assessed using proteasome inhibitor MG132. Treatment with MG132 completely abrogated MZ1-induced degradation of all BET proteins (compare lanes 3 and 6 in Figure 3b), establishing the expected proteasome-dependence of the approach. Interestingly, MG132 treatment in the absence of PROTAC showed no significant accumulation in BET protein levels, either alone or in combination with JQ1 (compare lanes 1 and 2 with 4 and 5 in Figure 3b, respectively), suggesting that basal proteasome activity level against BET proteins is negligible under those conditions and only becomes significant as a result of PROTAC treatment.

To further evaluate the biological activity of our compounds, we asked whether PROTAC treatment had any effect on the levels of its target E3 ligase (VHL) and on the level of HIF-1α, the natural substrate of VHL. VHL levels in the presence of MZ1 (1 μM) remained unaffected over the course of up to 36 h, thus indicating that the amount of E3 ligase is not influenced by MZ1 binding (Figure 3c). On the other hand, as the VHL ligand portions of our PROTACs occupy the same binding site on VHL that is used to recruit HIF-1α, PROTACs could block HIF-1α binding to VHL to an extent that it may lead to potential stabilization of HIF-1α within cells. For the approach herein described, this effect would not be desirable as upregulation of HIF-1α transcriptional activity would confound the effects resulting from degradation of BET proteins and would be expected to result in induction of the hypoxic response, potentially giving rise to unwanted side effects. To assess whether any HIF-1α stabilization could be observed, HeLa cells were treated with MZ1 and with cobalt(II) chloride as a hypoxia mimicking positive control. Reassuringly, we could not observe any evidence of HIF-1α stabilization even at concentrations of MZ1 up to 10 μM, while clear HIF-1α stabilization is observed in the presence of CoCl₂ (Figure 3d).

A number of non-BET potential off-targets of JQ1 have been recently reported, among which proteins DDB1 and RAD23B (hHR23b) were validated by proteome labeling and Western blotting. To assess whether MZ1 causes degradation of these off-targets, protein levels were examined in HeLa cells treated with MZ1 at 1 μM and 100 nM over a time course of 36 h, and no degradation was observed (Figure S6). Next, to determine whether the removal of BET proteins by PROTAC treatment is reversible, and to establish how long it would take for cells to reverse the effect, we treated HeLa cells for 4 h with 1 μM of MZ1, removed the compound from the media, and then monitored protein levels over a period of 48 h. The washed cells showed detectable recovery of intracellular BRD4 only by 20 h after washout, while in the absence of the wash step, no protein could be detected even after 48 h (Figure 3e, see Figure S7 for the same experiment monitoring time-dependent levels of BRD2 and BRD3). Taken together, these results demonstrate that PROTAC-induced protein degradation is strictly dependent on binding to VHL, on proteasome activity, and does not interfere with the normal endogenous levels of both VHL and HIF-1α. Furthermore, the degradation effect is not only rapid but also sustained and long lasting even upon removal of the compound.

BET inhibitors such as JQ1 influence the expression of an assortment of genes. Selective targeting of individual BET family members would be predicted to elicit distinct and more limited transcriptional responses, because the genome occupancy patterns of BET proteins are not identical. To evaluate the functional consequences of removing BET proteins using PROTACs, and in particular of inducing selective degradation of BRD4 over BRD2 and BRD3, we next monitored the mRNA expression profiles of a selection of cancer-related genes which respond to JQ1 treatment and BET protein inhibition: MYC, P21, AREG, FAS, TYRO3, and FGFR1. The dependence of MYC and P21 expression on BRD4 activity is well characterized. MYC stimulates cell cycle progression and is constantly expressed upon misregulation in cancer, thus leading to continuous overexpression of downstream MYC-dependent genes. In bone associated tumors as well as leukemia and lymphoma cell lines, JQ1 treatment or silencing of BRD4 resulted in downregulation of MYC. MYC represses transcription of the cell cycle CDK inhibitor P21, a tumor suppressor. Downregulation of MYC and consequent derepression of P21 promotes cell cycle arrest. In contrast to the well-characterized BRD4 dependency of MYC and P21, FAS, which encodes a proapoptotic protein belonging to the tumor necrosis factor receptor family, is downregulated by depletion of BRD2, while for the growth factors AREG and FGFR1 as well as the protein tyrosine kinase TYRO3 little is known about any BET protein specific regulation. However, these four genes are known to strongly respond to treatment with JQ1 and therefore were included as a representative set of genes to compare between the pan-BET inhibitory effect caused by JQ1 and a selective BRD4 degradation caused by MZ1. Treatment with MZ1 at 100 nM for 24 h was chosen as this provided an optimal condition and the lowest effective concentration for achieving selective degradation of BRD4 over BRD2 and BRD3 and at the same time minimizing potential interference due to BET bromodomain inhibition (Figure 2a panel I and Figure 2b panel II). In addition, treatments with negative control compound VHL-1’ (Figure S8) lacking the JQ1 moiety, as well as with JQ1 itself, were also conducted to provide comparisons. Treatment of MZ1 resulted in downregulation of MYC, similar to JQ1, after 12 h (Figure S9), although MYC levels recovered after 24 h. Treatment with MZ1 and JQ1 resulted in similar upregulation of P21 and AREG both after 12 h (Figure S9) and 24 h (Figure 4a). Interestingly, in
contrast to JQ1, which resulted in significant changes on FAS, TYRO3, and FGFR1, MZ1 showed more subtle and less significant effects on these genes relative to VHL-1′ (Figure 4a and Figure S9). We hypothesize that such differences observed in gene modulation may be the result of preferential degradation of BRD4 over the other two BET proteins caused by MZ1. To test this hypothesis, we suppressed individual BRD2, BRD3, or BRD4 genes using siRNA to mimic the protein removal effect (Figure S10) and analyzed the gene expression level of the target genes of interest (Figure 4b). While MYC, P21, and AREG levels were confirmed to be affected by suppression of BRD4, we found that FAS was downregulated upon suppression of BRD2 only but not BRD4 (Figure 4b), while FGFR1 is upregulated upon suppression of either BRD3 or BRD4. These results are consistent with preferential degradation of BRD4 over BRD2 and BRD3 by MZ1 and point to a more BRD4-selective pharmacological profile of MZ1 compared with pan-selective inhibitor JQ1.

In summary, we report a small molecule PROTAC approach achieving rapid, effective, and prolonged intracellular degradation of BET bromodomain proteins. The PROTAC-induced protein degradation is dependent on binding to VHL, is reversed upon blocking proteasome activity, and does not interfere with the endogenous, physiological levels of VHL and of its natural substrate HIF-1α. All investigated compounds showed preferential degradation of BRD4 over BRD2 and BRD3 at low concentrations. The downstream gene expression pattern resulting from treatment with our potent and selective PROTAC MZ1 is similar to JQ1 inhibition for BRD4-dependent genes MYC, P21, and AREG but not for FAS, FGFR1, and TYRO3. Our results suggest a different pharmacological response resulting from selectively depleting BRD4 with MZ1 compared to inhibiting the whole BET protein subfamily with JQ1. Given that no preference for binding the bromodomains of BRD4 over the highly homologous bromodomains of BRD2 and BRD3 was observed by ITC within the context of the purified proteins, we speculate that the observed selectivity could arise from preferential and more efficient polyubiquitination of lysine residues on the surface of BRD4 compared to those of BRD2 and BRD3. Alternatively or in addition, preferential direct interaction or reduced steric constraints between VHL and BRD4 compared to BRD2/3 may occur as a result of PROTAC binding, triggering a more productive formation of a VHL:PROTAC:BRD4 ternary complex. Elucidation of the molecular basis for the BRD4-selective activity of PROTACs will warrant further mechanistic investigation in the future. Our findings demonstrating effective and selective degradation of BRD4 with a PROTAC approach open up unprecedented opportunities to study the downstream physiological and pathological consequences of BRD4 modulation. It will allow determination of whether more selective pharmacological perturbations of BET
protein function will have improved therapeutic efficacy, potentially leading to more efficient and specific new drugs in the future. Finally, potent chemical probes that bind to human bromodomains outside the BET subfamily are beginning to emerge, which could be similarly conjugated to a VHL ligand to induce selective intracellular degradation of their respective target bromodomain-containing proteins.

**METHODS**

For detailed descriptions of synthetic and biological methods, see the Supporting Information.

**ASSOCIATED CONTENT**

† Supporting Information

Additional figures, tables, materials and methods, and .avi videos. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acschembio.5b00216.

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**Authors Contributions**

A.C. conceived and supervised the project; M.Z. and K.-H.C. performed the experiments; M.Z., K.-H.C. and A.C. analyzed the data; and M.Z., K.-H.C. and A.C. wrote and edited the manuscript.

**Notes**

The authors declare no competing financial interest.

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**REFERENCES**


■ NOTE ADDED IN PROOF

Around the time the "Just Accepted" version of this paper was published online, two independent reports have come out online describing pan-BET selective PROTAC compounds dBET1 (DOI: 10.1126/science.aab1433) and ARV-825 (DOI: 10.1016/j.chembiol.2015.05.009) that conjugate the same BET bromodomain recruiting ligand JQ1 to a ligand for a different E3 ligase, cereblon.