

Mannose binding lectin (*MBL*) 2 gene polymorphism & its association with clinical manifestations in systemic lupus erythematosus (SLE) patients from western India

Vandana Pradhan, Prathamesh Surve, Anjali Rajadhyaksha*, Vinod Rajendran, Manisha Patwardhan, Vinod Umare, Kanjaksha Ghosh & Anita Nadkarni

*National Institute of Immunohaematology (ICMR) & *Department of Medicine, King Edward Memorial Hospital, Mumbai, India*

Received December 24, 2012

Background & objectives: Systemic lupus erythematosus (SLE) is a prototypical autoimmune disease characterized by production of autoantibodies. Mannose binding lectin (MBL) is an important element of the innate defense system. The present study was undertaken to determine whether variant alleles in *MBL2* gene were associated with disease severity in SLE patients.

Methods: The *MBL* alleles [-550, -221, +4, Codon 52, Codon 54 and Codon 57] were studied by PCR-RFLP (restriction fragment length polymorphism) method in 100 SLE patients fulfilling ACR (American College of Rheumatology) criteria along with 100 healthy controls. SLE disease activity was evaluated using SLE Disease Activity Index (SLEDAI) score.

Results: Homozygosity for *MBL* variant allele (O/O) was observed in 24 per cent of the SLE patients compared to 16 per cent of the normal controls, while no difference was found for heterozygosity (A/O) (37 vs 35%). A significant difference was reported in incidence of double heterozygosity for mutant allele B and D (B/D) among SLE patients as against control group ($P = 0.015$). *MBL* genotypes did not show any association with renal involvement.

Interpretation & conclusions: In this study from western India, *MBL* gene polymorphism showed an influence as a possible risk factor for susceptibility to SLE, but had no direct effect on disease characteristics. Further studies need to be done on a larger number of SLE patients in different regions of the country.

Key words Allele - gene polymorphism - heterozygosity - mannose binding lectin (MBL) - systemic lupus erythematosus (SLE)

Systemic lupus erythematosus (SLE) is a complex trait characterized by the production of a range of autoantibodies and a diverse set of clinical phenotypes¹. Wide-ranging clinical phenotypes include skin rash,

neuropsychiatric and musculoskeletal symptoms and in some patients leading to lupus nephritis, are observed in SLE patients. Aberrant complement activation leads to inflammation resulting in tissue injury of multiple

organs. Mannose binding lectin (MBL) is a calcium-dependent serum protein that plays a role in the innate immune response by binding to carbohydrates on the surface of a wide range of pathogens, where it can activate the complement system or act directly as an opsonin². The complement system is a collection of blood and cell surface proteins that are involved in primary host defense and act as a clearance components of innate and adaptive immune responses. There are three different complement pathways, the classical complement pathway, the alternative complement pathway, and the mannose-binding lectin (MBL) pathway². The lectin pathway is stimulated when the MBL binds to mannose residues on the pathogen surface. The MBL-associated serine proteases, MASP-1, and MASP-2, are activated and cleave C4 and C2, which then form the C3 convertase and leads to the formation of membrane attack complex (MAC) which ultimately initiates cell lysis. MBL has an oligomeric structure (400-700 kDa), built of subunits that contain three identical peptide chains of 32 kDa each. Each subunit is characterized by a lectin domain, an α -helical coiled-coil, a hydrophobic neck region, a collagenous region and a cysteine rich N-terminal region^{3,4}.

MBL2 gene is present on chromosome 10 in the region 10q21-24. The normal structural MBL allele is named A, while the three variant structural alleles namely 'B' (codon 54), 'C' (codon 57) and 'D' (codon 52) are designated as 'O' allele⁵. The MBL expression is influenced by the three promoter polymorphic variations at position -550 (H/L), -221 (X/Y) and +4 (P/Q). The promoter and coding variants are in strong linkage disequilibrium which give rise to different haplotypes. Seven such haplotypes (HYPA, LYQA, LYPA, LXPA, HYPD, LYQC and LYPB) linked to *MBL2* gene have been defined⁶.

The aim of the present study was to investigate the role of MBL and its association with susceptibility and clinical expression of SLE through the analysis of promoter region and exonic polymorphism of *MBL2* gene.

Material & Methods

This retrospective study was conducted in 100 consecutive SLE patients from Mumbai, western India, collected over a span of three years (January 2010-December 2012). Their age ranged from 5-53 yr (mean \pm SD; 28.14 \pm 9.99) in females and 7-45 yr (mean \pm SD; 25.75 \pm 13.76) in males. These patients were referred from Rheumatology and Nephrology departments of KEM hospital, Mumbai, Maharashtra, India, to National

Institute of Immunohaematology, (NIIH), Mumbai. All these patients were diagnosed according to the American College of Rheumatology (ACR) criteria⁷. Disease activity was assessed at the time of evaluation using Systemic Lupus Erythematosus Disease Activity Index (SLEDAI)⁸. Pregnant and post-menopausal women, smokers, patients with diabetes and patients with significant hyperlipidaemia were excluded. The source of normal controls (n=100) was volunteer blood bank donors and healthy women staff members of the NIIH. Blood samples (5 ml) were collected after obtaining written informed consent from all the individuals. The study protocol was approved by the Institutional Ethical Committee (IEC) of NIIH.

The patients were categorized into two groups, SLE with lupus nephritis (LN) and SLE without lupus nephritis (non LN). Renal biopsies were examined by light microscopy using hematoxylin, eosin, periodic Schiff (PAS) staining. Immunofluorescence microscopy was done using anti-IgG, anti-IgM, anti-IgA, anti-C3, anti-C4 and anti-fibrinogen fluorescein isothiocyanate conjugate (FITC). The renal histology was classified according to WHO criteria⁹.

Molecular analysis: Genomic DNA was extracted by using conventional phenol-chloroform extraction method¹⁰. The genotyping of six single nucleotide polymorphisms (SNPs) in the *MBL2* gene was performed as previously described¹¹. The promoter region polymorphism +4 (P/Q) and -550 (H/L) was studied using allele specific oligonucleotide amplification¹¹. Another promoter region polymorphism at -221 (X/Y) and exonic polymorphisms R52H, G54D and G57E were studied by restriction fragment length polymorphism (RFLP) method using the restriction enzyme BsaI, HhaI, BanI, MboII respectively¹²⁻¹⁴. All the PCRs were initiated by 5 min denaturation step at 94°C and completed by 7 min extension step at 72°C.

Statistical analysis: Continuous variables were expressed as mean \pm SD. Pairs of groups were compared using student 't' test for normally distributed continuous distribution. The ' χ^2 ' test was used for the categorical variables as needed. Statistical analysis was carried out using Graph Pad In Stat 2 software (Graph Pad Software Inc., USA). Chi-square test, with Yates's correction was used for analysis¹⁵.

Results

Among 100 SLE patients included in the study, 92 were females. At the time of evaluation antinuclear antibody (ANA) positivity was 100 per cent and anti-

dsDNA (anti-double stranded DNA) positivity was 88.9 per cent. Age at onset of the disease was 4-46 yr (24.7+8.79 yr) and the age at evaluation was 5-53 yr (27.94+10.2 yr). The SLEDAI scores ranged between 6 - 53 (mean + SD; 17.53 + 9.2), the average SLEDAI score of LN patients was higher (20.60 ± 9.93) than non LN patients (12.80 ± 6.97) and overall SLE patients (16.94 ± 9.47). The disease severity of the patients was categorized into mild, moderate and severe based on their SLEDAI scores (mild <8, moderate 8-18, severe >18). Moderate disease severity was noted in 52 per cent of SLE patients, while 35 and 13 per cent patients had severe and mild disease, respectively.

Table I shows the distribution of *MBL2* structural gene polymorphisms in SLE patients among LN and non LN groups as compared with healthy controls. It was observed that 39 per cent of SLE patients showed A/A genotype, 37 per cent had A/O genotype and 24 per cent had O/O genotype. Overall distribution did not differ significantly between SLE patients and controls. However, increased frequency of O/O homozygosity was observed among SLE patients when compared to

controls. Similar tendency for loss of wild type allele was noted higher in LN patients as against non LN group. The total frequency of ‘O’ alleles was 0.4 in patients and 0.3 in the controls. The double heterozygosity for B/D allele was significantly higher in SLE patients (62.5%) as against control group (25%). [OR 3.973, CI (1.3-12.19), *P*= 0.015]. The frequency of promoter variants and position +4 is shown in Table II. There was no significant difference reported between SLE and control group. Only -550 region allele ‘L’ was found to be significantly higher among LN patients as compared to non LN group (*P*=0.004). *MBL* down-regulating promoter allele ‘X’ did not contribute to SLE susceptibility in our study.

Table III shows frequency of promoter haplotypes between SLE patients and controls. Significant difference was reported in the frequency of LY haplotype between SLE patients and controls [OR – 4.1687, CI -2.30-7.54), *P*=0.0001]. Similarly, LX haplotype that leads to lower *MBL* levels also showed significant difference between SLE patients and control group (*P*=0.03). Table IV summarizes details of clinical

Table I. Frequencies of exon 1 *MBL2* polymorphisms among the study group and controls

Exon 1 genotype	SLE			
	LN N (Frequency)	Non LN N (Frequency)	SLE N (Frequency)	Controls N (Frequency)
A/A	17 (0.27)	22 (0.41)	39 (0.32)	49 (0.45)
A/O	21 (0.50)	16 (0.46)	37 (0.50)	35 (0.44)
O/O	15 (0.23)	9 (0.13)	24 (0.18)	16 (0.11)
Total	53	47	100	100

LN, lupus nephritis; ‘A’, wild type allele; ‘O’, variant allele

Table II. Frequency of *MBL2* gene polymorphism in study group (n=100) and controls (n=100)

Position	Variant allele	SLE N (%)	Control N (%)	<i>P</i> value	LN N (%)	Non LN N (%)	<i>P</i> value
-550	H	78(39)	96(48)	0.086	31(29)	47(50)	0.004
	L	122(61)	104(52)		75(71)	47(50)	
-221	X	65(32.5)	80(40)	0.145	32(30.2)	30(35.1)	0.791
	Y	135(67.5)	120(60)		74(69.8)	61(64.9)	
+4	P	109(54.5)	101(50.5)	0.483	56(53)	53(56)	0.717
	Q	91(44.5)	99(49.5)		50(47)	41(44)	

A total of 200 chromosomes were studied.
N represents the number of chromosomes showing positivity of that particular allele.

Table III. Promoter haplotype frequency in the study group and controls

Haplotype	SLE N (%)	Control N (%)
HY	64(32)	83(41.5)
LY	71(35.5)**	37(18.5)
LX	51(25.5)*	67(33.5)
HX	14(7)	13(6.5)

*P**<0.05 **<0.001 compared with controls

manifestations and MBL genotypes. Approximately 50 per cent of the SLE patients showed renal involvement and arthritis, followed by 37 per cent of the patients showed positivity with immunological parameters like ANA, low C3-C4 levels and 26 per cent showed the presence of malar and/or discoid rash. Significantly higher prevalence of variant allele was reported in SLE patients having renal involvement ($P<0.001$) and neurological disorders ($P<0.05$). Histological subtypes of lupus nephritis (like MPGN, FPGN, DPGN and RPGN) were studied, but they did not exhibit any notable difference and there were no significant difference found (results not shown).

The regulatory and coding variants are in strong linkage disequilibrium; with seven most common haplotypes described HYP A, LYPA, LYQA, LXPA, HYPD, LYPB and LYQC. The distribution of seven common haplotypes was studied among SLE patients and control group. All seven haplotypes were equally distributed among patients and control group. There was no linkage association of any haplotype noted with disease severity. The haplotype LYPA (48%), LYQA

(46%) and HYP A (41%) showed the highest prevalence among study population.

Discussion

It has been shown that *MBL2* gene polymorphisms influence susceptibility to SLE and could be associated with some clinical and laboratory features, disease progression, cardiovascular disease and increased risk of infection¹⁶. Many groups have previously investigated the possibility of interaction among SLE candidate genes. Multiple genes are known to be involved in susceptibility of SLE¹⁷. Several studies have shown *MBL* gene variation at exon 1 as being additive risk factor for susceptibility in different populations¹⁸⁻²¹. In our study, slightly increased frequency of exonic variant allele was observed among SLE patients when compared to controls. The overall frequency for *MBL* structural mutant alleles in exon 1 region did not differ significantly between SLE and control groups but there was a decreased tendency of wild type allele 'A' in SLE patients. Similar findings have been reported by Asgharzadeh *et al*¹¹ in their study on *MBL2* gene polymorphism, susceptibility to renal dysfunction among SLE patients. Another study on white Danish population has demonstrated that complicating infections in SLE patients are strongly associated with homozygosity (O/O) for *MBL* variant allele⁵. In our study also, the variant allele 'O' contributed significantly in infections.

A recent study carried out in the eastern India showed higher frequency of B/B genotype in SLE patients as compared to healthy controls²². In our study from western India higher frequency of B/D genotype was observed in SLE patients. A meta-analysis involving

Table IV. Association of clinical manifestations (as per ACR criteria) with structural *MBL* genotypes

Clinical manifestations	Number positivity	A/A(N.)	A/O & O/O (N)	<i>P</i> value
Malar and/or Discoid rash	26	10	16	0.0968
Photosensitivity	25	10	15	0.1646
Oral ulcers	23	9	14	0.1436
Arthritis	49	20	29	0.0769
Serositis	17	8	9	0.7317
Renal disorders*	53	17	36	0.0003
Neurologic disorders*	8	0	8	0.0059
Hematologic disorders	16	7	9	0.4807
Immunologic parameters (ANA, low C3, low C4)	37	20	17	0.4860

* $P<0.005$

studies with three ethnic populations namely European derived, African derived and Asian derived concluded that allele 'B' and the allelic variant at promoter region of *MBL2* gene, specifically those found at position -550 and -221 were risk factors for SLE development²³. In our study the promoter variant analysis did not show any difference among SLE patients and control group except that -550 region allele 'L' was found to be significantly higher among LN patients as compared to non LN group. Another meta-analysis by Xu *et al*²⁴ of five European and three American studies indicated a significant association between the polymorphism and SLE in allelic contrast. While stratified by ethnicity in European population, no significant association was found, therefore, they concluded that the *MBL2* A/O polymorphism might be associated with SLE²⁴. In our study, non significant association of *MBL2* A/O polymorphism with susceptibility to SLE was observed.

Glesse *et al*²⁶ observed a significant difference among the frequency of both promoter haplotype and haplotypic combination in African derived patients, with a higher incidence of HY haplotype and LY/HY combinations in SLE patients when compared with controls. The studies among Americans, Caucasians and Chinese have shown complete absence of HX haplotype showing complete linkage disequilibrium between alleles H/L and X/Y²⁷. We observed equal frequency of HX haplotype among our SLE patients and control group. Similar results have been reported by Navarra *et al*²⁸ among Filipinos population showing higher frequency of HX alleles (SLE - 22.41%, control - 25.4%) suggesting genetic diversity. The regulatory and coding variants are in strong linkage disequilibrium; only seven haplotypes have been so far reported in human population and, therefore, thought to be resulted due to founder mutational events. The haplotype HYPA was reported to be associated with wild type allele 'A' showing higher MBL levels, whereas haplotype LXPA was associated with most severe defects^[29]. In our study, we did not find such associations as both the haplotype HYPA and LXPA showed nearly equal distribution among SLE patients and controls.

In conclusion, our study indicated that frequency of double heterozygosity for B/D allele was increased in SLE patients as compared to normal healthy individuals. Similarly, allele 'L' of the -550 region in the promoter site showed susceptibility for renal involvement suggesting that these alleles could be

an important risk factor for SLE development in our population. LX haplotype that leads to lower MBL levels might have contributed to disease susceptibility in our SLE patients from western India. Thus, the overall effect and role of associated risk factors may vary according to the ethnic and genetic background of the study population.

Acknowledgment

Authors acknowledge the Department of Biotechnology (DBT), Government of India, New Delhi, for the financial support.

References

1. Ippolito A, Wallace DJ, Gladman D, Fortin PR, Urowitz M, Werth V, *et al*. Autoantibodies in systemic lupus erythematosus: comparison of historical and current assessment of seropositivity. *Lupus* 2011; 20 : 250-5.
2. Manderson AP, Botto M, Walport MJ. The role of complement in the development of systemic lupus erythematosus. *Annu Rev Immunol* 2004; 22 : 431-56.
3. Pradhan V, Surve P, Ghosh K. Mannose binding lectin (MBL) in autoimmunity and its role in systemic lupus erythematosus (SLE). *J Assoc Physicians India* 2010; 58 : 688-90.
4. Monticciolo OA, Mucenic T, Xavier RM, Brenol JC, Chies JA. The role of mannose-binding lectin in systemic lupus erythematosus. *Clin Rheumatol* 2008; 27 : 413-9.
5. Garred P, Madsen HO, Halberg P, Petersen J, Kronborg AS, Svejgaard A, *et al*. Mannose-binding lectin polymorphisms and susceptibility to infection in systemic lupus erythematosus. *Arthritis Rheum* 1999; 42 : 2145-52.
6. Casanova JL, Abel L. Human mannose-binding lectin in immunity: friend, foe or both? *J Exp Med* 2004; 199 : 1295-9.
7. Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* 1997; 40 : 1725.
8. Bombardier C, Gladman DD, Urowitz MB, Caron D, Chang CH. Derivation of SLEDAI: a disease activity index for lupus patients. The Committee on Prognosis Studies in SLE. *Arthritis Rheum* 1992; 35 : 630-40.
9. Weening JJ, D'Agati VD, Schwartz MM, Seshan SV, Alpers CE, Appel GB, *et al*. The classification of glomerulonephritis in systemic lupus erythematosus revisited. *J Am Soc Nephrol* 2004; 15 : 241-50.
10. Ausubel F, Brent R, Kingston RE, Moore DD, Seidman JG, Smith J, *et al*, editors. *Protocol adapted from short protocols in molecular biology*, 3rd ed. New York: John Wiley; 1995 Unit 2.1: p. 2-3.
11. Asgharzadeh M, Kafil HS, Ebrahimzadeh ME, Bohlolvi A. Mannose-binding lectin gene and promoter polymorphism and susceptibility to renal dysfunction in systemic lupus erythematosus. *J Biol Sci* 2007; 7 : 801-5.
12. Roelofs RW, Sprong T, de Kok JB, Swinkels DW. PCR-restriction fragment length polymorphism method to detect

- the X/Y polymorphism in the promoter site of the mannose binding lectin gene. *Clin Chem* 2003; 49 : 1557-8.
13. Ramaswami R, Spina GS, Fae KC, Pereira AC, Nisihara R, Messias Reason II, *et al.* Association of mannose-binding lectin gene polymorphism but not of mannose serine protease 2 with chronic severe aortic regurgitation of rheumatic etiology. *Clin Vaccine Immunol* 2008; 15 : 932-6.
 14. Onay H, Pehlivan M, Alper S, Ozkinay F, Pehlivan S. Might there be a link between mannose binding lectin and vitiligo. *Eur J Dermatol* 2007; 17 : 146-8.
 15. Woolf B. On estimating the relation between blood group and disease. *Ann Hum Genet* 1955; 19 : 251-3.
 16. Monticielo OA, Mucenic T, Xavier RM, Brenol JC, Chies JA. The role of mannose-binding lectin in systemic lupus erythematosus. *Clin Rheumatol* 2008; 27 : 413-9.
 17. Sullivan KE, Jawad AF, Piliero LM, Kim N, Luan X, Goldman D, *et al.* Analysis of polymorphisms affecting immune complex handling in systemic lupus erythematosus. *Rheumatology (Oxford)* 2003; 42 : 446-52.
 18. Selvaraj P, Narayanan PR, Reetha AM. Association of functional mutant homozygotes of the mannose binding protein gene with susceptibility to pulmonary tuberculosis in India. *Tuber Lung Dis* 1999; 79 : 221-7.
 19. Selvaraj P, Jawahar MS, Rajeswari DN, Alagarasu K, Vidyarani M, Narayanan P. Role of mannose binding lectin gene variants on its protein levels and macrophages phagocytosis with live mycobacterium tuberculosis in pulmonary tuberculosis. *FEMS Immunol Med Microbiol* 2006; 46 : 433-7.
 20. Singla N, Gupta D, Joshi A, Batra N, Singh J, Birbian N. Association of mannose-binding lectin gene polymorphism with tuberculosis susceptibility and sputum conversion time. *Int J Immunogenet* 2012; 39 : 10-4.
 21. Vaid M, Kaur S, Taruna M, Singh H, Gupta VK, Murthy KJ, *et al.* Association of SP-D, MBL, I-NOS genetic variants with pulmonary tuberculosis. *Indian J Hum Genet* 2006; 12 : 105-10.
 22. Panda AK, Parida JR, Tripathy R, Pattanaik SS, Ravindran B, Das BK. Low producer *MBL* genotypes are associated with susceptibility to systemic lupus erythematosus in Odisha, India. *Hum Immunol* 2013; 74 : 114-9.
 23. Lee YH, Witte T, Momot T, Schmidt RE, Kaufman KM, Harley JB, *et al.* The mannose-binding lectin gene polymorphisms and systemic lupus erythematosus; two case-control studies and a meta-analysis. *Arthritis Rheum* 2005; 52 : 3966-74.
 24. Xu WD, Peng H, Zhou M, Zhang M, Li BZ, Pan HF, *et al.* Association of *RANTES* and *MBL* gene polymorphisms with systemic lupus erythematosus: a meta analysis. *Mol Biol Rep* 2013; 40 : 941-8.
 25. Ip WK, Chan SY, Lau CS, Lau YL. Association of systemic lupus erythematosus with promoter polymorphisms of the mannose-binding lectin gene. *Arthritis Rheum* 1998; 41 : 1663-8.
 26. Glesse N, Monticielo OA, Mattevi VS, Brenol JC, Xavier RM, da Silva GK, *et al.* Association of mannose-binding lectin 2 gene polymorphic variants with susceptibility and clinical progression in systemic lupus erythematosus. *Clin Exp Rheumatol* 2011; 29 : 983-90.
 27. Chies JA. On the haplotypic frequencies of the *MBL2* gene among human populations. *Lupus* 2007; 16 : 838.
 28. Navarra SV, Villamin CA, Baes RP, Pimenta L, Nicdao JL, Bernas GD. Increased frequency of mannose-binding lectin promoter LX haplotype among Filipinos with systemic lupus erythematosus. *Lupus* 2007; 16 : 147-8.
 29. Madsen HO, Garred P, Thiel S, Kurtzhals JA, Lamm LU, Ryder LP, *et al.* Interplay between promoter and structural gene variants control based serum level of mannan-binding protein. *J Immunol* 1995; 155 : 3013-20.

Reprint requests: Dr Anita Nadkarni, National Institute of Immunohaematology (ICMR), 13th Floor, King Edward Memorial Hospital, Parel, Mumbai 400 012, Maharashtra, India
e-mail: anitahnadkarni@yahoo.com