



A REVIEW: ISOLATION OF ACANTHAMOEBA SPECIES IN SURFACE WATERS OF YASUJ DISTRICT SOUTH OF IRAN

Abdolali Moshfe^{1*}, Mohammad ali Kohansal², Najmeh Parhizgari¹, Seyed Sajjad Khoramrooz¹, Mojtaba Abbasi³, Maryam Niyiyati⁴, Ashkan Akbarzadeh⁵, Ali Jamshidi¹

1. Cellular and Molecular Research Center, Yasuj University of Medical Sciences, Yasuj, Iran
2. Student Research Committee, Yasuj University of Medical Sciences, Yasuj, Iran
3. Faculty of Veterinary Medicine, Shahrekord Branch, Islamic azad University, Shahrekord, Iran
4. Department of Parasitology and Mycology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran
5. General Practitioner

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ABSTRACT

Background: *Acanthamoeba* spp. are opportunistic amphizoic protozoans, which are distributed in the natural and artificial environment water sources. This ubiquitous amoeba is the causative agent of Amoebic Keratitis(AK) and Granulomatous Amoebic Encephalitis(GAE). The main aim of the current study was to identify the presence of *Acanthamoeba* spp. in surface waters of Yasuj district south of Iran.

Methods: In this descriptive cross-sectional study, a total of 30 surface water samples were collected from environmental sources, including natural rivers, springs, waterfall and freshwater source in 2013. All samples were collected using 500 ml sterile plastic bottles during two month. After filtration through millipore nylon membrane, samples were cultured on non-nutrient bacto agar medium enriched with *Escherichia coli* and incubated for 3 to 14 days at room temperature. Identification of the *Acanthamoeba* spp. was based on morphological criteria of cysts and trophozoites. Following DNA extraction, PCR was used to confirm the microscopically identification.

Results: A total of 11 out of 30 samples (36.6%) were positive for *Acanthamoeba* species based on the morphological criteria. Five out of 11 positive samples (45.46%) were confirmed by PCR method. In total, 5 (16.6%) samples out of 30 samples were positive for *Acanthamoeba* species based on PCR method.

Conclusion: High frequency of *Acanthamoeba* spp. in different surface water sources specially promenades in that region is an alert for the public health and highlights the needs for more awareness of health professionals and for the related risks.

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Introduction

Acanthamoeba spp. are free-living opportunistic protozoan parasites that pervade the entire environment, and they can be found in tap, fresh, coastal and bottled mineral water, sewage, soil, dust and air, heating or air-conditioning units and contact lens solutions, eyewash stations, and gastrointestinal washings (1,2,3). These amoebae are the causative agents of multifocal encephalitis called granulomatous amebic encephalitis, a chronic central nervous system disease that usually occurs in

immunocompromised hosts, amoebic keratitis (AK) and pneumonitis(4). Most episodes of keratitis occur after water exposure or a history of swimming in lakes and ponds while wearing contact lenses and the infection is also linked to non-sterile home-made saline solutions for contact lenses (3,5).

The importance of *Acanthamoeba* more revealed according to recently studies in some parts of Iran. Positive rate of *Acanthamoeba* in drinking waters in several hospitals of Iran was 48% (6) and in hospital wards with immunodeficient patients in Tehran, was 52.9% (7) and in surface waters consist of rivers, lakes, springs, and lagoon of Gilan province-north of Iran was 70.3% (8), In Sarein, Ardebil province, north west of Iran, in hot springs were 42.9% (9) and in river recreation areas in Tehran province was 27.3% (10) and in environmental sources in ahvaz city, khuzestan province, southern Iran was 71.6% of samples of water and 26% soil samples (11)

Otherwise new reported cases of diseases show the importance role of *Acanthamoeba* in producing illness. In recent published paper the authors reported a 5-year-old Iranian immunocompetent girl who died of fulminant *Acanthamoeba* meningoencephalitis. To the authors' knowledge, that was the first case of *Acanthamoeba* meningoencephalitis in Iran(12)

Also, a case of young immunocompetent male adult with autopsy proven *Acanthamoeba* meningoencephalitis was reported by Chandra and etal. From India. (13).

New studies in Iran revealed the genotype of *Acanthamoeba* spp. Isolated *Acanthamoeba* from water in recreational areas of Tehran, was belonged to the T4 and T5 genotypes(14). and in another study The percentage of positive FLA isolates was 27.3%, of which 80% were *Acanthamoeba*, assigned to the T4 and T15 genotype, and 20% were *Naegleria*(10). In a study, Sequence analysis of the single isolate of *Acanthamoeba* revealed potentially pathogenic T(4) genotype corresponding to *A. castellanii*(9). In Ahvaz. Genotyping of positive samples proved that *Acanthamoeba* belonged to T4, T2, and T5 genotypes(11). In hospital research *Acanthamoeba* belonged to the T4 genotype was the most prevalent isolate. Presence of the T4 genotype on medical instruments, including an oxygen mask in an isolation room of an immunodeficiency pediatric ward, should be of concern for health authorities. *Acanthamoeba* T5 genotypes, *Hartmannella vermiformis*, and *Vahlkampfia avara* were also present(7). In therapeutic hot springs in Iran *Acanthamoeba* belonging to the potentially pathogenic T4 and T3 genotypes(15). Results of the first study for presenting the identification of pathogenic genotypes of *Acanthamoeba* in dust samples in Iran revealed the presence of T4, T5 and T11 genotypes within those samples(16).

In another study in Iran, The obtained results revealed that most of *Acanthamoeba* strains belonged to genotype T4 both in clinical and environmental(water, soil and animal-origin) samples and T11 genotype in clinical samples was also found after the genotyping analysis. Moreover, the isolation of T4 genotype from cow faeces in that study highlights a possible transmission of *Acanthamoeba* through animal faeces in Iran(17).

Furthermore, reports of new diseases due to *Acanthamoeba* is remarkable for researchers. Such as First report of a mixed infection due to *Acanthamoeba* genotype T3 and *Vahlkampfia* in a cosmetic soft contact lens wearer in Iran(18) and In a study authors results support the hypothesis that some parasitic microorganisms such as *Acanthamoeba* can involve and contribute toward the development of rheumatoid syndromes(19).

According to above evidences, this study was conducted to isolation of *Acanthamoeba* species in surface waters of Yasuj district south of Iran for use the results to identification areas with *Acanthamoeba* contamination.

2. Materials and methods

2.1. General information on geography

Yasuj is an industrial city in the Zagros Mountains of southwestern Iran, and is the capital of Kohgiluyeh and Boyer-Ahmad Province. Yasuj has an estimated population of 140,000 and famous to capital of nature of Iran.

The province is mostly mountainous in terrain, part of the Zagros range. The highest point is the Dena summit with a height of 4,409 meters.

The mountain range, which is located in Kohgiluyeh and Buyer-Ahmad province, is covered with oak forests. Natural springs, singing of the birds and fresh air fascinate all lovers of nature. The snow and rain falling is quite sizable in the city. Yasuj has a humid temperate climate with plenty of annual rainfall and is known for its moderate, mild, and Mediterranean-like climate. Large parts of the province are mountainous, green, and forested. Thousands of domestic and foreign tourists come to the seashore river for swimming and camping.

2.2. Sampling

During May to November 2013, 30 surface water samples were collected from environmental sources, including natural (rivers, lakes and springs) source from different parts of Yasuj, south of Iran. From each sampling point, one to three water samples were collected in 1000-ml sterile bottles and transported immediately to the laboratory of medicine school for further processing.

2.3. Isolation of *Acanthamoeba* species and culture

For the isolation of *Acanthamoeba* species, approximately 1000 ml of the collected water samples were filtered through a cellulose nitrate membrane with pore size 0.45 μ . Filter was transferred on bacto agar plates seeded with Gram-negative bacteria (*E. coli*) as a food source. Plates were incubated at room temperature, and after 3 days later, they were microscopically examined for the presence of *Acanthamoeba* trophozoites. However, in the absence of amoebae, plates were

monitored for up to 14 days. *Acanthamoeba* were identified at the genus level, based on morphological characteristics of trophozoites and cysts using phase-contrast microscopy.

2.4. DNA extraction and PCR

Amoeba cells were harvested from culture plates, concentrated by centrifugation, and then lysed by treatment with lysozyme (100 mg/ml). The samples were then treated with 2–5 µl proteinase K (18.9 mg/ml), and DNA extraction was performed by phenol–chloroform method. PCR assay: the *Acanthamoeba*-specific primer pairs JDP1 (5'-GGCCCAGATCGTTTACCGTGAA) and JDP2 (5'-TCTCACAAAGCTGCTAGGGAGTCA) as described by Schroeder et al. (2001) were used for the amplification of the 500 bp of 18S rDNA gene (20,21).

Standard PCRs were performed in 50-µl volumes, containing 5 µl of 10× PCR buffer, 20 pM of each of the primers, 4 mM MgCl₂, 0.2 mM dNTP, 1.25 U Taq polymerase (Cinnagen), and 1–10 ng of template DNA. Thermal cycling conditions were 94°C for 5 min; 32 cycles of 94°C for 30 s, 57°C for 30 s, 72°C for 40 s; followed by a final extension at 72°C for 5 min.

3. Results

A total of 11 out of 30 samples (36.6%) were positive for *Acanthamoeba* species based on the morphological criteria. Five out of 11 positive samples (45.46%) were confirmed by PCR method. In total, 5 (16.6%) samples out of 30 samples were positive for *Acanthamoeba* species based on PCR method (fig1).

The name of five positive area are:

- 1- Tange Gange ei
- 2- Sadde Shah Ghasem (arrival river)
- 3- Beshar River (Ali Abad)
- 4- Sadde Shah Ghasem (going out river)
- 5- Tange Mehrian



Fig1) Gel electrophoresis of PCR products

A: marker, 1-6: negative cases, 7-11: positive cases (500 bp)

4. Discussion

The present study highlight the presence of *Acanthamoeba* in surface waters of Yasuj city. All the waters included in the present research was in contact with human activity and thus this could be a hazard for high risk people including soft contact lens wearers and immunosuppressed patients especially those with frequent contact with such waters (22). On the other hand, there is a high risk for users, including children who have weak immune system, related to the high resistance of *Acanthamoeba* cysts to harsh environmental situation. There are various studies on surface waters and their contamination to *Acanthamoeba* in Iran and worldwide (23,24). In a study conducted on the surface waters by Rezaeian et al. there were a high occurrence of *Acanthamoeba* in environmental sources including water, dust, soil and animal feces (24). Their study showed that out of 80 samples of different environments in Tehran, 37 samples were contaminated with *Acanthamoeba* (46.25%).

The difference between Rezaeian et al study and the present research was the source of samples. The present study focused on only recreational water sources.

Another study conducted by Badirzadeh et al. showed the presence of free living amoeba including *Vahlkampfiids* (11) and *Acanthameba* (1) in hot springs of Sarein county, Ardebil province. The low occurrence of *Acanthamoeba* in hot spring samples in the mentioned research could be attributed to high temperature of waters in their study (9). However, despite

Badirzadeh et al research, Solgi et al. study in 2012 on all hot springs in Ardebil revealed that 20% of the hot springs were contaminated with *Acanthamoeba* (25). It should be mention that they also isolated other FLAs including *Hartmannella* and *Naegleria* (25).

The present study isolated FLA in recreational waters in order to find more detailed data about distribution of these amoebae in Yasuj city. It should be mention that no study has been performed on *Acanthamoeba* in this region and the present research was the first to investigate the presence of Amoebae in the region .

In Coşkun et al. research in 2013 in Turkey, 150 water samples were collected from six region of Sivas, Turkey The samples were cultured and of the 150 samples, 33 samples (22%) were positive for FLA. It should be mention that amoebae were detected using morphological keys, and most isolated amoebae were *Acanthamoeba* (26). Shoff et al. also conducted a study (2008) to examine the distribution of *Acanthamoeba* and other FLA in household waters. In their study, 283 water samples were collected from houses, and amoebae were detected using morphological criteria in the culture medium. Of 283 samples, 80 were positive for amoebae, and *Acanthamoeba* were detected in 8 samples (2.8%)(27).

One of the finding of the present research was the low number of PCR positive versus morphological identification. This is may be due to contamination of the plates to various microorganisms such as funji and bacteria. Another limitations of the present study was the impossibility of sequencing that could not be supplied due to financial supports, however it is hoped that we could achieve the goal in near future.

Overall, the present research reflect that warning signs should be implicated in recreational areas such as near water sources and education to high risk people are necessary for amoeba disease prevention.

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References

- 1- Mergeryan, H., 1991. The prevalence of *Acanthamoeba* in the human environment. *Rev. Infect. Dis.* 5, 390–391.
- 2- Guy, A. Cabral, Francine, Marciano-Cabral., 2004. Cannabinoid-mediated exacerbation of brain infection by opportunistic amebae. *J. Neuroimmunol.* 147(1–2), 127–130.
- 3- Khan, NA., 2003. Pathogenesis of *Acanthamoeba* infections. *Microb. Pathog.* 34(6), 277-85.
- 4- Govinda, S. Visvesvara, Hercules, Moura&Frederick, L., 2007. Schuster. Pathogenic and opportunistic free-living amoebae: *Acanthamoeba* spp., *Balamuthia mandrillaris*, *Naegleria fowleri*, and *Sappinia diploidea*. *FEMS. Immunol. Med. Microbiol.* 50, 1-26.
- 5- Radford, CF., Minassian, DC., Dart, JKG., 2002. *Acanthamoeba* keratitis in England and Wales: incidence, outcome, and risk factors. *Br. J. Ophthalmol.* 86(5), 536–542.
- 6- Bagheri, H., Shafiei, R., Shafiei, F., Sajjadi, S., 2010. Isolation of *acanthamoeba* spp. From drinking waters in several hospitals of iran. *Iran. J. Parasitol.* 5(2), 19-25.
- 7- Lasjerdi, Z., Niyiyati, M., Haghghi, A., Shahabi, S., Biderouni, FT., Taghipour, N., Eftekhar, M., Nazemalhosseini Mojarad, E., 2011. Potentially pathogenic free-living amoebae isolated from hospital wards with immunodeficient patients in Tehran, Iran. *Parasitol. Res.* 109(3), 575-80.
- 8- Mahmoudi, MR., Taghipour, N., Eftekhar, M., Haghghi, A., Karanis, P., 2012. Isolation of *Acanthamoeba* species in surface waters of Gilan province-north of Iran. *Parasitol. Res.* 110(1), 473-7.
- 9- Badirzadeh, AL., Niyiyati, M., Babaei, Z., Amini, H., Badirzadeh, H., Rezaeian, M., 2011. Isolation of free-living amoebae from sarein hot springs in ardebil province, iran. *Iran. J. Parasitol.* 6(2), 1-8.
- 10- Niyiyati, M., Lasjerdi, Z., Nazar, M., Haghghi, A., Nazemalhosseini Mojarad, E., 2012. Screening of recreational areas of rivers for potentially pathogenic free-living amoebae in the suburbs of Tehran, Iran. *J. Water. Health.* 10(1), 140-6.
- 11- Rahdar, M., Niyiyati, M., Salehi, M., Fegghi, M., Makvandi, M., Pourmehdi, M., Farnia, S., 2012. Isolation and genotyping of *acanthamoeba* strains from environmental sources in ahvaz city, khuzestan province, southern iran. *Iran. J. Parasitol.* 7(4), 22-6.
- 12- Binesh, F., Karimi, M., Navabii, H., 2011. Unexpected postmortem diagnosis of *acanthamoeba* meningoencephalitis in an immunocompetent child. *BMJ. Case. Rep.* 2011 Oct 4;2011. pii: bcr0320113954. doi: 10.1136/bcr.03.2011.3954

- 13- Chandra, SR., Adwani, S., Mahadevan, A., 2014. Acanthamoeba meningoencephalitis. *Ann. Indian. Acad. Neurol.* 17(1), 108-12.
- 14- Nazar, M., Haghghi, A., Niyyati, M., Eftekhar, M., Tahvildar-Biderouni, F., Taghipour, N., Abadi, A., Nazemalhosseini Mojarad, E., Athari, A., 2011. Genotyping of Acanthamoeba isolated from water in recreational areas of Tehran, Iran. *J. Water. Health.* 9(3), 603-8.
- 15- Solgi, R., Niyyati, M., Haghghi, A., Taghipour, N., Tabaei, SJ., Eftekhar, M., Nazemalhosseini Mojarad, E., 2012. Thermotolerant Acanthamoeba spp. isolated from therapeutic hot springs in Northwestern Iran. *J. Water. Health.* 10(4), 650-6.
- 16- Niyyati, M., Lorenzo-Morales, J., Rahimi, F., Motevalli-Haghi, A., Martín-Navarro, C.M., Farnia, S., Valladares, B., Rezaeian, M., 2009. Isolation and genotyping of potentially pathogenic Acanthamoeba strains from dust sources in Iran. *Trans. R. Soc. Trop. Med. Hyg.* 103(4), 425-7.
- 17- Niyyati, M., Lorenzo-Morales, J., Rezaie, S., Rahimi, F., Mohebbi, M., Maghsood, A.H., Motevalli-Haghi, A., Martín-Navarro, C.M., Farnia, S., Valladares, B., Rezaeian, M., 2009. Genotyping of Acanthamoeba isolates from clinical and environmental specimens in Iran. *Exp. Parasitol.* 121(3), 242-5.
- 18- Niyyati, M., Lorenzo-Morales, J., Rezaie, S., Rahimi, F., Martín-Navarro, C.M., Mohebbi, M., Maghsood, A.H., Farnia, S., Valladares, B., Rezaeian, M., 2010. First report of a mixed infection due to Acanthamoeba genotype T3 and Vahlkampfia in a cosmetic soft contact lens wearer in Iran. *Exp. Parasitol.* 126(1), 89-90.
- 19- Eftekhar, M., Athari, A., Haghghi, A., Mosaffa, N., Shahram, F., Abadi, A., 2010. Seroprevalence of Acanthamoeba Antibodies in Rheumatoid Arthritis Patients by IFAT, Tehran, Iran 2007. *Iran. J. Parasitol.* 5(1), 35-40.
- 20- Schroeder, J.M., Booton, G.C., Hay, J., Niszl, I.A., Seal, D.V., Markus, M.B., Fuerst, P.A., Byers, T.J., 2001. Use of subgenetic 18S ribosomal DNA PCR and sequencing for genus and genotype identification of acanthamoebae from humans with keratitis and from sewage sludge. *J. Clin. Microbiol.* 39(5), 1903-11.
- 21- Edagawa, A., Kimura, A., Kawabuchi-Kurata, T., Kusuhara, Y., Karanis, P., 2009. Isolation and genotyping of potentially pathogenic Acanthamoeba and Naegleria species from tap-water sources in Osaka, Japan. *Parasitol. Res.* 105(4), 1109-17.
- 22- Niyyati, M., Lasjerdi, Z., Nazar, N., Haghghi, A., Nazemalhosseini Mojarad, E., 2012. Screening of recreational areas of rivers for potentially pathogenic free-living amoebae in the suburbs of Tehran, Iran. *J. Water. Health.* 10(1), 140-6.
- 23- Khan, N.A., Siddi, 2009. Acanthamoeba affects the integrity of human brain microvascular endothelial cells and degrades the tight junction proteins. *Int. J. Parasitol.* 39(14), 1611-6.
- 24- Rezaeian, M., Niyyati, M., Farnai, Sh., Rahimi, F., Motevalli haghi, A., 2008. Isolation of Acanthamoeba Spp. from Different Environmental Sources. *Iran. J. Parasitol.* 3, 44-47.
- 25- Solgi, R., Niyyati, M., Haghghi, A., Mojarad, E.N., 2012. Occurrence of Thermotolerant Hartmannella vermiformis and Naegleria Spp. in Hot Springs of Ardebil Province, Northwest Iran. *Iran. J. Parasitol.* 7(2), 47-52.
- 26- Coşkun, K.A., Özçelik, S., Tutar, L., Elaldı, N., Tutar, Y., 2013. Isolation and identification of free-living amoebae from tap water in Sivas, Turkey. *Biomed. Res. Int.* 67-145.
- 27- Shoff, M.E., Rogreson, A., Kessler, K., Schatz, S., Seal, D.V., 2008. Prevalence of Acanthamoeba and other naked amoebae in South Florida domestic Water. *J. water. Health.* 6(1), 99-104.