

The Sanitation Hygiene Infant Nutrition Efficacy (SHINE) Trial: Rationale, Design, and Methods

The Sanitation Hygiene Infant Nutrition Efficacy (SHINE) Trial Team^a

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Child stunting and anemia are intractable public health problems in developing countries and have profound short- and long-term consequences. The Sanitation Hygiene Infant Nutrition Efficacy (SHINE) trial is motivated by the premise that environmental enteric dysfunction (EED) is a major underlying cause of both stunting and anemia, that chronic inflammation is the central characteristic of EED mediating these adverse effects, and that EED is primarily caused by high fecal ingestion due to living in conditions of poor water, sanitation, and hygiene (WASH). SHINE is a proof-of-concept, 2 × 2 factorial, cluster-randomized, community-based trial in 2 rural districts of Zimbabwe that will test the independent and combined effects of protecting babies from fecal ingestion (factor 1, operationalized through a WASH intervention) and optimizing nutritional adequacy of infant diet (factor 2, operationalized through an infant and young child feeding [IYCF] intervention) on length and hemoglobin at 18 months of age. Within SHINE we will measure 2 causal pathways. The program impact pathway comprises the series of processes and behaviors linking implementation of the interventions with the 2 child health primary outcomes; it will be modeled using measures of fidelity of intervention delivery and household uptake of promoted behaviors and practices. We will also measure a range of household and individual characteristics, social interactions, and maternal capabilities for childcare, which we hypothesize will explain heterogeneity along these pathways. The biomedical pathway comprises the infant biologic responses to the WASH and IYCF interventions that ultimately result in attained stature and hemoglobin concentration at 18 months of age; it will be elucidated by measuring biomarkers of intestinal structure and function (inflammation, regeneration, absorption, and permeability); microbial translocation; systemic inflammation; and hormonal determinants of growth and anemia among a subgroup of infants enrolled in an EED substudy. This article describes the rationale, design, and methods underlying the SHINE trial.

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Globally, stunting affects 26% (165 million) of children aged <5 years, underlies 15%–17% of their mortality [1], and leads to long-term cognitive deficits, fewer years and poorer performance in school, lower adult

economic productivity, and a higher risk that their own children will also be stunted, perpetuating the problem into future generations. Stunting begins antenatally and peaks at 18–24 months of postnatal life, when mean length-for-age z score (LAZ) is approximately –2.0 among children living in Africa and Asia [2]. Inadequate diet and recurrent illness (especially diarrhea) have been the 2 most commonly implicated causes of stunting, and therefore the focus of most research designed to inform prevention strategies.

Dietary studies have tested a myriad of nutrient-dense foods including high-energy milks [3, 4], nutrient-dense porridges [5–8], and bacterial-resistant, micronutrient-fortified lipid pastes [9–11]. Studies have also tested different combinations of nutrition education without

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provision of food [12, 13], food without education [3, 5, 8], and food given together with education [14, 15]. Behavior change strategies have been undertaken to persuade mothers to feed their children more responsively to hunger cues in a physically comfortable and psychologically supportive environment [16]. A systematic review of 38 of the best studies revealed that children who received these interventions grew 0–1.7 cm taller (LAZ, 0.0–0.64) by 12–24 months compared with control children [17], indicating that the growth effect of the most efficacious dietary interventions is equivalent to about one-third of the average deficit experienced by Asian and African children (LAZ score approximately –2.0). This suggests that, at best, optimizing infant diet can solve about one-third of the stunting problem.

Studies estimating the effect of diarrhea on linear growth have reached inconsistent conclusions. In a pooled analysis of 9 studies that together collected diarrhea and growth data on 1393 children, the odds of stunting at 24 months increased multiplicatively by 2.5% per episode of diarrhea, and 25% of all stunting among 24-month-old children was attributable to having 5 or more episodes of diarrhea in the first 2 years of life [17]. In other studies, the observed effect of diarrhea on long-term linear growth was small or absent because children grew at higher-than-average “catch-up” growth rates between illness episodes [18, 19]. Most recently, an analysis of 7 longitudinal cohort studies of children aged <2 years, conducted in 4 low-income countries, estimated that the average child’s diarrhea burden (46 days with diarrhea between birth and 2 years) was associated with a 0.38-cm height deficit at 2 years of age [18]. Because a change in LAZ of 1.0 at 2 years is equivalent to approximately 3 cm [19], their analysis suggests that the growth effect of reducing diarrhea during the first 2 years of life from the average burden currently experienced in low-income countries to zero is a statistically significant but clinically modest LAZ score of approximately 0.13, or about 7% (one-fifteenth) of the average height deficit of 2-year-old Asian and African children.

Like stunting, childhood anemia is a major public health problem in Africa and Asia [1] and a primary cause of cognitive and behavioral developmental delay throughout childhood and adolescence [20]. The 2010 Global Burden of Disease ranks iron deficiency anemia as the 15th leading cause of lost disability-adjusted life-years globally [21]. Anemia becomes prevalent in the second half of infancy and peaks between 12 and 24 months of age [22], affecting well over half of all children in Africa and Asia [23]. Iron deficiency causes about half of the anemia among children under 2 [24]. Accordingly, increasing iron intake by young children has been the focus of most research and public health programming, and, in randomized trials, iron supplements or iron-fortified foods reduce anemia in young children by about 37%–62% [25], leaving a substantial portion of child anemia unaddressed.

The Sanitation Hygiene Infant Nutrition Efficacy (SHINE) trial is motivated by a 2-part premise [26]:

- A major cause of child stunting and anemia is environmental enteric dysfunction (EED). EED is a subclinical disorder of the small intestine, which is virtually ubiquitous among asymptomatic people living in low-income settings throughout the world [27–38]. EED is characterized by increased permeability, which facilitates microbial translocation into the systemic circulation and triggers chronic immune activation [39, 40].
- The primary cause of EED is infant ingestion of fecal microbes due to living in conditions of poor quality and quantity of water, sanitation, and hygiene (WASH).

EED Develops During Infancy, Is Strongly Associated With Poor Growth, and May Cause Anemia

Studies of stillborn fetuses have established that EED is not present at birth [41]. Seminal research conducted over 4 decades in The Gambia demonstrated that EED develops during early infancy and is strongly associated with poor growth [42–52]. These studies showed that stunting was not explained by diarrheal burden or inadequate dietary quantity or quality [45, 46], but was associated with gut permeability (assessed by the lactulose-mannitol [LM] test) and immune activation (assessed by plasma concentrations of immunoglobulin G [IgG] and endotoxin core antibody) [51]. Among children studied longitudinally from 2 to 15 months of age, these biomarkers increased as LAZ plummeted and, in a regression model, IgG and IgG endotoxin core antibody concentrations explained 51% of the variability in linear growth during this age interval [48]. In a more recent study (The Interactions of Malnutrition & Enteric Infections: Consequences for Child Health and Development), in which 8 longitudinal cohorts of children were followed from birth to 2 years, an index of 3 fecal biomarkers of intestinal inflammation (neopterin, myeloperoxidase, and α -1 antitrypsin) predicted change in LAZ over the subsequent 6 months [53]. Finally, in a retrospective case-control study in Zimbabwe, cases (children who were stunted at 18 months of age) had higher concentrations of inflammatory markers (serum C-reactive protein and α -1 acid glycoprotein) and lower concentrations of plasma insulin-like growth factor 1 (IGF-1) between 6 weeks and 12 months of age compared to controls (children with normal LAZ at 18 months) [24]. These findings suggested that growth hormone resistance, characterized by inflammatory suppression of plasma IGF-1 (an already well-established mechanism of linear growth failure in children with juvenile idiopathic arthritis [54] and Crohn disease [55]) may also underlie stunting among otherwise healthy children living in resource-constrained environments.

The chronic immune activation that is characteristic of EED also may cause anemia through 2 possible pathways. First, elevated proinflammatory cytokines trigger increased hepatic synthesis of hepcidin, which suppresses iron absorption and utilization [56, 57], leading to iron deficiency anemia [58]. Second, cytokines may act directly on differentiation of bone

marrow stem cells, suppressing erythropoiesis and leading to anemia of inflammation [59].

Conditions of Poor WASH Are Associated With EED in Humans

Evidence linking poor WASH with EED in humans is limited to observational studies. Among 3- to 5-year-old children in Malawi, those in households with latrines and a greater quantity of water had lower (more normal) LM ratios, and children who washed their hands more often or washed their hands with soap had a more normal sucrose-lactulose ratio (an indicator of gastric permeability) [60]. Similarly, Bangladeshi children living in clean compared to dirty households (based on water quality, facilities for hand washing, and sanitation) had lower urinary LM ratios and lower plasma IgG concentrations [61]. A small intestinal biopsy study of adult volunteers in Lusaka, Zambia and Soweto, South Africa found that all the Lusaka residents but none of the Soweto residents had morphometric changes of EED; at the time of the study, sanitation coverage was 59% in Lusaka [62] and 99.9% in Soweto [63].

WASH May Be More Strongly Associated With Child Stunting Than With Diarrhea

Numerous observational studies have reported a strong association between the level of environmental WASH and child linear growth [61, 64–70]; indeed, in some, conditions of poor WASH were more strongly associated with stunting than with diarrhea. Esrey analyzed Demographic Health Survey data on nearly 12 000 rural children from 8 developing countries and found that neither improved sanitation nor water was significantly associated with diarrhea, but both were associated with increases of 0.05–0.6 in LAZ [64]. In a prospective cohort study evaluating the child health benefits of water and sanitation services in Peru, diarrhea prevalence explained only 16% of children's height deficit at 2 years of age, while differences in water and sanitation services explained 40% of this deficit [22]. These findings imply that the primary pathway linking WASH to growth may be largely independent of diarrhea.

Among WASH Interventions, Sanitation May Be Central

Adequate sanitation may be a particularly important requirement for normal growth. In the Esrey analysis, the association with growth was stronger for sanitation than water services [64]. In the Peru study, the growth effects of improved water source were magnified when concurrent with improved sanitation [22]. In a large ecological analysis of data from India (where about half the population lacks sanitation facilities), Spears estimated that differences in open defecation rates explained 35%–55% of the average difference in stunting between districts [71]. This apparent importance of sanitation could partly explain the relatively small magnitude of impact of WASH on linear growth (LAZ, +0.08) observed in a recent systematic review of 5 randomized controlled trials that tested improved drinking water quality

and/or hand washing, but did not include improved sanitation [72]. Finally, an econometric analysis of data from 26 low-income countries estimated that a 15-minute reduction in walking time to collect water (an indicator of water access) is associated with an increase in height-for-age *z* score (HAZ) of 0.26 and that this effect was stronger among households with sanitation (HAZ, +0.35) vs without sanitation (HAZ, +0.21) [73].

Unhygienic Living Conditions Restrict Growth in Animals and This Effect Is Mediated by Inflammation

Controlled trials in animal husbandry provide strong evidence that poor WASH restricts animal growth and that this effect is mediated by inflammation. In a factorial trial in which chicks were randomized to live either in cages with accumulated feces, dust, and dander (“dirty chicks”) or steam-cleaned cages (“clean chicks”), and to receive either antibiotics or no antibiotics, the clean chicks grew better than dirty chicks. However, the dirty chicks receiving antibiotics grew as well as the clean chicks not receiving antibiotics; overall, antibiotics had no growth effect in clean chicks [74]. The poor growth in dirty chicks was accompanied by high plasma concentrations of interleukin 1 (IL-1), a key proinflammatory cytokine. Levels of IL-1 were significantly lower in clean chicks and in dirty chicks fed antibiotics, suggesting that antibiotics may promote growth by reducing immune stress.

High Exposure to Fecal Microbes May Cause EED

The pathogenesis of EED, similar to other inflammatory intestinal diseases, results from unrestrained enteric T-cell activation: biopsy studies of the human small intestine demonstrate rapid development of crypt hyperplasia, villous atrophy, and mucosal destruction in response to chronic T-cell stimulation [75–77]. However, unlike celiac disease, in which *abnormal T cells* are hyperreactive to a *normal exposure* (ie, dietary gluten [78]), EED may be the result of *normal T-cells* appropriately reactive to *excessive exposure* of fecal bacteria [79]. Bacterial colonization has long been recognized to influence intestinal morphology. In an animal study from 1961 [80], germ-free guinea pigs were found to have an immature intestinal structure. Controlled introduction of bacteria to germ-free animals led to a progressive change in mucosal architecture, but to a lesser degree than guinea pigs reared in conventional conditions. These animals, exposed to environmental bacteria and other insults, had substantial villous shortening and thickening and crypt elongation. It is therefore plausible that exposure to large quantities of fecal microbes among children living in conditions of poor WASH drives the profound mucosal changes characteristic of EED. This may be particularly true in the context of hypochlorhydria, where gastric barrier function is reduced, leading to extensive colonization of the small intestine. We hypothesize that, while only pathogenic bacteria cause

diarrhea, excessive exposure to all fecal microbes (including those from healthy people and animals) may contribute to EED. If true, EED prevention may require more comprehensive interventions than those designed to reduce diarrhea.

Summarizing the Background and Linking It to the SHINE Trial Design

Childhood stunting and anemia are 2 highly prevalent, frequently overlapping problems among children aged <2 years in Africa and Asia, which result in immediate and long-term morbidity and cognitive deficits, lost human capacity, and reduced adult economic productivity. Moreover, both problems are only partially responsive to current public health interventions. SHINE is motivated by the premise that EED is a major underlying cause of stunting and anemia, that chronic inflammation is the central characteristic of EED mediating these adverse effects, and that EED is primarily caused by high fecal ingestion due to living conditions of poor WASH. SHINE will also investigate whether these hypothesized effects of improved WASH will be additive to those of concurrently improved infant feeding in reducing child stunting and anemia. Accordingly, SHINE is designed as a proof-of-concept, 2 × 2 factorial trial to assess the independent and combined effects of protecting babies from fecal ingestion (factor 1, operationalized through a WASH intervention) and optimizing nutritional adequacy of infant diet (factor 2, operationalized through an infant and young child feeding [IYCF] intervention) on length and hemoglobin at 18 months of age. Within SHINE we are measuring 2 causal pathways. The program impact pathway comprises the series of processes and behaviors linking implementation of the interventions with the 2 child health primary outcomes; it will be modeled using measures of fidelity of intervention delivery and household uptake of promoted behaviors and practices [81]. We are also measuring a range of household and individual characteristics, social interactions, and maternal capabilities for childcare [82], which we hypothesize will explain heterogeneity along these pathways. The biomedical pathway comprises the infant biologic responses to the WASH and IYCF interventions that ultimately result in attained stature and hemoglobin concentration at 18 months of age; it will be elucidated by measuring biomarkers of intestinal structure and function (inflammation, regeneration, absorption, and permeability); microbial translocation; systemic inflammation; and hormonal determinants of growth and anemia among a subgroup of infants enrolled in an EED substudy [83].

The WASH Benefits Trials in Bangladesh and Kenya [84] are ongoing trials similar to SHINE in the interventions being tested, outcomes measured, and research questions addressed. Regular meetings between the trial groups have enabled the sharing of preliminary findings and alignment of methods and outcomes, to facilitate comparison of trial results and enhance the joint contributions of the trials to global health.

STUDY DESIGN OVERVIEW

The SHINE trial addresses one primary objective and multiple secondary objectives (Table 1). The 4 treatment arms of the trial are described in Table 2.

Clusters are defined as the catchment area of 1–4 community-based village health workers (VHWs) of the Zimbabwe Ministry of Health and Child Care (MoHCC). More than 5000 pregnant women were identified in a surveillance system of early pregnancy detection using urine pregnancy tests and enrolled into the trial between November 2012 and March 2015. All enrolled women receive the same number (15) of visits from their VHW between enrollment and 18 months postpartum, during which standard-of-care (SOC) primary healthcare messages are delivered. During some of these 15 visits, VHWs in clusters randomized to WASH, IYCF, and WASH + IYCF also deliver lessons specific to their arm (Table 1 of Supplementary Appendix) so that women in active arms receive more information than women in the SOC arm. Each randomized intervention comprises 5 core modules, focused on a key message and delivered to the mother at a specified relevant fetal or infant age. A separate team of research staff employed by the trial make data collection home visits at baseline (approximately 14 weeks' gestation), at 32 weeks' gestation, and at 1, 3, 6, 12, and 18 months postpartum to measure fidelity of intervention delivery, uptake of the interventions, and trial outcomes (Table 3). Women who move from their primary residence (their home at the time of recruitment) are not followed up at interim data collection visits, but extensive effort is made to trace and follow all women residing anywhere within Zimbabwe at the 18-month visit to collect trial endpoints for the intent-to-treat analyses.

DEVELOPMENT OF THE INTERVENTIONS

The underlying biologic mechanisms of child stunting are poorly understood. For this reason, the hypothesized causal pathway was the primary principle guiding development of the 2 interventions designed to optimize the 2 biologic objectives for enrolled children between birth and 18 months of age (WASH intervention: children are protected from ingesting human and animal fecal microbes; IYCF intervention: children consume a nutritionally adequate diet). As much as possible, we chose messages, commodities, and delivery systems that could be economically sustainable at scale, but we prioritized potential biological efficacy over cost.

SHINE interventions were based on formative research (detailed in companion articles [85, 86]) and incorporated adult education evidence-based behavior change strategies, including motivational interviewing, repetition of key messages, elicitation of emotional triggers, culturally adapted visual aids and stories, interactive games, and respect for the circumstances and autonomy

Table 1. Objectives of the Sanitation Hygiene Infant Nutrition Efficacy (SHINE) Trial

Primary objectives
To determine the independent and combined effects of improved household WASH and improved IYCF on length and hemoglobin concentration among children at 18 mo of age who are born to HIV-negative women in rural Zimbabwe.
Secondary objectives ^a
To examine the effects of the 2 randomized interventions (WASH and IYCF) on stunting (LAZ <−2) and anemia (hemoglobin <105 g/L) among children at 18 mo of age who are born to HIV-negative women in rural Zimbabwe.
To examine differential effects on length and hemoglobin concentration at 18 mo of the 2 randomized interventions (WASH and IYCF) in these prespecified subgroups:
(a) Children born to HIV-positive compared to HIV-negative mothers
(b) Male and female children
(c) Subgroups formed by categorizing household wealth, household distance from a water point, and maternal capabilities.
To examine the independent and combined effects of the 2 randomized interventions on body weight, mid-upper arm circumference and head circumference at 18 mo, and on all anthropometric measures at intermediate ages (1, 3, 6, and 12 mo).
To describe the PIP linking implementation of each randomized intervention (WASH and IYCF) with length and hemoglobin concentrations by assessing:
(a) The quality of VHW training and supervision
(b) VHW capacity, defined as a composite of attained knowledge, goal-setting capacity, and achieved performance;
(c) Fidelity of intervention implementation, defined as degree of conformance with protocol specifications for both VHW and mother;
(d) Attained maternal knowledge and skills assessed by questionnaire and observation;
(e) Uptake or adoption of promoted behaviors by mothers and their households assessed by questionnaire and observation.
To assess potential effect modifiers along the PIP:
(a) Individual VHW characteristics (age, time in post, intrinsic, and extrinsic motivational characteristics);
(b) Maternal capabilities, defined by a woman's physical and mental health, stress, time allocation, maternal self-efficacy, and autonomy;
(c) Household socioeconomic status;
(d) Intervention complexity (WASH and IYCF implemented together vs each intervention implemented alone);
(e) For the WASH intervention only, access to water, defined as distance to water in each season of the year.
To describe the prevalence of exclusive breastfeeding among all infants enrolled in the trial by maternal/infant HIV status.
To evaluate the effect of the IYCF intervention on uptake of improved infant feeding practices by maternal/infant HIV status, specifically:
(a) Infant diet quality as assessed by World Health Organization IYCF indicators
(b) Infant nutrient intake from complementary foods assessed by 24-h dietary recall
(c) Appropriate use of Nutributter from 6 to 18 mo
To evaluate the effect of the WASH intervention on the 5 key behaviors it promotes by maternal/infant HIV status:
(a) Proper disposal of animal and human feces
(b) Handwashing with soap after fecal contact
(c) Point-of-use chlorination of drinking water
(d) Protecting children from ingestion of dirt and feces
(e) Feeding baby freshly prepared foods, or reheating leftover food
To elucidate the biological pathways linking WASH and IYCF with linear growth and hemoglobin concentration by measuring domains of EED, stratified by maternal/infant HIV status ^b :
(a) The composition and function of the infant intestinal microbiota;
(b) Intestinal permeability and absorptive capacity (assessed by LM ratio urine test), inflammation (assessed by fecal myeloperoxidase, α -1-antitrypsin and neopterin), epithelial damage (assessed by plasma I-FABP), and regeneration (assessed by fecal REG1B);
(c) Microbial translocation (assessed by plasma soluble CD14 and soluble CD163);
(d) Systemic inflammation (assessed by LPS, EndoCAb, plasma soluble CD14, and soluble CD163);
(e) Two hormonal responses to immune activation (IGF-1 and hepcidin);
(f) Hemoglobin concentration, sTFR, and hepcidin.
To measure the impact of the 2 randomized interventions (WASH and IYCF) on incidence, prevalence, and severity of diarrheal disease in infants, stratified by maternal/infant HIV status.
To model the relative contributions of diarrheal disease and EED in mediating the effects of improved WASH on child length and hemoglobin concentrations, stratified by maternal/infant HIV status.
To measure the strength of association between severity of maternal EED and systemic inflammation during pregnancy with the risk of 6 adverse birth outcomes (miscarriage, stillbirth, premature delivery, fetal stunting, low birth weight and neonatal death), stratified by maternal HIV status.
To measure the strength of association between other potential causes of stunting and anemia (other than poor WASH or IYCF) with linear growth and hemoglobin:
• Maternal schistosomiasis infection during pregnancy;
• Maternal HIV infection together with adherence to antiretroviral and cotrimoxazole regimens during pregnancy and lactation;
• Infant HIV infection or exposure, together with adherence to antiretroviral and/or cotrimoxazole regimens;
• Exposure to dietary mycotoxin contamination by the mother during pregnancy and lactation, and by the infant during complementary feeding.

Abbreviations: AGP, α -1 acid glycoprotein; CRP, C-reactive protein; EED, environmental enteric dysfunction; EndoCAb, Endotoxin core antibody; HIV, human immunodeficiency virus; I-FABP, intestinal fatty acid binding protein; IGF-1, insulin-like growth factor 1; IYCF, infant and young child feeding; LAZ, length-for-age z score; LM, lactulose-mannitol; LPS, lipopolysaccharide; PIP, program impact pathway; REG1B, regenerating protein 1B; SHINE, Sanitation Hygiene Infant Nutrition Efficacy; sTFR, soluble transferrin receptor; VHW, village health worker; WASH, water, sanitation, and hygiene.

^a A complete list of trial secondary endpoints is prespecified in the SHINE Statistical Analysis Plan.

^b The final choice of biomarkers to define each domain of EED may change depending on emerging data.

Table 2. Randomized Arms in the Sanitation Hygiene Infant Nutrition Efficacy (SHINE) Trial

Standard of Care: ^a <ul style="list-style-type: none"> • Exclusive breastfeeding promotion for all infants, birth to 6 mo • Strengthened PMTCT services • Strengthened village health worker system 	WASH: <ul style="list-style-type: none"> • Standard-of-care interventions • Provide household ventilated pit latrine, Tippy Taps, monthly liquid soap, water treatment solution, and protective play space • Provide interpersonal communication interventions promoting feces disposal in a latrine, handwashing with soap, drinking water treatment, hygienic weaning, food preparation, and preventing babies from putting dirt and animal feces in their mouths.
IYCF: ^a <ul style="list-style-type: none"> • Standard of care interventions • Provide 20 g/d Nutributter from 6–18 mo • Provide interpersonal communication interventions promoting optimal use of locally available foods for complementary feeding after 6 mo, continued breastfeeding, and feeding during illness. 	Sanitation/Hygiene AND Nutrition: <ul style="list-style-type: none"> • Standard of care interventions • All WASH interventions • All IYCF interventions

Abbreviations: IYCF, infant and young child feeding; PMTCT, prevention of mother-to-child transmission of HIV; WASH, water, sanitation, and hygiene.

^a Women in standard-of-care and IYCF arms are provided with a latrine at the end of the trial.

of participants [87–90]. Interventions target the mother, but all household members are encouraged to participate.

We chose to implement interventions through the existing MoHCC VHWs rather than creating a separate intervention delivery system. This decision was motivated partly by Zimbabwe's historically strong community health system, but primarily taken because VHWs (who are nominated by and live in the communities they serve) could achieve the greatest coverage and acceptability within these rural communities. Beginning in 2010, vacant VHW posts were filled (total staffing = 180 per district, 360 VHWs in the trial area) and new VHWs completed the standard MoHCC 5-month training program (5 weeks residential didactic sessions interspersed with on-the-job training) [62]. VHWs were trained in SHINE-specific modules in separate groups according to the randomized allocation of the cluster in which they live and work. Residential training in MoHCC district-level training facilities lasted 20 days for SOC, 30 days for WASH, 32 days for IYCF, and 35 days for WASH + IYCF. Throughout trial implementation, VHWs receive supportive supervision from intervention-specific SHINE nurses at least monthly. VHWs receive a \$14 monthly allowance from the MoHCC. To compensate them for the additional time they spend on SHINE activities, the trial provides a grocery basket every 3 months and modest monetary performance-based incentives.

All interventions (including latrine construction) were pilot-tested in a district adjacent to the study area among 40 households with children <18 months. VHWs in the pilot district were trained to implement the study to assess the training requirements and support needs of this cadre before training the 360 VHWs in the SHINE districts.

Standard-of-Care Intervention

VHWs are trained through the MoHCC curriculum to respond to a broad spectrum of health problems. The curriculum instructs VHWs to visit pregnant women and infants frequently,

but the precise content or purpose of each visit is not specified. Consequently, the SHINE SOC intervention was designed to standardize the number of visits (3 antenatal and 12 postnatal visits) and the content of primary healthcare messages across treatment arms. Four of these visits promote exclusive breastfeeding (EBF) from birth to 6 months using modules designed to overcome contextual barriers to EBF [91, 92]. Other SOC modules include prevention of mother-to-child human immunodeficiency virus (HIV) transmission (PMTCT), antenatal care, hospital-based delivery, family planning, and immunizations.

WASH Intervention

Within 6 weeks of enrollment (approximately 20 weeks' gestation) into the WASH and WASH + IYCF arms of the trial (see companion article [85]), a Blair ventilated improved pit (VIP) latrine [93] is constructed at the participant's household and 2 "Tippy Tap" handwashing stations (locally manufactured, and adapting the model piloted by the Kenya WASH Benefits Trial [92]) are installed near the latrine and kitchen. WASH module 1 (delivered at 24 gestational weeks) and module 2 (at 32 gestational weeks) promote safe disposal of feces, and handwashing with soap after fecal contact and before food preparation and eating, respectively. Our intention was for the baby to be born into a household in which latrine use and household handwashing behaviors were normalized and habitual. WASH module 3 (protecting babies from fecal ingestion during explorative play) is delivered when the baby is 2 months old; a washable 2.8 × 3.0-m locally manufactured mat and plastic play yard (North States, Minneapolis, Minnesota) are provided at 2 months and 6 months, respectively. WASH module 4 (treat all drinking water given to babies after 6 months of EBF) is delivered at 4 months of age, along with point-of-use chlorination (WaterGuard: a dilute sodium hypochlorite solution, manufactured locally by Nelspot). Liquid soap and WaterGuard are regularly replenished from time of introduction (modules 2 and 5,

Table 3. Interview, Observation, Measurement, Test Administration, Health Record Transcription, and Biologic Specimen Collection According to Fetal/Infant Age

Data Collected	Research Visit							
	Antenatal			Postnatal, mo				
	Baseline	32 wk	Birth	1	3	6	12	18
Household composition, socioeconomic status	I					I		I
IYCF and WASH knowledge and practices	I/O			I/O	I/O	I/O	I/O	I/O
Maternal factors								
Antenatal care, pregnancy exposures, PMTCT	I	I		I				
Sick clinic visits and hospitalizations	I	I		I	I	I	I	I
Maternal capabilities	I					I		I
Maternal depression		T				T	T	
Height	M							
Weight, mid-upper arm circumference	M	M		M	M	M	M	M
Blood pressure	T	T						
Biologic specimen collection, testing, and archival								
Blood: HIV, CD4 (baseline), Hb, EED panel in substudies ^a	All	All						All
Urine: Pre- and post-LM; spot for urinalysis/schistosomiasis ^b	All	All-spot						
Stool: EED panel in substudies	All			EED				
Saliva: cortisol	All							
Infant factors								
Delivery date, time, mode, place, complications, care, sex			R					
Weight, length, head circumference			R	M	M	M	M	M
Immunizations and vitamin A			R	R/I	R/I	R/I	R/I	R/I
7-day and long-term morbidity, sick clinic visits, hospitalizations				I	I	I	I	I
Biologic specimen collection, testing, and archival								
Blood: EED panel, HIV DNA PCR, Hb ^c				EED	EED	EED	EED	All
Urine: Pre- and post-LM test					EED	EED	EED	EED
Stool: EED panel				EED	EED	EED	EED	EED
Saliva					EED	EED	EED	EED

For specimen collections, All refers to all women and infants enrolled in the trial; EED refers to mother–infant dyads selected for EED substudy.

Abbreviations: EED, environmental enteric dysfunction; Hb, hemoglobin; HIV, human immunodeficiency virus; I, interview; IYCF, infant and young child feeding; LM, lactulose-mannitol; M, measurement; O, observation; PCR, polymerase chain reaction; PMTCT, prevention of mother-to-child transmission of HIV; R, transcription from medical record; T, test; WASH, water, sanitation, and hygiene.

^a EED Panel includes assays which assess causal pathways of EED and anemia. Planned assays are listed below though final choice of biomarkers may change depending on emerging data. Blood/Serum/Plasma: α -1 acid glycoprotein; C-reactive protein; intestinal fatty acid binding protein; insulin-like growth factor 1; activation levels of T-cells by flow cytometry; soluble transferrin receptor; hemoglobin; Urine: lactulose-mannitol ratio; Feces: myeloperoxidase, α -1-antitrypsin; neopterin; regenerating protein 1B; composition and function of intestinal microbiota.

^b A single-void urine sample (spot sample) is collected from all women at baseline and at 32 gestational weeks for urinalysis using Multistix test strips in the homestead and for urinary microscopy for ova of *Schistosoma hematobium* in the field laboratory. At baseline, an LM test is additionally conducted in all women. Prior to ingestion of LM solution, a pre-LM single-void urine sample is collected to measure baseline mannitol by mass spectrometry; after ingestion of LM solution, all urine is collected over a 2-hour period (LM urine), preserved with chlorhexidine and measured, then aliquots frozen at -80°C for subsequent analysis of lactulose and mannitol recovery by mass spectrometry.

^c Hemoglobin is not measured at 1 month of age.

respectively) until the infant is 18 months old. WASH module 5, delivered at 5 months of age, stresses the importance of freshly preparing or fully reheating all foods fed to infants. A review module is delivered at 12 months.

IYCF Intervention

For the IYCF intervention (see companion paper [86]), IYCF module 1 (delivered at 5 months) links good infant feeding

to child growth, health, and intelligence. IYCF module 2 (6 months) promotes feeding nutrient-dense food, including 20 g per day of the lipid-based nutrient supplement developed by the International Lipid-Based Nutrients Supplements Project [94], provided monthly when the baby is 6–18 months of age. Module 3 (7 months) is a participatory cooking demonstration in which any available household food is prepared and fed to the baby, stressing 3 messages from formative research [37]:

(1) An infant can eat any food that an adult eats; (2) food should be ground so that the infant can swallow and digest it; (3) food that is locally available is important for the infant. Module 4 (8 months) promotes responsive feeding during illness, module 5 (9 months) promotes diet diversity, and a review module is delivered at 12 months.

STUDY SITE AND ITS PREPARATION FOR THE TRIAL

Study Site Selection and Description

The study site comprises the contiguous districts of Chirumanzu and Shurugwi (combined population of 158 821 in 2010) [95], 300 km south of Harare. Districts were chosen for SHINE based on the following criteria: predominantly rural, low sanitation coverage (53% for these districts, with 45% open defecation), high prevalence of child stunting (mean LAZ at 18 months and 24 months, -1.89 and -1.97 , respectively), reasonable road access throughout the year, availability of primary healthcare services at the district level offering facility-based antenatal care and deliveries, geographical contiguousness, and stakeholder receptivity. There were no district-level data available on childhood anemia in the study areas; nationally, 74% of children aged 9–17 months were anemic in 2011, with no change from the previous 5-year period [96]. Among school-aged children, both study districts have a high prevalence of *Schistosoma hematobium* infection (23.5% in Chirumanzu, 55.9% in Shurugwi), but very low prevalence of soil-transmitted helminth infections (0% in Chirumanzu, 1.5% in Shurugwi) [97].

Cluster Formation

The study area was divided into 212 clusters, defined as the catchment area of 1–4 VHWs. Digital maps of administrative boundaries of the study area were obtained from the Central Statistics Office and, using Google Earth, all homesteads and key landmarks (roads, clinics, schools, rivers) were plotted. Using large-scale maps, each of the 360 VHWs circled the houses they serve. Cluster-specific numbers of households, reproductive-age women, and <2-year-old children were gathered from VHW registers. Boundaries were finalized by grouping up to 4 VHWs into the same cluster if (1) they were working habitually as a team; (2) their catchment areas included homesteads within close proximity of each other; or (3) to achieve approximately the same number of households, pregnant women, and young children per cluster. About half of the clusters comprise the catchment area of a single VHW; on average, there are 1.6 VHWs per cluster.

Cluster-Specific Sanitation and Water Surveys

In 2011–2012, we assessed cluster-specific water access and sanitation coverage so that these factors could be balanced across treatment arms during randomization (see companion article

[98]). A georeferenced distribution dataset was constructed of all households in the 2 study districts using Google Earth satellite imagery. The spatial distribution of all water points was mapped through a comprehensive survey. Spatial analyses were conducted to calculate cluster-specific mean distance from households to the nearest functional perennial protected water source, and the proportion of households <500 m and >1500 m from such a water source. Sanitation facilities were surveyed in 13 households randomly selected from each cluster. The cluster-specific proportions of households with a VIP latrine, a VIP latrine less than one-half full, and a VIP latrine less than one-half full also accompanied by a nearby handwashing station were calculated. These cluster-specific water access and sanitation indicators were included as covariates in the constrained randomization process (see section 7).

Community Engagement

Community engagement began 5 years prior to trial enrollment through discussion with leaders, strengthening of the PMTCT of HIV program, and VHW network strengthening. During a 3-month period before launching SHINE, we held 45 meetings throughout the study area, each attended by 50–75 influential community members and led by local government and district-level Zimbabwe MoHCC authorities. Following these consultations, Memoranda of Understanding were signed between each district, the MoHCC and Zvitambo, detailing SHINE activities and defining the tripartite roles.

Hub Staffing and Facilities

We established 4 study hubs in the study area, strategically sited such that all households in the study area could be reached by motorbike in <2 hours from the nearest hub; all were established within MoHCC facilities. Each hub is staffed with approximately 25 people and includes specimen processing laboratories with -80°C archiving freezers, internet connection, offices, and warehousing.

SAMPLE SIZE AND POWER

The enrollment goal was a total sample size of 4800 pregnant women, 1200 in each of the 4 trial arms, each of which is comprised of 53 clusters. The primary outcomes are LAZ and hemoglobin level at 18 months of age, stratified by maternal HIV status. The main trial inference will be based on outcomes of infants of HIV-negative women, of whom we expect at least 4080 based on an estimated 15% HIV prevalence among pregnant women at the time of designing the trial. Although the primary outcomes are continuous, the sample size is based on dichotomous versions of these, as we wanted to provide answers regarding stunting and anemia per se. Allowing for 20% loss of evaluable infants at 18 months (including pregnancy losses,

infant mortality, and loss to follow-up), we expect about 816 measurements in each of the 4 study arms. With type I error of 5% (2-sided test), and power of 90%, and control group stunting ($\text{HAZ} < -2$) of 30%, we will be able to detect a reduction of about 8 percentage points (ie, from 30% to 22%) for the marginal effect of either of the interventions, even in the absence of an effect of the other intervention [99].

Calculations were based on an assumed coefficient of variation of the true proportion stunted at 18 months of 0.43 (derived from the 2011 Demographic Health Survey), and an effective loss of 33% of sample size due to cluster size variability, resulting in a design effect of 2.5. To be conservative, we used the same coefficient of variation for anemia. The largest design effect reported by Katz for stunting in several countries was 2.6 [100]. Using the design effect of 2.5, we have 89.7% power to detect a shift of 0.2 in weight-for-age and length-for-age z scores, assuming a within-randomized cluster variance of 1.25. For hemoglobin, assuming a standard deviation at 18 months of 128 g/L [101], we will be able to detect a shift of 26 g/L for either of the interventions with 95% power.

RANDOMIZATION

A highly constrained randomization technique was used to allocate clusters (stratified by district) to treatments [102]. A GAUSS computer program generated millions of random permutations, enumerating those that met specified balance criteria for 14 variables related to geography, demography, water access, and sanitation coverage; a check was made for validity of the scheme on 5000 such acceptable allocations. Then, from the acceptable allocations that also met bias and validity specifications, 10 allocations were randomly selected. Each randomization scheme divided the 212 randomization units into 4 groups of approximately 53 units. Each scheme's corresponding color-coded map was printed on a separate sheet and displayed at a public forum attended by all elected councilors from the study area, district and provincial administrators, and MoHCC authorities. In their presence, 10 plastic balls (numbered 1–10) were placed in an opaque sack. A community representative selected one ball from the sack, thereby identifying which of the 10 numbered allocations would be used. Then, 4 balls (labeled A, B, C, and D) were placed in one sack, and 4 balls (labeled with the 4 treatment arms) were placed in a second sack. Representatives drew a ball from the first sack and a ball from the second sack, pairing a group of clusters with 1 of the 4 treatment arms, thereby mimicking a widely known World Cup draw procedure. This was repeated twice more to pair the next 2 groups of clusters with 2 more treatment arms. The remaining balls formed the final pairing. This second stage was included to provide an additional assurance of impartiality/randomness and a further opportunity for participation

of the community leadership. The resulting map is shown in Figure 1, where the lines represent cluster boundaries and each color represents 1 of the 4 trial arms.

The randomization was stratified by district to achieve the same approximate number of clusters within each of the 4 study arms within each of the 2 districts. Although overall there were 53 clusters in each study arm, exact balance within each district was not possible, because 111 clusters were formed in one district and 101 in the other, neither of which was a multiple of 4. Forming appropriate clusters was deemed more important than achieving exact balance. The constraint criteria and the achieved values of the constraining variables by study arm are shown in Table 2 in the Supplementary Appendix.

RECRUITMENT OF SUBJECTS

VHWs were trained to visit all 15- to 49-year-old women in their catchment area once every 5 weeks, to offer a urine pregnancy test to those who had missed a menses, and to refer pregnancies to antenatal clinic and (with the woman's assent) to SHINE. We enrolled all consenting pregnant women permanently residing in the rural parts of the study districts during the period of trial recruitment; women living in employer-provided housing or renting accommodation were excluded because their residence was unlikely to be permanent and they could not authorize construction of a latrine at the household. We strove to enroll women after gestational week 8 (to minimize miscarriage risk after enrollment) but before week 14 (to provide a substantial period for WASH households to take up optimal fecal disposal and handwashing behaviors before the baby was born).

OUTCOME MEASUREMENT

Research Visits

All research visits are conducted in study households by hub-based data collectors (DCs) who travel by motorcycle. Forty-three primary care nurses were recruited as DCs and given a 14-week training course in research ethics and Good Clinical Practice (GCP) (1 week); motorcycle riding (3 weeks); computing and data entry (2 weeks); HIV testing and counseling (3 weeks); SHINE questionnaire content and interviewing techniques (2 weeks); maternal and child anthropometry (1 week); clinical assessment and referral of infants according to Integrated Management of Childhood Illness [103] guidelines, and specimen collection and study procedure SOPs (2 weeks). Every 6 months, all DCs are standardized in maternal and infant anthropometry; a coefficient of reliability (R) value against a gold standard anthropometrist is calculated and DCs with R values < 0.95 for intra- and interobserver accuracy are given further training and additional supervision in the field. Supervisors observe every

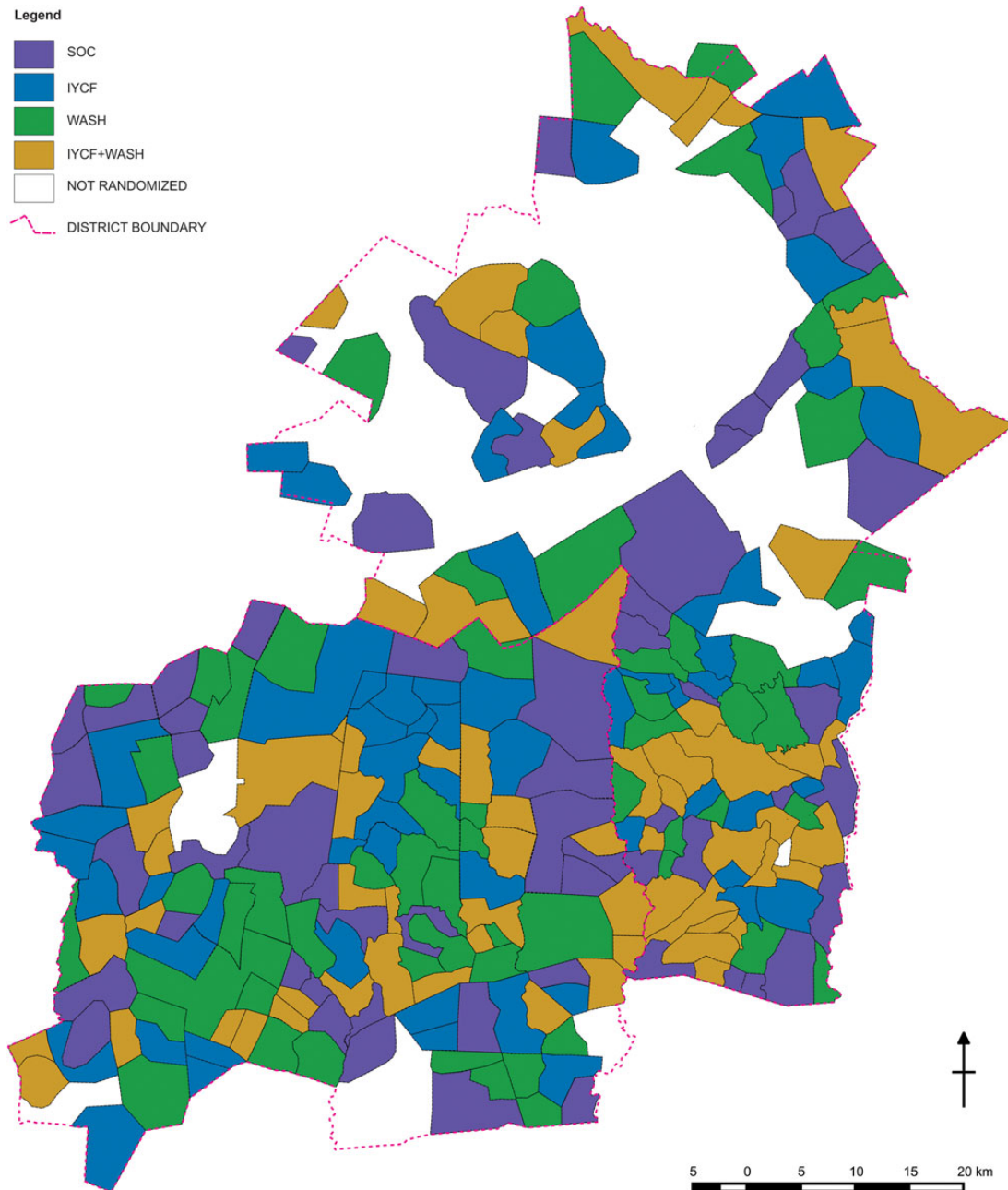


Figure 1. Randomized clusters of the Sanitation Hygiene Infant Nutrition Efficacy (SHINE) trial, Chirumanzu and Shurugwi districts, Zimbabwe. Uncolored areas were not randomized because at the time of mapping they were urban; commercial; not covered by a Ministry of Health and Child Care Village Health Worker; or uninhabited. Abbreviations: IYCF, infant and young child feeding; SOC, standard of-care; WASH, water and sanitation/hygiene.

DC conducting a research visit at least quarterly and conduct spot checks with study participants to evaluate DC performance.

Following referral of a pregnant woman to SHINE, a DC conducts a visit to confirm the woman’s pregnancy (by repeat urine test) and eligibility for the trial, and to obtain written informed consent. Illiterate women provide a thumbprint in the presence of a witness. Subsequently, DCs make 2 antenatal visits, at

approximately 10–20 weeks (baseline) and 32 weeks, and 5 post-natal visits when the infant is 1, 3, 6, 12, and 18 months of age (Table 3).

Maternal and Household Data

Maternal and household data collected at baseline include household composition, socioeconomic status, current WASH

facilities and practices, and current IYCF practices if the household has a child <2 years old. The woman is interviewed regarding her antenatal care for the current pregnancy and recent morbidity; maternal capacities and depression are assessed (see companion article [82]). Information pertaining to maternal exposure to study messages and the quality of their interaction and relationship with the VHW is ascertained. (The collection of information on fidelity of intervention delivery is detailed in a companion article [81]). Maternal height, weight, mid-upper arm circumference, and blood pressure are measured; HIV status, hemoglobin, and urinalysis are tested by point-of-care methods; biospecimens are collected as described below (and in companion article [83] and Table 3; equipment used is listed in Table 3 in the Supplementary Appendix). Maternal and household factors are reassessed at subsequent research visits as detailed in Table 3.

Birth Information

Birth information including date, mode, complications, weight, length, and head circumference are transcribed from the institutional delivery register (or the “born before arrival” register for home deliveries, who generally present for birth certification within 2 days) and also from the mother’s handheld antenatal care card. The same models (Table 3 of Supplementary Appendix 1) of infant scales and stadiometers used in trial research visits were installed by the trial in all 43 health institutions offering delivery services to women in the study area; anthropometry training is repeated every 6 months for health institution staff. For home deliveries, a visit is conducted to collect infant anthropometric measurements if SHINE learns of the delivery within 7 days. Birth dates are combined with last menstrual period date to calculate gestational age at birth and to categorize preterm birth.

Infant Data

Infant data are collected at ages 1, 3, 6, 12, and 18 months as detailed in Table 3. Infant weight, length, head circumference, and mid-upper arm circumference are measured at every postnatal visit. At 18 months of age, infant length (trial primary endpoint) is recorded 3 times. Equipment is calibrated weekly. Other infant information collected includes delivery details and birth outcomes; vaccination history; PMTCT care; 7-day morbidity history; and sick clinic and hospitalization history since last visit (eg, tuberculosis, malaria). Infant diarrhea (≥ 3 loose watery stools in 24 hours; or blood or mucus in stools) is assessed by 7-day recall and long-term recall (since prior research visit) in all infants at each postnatal visit, and by daily diary completion in a subgroup of infants (see EED substudy, section 10). Infant feeding knowledge and practice and household WASH knowledge and behaviors are reassessed by questionnaire and structured observation at every postnatal visit.

Biospecimens Collected From All Trial Participants

At baseline, maternal stool and blood specimens are obtained; urine is collected over 2 hours following ingestion of an oral dose of lactulose and mannitol, to enable measurement of the urinary recovery of sugars by mass spectrometry and computation of the LM ratio, which provides a noninvasive indicator of intestinal absorptive capacity and permeability [104]. At 32 gestational weeks, maternal blood and stool are collected. At 18 months after delivery, blood is collected from all infants and mothers. Hemoglobin concentration is measured using the point-of-care HemoCue Hb 301 hemoglobinometer (HemoCue, Angelholm, Sweden) in all infants at 18 months of age (primary endpoint).

Samples are transported in cooler boxes by motorcycle to 1 of the 4 hub laboratories where they are processed and archived in -80°C freezers; every 2 weeks, samples are transferred from hub laboratories for long-term archiving in the Zvitambo Institute for Maternal and Child Health Research (Zvitambo) laboratory in Harare. Because both districts are known to have a high prevalence of schistosomiasis among school-aged children, women are tested for *Schistosoma haematobium* infection at baseline and at 32 weeks by urinary microscopy; women testing positive are referred for postnatal praziquantel treatment per MoHCC policy. Women are offered home HIV testing by DCs at baseline and 32 gestational weeks using a rapid test algorithm approved by the MoHCC and, if positive, urgently referred to antenatal clinics to access PMTCT interventions. Women who do not wish to know their HIV status have blood tested in the field laboratory using the same rapid test algorithm, but the result is not returned to them. Women are offered a repeat HIV test at 18 months postpartum to detect seroconversion (incidence <1% in Zimbabwe). HIV-infected women are advised to take their infants to the local clinic to undertake early infant diagnosis and commence cotrimoxazole prophylaxis at 6 weeks of age; blood samples from HIV-exposed infants collected from 3 months of age onward are additionally tested for HIV by DNA polymerase chain reaction in the Zvitambo laboratory and results returned to women with appropriate counseling, unless the mother does not wish to learn her infant’s HIV status. In the case of maternal or infant deaths, a verbal autopsy is conducted with a family member and cause of death adjudicated by 2 study clinicians according to World Health Organization criteria [90].

EED SUBSTUDY OF THE BIOLOGIC CAUSAL PATHWAY

A subsample of at least 1000 HIV-unexposed infants (250 per trial arm) and all consenting HIV-exposed infants are being recruited into the EED substudy (see companion article [85]). The substudy population comprises infants of women reaching their 32 weeks’ gestation visit from 1 May 2014 through the end of

the trial. We will undertake analysis of the causal pathway separately for HIV-exposed and HIV-unexposed infants because maternal HIV infection and cotrimoxazole prophylaxis in HIV-exposed infants will likely modify the microbiota, intestinal inflammation, intercurrent infections, and systemic inflammation. HIV-unexposed infants are defined as those born to women who tested HIV-negative at baseline and 32 gestational weeks. For all dyads selected into the EED substudy, a paired maternal–infant stool and blood sample is collected at 1 month postpartum, and infant biospecimens are collected at 3, 6, 12, and 18 months of age to assess the EED pathway, as described elsewhere [83]. Mothers are asked to complete a daily morbidity diary using stickers to record episodes of infant illness (diarrhea; blood or mucus in stools; cough; fast or difficult breathing; fever; or lethargy interfering with feeding), for which they are sent a weekly cellphone reminder (to eliminate bias due to cellphone ownership, the trial provides a cellphone, solar charger, and airtime to mothers in the EED substudy). In a subgroup of 800 EED infants (all consenting EED infants reaching 3 months of age from 1 June 2015 to the end of the trial), additional specimens are collected during diarrheal episodes to evaluate the pathogens causing diarrhea in this setting using the TaqMan Array Card, and to compare pathogen-specific diarrhea rates across trial arms [105].

OBSERVATIONAL STUDIES OF OTHER EXPOSURES AND OUTCOMES WITHIN SHINE

Several observational studies are being pursued within the SHINE study population, drawing upon the infrastructure, dataset, and sample archive. Maternal schistosomiasis (assessed by urine microscopy) and maternal and infant biomarkers of mycotoxin exposure (see companion article [106]) are being measured and will be analyzed in a longitudinal observational design as potential contributors to EED and inflammation.

We also will investigate the possibility that maternal EED and/or mycotoxin exposure during pregnancy contributes to adverse birth outcomes, including miscarriage, stillbirth, preterm delivery, fetal stunting, low birth weight, and neonatal death. Pregnancy losses are ascertained prospectively and data collected on risk factors and outcomes in a specific questionnaire. These studies are planned as case-control designs; the randomized interventions will be treated as covariates in analysis.

DATA FLOW AND MANAGEMENT

DCs electronically collect questionnaire and observation data at all research visits using netbooks; questionnaires were programmed in Microsoft Access and SQL database and include quality control functions (eg, skip functions, permissible data range checks). Paper forms are available if technology fails in

the field. At the close of each working day, all new data in the netbooks are copied onto 2 computers in the hub office: one accumulates all research data and the other is linked to the internet and automatically transfers data overnight to the main server in the Zvitambo office in Harare.

Blood, fecal, saliva, and urine samples are labeled with pre-printed bar-code labels at the time of collection and transported to hub laboratories. After transfer to the Zvitambo laboratory in Harare, specimens are logged into laboratory information system prior to further processing, analysis, and archival.

STATISTICAL ANALYSIS

General Considerations

As a cluster-randomized trial, SHINE outcomes, in general, will be analyzed using statistical methods that account for within-cluster correlation. With a relatively large number of clusters (212) and for some analyses only a few events or prevalent cases per cluster, generalized estimating equations (GEEs) will be used, specifically the Stata GEE regression procedure, with exchangeable correlation structure within each cluster, for both continuous and discrete response variables. Primary analyses will contain 2 dummy variables representing each intervention; they will be unadjusted for other covariates. A log link function with Bernoulli response will be used for prevalence outcomes, to aid in interpretability by directly producing risk ratio estimates. Although the study is not powered to detect a statistical interaction effect between the 2 interventions, it will be estimated. If there is a statistically significant interaction, defined as $P < .05$ from a Wald test of the product term, we will also present results using a regression model with 3 terms to represent the 4 study arms.

In addition to these (unadjusted) primary analyses, we will report a set of adjusted estimates, conditional on baseline covariates described elsewhere [81]. The goals of such adjustment include accounting for baseline imbalances, reducing variance, and mitigating the effects of potential differential enrollment. We will also compare loss to follow-up across arms and conduct sensitivity analyses of such differential attrition [107]. We will test for effect modification by a limited number of hypothesized variables that might influence the effects of the randomized interventions in important ways. The main ones are sex of the child, and maternal HIV status (women testing HIV-positive at baseline or 32 gestational weeks vs no positive antenatal tests); women who seroconvert during the trial will be removed from the HIV-negative group in a sensitivity analysis. The theoretical basis for the additional hypotheses and our approach to assessment are described in companion papers [81, 82]. In the effect modification analyses, statistical significance will be assessed by multiplying the dummy variable for whichever intervention is being considered by the continuous/scale version of the potential modifier; the latter may then be categorized for communication purposes. For the

EED substudy, questions that address the impact of the WASH intervention will use analyses that adjust for clustering, as per the GEE regression approach described above, and path analysis, as described in a companion article [83].

Reporting of results will follow the guidelines established in the extended CONSORT guidance for cluster-randomized trials [108].

Analysis Populations

Most analyses, including the primary ones, are modified intent-to-treat at the child level. “Modified” refers to the requirement of an 18-month follow-up visit to determine the child’s anthropometric status. Loss to follow-up is anything that prevents a child’s 18-month visit from being completed, including pregnancy loss, infant death, refusal for further participation, and a permanent move outside of Zimbabwe. The analyses will be conducted according to the mother’s assigned study arm based on her residence at the time of her enrollment, regardless of subsequent migration.

Secondary analyses may require varying levels of contact and/or adherence with a focus on the following modified per-protocol population [81]: 18-month length (and hemoglobin concentration) among those who participated in at least two-thirds (10 of 15) of the VHW visits, starting at 24 gestational weeks (visit defined as having contact at a scheduled visit, with 2 attempts made per visit).

OVERSIGHT OF ETHICAL AND GCP COMPLIANCE

All research procedures are conducted in accordance with the Declaration of Helsinki and according to International Conference on Harmonisation GCP guidelines. The Medical Research Council of Zimbabwe and the Institutional Review Board of the Johns Hopkins Bloomberg School of Public Health provided initial and ongoing review and approval of the study protocol. Adverse events and serious adverse events are reported to both boards, according to their respective reporting requirements. An independent data safety and monitoring board (DSMB) was formed, comprising 2 physicians from the University of Zimbabwe and a statistician from the United Kingdom. The DSMB oversees implementation of the trial and monitors the safety and efficacy of interventions during quarterly reviews of interim data. A monitoring plan outlines the scope of internal and external monitoring for all trial activities. Internal monitoring is conducted quarterly at each field hub, and findings requiring corrective action are categorized as critical, major, or minor, with appropriate timelines for resolution. An external monitor appointed by the trial visits each hub every 6 months to conduct an inspection using his or her own monitoring and reporting tools.

Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online (<http://cid.oxfordjournals.org>). Supplementary materials consist of data provided by the author that are published to benefit the reader. The posted materials are not copyedited. The contents of all supplementary data are the sole responsibility of the authors. Questions or messages regarding errors should be addressed to the author.

Notes

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References

1. Black RE, Victora CG, Walker SP, et al. Maternal and child undernutrition and overweight in low-income and middle-income countries. *Lancet* **2013**; 382:1–25.
2. Victora CG, De Onis M, Hallal PC, Blossner M, Shrimpton R. Worldwide timing of growth faltering: revisiting implications for interventions. *Pediatrics* **2010**; 125:e473–80.
3. Beckett C, Durnin J, Aitchison TC, Pollitt E. Effects of an energy and micronutrient supplement on anthropometry in undernourished children in Indonesia. *Eur J Clin Nutr* **2000**; 54(suppl 2):S52–9.
4. Santos IS, Gigante DP, Coitinho DC, Haisma H, Valle NC, Valente G. Evaluation of the impact of a nutritional program for undernourished children in Brazil. *Cadernos de Saude Publica* **2005**; 21:776–85.
5. Lartey A, Manu A, Brown KH, Pearson JM, Dewey KG. A randomized, community-based trial of the effects of improved, centrally processed complementary foods on growth and micronutrient status of Ghanaian infants from 6 to 12 mo of age. *Am J Clin Nutr* **1999**; 70:391–404.
6. Hossain MI, Wahed MA, Ahmed S. Increased food intake after the addition of amylase-rich flour to supplementary food for malnourished children in rural communities of Bangladesh. *Food Nutr Bull* **2005**; 26:323–9.
7. Mamiro PS, Kolsteren PW, van Camp JH, Roberfroid DA, Tatala S, Opsomer AS. Processed complementary food does not improve growth or hemoglobin status of rural Tanzanian infants from 6–12 months of age in Kilosa district. *J Nutr* **2004**; 134:1084–90.
8. Owino VO, Kasonka LM, Sinkala MM, Wells JK, Eaton S, Darch T. Fortified complementary foods increases growth and hemoglobin independently of a-amylase treatment, without reducing breastmilk intake of 9-month old Zambian infants. *Am J Clin Nutr* **2007**; 86:1094–103.

9. Adu-Afarwuah S, Lartey A, Brown KH, Zlotkin S, Briend A, Dewey KG. Randomized comparison of 3 types of micronutrient supplements for home fortification of complementary foods in Ghana: effects on growth and motor development. *Am J Clin Nutr* **2007**; 86:412–20.
10. Phuka JC, Maleta K, Thakwalakwa C, et al. Complementary feeding with fortified spread reduces the incidence of severe stunting among 6- to 18-month-old rural Malawian infants. *Archives Pediatr Adolesc Med* **2008**; 162:619–26.
11. Kuusipalo H, Maleta K, Briend A, Manary M, Ashorn P. Growth and change in blood haemoglobin concentration among underweight Malawian infants receiving fortified spreads for 12 weeks: a preliminary trial. *J Pediatr Gastroenterol Nutr* **2006**; 43:525–32.
12. Ruel MT, Menon P, Habicht J-P, et al. Age-based preventive targeting of food assistance and behaviour change and communication for reduction of childhood undernutrition in Haiti: a cluster randomised trial. *Lancet* **2008**; 371:588–95.
13. Penny ME, Creed-Kanashiro HM, Robert RC, Narro MR, Caulfield LE, Black RE. Effectiveness of an educational intervention delivered through the health services to improve nutrition in young children: a cluster-randomised controlled trial. *Lancet* **2005**; 365:1863–72.
14. Roy SK, Fuchs GJ, Mahmud S, Ara G, Islam S, Shafique S. Intensive nutrition education with or without supplementary feeding improves the nutritional status of moderately-malnourished children in Bangladesh. *J Health Popul Nutr* **2005**; 23:320–30.
15. Bhandari N, Bahl R, Nayyar B, Khokhar P, Rohde JE, Bhan MK. Food supplementation with encouragement to feed it to infants from 4 to 12 months of age has a small impact on weight gain. *J Nutr* **2001**; 131:1946–51.
16. Pelto GH, Levitt E, Thairu L. Improving feeding practices: current patterns, common constraints, and the design of interventions. *Food Nutr Bull* **2003**; 24:45–82.
17. Dewey KG, Adu-Afarwuah S. Systematic review of the efficacy and effectiveness of complementary feeding interventions in developing countries. *Matern Child Nutr* **2008**; 4:24–85.
18. Richard SA, Black RE, Gilman RH, et al. Diarrhea in early childhood: short-term association with weight and long-term association with length. *Am J Epidemiol* **2013**; 178:1129–38.
19. WHO Multicentre Growth Reference Study Group. WHO Child Growth Standards based on length/height, weight and age. *Acta Paediatr* **2006**; 450(Suppl):76–85.
20. Walker SP, Wachs TD, Grantham-McGregor S, et al. Inequality in early childhood: risk and protective factors for early child development. *Lancet* **2011**; 378:1325–38.
21. Lozano R, Naghavi M, Foreman K, et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* **2013**; 380:2095–128.
22. Siegel EH, Stoltzfus RJ, Khatri SK, LeClerq SC, Katz J, Tielsch JM. Epidemiology of anemia among 4- to 17-month-old children living in south central Nepal. *Eur J Clin Nutr* **2006**; 60:228–35.
23. World Health Organization. Worldwide prevalence of anaemia 1993–2005: WHO global database on anaemia. Geneva, Switzerland: WHO, **2008**.
24. World Health Organization. Iron deficiency anaemia: assessment, prevention and control: a guide for programme managers. Geneva, Switzerland: WHO, **2001**.
25. Gera T, Sachdev HPS, Nestel P, Sachdev SS. Effect of iron supplementation on haemoglobin response in children: systematic review of randomised controlled trials. *J Pediatr Gastroenterol Nutr* **2007**; 44:468–86.
26. Humphrey JH. Child undernutrition, tropical enteropathy, toilets, and handwashing. *Lancet* **2009**; 374:1032–5.
27. Sprinz H, Sribhibhadh R, Gangarosa EJ, Benyajati C, Kundel D, Halstead S. Biopsy of small bowel of Thai people: with special reference to recovery from Asiatic cholera and to an intestinal malabsorption syndrome. *Am J Med* **1962**; 38:43–51.
28. Lindenbaum J, Alam AKMJ, Kent TH. Subclinical small-intestinal disease in East Pakistan. *Brit Med J* **1966**; 2:1616–9.
29. Walravens PA, Chakar A, Mokni R, Denise J, Lemmonier D. Zinc supplements in breastfed infants. *Lancet* **1992**; 340:683–5.
30. Baker SJ, Mathan VI. Syndrome of tropical sprue in South India. *Am J Clin Nutr* **1968**; 21:984–93.
31. Chaves FJZC, Veloso FT, Cruz I, et al. Subclinical tropical enteropathy in Angola: peroral jejunal biopsies and absorption studies in asymptomatic healthy men. *Mt Sinai J Med* **1981**; 48:47–52.
32. Cook GC, Kajubi SK, Lee FD. Jejunal morphology of the African in Uganda. *J Pathol* **1969**; 98:157–71.
33. Falaiye JM. Present status of subclinical intestinal malabsorption in the tropics. *BMJ* **1971**; 4:454–8.
34. Thomas G, Clain DJ, Wicks ACB. Tropical enteropathy in Rhodesia. *Gut* **1976**; 17:888–94.
35. Bayless TM, Wheby MS, Swanson VL. Tropical sprue in Puerto Rico. *Am J Clin Nutr* **1968**; 21:1030–41.
36. Schenk EA, Klipstein FA, Tomasini JT. Morphologic characteristics of jejunal biopsies from asymptomatic Haitians and Puerto Ricans. *Am J Clin Nutr* **1972**; 25:1080–3.
37. Chacko CJG, Paulson KA, Mathan VI, Baker SJ. The villus architecture of the small intestine in the tropics: a necropsy study. *J Pathol* **1969**; 98:146–51.
38. Rhodes AR, Shea N, Lindenbaum J. Malabsorption in asymptomatic Liberian children. *Am J Clin Nutr* **1971**; 24:574–7.
39. Haghighi P, Wolf PL. Tropical sprue and subclinical enteropathy: a vision for the nineties. *Crit Rev Clin Lab Sci* **1997**; 34:313–41.
40. Prendergast A, Kelly P. Enteropathies in the developing world: neglected effects on global health. *Am J Trop Med Hyg* **2012**; 86:756–63.
41. Baker SJ. Subclinical intestinal malabsorption in developing countries. *WHO Bull* **1976**; 54:485–94.
42. Lunn PG, Northrop-Clewes CA, Downes RM. Growth in rural Gambian infants is closely related to small-intestinal permeability: why? *Proc Nutr Soc* **1997**; 56:280A.
43. Hutton G, Haller L. Evaluation of the costs and benefits of water and sanitation improvements at the global level. Geneva, Switzerland: WHO, **2004**. WHO/SDE/WSH/04.04.
44. Lunn PG, Northrop-Clewes CA, Downes RM. Intestinal permeability, mucosal injury, and growth faltering in Gambian infants. *Lancet* **1991**; 338:907–10.
45. Prentice A. Nutrient requirements for growth, pregnancy and lactation: the Keneba experience. *S Afr J Clin Nutr* **1993**; 6:33–8.
46. Sullivan PB, Mascie-Taylor CGN, Lunn PG, Northrop-Clewes CA, Neale G. The treatment of persistent diarrhoea and malnutrition: long-term effects of in-patient rehabilitation. *Acta Paediatr Scand* **1991**; 80:1025–30.
47. Campbell DI, McPhail G, Lunn PG, Elia M, Jeffries DJ. Intestinal inflammation measured by fecal neopterin in Gambian children with enteropathy: association with growth failure, *Giardia lamblia* and intestinal permeability. *J Pediatr Gastro Nutr* **2004**; 39:153–7.
48. Campbell DI, Elia M, Lunn PG. Growth faltering in rural Gambian infants is associated with impaired small intestinal barrier function, leading to endotoxemia and systemic inflammation. *J Nutr* **2003**; 133:1332–8.
49. Campbell DI, Lunn PG, Elia M. Age-related association of small intestinal mucosal enteropathy with nutritional status in rural Gambian children. *Br J Nutr* **2002**; 88:499–505.
50. Behrens RH, Lunn PG, Northrop CA, Hanlon PW, Neale G. Factors affecting the integrity of the intestinal mucosa of Gambian children. *Am J Clin Nutr* **1987**; 45:1433–41.
51. Lunn PG, Northrop-Clewes CA, Downes RM. Recent developments in the nutritional management of diarrhoea: 2. Chronic diarrhoea and malnutrition in the Gambia: studies on intestinal permeability. *Trans R Soc Trop Med Hyg* **1991**; 85:8–11.
52. Weaver LT. The impact of milk and weaning diet on gastro-intestinal permeability in English and Gambian infants. *Trans R Soc Trop Med Hyg* **1998**; 82:784–9.

53. Kosek M, Haque R, Lima A, et al. Fecal markers of intestinal inflammation and permeability associated with the subsequent acquisition of linear growth deficits in infants. *Am J Trop Med Hyg* **2013**; 88:390–6.
54. de Benedetti F, Alonzi T, Moretta A, et al. Interleukin 6 causes growth impairment in transgenic mice through a decrease in insulin-like growth factor-I: a model for stunted growth in children with chronic inflammation. *J Clin Invest* **1997**; 99:643–50.
55. Walters TD, Griffiths AM. Mechanisms of growth impairment in pediatric Crohn's disease. *Nat Rev Gastroenterol Hepatol* **2009**; 6:513–23.
56. Nemeth E, Rivera S, Gabayan V, et al. IL-6 mediates hypoferrremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J Clin Invest* **2004**; 113:1271–6.
57. Nemeth E, Tuttle MS, Powelson J, et al. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* **2004**; 306:2090–3.
58. Prentice AM, Doherty CP, Abrams SA, et al. Hepcidin is the major predictor of erythrocyte iron incorporation in anemic African children. *Blood* **2012**; 119:1922–8.
59. Jelkmann W, Wolff M, Fandrey J. Modulation of the production of erythropoietin by cytokines: in vitro studies and their clinical implications. *Contrib Nephrol* **1989**; 87:68–77.
60. Galpin L, Manary MJ, Fleming K, Ou C-N, Ashorn P, Shulman RJ. Effect of *Lactobacillus* GG on intestinal integrity in Malawian children at risk of tropical enteropathy. *Am J Clin Nutr* **2005**; 82:1040–5.
61. Lin A, Arnold BF, Afreen S, et al. Household environmental conditions are associated with enteropathy and impaired growth in rural Bangladesh. *Am J Trop Med Hyg* **2013**; 89:130–7.
62. Omer-Salim A, Persson LÅK, Olsson P. Whom can I rely on? Mothers' approaches to support for feeding: an interview study in suburban Dar es Salaam, Tanzania. *Midwifery* **2007**; 23:172–83.
63. Beall J, Crankshaw O, Parnell S. Victims, villains and fixers: the urban environment and Johannesburg's poor. *J South Afr Stud* **2000**; 26:803–55.
64. Esrey SA. Water, waste, and well-being: a multicountry study. *Am J Epidemiol* **1996**; 143:608–23.
65. Checkley W, Gilman RH, Black RE, et al. Effect of water and sanitation on childhood health in a poor Peruvian peri-urban community. *Lancet* **2004**; 363:112–8.
66. Daniels DL, Cousens SN, Makoae LN, Feachem RG. A study of the association between improved sanitation facilities and children's height in Lesotho. *Eur J Clin Nutr* **1991**; 45:23–32.
67. Magnani RJ, Mock NB, Bertrand WE, Clay DC. Breast-feeding, water and sanitation, and childhood malnutrition in the Philippines. *J Biosoc Sci* **1993**; 25:195–212.
68. Merchant AT, Jones C, Kiure A, et al. Water and sanitation associated with improved child growth. *Eur J Clin Nutr* **2003**; 57:1562–8.
69. Ngure FM, Humphrey JH, Menon P, Stoltzfus R. Environmental hygiene, food safety and growth in less than five year old children in Zimbabwe and Ethiopia. *FASEB J* **2013**; 27:243.2.
70. Fenn B, Bulti AT, Nduna T, Duffield A, Watson F. An evaluation of an operations research project to reduce childhood stunting in a food-insecure area in Ethiopia. *Public Health Nutr* **2012**; 15:1746–54.
71. Spears D, Ghosh A, Cummings O. Open defecation and childhood stunting in India: an ecological analysis of new data from 112 districts. *PLoS One* **2013**; 8:1–9.
72. Dangour AD, Watson L, Cumming O, et al. Interventions to improve water quality and supply, sanitation and hygiene practices, and their effects on the nutritional status of children. *Cochrane Database Syst Rev* **2013**; 8.
73. Pickering AJ, Davis J. Freshwater availability and water fetching distance affect child health in sub-Saharan Africa. *Environ Sci Technol* **2012**; 46:2391–7.
74. Roura E, Homedes J, Klasing KC. Prevention of immunologic stress contributes to the growth-permitting ability of dietary antibiotics in chicks. *J Nutr* **1992**; 122:2383–90.
75. MacDonald TT, Spencer J. Evidence that activated mucosal T cells play a role in the pathogenesis of enteropathy in human small intestine. *J Exp Med* **1988**; 167:1341–9.
76. Lionetti P, Breese E, Braegger CP, Murch SH, Taylor J, MacDonald TT. T-cell activation can induce either mucosal destruction or adaptation in cultured human fetal small intestine. *Gastroenterology* **1993**; 105:373–81.
77. Ferreira RC, Forsyth LE, Wells C, Spencer J, MacDonald TT. Changes in the rate of crypt epithelial cell proliferation and mucosal morphology induced by a T-cell-mediated response in human small intestine. *Gastroenterology* **1990**; 98:1255–63.
78. Halstensen TS, Brandtzaeg P. Activated T lymphocytes in the celiac lesion: non-proliferative activation (CD25) of CD4+ a/b cells in the lamina propria but proliferation (Ki-67) of a/b and g_d cells in the epithelium. *Euro J Immunol* **1993**; 23:505–10.
79. Fagundes Neto U, Martins MC, Lima FL, Patricio FR, Toledo MR. Asymptomatic environmental enteropathy among slum-dwelling infants. *J Am Coll Nutr* **1994**; 13:51–6.
80. Sprinz H, Kundel DW, Dammin GJ, Horowitz RE, Schneider H, Formal SB. The response of the germ-free guinea pig to oral bacterial challenge with *Escherichia coli* and *Shigella flexneri*: with special reference to lymphatic tissue and the intestinal tract. *Am J Pathol* **1961**; 39:681–95.
81. Mbuya MNN, Jones AD, Ntozini R, et al. Theory-driven process evaluation of the SHINE Trial using a program impact pathway approach. *Clin Infect Dis* **2015**; 61(suppl 7):S752–8.
82. Matare CR, Mbuya MNN, Pelto G, Dickin KL, Maluccio JA, Stoltzfus RJ, for the Sanitation Hygiene Infant Nutrition Efficacy (SHINE) Trial Team. Assessing maternal capabilities in the SHINE Trial: highlighting a hidden link in the causal pathway to child health. *Clin Infect Dis* **2015**; 61(suppl 7):S745–51.
83. Prendergast AJ, Humphrey JH, Mutasa K, et al. Assessment of environmental enteric dysfunction in the SHINE trial: methods and challenges. *Clin Infect Dis* **2015**; 61(suppl 7):S726–32.
84. Arnold BF, Null C, Luby SP, et al. Cluster-randomised controlled trials of individual and combined water, sanitation, hygiene and nutritional interventions in rural Bangladesh and Kenya: the WASH Benefits study design and rationale. *BMJ Open* **2013**; 3:e003476.
85. Mbuya MNN, Tavengwa NV, Stoltzfus RJ, et al. Design of an intervention to minimize ingestion of fecal microbes by young children in rural Zimbabwe. *Clin Infect Dis* **2015**; 61(suppl 7):S703–9.
86. Desai A, Smith LE, Mbuya MNN, et al. The SHINE Trial infant feeding intervention: pilot study of effects on maternal learning and infant diet quality in rural Zimbabwe. *Clin Infect Dis* **2015**; 61(suppl 7):S710–5.
87. Britt E, Hudson SM, Blampied NM. Motivational interviewing in health settings: a review. *Patient Educ Couns* **2004**; 53:147–55.
88. Auger R, Curtis V. The Evo-Eco approach to behaviour change. *Applied evolutionary anthropology*. New York: Springer, **2014**:271–95.
89. Knowles MS. The modern practice of adult education. From pedagogy to andragogy. Englewood Cliffs, NJ: Prentice Hall/Cambridge, **1980**.
90. Bang AT, Bang RA. Diagnosis of causes of childhood deaths in developing countries by verbal autopsy: suggested criteria. The SEARCH Team. *Bull WHO* **1992**; 70:499.
91. Desai A, Mbuya MN, Chigumira A, et al. Traditional oral remedies and perceived breast milk insufficiency are major barriers to exclusive breastfeeding in rural Zimbabwe. *J Nutr* **2014**; 144:1113–9.
92. Matare C, Mbuya M, Stoltzfus R, Dickin K, Humphrey J. A village health worker-led intervention to promote and support exclusive breastfeeding in rural Zimbabwe is feasible, acceptable and effective. *FASEB J* **2014**; 28:1015.8.
93. Morgan PR. A ventilated pit privy. *Appropriate Technol* **1979**; 6:10–1.
94. Arimond M, Zeilani M, Jungjohann S, et al. Considerations in developing lipid-based nutrient supplements for prevention of undernutrition:

- experience from the International Lipid-Based Nutrient Supplements (iLiNS) Project. *Matern Child Nutr* **2013**; doi:10.1111/mcn.12049.
95. Zimbabwe National Statistics Agency. Census 2012: preliminary report, **2012**. Available at: http://www.zimstat.co.zw/index.php?option=com_content&view=article&id=65:census. Accessed 12 October 2015.
 96. Zimbabwe National Statistics Agency and ICF International Inc. Zimbabwe Demographic and Health Survey 2010–2011. Available at: <http://www.measuredhs.com/publications/publication-FR254-DHS-Final-Reports.cfm>. Accessed 15 December 2012.
 97. Ministry of Health and Child Welfare, Ministry of Education, Sports and Culture. Report on the national soil transmitted helminthiasis and schistosomiasis survey. Harare, Zimbabwe: MOHCW, MOESC, **2010**.
 98. Ntozini R, Marks SJ, Mangwando G, et al. Using geographic information systems and spatial analysis methods to assess household water access and sanitation coverage in the SHINE trial. *Clin Infect Dis* **2015**; 61(suppl 7):S716–25.
 99. Hayes RJ, Bennett S. Simple sample size calculation for cluster-randomized trials. *Int J Epidemiol* **1999**; 28:319–26.
 100. Katz J. Sample-size implications for population-based cluster surveys of nutritional status. *Am J Clin Nutr* **1995**; 61:155–60.
 101. Siegel JD, Rhinehart E, Jackson M, Chiarello L; Healthcare Infection Control Practices Advisory Committee. Management of multidrug-resistant organisms in health care settings, 2006. *Am J Infect Control* **2007**; 35:S165–93.
 102. Moulton LH. Covariate-based constrained randomization of group-randomized trials. *Clin Trials* **2004**; 1:297–305.
 103. World Health Organization. IMCI guidelines, **2015**. Available at: http://www.who.int/maternal_child_adolescent/topics/child/imci/en/. Accessed 15 September 2015.
 104. Denno DM, VanBuskirk K, Nelson ZC, Musser CA, Hay Burgess DC, Tarr PI. Use of the lactulose to mannitol ratio to evaluate childhood environmental enteric dysfunction: a systematic review. *Clin Infect Dis* **2014**; 59(suppl 4):S213–9.
 105. Liu J, Kabir F, Manneh J, et al. Development and assessment of molecular diagnostic tests for 15 enteropathogens causing childhood diarrhoea: a multicentre study. *Lancet Infect Dis* **2014**; 14:716–24.
 106. Smith LE, Stoltzfus RJ, Turner P, et al. The potential role of mycotoxins as a contributor to stunting in the SHINE trial. *Clin Infect Dis* **2015**; 61(suppl 7):S733–7.
 107. Seaman SR, White IR. Review of inverse probability weighting for dealing with missing data. *Stat Methods Med Res* **2013**; 22: 278–95.
 108. Campbell MK, Piaggio G, Elbourne DR, Altman DG. Consort 2010 statement: extension to cluster randomised trials. *BMJ* **2012**; 345: e5661.

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