

# Association of Betel Nut with Carcinogenesis: Revisit with a Clinical Perspective

Rajeshwar N. Sharan<sup>1\*</sup>, Ravi Mehrotra<sup>2</sup>, Yashmin Choudhury<sup>3</sup>, Kamlesh Asotra<sup>4</sup>

**1** Radiation and Molecular Biology Unit, Department of Biochemistry, North-Eastern Hill University, Shillong, Meghalaya, India, **2** Division of Cytopathology, Center for Oral Cancer and Precancer, Moti Lal Nehru Medical College, Allahabad, Uttar Pradesh, India, **3** Department of Biotechnology, Assam University, Silchar, Assam, India, **4** Ganeshiva Consultants, Los Angeles, California, United States of America

## Abstract

Betel nut (BN), betel quid (BQ) and products derived from them are widely used as a socially endorsed masticatory product. The addictive nature of BN/BQ has resulted in its widespread usage making it the fourth most abused substance by humans. Progressively, several additives, including chewing tobacco, got added to simple BN preparations. This addictive practice has been shown to have strong etiological correlation with human susceptibility to cancer, particularly oral and oropharyngeal cancers. The PUBMED database was searched to retrieve all relevant published studies in English on BN and BQ, and its association with oral and oropharyngeal cancers. Only complete studies directly dealing with BN/BQ induced carcinogenesis using statistically valid and acceptable sample size were analyzed. Additional relevant information available from other sources was also considered. This systematic review attempts to put in perspective the consequences of this widespread habit of BN/BQ mastication, practiced by approximately 10% of the world population, on oral cancer with a clinical perspective. BN/BQ mastication seems to be significantly associated with susceptibility to oral and oropharyngeal cancers. Addition of tobacco to BN has been found to only marginally increase the cancer risk. Despite the widespread usage of BN/BQ and its strong association with human susceptibility to cancer, no serious strategy seems to exist to control this habit. The review, therefore, also looks at various preventive efforts being made by governments and highlights the multifaceted intervention strategies required to mitigate and/or control the habit of BN/BQ mastication.

**Citation:** Sharan RN, Mehrotra R, Choudhury Y, Asotra K (2012) Association of Betel Nut with Carcinogenesis: Revisit with a Clinical Perspective. PLoS ONE 7(8): e42759. doi:10.1371/journal.pone.0042759

**Editor:** Sudha Agarwal, Ohio State University, United States of America

**Received:** April 5, 2012; **Accepted:** July 11, 2012; **Published:** August 13, 2012

**Copyright:** © 2012 Sharan et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** Part funding from North Eastern Hill University, UPE (NEHU) and DST, which were utilized for some of the works mentioned in the manuscript, are acknowledged. No additional external funding was received for this study. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: msharan@nehu.ac.in

## Introduction

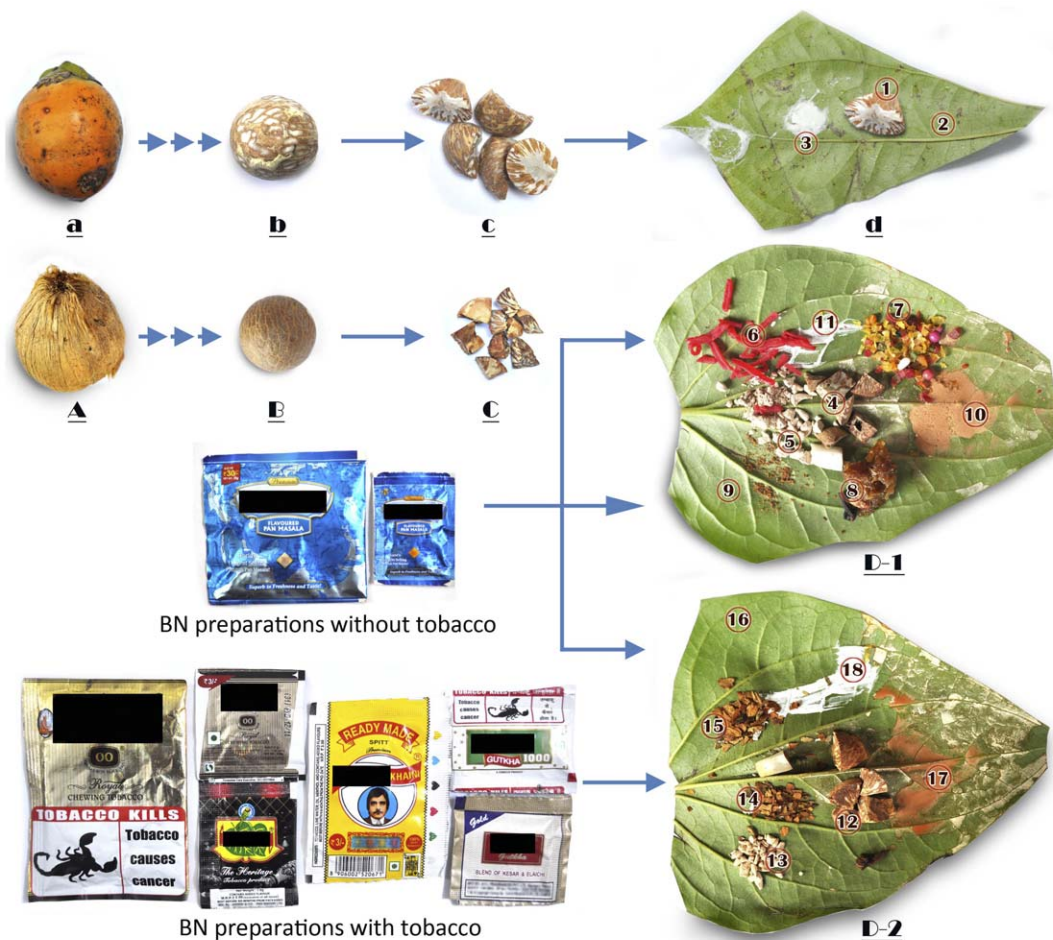
*“They are always chewing Arecca, a certaine Fruit like a Peare, cut in quarters and rolled up in leaves of a Tree called Bette (or Vettele), like Bay leaves; which having chewed they spit forth. It makes the mouth red. They say they do it to comfort the heart, nor could live without it.”—Pigafetta, in Purchas, i. 38. [Circa 1521] [1]*

The above quotation gives a vivid description of the *Areca* nut (AN), which is more commonly referred to as betel nut (BN). BN is masticated or chewed either alone or in combination with a wide variety of additives, which are often wrapped in the leaf of *Piper betle* (popularly called as betel leaf), giving it the more common name, betel quid (BQ) [2,3]. More significantly, the verse also alludes to the rampant use of BN or BQ as a masticatory product and its probable addictive nature.

BN is normally harvested as unripe (yellow-green) or ripe (orange/red) fruit from the tropical palm, *Areca catechu*. The *Areca* fruits may be sun dried for several weeks, fibrous shells removed and the hard, dry nuts, commonly called ‘*supari*’ in India, are ready for use. Such sun dried variety of BN is very hard, and is cut into small pieces to make it easier to masticate. Alternatively, the ripe

BN are boiled for several hours in an aqueous solution containing the bark of the plant *Eugenia jambolana*, jaggery or brown sugar, and various edible oils, to ‘cure’ it. The cured fruits are sun dried for several weeks, fibrous shell removed and very hard, brown nuts, another variety of ‘*supari*’, are ready for use. In contrast, ripe, partly ripe or unripe *Areca* fruits are freshly picked, fibrous shells removed and the relatively soft nuts are ready for mastication (Figure 1). Occasionally, the fruits can be cured by burying them into moist pits for 1–2 weeks for fermentation (maturation) before shelling and use. Such raw and wet variety of BN, widely used particularly in the northeastern part of India, is locally called ‘*kawai*’ or ‘*tambul*’. These are relatively soft, and hence, larger pieces of the nut are masticated. Aged people may, at times, even masticate powdered form of the raw/wet or dry variety of BN [2–5].

The BN is consumed either alone or as BQ in which case it is wrapped in a betel leaf along with slaked lime (Calcium oxide and Calcium hydroxide) and additives (Figure 1). The BQ with a variety of additives is commonly referred to as ‘*paan*’ in India. The components of BQ can vary widely between countries, regions, communities and individuals. However, the major constituents are BN, catechu (*Acacia catechu*), a resinous extract from the wood of the *Acacia* tree, slaked lime, various additives, such as grated coconut, aniseed, pepper mint, cardamom, cloves, perfumes and stimulants wrapped in betel leaf (Figure 1) [2–5]. In India, most



**Figure 1. Betel nut (BN), betel quid (BQ) and different preparations associated with its mastication, including their commercial reincarnations.** The unripe *Areca* fruit (a), either directly or after short curing is shelled to get wet and soft BN (b) (*tambul* or *kwai*), which after cutting into 4–5 pieces (c & 1) is normally consumed with a piece of betel leaf (2) and slaked lime (3) making a simple BQ (d). The ripe *Areca* fruit (A), after drying and curing is shelled to get dry and hard nut (B), which is cut into smaller pieces (C) (*supari*) for mastication. The dry pieces of BN (4 & 12) are usually masticated with a variety of additives (5–8), all of which usually contain BN, on a betel leaf (9) supplemented with catechu (10) and slaked lime (11) in a complex BQ (D-1). A variant of the complex BQ (D-2) may include all of the above plus a variety of chewing tobacco additives (13–15). Commercialization of this widespread practice of BQ mastication has led to mushrooming production of convenient and inexpensive alternate forms of BN preparations without (*paan masala*) or with tobacco (*gutkha*). Few of these products, packages in sachets (shown) or containers of various sizes (not shown), which are widely available in markets in India are shown here. All these products have no standardized production frame or declaration of nutritional components. See text for details. doi:10.1371/journal.pone.0042759.g001

habitual chewers of BQ add tobacco, while in some countries, such as Papua New Guinea and China, tobacco is normally not added [6]. In northeastern India, *kwai* or *tambul* is primarily consumed only with betel leaf and slaked lime (Figure 1) [5]. Betel leaf is perishable and the preparation of BQ is somewhat complex. Hence, over the past few decades, commercial BQ substitutes, a flavored and sweetened dry mixture of BN, catechu and slaked lime either with tobacco (*gutkha* or *khaini*) or without tobacco (*paan masala*), have become increasingly popular [6]. These products are packaged in small, attractive and inexpensive sachets, and are easily advertised and marketed, often claimed to be safe products (Figure 1). Use of *gutkha* often begins at a very young age. *Gutkha* contains large amounts of sweeteners to conceal the bitterness of tobacco, and children often consider it as a type of candy. Many people take *gutkha* to be harmless and mere ‘mouth freshener’ [7]. *Gutkha* and *paan masala* are consumed by very young and old alike, particularly in India, and also among migrant populations from these areas worldwide [6]. It has been reported that in the Hunan

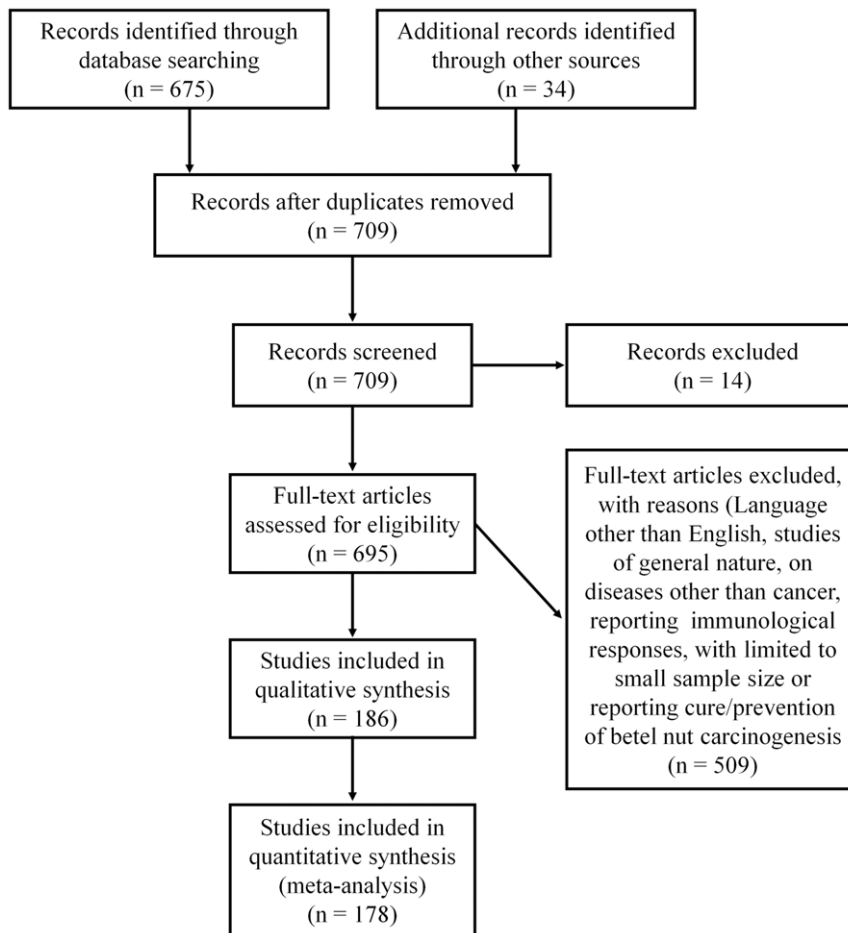
province of China, the shell of the *Areca* fruit is not removed before consumption. Three main variants of BQ are prepared - husk without BN (kernel, seed, endosperm) being the most common, husk with other substances e.g. dried grapes, and husk with BN [8].

Several studies have reported the effect of BN and its constituents on human health, especially as a possible cause of oral cancer (OC) [2–5,9]. This review focuses on the consequences of BN chewing with a clinical perspective and seeks to bring into perspective the strategies required as well as those adopted by different countries in order to curb the growing hazards supposed to be ensuing from their usage.

## Methods

### 1. SEARCH STRATEGY

The studies included in this review have been retrieved from the PUBMED database of the National Library of Medicine, National



**Figure 2. Flow chart of included studies.** The flow chart depicts the number of citations and resource materials that have been screened, excluded and/or included in the systematic review.  
doi:10.1371/journal.pone.0042759.g002

Institutes of Health, Bethesda, Maryland, by setting limits for papers published only from 1985 to 2012, using the keywords “Betel nut AND cancer”. The search was refined using the operator ‘AND’ in order to retrieve the desired results. This search yielded 675 records (see Figure 2; Flow Chart of included studies).

## 2. EXCLUSION AND INCLUSION CRITERIA

Criteria for exclusion were reports in languages other than English, reports for which abstract was not available, studies pertaining to association of *Arecia* with studies other than cancer, studies which investigated the association between cancer or cancer risk with other factors such as tobacco chewing, smoking and alcohol consumption only, without considering BN exposure, studies which focused on single or limited cases, studies which delved into the immunological responses to BN chewing, studies which investigated the treatment of OC caused by BN chewing, and studies of general interest featuring BN without showing a clear role of BN in the pathogenesis of cancer/pre-cancerous conditions. Ultimately, a total number of 148 reports indexed in PUBMED were found to satisfy the criteria for inclusion. Several pertinent reports not indexed in PUBMED were obtained by manual searches, and 30 such reports which satisfied the criteria for inclusion were further retrieved. Thus, the total number of reports included in this review is 178 (see details in Figure 2; Flow chart of included studies).

## Results

### 1. HISTORY OF BETEL NUT AND BETEL QUID USAGE

The earliest use of BN as a masticatory object by humans has been mentioned by Theophrastus in scripts dating around 430 BCE (Before Common Era), which described it as a component of the betel morsel. Chinese texts of 150 BCE, also mention BN as “*pinlang*”. In Persia (modern Iran), it is believed that around 30,000 shops sold BN in the capital town during the reign of Khosrau II, the King of Persia during 590–628 AD. The use of BN by humans has been known since the 4<sup>th</sup> century AD in different parts of the world, including the South and South-East Asia, several Pacific islands, many regions of the former Soviet Union, parts of North America and Europe [2,5], and is deeply ingrained in many socio-cultural and religious activities [2–5]. A study of the ‘Skull from Bangkok’ collected by Rudolf Virchow (Berlin, Germany) in the late 19<sup>th</sup> century, as part of an extensive anthropological collection of skeletons and skulls from all over the world, shows brown black stains because of BQ chewing, on the few remaining teeth of the maxilla. In fact, an extensive number of skulls from the South- and Southeast Asia in the collection have been found to show similar betel stains. The Skull from Bangkok is a proof that BQ chewing was prevalent in Siam of the late 19<sup>th</sup> century [10]. BN is used by both men and women though in some societies the latter predominate, across all age groups, and social

classes [3]. In fact, only three other ‘addictive’ substances, namely nicotine, alcohol and caffeine, are used more widely than BN/BQ in the world today [11].

## 2. DEMOGRAPHICS

It is estimated that currently 10% of the world population or nearly 700 million individuals might be consuming BN in different forms across the globe [2,4,5]. Epidemiological surveys show that in the past 2 to 3 decades, 20–40% of the population in India, Nepal and Pakistan have used BQ. India has the largest BN chewing population in the world. In fact, aggressive marketing, easy availability and reasonable price of *guthka* have made it very attractive to youth, and an alarmingly high number of children and teenagers in India. According to one estimate, as many as one in three individuals regularly or occasionally chew *guthka* [7]. Although a decreasing trend in the consumption of BQ has been observed in certain countries or regions, such as in Thailand, an alarmingly high chewing prevalence has been found among the Palauans of the West Pacific (72–80% use of BQ, with 80% found to be consumers of tobacco-added mixtures) [12]. In China, BQ chewing is largely prevalent in the Hunan province [8]. An inter-country Asian Betel-quinid Consortium study (the ABC study) was conducted for East Asia: Taiwan, Mainland China, Malaysia, Indonesia and South Asia: Nepal and Sri Lanka [12]. Chewing rates among men (10.7–43.6%) were significantly higher than women (1.8–34.9%) in Taiwan, Mainland China, Nepal and Sri Lanka, while women’s rates (29.5–46.8%) were higher than that in men (9.8–12.0%) in Malaysia and Indonesia. An emerging, large group of new users has been identified in Hunan province in the Mainland China (11.1–24.7%), where chewers have the unique practice of using the dried husk of *Areca* fruit rather than the solid nut used by others. Although the raw *Areca* used in Hunan province is imported from Hainan in Mainland China as well as from Thailand, the vast majority of BQ products in China, including commercial forms, are manufactured locally. The Xiangtan city in Hunan province, where BQ chewing was reported to be very common (prevalence 64.5–82.7%), is also where most BQ production factories and workshops are located. The prevalence of BQ chewing in Hunan men was higher in the younger age groups, suggesting that Hunan province is an emerging region of BQ usage. The improving economy and easy access to BQ products there, supplemented by aggressive advertising campaigns, could be the factors responsible for the widespread use of this substance, particularly among young people. Men in the Eastern and South Asian study communities were deemed likely to combine chewing with smoking and drinking (5.6–13.6%). Low level of school education, alcohol drinking and tobacco smoking were identified as factors associated with BQ chewing [12].

South Asian immigrants in Australia, Europe, the United Kingdom, South and East Africa and the Malay Peninsula continue using BN products, including *paan* and *guthka*, long after immigration [7]. The United Kingdom is the leading importer of *guthka* outside of Asia, with imports having doubled in the last three decades. The sale and use of *paan* and *guthka* are legal in the United States and they are readily available in ethnic enclaves, widely used by the large and rapidly growing South Asian communities [7].

## 3. CONSTITUENTS OF BETEL NUT AND ITS ACTIVE PRINCIPLES

The constituents of BN include crude fiber, carbohydrates, fats, polyphenols, alkaloids, tannins, proteins and water. Trace amounts of fluorine, saponinins (glycosidic derivatives of steroids

and triterpenoids) and free amino acids have also been reported in some forms. The relative amounts of these constituents are highly variable in produce of different regions as well as in the dry or raw/wet variety of BN. Geographical and climatic conditions of growth of the *Areca* palm tree and the methods of curing BN are main factors that contribute to the observed variation in the constituents [5]. The raw and wet variety of BN is relatively rich in all constituents as compared to the dry variety [2,5]. Notwithstanding these variations, the active components of both forms of BN, which produce BN associated effects, are primarily the alkaloids, polyphenols, and tannins (Figure 3). Figure 3 also highlights the outlines of the main events triggered in a living cell upon exposure to BN and/or its components that eventually lead to carcinogenic transformation of the cell (for details see reviews 2–5).

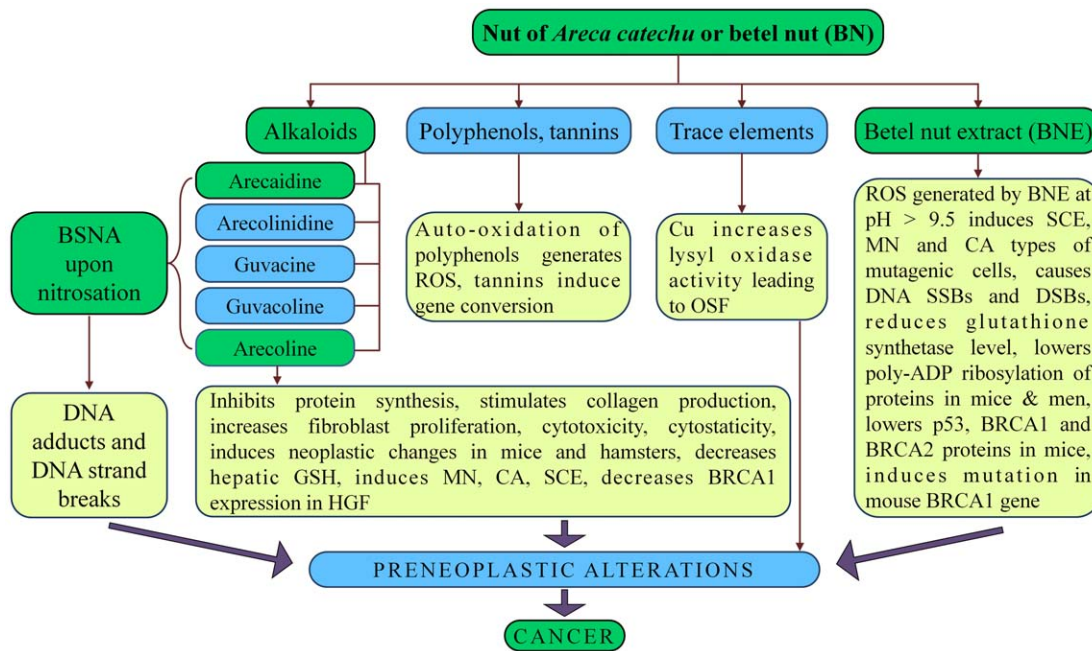
**(a) Alkaloids.** Alkaloids are reduced pyridines. BN contains several alkaloids, of which arecoline and arecaidine are biologically highly relevant. Arecoline (1,2,4,5-tetrahydro-1-methyl-pyridinecarboxylic acid; molecular weight 155.19 Da) is the most abundant alkaloid of BN followed by arecaidine (1,2,5,6-tetrahydro-1-methyl-3-pyridinecarboxylic acid; molecular weight 141.17 Da). Other alkaloids, such as guvacine (methyl ester of arecaidine), guvacoline (methyl ester of guvacine) and arecolinidine are also present in small to very small or trace amounts [5]. The amount of alkaloid in BN varies with seasonal and geographical variations. In an aqueous extract of Taiwanese BQ composed of fresh BN, betel inflorescence and red lime paste (80.5:12.5:7 by weight), arecaidine was the most abundant alkaloid (7.53 mg/g dry weight) and guvacoline the least abundant (0.26 mg/g dry wt.). Cold storage or freeze drying does not bring about alterations in the amount of alkaloids. However, arecoline content is reduced variably following processing of the nut by different methods in different regions of the world. The alkaloids may be converted to several derivatives, each of which can potentially produce even more diazohydroxide derivatives. Presence of most of these derivatives has been demonstrated in the saliva of BQ chewers [4,9,13].

**(b) Polyphenols.** Catechin, flavanoids, flavan-3:4-diols, leucocyanidins and hexahydroxyflavans are the prominent polyphenols found in BN [2,5]. Huang *et al.* have reported that betel nut extract (BNE) contains catechin based procyanidins which range from dimers to decamers and polymers [14]. During mastication, either as BN or BQ, they get oxidized and confer the characteristic red color to saliva, teeth and lips of BN/BQ masticator.

**(c) Tannins.** Specific types of polyphenols that are capable of precipitating proteins are tannins. The predominant tannin of BN is gallotannic acid, which is present in the outer part of the nut. In addition, minor amounts of gallic acid, D-catechol and phlobatannin are also present in the inner part of the nut [2,5].

**(d) Trace elements.** BN and *paan masala* have been reported to contain sodium, magnesium, chlorine calcium, vanadium, manganese, copper and bromine. The copper content in samples of raw and processed BN was analysed and reported to be much higher than that found most frequently in other nuts consumed by humans. The mean concentration of copper in samples of processed, commercially available BN was  $18 \pm 8.7 \mu\text{g/g}$ . In an Indian Food Report, the copper content of processed BN was found to be 2.5 times that of the raw BN [9].

**(e) Reactive oxygen species.** Cellular metabolism of BN or BQ components may also generate reactive oxygen species (ROS), such as superoxide anion radicals ( $\text{O}_2^-$ ) and hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) at pH greater than 9.5 [15]. While saliva was found to inhibit both  $\text{O}_2^-$  and  $\text{H}_2\text{O}_2$  formation from BQ ingredients, ROS



**Figure 3. Simplified flow chart of main events of BN induced carcinogenesis.** The simplified flow chart is intended to highlight the complexity of BN and its constituents, and how they affect different metabolic components and systems of a cell to eventually lead to carcinogenic transformation. For more details see reviews in references 2–5. doi:10.1371/journal.pone.0042759.g003

are formed in the alkaline chewing mixture within the saliva of a chewer due to the addition of slaked lime [16,17].

#### 4. GENERAL EFFECTS AND ADDICTIVE POTENTIAL OF BETEL NUT

In old Indian scripts such as *Vagbhata* (4<sup>th</sup> century), and *Bhavamista* (13<sup>th</sup> century), BN has also been described as a ‘therapeutic agent’ [5]. BN users report increased well-being and stamina, a soothing effect on the digestion, protection of the mouth and gums, and some euphoria. Its use was recommended in wide ranging human diseases and other disorders, which included vitiligo or leucoderma, leprosy, anemia, digestive disorders and infections, urinary and dental infections as well as obesity. BN is also reported to have aphrodisiac property and has been recommended as a general stimulant. In China, it has been used as a vermifuge since the 6<sup>th</sup> century [5].

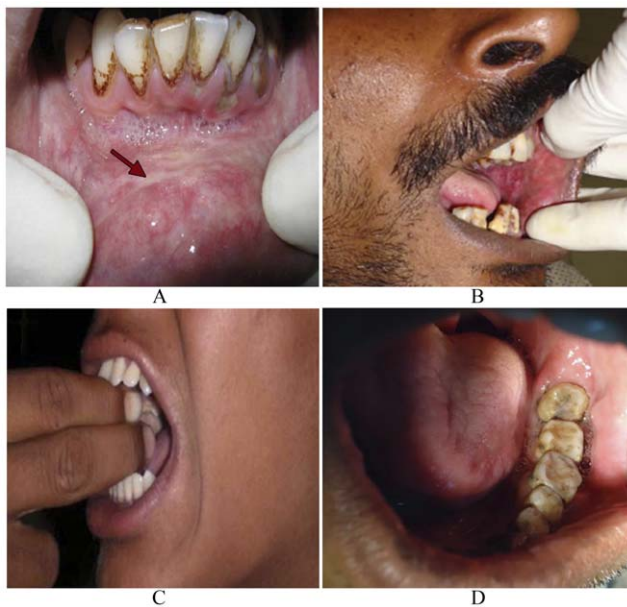
BQ chewing has been claimed to produce a sense of well-being, euphoria, warm sensation of the body, sweating, salivation, palpitations, heightened alertness, improved concentration and relaxation, diminished hunger, improved digestion and an increased capacity to work [2,18]. BN is also reported to have varied and widespread stimulating effects [2,19,20]. Small scale studies suggest that BN use may result in a dependence syndrome, though large scale studies testing this hypothesis are lacking [21,22]. Using a modified version of the Fagerstrom Test for nicotine dependence, it was found that 7% of the patients exhibited one of the following characteristics: daily chewing, chewing within one hour of awakening, difficulty in avoiding chewing, and increasing the quantity chewed. Patients with these symptoms are categorized as ‘severe’ or ‘heavy’ users of BN. Winstock *et al.* reported that 10 out of 11 current and former heavy BN users reported cessation withdrawal effects with the mean severity of Dependence Score of 7.3 consistent with the existence of a dependence syndrome among those who use BN products

[22]. However 55% among them used tobacco and BN in combination. Benegal *et al.* reported that about two out of five persons using BN preparations without tobacco additives developed a recognizable pattern of dependent use, satisfying both Diagnostic and Statistical Manual of Mental Disorder, 4<sup>th</sup> edition (DSM-IV; 38.8% of BN users) as well as International Classification of Diseases, 10<sup>th</sup> revision (ICD-10; 40.8% of BN users) criteria for current dependence [23]. Given the addiction potential of nicotine, the prevalence of dependence among those using BN preparations with tobacco additives was much higher than among persons using BN alone. Their findings provide support for the concept of an identifiable BN dependence syndrome, which can be diagnosed using criteria very similar to the ones currently used for other substances of abuse.

#### 5. BETEL NUT CONSUMPTION AND ORAL MALIGNANCY

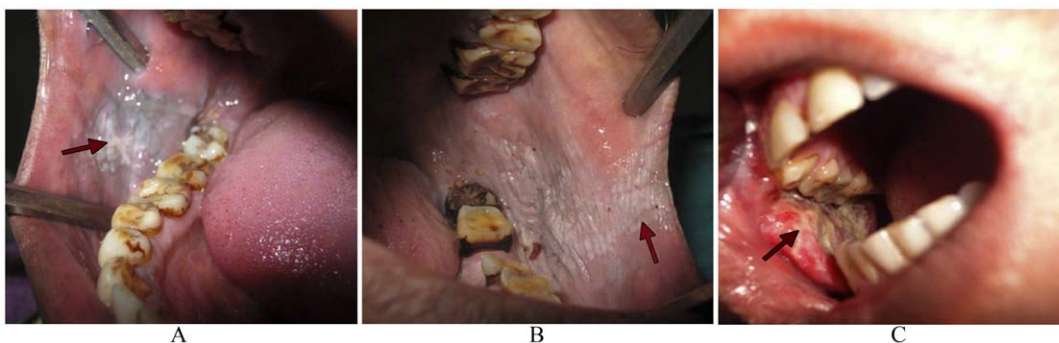
Prolonged as well as excessive usage of BN has been reported to exert significantly adverse effects on human health. There is enough evidence to suggest that BN products, even without tobacco, are associated with increased risk for the development of oral malignancy, such as oral squamous cell carcinoma (OSCC). A vast majority of BN users show precancerous clinical conditions, such as oral leukoplakia (OL) (Figure 4A) as well as its variant, oral erythroplakia (Figure 4B) or oral submucous fibrosis (OSF) (Figure 4C) among others. The risk is reported to be higher for *paan masala* chewers [24].

*In vitro* studies have demonstrated that BN extracts containing arecoline inhibit growth and protein synthesis in cultured human periodontal fibroblasts. These findings suggest that BN may be cytotoxic to periodontal fibroblasts and may exacerbate preexisting periodontal disease as well as impair periodontal reattachment [24]. The use of BQ was also found to be associated with the appearance of lichenoid lesions on the buccal mucosa and, occasionally, on the tongue (Figure 5A). These lesions are found at the site of quid placement in BN chewers. Fine wavy keratotic lines



**Figure 4. Clinical conditions associated with BN mastication.** Mastication of BN/BQ, even without tobacco, manifests itself in some preneoplastic alterations in the oral cavity of the masticator. This includes appearance of whitish patches or plaque (arrow) in the buccal mucosa, known as oral leukoplakia (OL) (A), or its variant with reddish patches/plaques, known as oral erythroplakia (OE) (B). In a third clinical manifestation, stiffening of oral mucosa leads to a clinical condition known as oral submucous fibrosis (OSF) characterized by inflammation and reduced fibro-elasticity which limits the opening of the mouth (C). Prolonged usage lead to a typical clinical manifestation known as the betel chewer's mucosa (BCM). This clinical condition is characterized by brownish-red discoloration of the oral mucosa, especially found in elderly BN chewing women (D).  
doi:10.1371/journal.pone.0042759.g004

are seen to radiate from a central red/atrophic area and the keratotic striae are parallel to each other. The histology is suggestive of a lichenoid reaction and the lesion is noted to resolve following cessation of BN use. Thus, such lichenoid lesions are considered to be type-IV contact hypersensitivity-type lesions, which clinically resemble oral lichen planus (OLP) (Figure 5B) [24].



**Figure 5. Potentially malignant and malignant conditions associated with BN mastication.** Prolonged mastication of BN/BQ eventually manifest itself in development of cancerous condition in the oral cavity of the masticator. Potentially malignant lesions in the oral cavity include lichenoid lesion(s) in the cheek (arrow) close of the site of mastication (A) or even tongue (not shown). At a late stage, lichenoid lesions lead to formation of Oral lichen planus (OLP), which is a type-IV contact hypersensitive type of potentially malignant lesion seen in the oral cavity of BN chewers (arrow) (B). A patient with history of prolonged use of BN alone (without tobacco) eventually shows development of a cancerous condition clinically known as Oral squamous cell carcinoma OSCC (arrow) in his right cheek (C), which was the primary site of BN mastication.  
doi:10.1371/journal.pone.0042759.g005

Another condition associated with prolonged BQ use, especially among elderly women, is betel chewer's mucosa (BCM), which is characterized by a brownish-red discoloration of the oral mucosa (Figure 4D). BCM is often accompanied by encrustation of the affected mucosa with quid particles, which are not easily removed and exhibit a tendency for desquamation and peeling [24]. Several epidemiological studies have shown that the prevalence of BCM varied widely between 0.2% in a Cambodian population in 1995 to 60.8% in the same population in 1996, while a prevalence of 21.9% was reported in a Malaysian population in 1995. Histologically, BCM shows epithelial hyperplasia, which is encrusted with an amorphous deposit. This reacts positively to von Kossa staining suggesting that these granules, which are both intra- and intercellular, may contain calcium from the calcium hydroxide of slaked lime. The presence of the human papilloma virus (HPV) subtypes 11, 16 and 18 have also been demonstrated in BCM but the significance of this is not fully understood. At present BCM is not considered to be potentially malignant, although the condition often coexists with other mucosal lesions such as OL (Figure 4A), OE (Figure 4B) and OSF (Figure 4C), which are well known for their potential for subsequent malignant changes [24].

**(a) Betel nut and oral leukoplakia.** OL can be defined as a predominantly white patch or plaque on the oral mucosa. Based on clinical appearance, leukoplakia can be divided into several subtypes: homogeneous (white), speckled (red/white), nodular or verrucous leukoplakia [24]. As an early sign of damage to the oral mucosa, chewers of BN or BQ with or without tobacco often develop clinically visible whitish (leukoplakia) (Figure 4A) or reddish (erythroplakia) (Figure 4B) lesions, which may or may not be accompanied by stiffening of the oral mucosa and OSF (Figure 4C). These manifestations are well-established precancerous lesions and are taken as early and important indicators of OC risk to an individual. Some 2–12% of these lesions have been reported to turn malignant over several years [3]. Although less common than leukoplakia (Figure 4A), erythroplakia (Figure 4B) poses a greater threat of cancer and lesions usually demonstrate significant epithelial dysplasia, carcinoma *in situ* or invasive squamous cell carcinoma. The presence and degree of epithelial dysplasia is generally accepted as the best indicator of malignant potential of leukoplakia, although some clinicians believe that ploidy analysis may be more reliable. There also appears to be an increased risk of transformation associated with a non-homoge-

neous leukoplakia, especially one that is clinically erythroplakic, verrucous or nodular [25]. One study in Taiwan indicated that the risks of developing OC after 20 years of follow-up were 42.2% for leukoplakia and 95% for erythroplakia [26]. Biopsies of leukoplakia reveal that in addition to the presence of an amorphous brown staining von Kossa positive layer on the surface, parakeratosis and atrophy of the covering oral epithelium were also observed in BN chewers. In another study, 14% of leukoplakia biopsies obtained from BN chewers demonstrated cellular atypia amounting to epithelial dysplasia [24].

It has been reported that the cessation of BN chewing resulted in resolution of 62% of leukoplakia, suggesting that BN on its own is a significant etiological factor in the development of leukoplakia. Further evidence of its relationship with BN chewing has come from the increased prevalence of this condition in subjects who suffer from OSF, which is associated strongly with the habit of BN chewing [24].

**(b) Betel nut and oral submucous fibrosis.** OSF is a chronic disorder characterized by fibrosis of the mucosa lining the upper digestive tract involving the oral cavity, oro- and hypopharynx and the upper third of the oesophagus. It was first described by Schwartz in 1952 as a fibrosing condition in five Indian women in Kenya and he called it as atrophica idiopathica atropica [27]. However, this condition is well established in medical literature since the time of *Sushruta*, a renowned Indian physician, who lived in 2500–3000 BC and described a condition resembling OSF which he referred to as '*Vidari*' [27]. There are also descriptions of similar conditions occurring in BN chewers in early texts dating back to 1908 [24]. The fibrosis is characterized by juxta-epithelial inflammatory reaction followed by chronic change in the fibro-elasticity of the lamina propria and is associated with epithelial atrophy. This leads to burning sensation in the oral cavity, blanching, and stiffening of oral mucosa and oropharynx, resulting in restricted mouth opening (Figure 4C). This condition, in turn, causes limited food consumption, difficulty in maintaining oral health, and impairs the ability to speak. The signs and symptoms depend on the evolution of lesions and number of affected sites. The malignant transformation rate of OSF has been reported to be around 7.6% over a 17-year period [24]. OSF has also been reported in several epidemiological studies mainly in the Southern states of India, among Indians living in South Africa, and among Chinese and Taiwanese [24,27]. Other occurrences are from Pakistan, Sri Lanka, Bangladesh, Malaysia, Singapore, Thailand and Saudi Arabia with reports of sporadic incidence among Europeans [27]. OSF is also described among Asians living in Europe and the United States but who continue to chew BN [24].

It is now well accepted that all BN products, even those without tobacco, are associated with OSF, which has been established as a precancerous condition. When a paste made out of an instant BN preparation was painted into the oral cavity of albino rats, biopsies taken from the oral mucosa revealed mild to moderate loss of nuclear polarity and increase in keratoses, parakeratoses, inflammatory cell infiltration and vascularity [28]. Submucosal collagen also increased steeply and steadily throughout the study period. At the end of six months, 88.23% of biopsies showed thickened and condensed submucosal collagen, indicating submucous fibrosis [28]. It has been reported that BQ chewing with or without tobacco is a major risk factor for high prevalence of oral potentially malignant disorders (OPMD) in rural Sri Lanka [29]. The relative risk of malignant transformation in the oral mucosa of OSF patients compared to tobacco users without any precancerous lesion or condition has been estimated at around 400. Thus, BN users are potentially more liable to develop OSF and cancer

over a relatively shorter duration and die earlier compared to smokers. Commercial products such as *paan masala*, *gukha*, and *mawa* have higher concentrations of BN and appear to cause OSF more rapidly than self prepared conventional BQ, which contains smaller amounts of BN [30,31]. Thus, the popularity of BN mixtures like *paan masala*, *gukha* and *mawa* has spawned an epidemic of OSF, particularly among young individuals in India [32,33].

A clear dose dependent relationship has been reported for both frequency and duration of chewing BN without tobacco in the development of OSF [34]. Only smoking and/or alcohol consumption were not found to influence the development of OSF [35,36]. But their addition to BN chewing habit can be a risk for OSF [36]. Although there is good evidence to support the role of BN as a major risk factor in the development of OSF, the mechanisms by which this occurs is not fully understood. Most studies on pathogenesis have concentrated on changes in extracellular matrix based on the premise that increased collagen synthesis or reduced collagen degradation is the possible mechanism for the development of this condition. Studies have revealed that OSF fibroblasts have marked deficiency in collagen phagocytosis, which may lead to fibrosis. In one study, arecoline was found to elevate the mRNA and protein expression of Cystatin C, a non-glycosylated basic protein consistently upregulated in a variety of fibrotic diseases, in a dose dependent manner in persons with OSF [37]. Another study showed an upregulation of Cystatin C in resident cells of buccal mucosa in OSF patients on exposure to BN. Cystatin C, in turn, inhibited the lysosomal cysteine proteases like Cathepsin B and H, resulting in decreased degradation of collagen [27]. Moreover, arecoline was also found to stimulate Cyr61 synthesis in human gingival epithelial S-G cells. Constitutive overexpression of Cyr61 protein in oral epithelial cells during BN chewing may play a role in the pathogenesis of oral cancer, since Cyr61 is associated with growth and progression of many types of tumors and is an independent poor prognostic indicator for oral cancer patients [38]. Lin *et al.* assessed the mRNA expression of histone methyltransferases, acetyltransferases, and demethylases in K-562 cells following exposure to arecoline. They observed that arecoline produced changes in the expression of several genes catalyzing histone methylation (Mll, Setdb1, and Suv39h2), acetylation (Atf2), and demethylation (JMJD6). Since H3K9 methylation is involved in maintaining the stability of heterochromatin structures and inactivating euchromatic gene expressions, this study indicates that arecoline-induced epigenetic changes play a role in the mechanisms underlying chemical-mediated cytotoxicity and genotoxicity [39].

In three separate but related studies, interleukin-6 [IL-6], keratinocyte growth factor-1 (KGF-1) and insulin-like growth factor-1 (IGF-1) expressions, which have all been implicated in tissue fibrogenesis, were significantly upregulated in persons with OSF due to BQ chewing and arecoline may be responsible for their enhanced expression [40–42]. Moreover, it was noticed that addition of slaked lime to BN in BQ facilitated hydrolysis of arecoline to arecaidine, which caused amplified fibroblastic proliferation and increased collagen formation. *In vitro* examination of effects of arecoline on both normal and OSF fibroblasts in culture revealed an augmented collagen synthesis by OSF fibroblasts as compared to normal fibroblasts. The reason for this elevation was thought to reflect the clonal selection of a particular cell population in the altered tissues or normal cells with somatic mutation that persists through several generations. This could be due to upregulation of pro-inflammatory and pro-fibrotic cytokines like interleukins (IL-1, IL-6, IL-8), tumor necrosis factors (TNF- $\alpha$ , TNF- $\beta$ ), platelet-derived growth factors (PDGF), fibro-

blast growth factors (FGF) and keratinocyte growth factor-1 (KGF-1), among others, and downregulation of interferon gamma (IFN- $\gamma$ ) level, resulting in fibrosis. Additionally, activation of pro-collagen genes like COL1A1, COL3A1, COL6A1 and COL7A1 has also been reported in OSF [27]. However, some studies have also shown that arecoline inhibits collagen synthesis and fibroblast proliferation *in vitro* suggesting that arecoline may have cytotoxic properties. The disparity of results from *in vitro* studies suggests that the BN may contain other agents in addition to arecoline, which are important in the pathogenesis of OSF through increased collagen synthesis [24]. It has also been reported that BQ chewing contributed to the pathogenesis of cancer and OSF by impairing T cell activation and by induction of prostaglandin E2 (PGE2), TNF- $\alpha$  and IL-6 production, which affect oral mucosal inflammation and growth of oral fibroblasts/oral epithelial cells [43].

Another mechanism envisions involvement of BN in the pathogenesis of OSF due to decreased collagen degradation through decreased obliteration, inhibition of phagocytosis or resistance to degradation [27]. Reduced collagenase activity and subsequently decreased degradation of collagen have been demonstrated in OSF. Polyphenols of BN, such as flavanoid, catechin and tannins cause collagen fibers to crosslink, making them less susceptible to collagenase degradation [44]. This results in increased fibrosis due to decreased collagen breakdown [45]. OSF remains active even after cessation of the chewing habit suggesting that components of the BN initiate OSF and then affect gene expression in the fibroblasts, which then produces greater amounts of collagen [46,47]. Chewing BQ may also activate nuclear factor-kappaB (NF- $\kappa$ B) expression, thereby stimulating collagen synthesis by human buccal mucosal fibroblasts and leading to further fibrosis in persons with OSF [48]. In fact, OECM-1 and SAS oral keratinocytes treated with BNE activated the NF- $\kappa$ B pathway in a biphasic manner, particularly for SAS cells, resulting in periods of significantly elevated activity interrupted by a plateau or period of decreased activity. BNE treatment did not activate epidermal growth factor receptor signaling system, but blockage of NF- $\kappa$ B activation rendered the suppression of BNE-modulated COX-2 upregulation in OECM-1. Both OECM-1 and SAS oral keratinocytes also exhibited a rapid increase in c-Jun N-terminal kinases (JNK1) activity, while extracellular signal-regulated kinase (ERK) was profoundly activated in OECM-1 cells. This study thus identified that BNE induced alterations in interactive signaling systems in oral keratinocytes could be a basis of the pathogenicity of BN [49]. Additionally, reduced level of main gelatinolytic proteinases secreted by buccal mucosal fibroblasts (BMF), namely matrix metalloproteinases MMP2, MMP9 and elevated levels of tissue inhibitor of metalloproteinase-1 (TIMP-1) have been reported in OSF as a possible means of loss of equilibrium of extracellular matrix (ECM) in OSF. This may result in increased and continuous deposition of ECM. In fact, arecoline and safrole significantly elevated TIMP-1 protein and mRNA expression in BMF, and this is a possible pathogenesis for OSF [50]. In contrast, MMP-2 and MMP-9 have been reported to be present in human OSCC and the activated MMP-2 could be the main enzyme for gelatinolysis in OSCC, facilitating invasion and metastasis [51]. One study assessed the change in salivary MMP-9 protein levels 2 hours after 5-minute BQ chewing stimulation (BQCS) in non-BQ users and the expression profile of this proteinase in saliva and tumor specimens of OSCC patients with a history of BQ use. MMP-9 was found to be upregulated in response to BQCS and MMP-9 expression was also associated with neck lymph node metastasis, thus implying a significant role of MMP-9 in the

progression of OSCC among patients with a history of BQ use in Taiwan [52].

Raised copper concentrations have been shown in products containing BN in comparison to other nut based snacks. It has also been observed that chewing BN for 5–30 min significantly raised the soluble Cu level in saliva. Study of buccal mucosal biopsies from patients with OSF indicated raised Cu level [53]. Addition of CuCl<sub>2</sub> increased the collagen synthesis by the oral fibroblasts. However, the addition of CuCl<sub>2</sub> neither increased the synthesis of non-collagenous proteins by the fibroblasts nor influenced their proliferation rate. These *in vitro* results support the hypothesis that Cu in BN acts as a mediator of OSF [54]. This has led to the hypothesis that the increased tissue Cu may increase the activity of the enzyme lysyl oxidase, which is a Cu-dependent enzyme that has been implicated in the pathogenesis of several fibrotic disorders, including OSF [24]. Cu salts significantly increased the production of collagen by oral fibroblasts *in vitro* supposedly by upregulation of activity of a Cu-dependent enzyme, lysyl oxidase, which catalyses the cross linking of collagens and elastin [6]. The collagen cross linked with lysyl oxidase is rendered insoluble and is shown to be ten times more resistant to digestion by mammalian collagenase [27]. Further, a significant gradual increase in serum Cu levels from pre-cancer to advanced cancer in patients has been documented [55], which may have a role in oral fibrosis to cancer pathogenesis (Figure 3).

## 6. LINK BETWEEN BETEL NUT AND CARCINOGENESIS

There exists an accumulated wealth of historical evidences suggesting that the BN may be involved in the development of OSCC (Figure 5C) [24]. Recent research has also generated sufficient evidence to implicate BN as well as BQ with or without tobacco, as a suspected carcinogen to humans [4,5,9]. In addition to OC, significant increase in the incidence of cancers of the esophagus, liver, pancreas, larynx and lung were seen among BN chewers [56]. A study of esophageal squamous cell carcinoma (ESCC) in Taiwan revealed that subjects who chewed between 1 and 495 BN or more than 495 BN per year had 3.6-fold and 9.2-fold higher risk, respectively, of developing esophageal cancer compared to those who did not chew BN [57].

A causal association between tobacco and BQ chewing habits and oral mucosal diseases such as OL, OSF and OC has been established and heavy users have a significantly increased mortality rate [6]. A study in Taiwan has suggested that elimination of BN may prevent 62% of OL and 26% of malignant transformation to OC in the underlying population [58]. Data on oral cavity cancer in Taiwan from the period between 1986 and 1997 also indicated that those who chewed BN had a higher risk for OC [59]. Studies in OC patients demonstrated that male cases were far more common than females, comprising 90–93% men and 7–10% women. It is proposed that this gender difference may be explained by the prevalence of lower proportion of BQ chewing habits among females. Concerns about the disfiguring effects of BQ chewing, including red staining of lips and teeth, and foul smelling breath, are frequently reported by females, which may account for gender differences in head & neck cancer (HNC) prevalence among females and males [60]. Chewing BQ independently was found to contribute to the risk of HNC and the estimated prevalence of BQ chewing in Taiwanese patients with HNC was found to be approximately 85% [60]. Chewing BQ without tobacco has also been implicated in the causation of OC among South Asian communities [61].

A study of OSCC and concomitant oral habits undertaken in South African Indians revealed that 68% women with cheek cancer and 84% with tongue cancer only chewed BN without



tobacco, snuff or smoking. The data show that the BN habit with or without tobacco is important in the development of OSCC and it has been suggested that elimination of this habit can reduce the risk in these women substantially (89–91%) if all other factors remained the same [46]. Time-trend analysis of cancers at all sites for the period 1990–1996 showed a decrease in cancers of the oral cavity in Indian population based registries, but an increase in the incidence of OC was reported among those aged <50 years between 1983–1987 and 1995, consistent with the hypothesis of an increase in OC among the young due to increased consumption of the alternative chewing products such as, *gutkha* and *paan masala* [6]. BN without tobacco was, thus, recognized as a group I carcinogen to human by the International Agency for Research on Cancer (IARC) and World Health Organization (WHO) in 2004 [9]. In addition to human data, there are also a large number of experimental studies, which have reported the carcinogenic potential of BN and its derivatives both *in vivo* and *in vitro* [24].

**(a) *In vivo* studies.** Several animal studies have confirmed that BN products and derivatives, such as arecoline and BN derived nitrosamines, also referred to as betel specific nitrosamines (BSNA), have the ability to induce neoplastic changes in experimental animals (Figure 3). Alkaloids of BN are suspected to be its main carcinogenic constituent [2–6,9,24,62]. Early studies found that the application of arecaidine to the oral mucosa of experimental animals failed to have any carcinogenic effects unless it was supplemented with a known promoter, such as croton oil [24]. However, arecoline administered by gavage produced lung adenocarcinoma, stomach SCC and liver haemangioma in male mice [9]. Cheek pouch application of arecoline following application of slaked lime produced an esophageal papilloma in female hamsters, while local application of arecaidine to the cheek pouch did not produce tumor in male hamsters [9]. To explain the variable observation, it is proposed that the alkaloids first required metabolic activation via nitrosation to develop its carcinogenicity [63]. *In vitro* data suggest that arecoline is metabolized by carboxylesterase in mouse liver and kidney. Male Swiss albino mice fed BN powder or arecoline showed enhanced levels of the hepatic cytochrome P450 and b5 and decreased levels of hepatic GSH [9]. In fact, human cytochrome P450 was found to be involved in the mutagenic activation of BSNA such as 3-(N-nitrosomethylamino)propionitrile (NMPN), 3-(N-nitrosomethylamino)propionaldehyde (NMPA) and N-nitrosoguvacoline (NG) using genetically engineered *Salmonella typhimurium* YG7108 expressing each form of human P450 together with NADPH-P450 reductase [64]. Exposure of Swiss albino mice to arecoline was found to lower poly-ADP-ribosylation (PAR) of most cellular and histone proteins and induce relaxation of chromatin, thereby allowing the N-nitrosamines of arecoline easy access to genomic DNA for interaction, while the absence of PAR mediated repair may favour the accumulation of DNA damage [65]. Arecoline induced micronuclei (MN), chromosomal aberrations (CA) and sister chromatid exchange (SCE) in bone marrow cells (BMC) of Swiss albino mice [66,67]. The arecoline induced DNA damage was found to be influenced by endogenous GSH levels with the frequency of CA and SCE increasing when arecoline was given to mice treated with buthioninestulfoximine (BSO), a GSH synthesis inhibitor (Figure 3).

The frequency of SCE was found to be elevated in mouse BMC when mice were exposed to the aqueous extract of betel nut (AEBN) and its tannin [67]. AEBN also induced micronucleated cells (MNC) in BMC of Swiss albino mice [5]. Hamsters fed with powdered diet containing BN or BQ showed significant decrease in the survival rate, body weight, and hyperkeratosis and acanthosis of cheek pouch indicating that BN and BQ components

may induce alterations in proliferation and differentiation of oral epithelial cells [68]. When the buccal mucosa of mice was treated regularly with a topical application of water based BNE, the oral epithelium showed progressive changes in epithelial thickness leading to atrophy, increased cellularity of fibroblasts, fibrosis of connective tissue, focal infiltration of inflammatory cells and muscle atrophy [69]. Frequency of all the three cytogenetic endpoints, viz. CA, SCE and MNC, were found to be elevated significantly in a dose dependent manner in cultures exposed to aqueous extracts of *paan masala* without metabolic activation [70]. The carcinogenic and tumor promoting potentials of an ethanolic *paan masala* extract (EPME) were determined using the hairless skin of S/RVCri-ba or Bare mice and the forestomach and esophagus of ICRC mice as the target tissues. EPME promoted skin papilloma formation and enhanced the rate of conversion of papilloma to carcinoma. Induction of mild epidermal hyperplasia, dermal edema, increase in epidermal mitotic activity and the rate of epidermal and dermal DNA syntheses by EPME correlated well with its skin tumor promoting potential. In ICRC mice, EPME was inactive as a complete carcinogen, but effectively promoted the development of forestomach and esophageal papilloma and carcinoma in a concentration dependent manner indicating that habitual *paan masala* use may exert carcinogenic and co-carcinogenic influences [71]. Exposure of male and female mice to *paan masala* revealed a significant dose dependent increase in lung adenocarcinoma but not in liver and stomach [72].

**(b) *In vitro* studies.** BNE was found to decrease cell survival, vital dye accumulation and membrane integrity of cultured human buccal epithelial cells in a dose dependent manner. BNE also caused formation of both DNA single strand breaks and DNA protein cross links [63,73,74]. Different BNE, such as aqueous extract of betel nut (AEBN), acetic acid extract of betel nut (AAEBN), HCl extract of betel nut (HEBN) and ethanol extract of betel nut (EEBN) as well as arecoline showed different extents of cytostatic and cytotoxic effects, and induced variable levels of dose dependent unscheduled DNA synthesis (UDS) in Hep2 cells *in vitro*. In manifestation of these effects arecoline, HEBN and EEBN were most potent [73,75]. Cultured normal human oral keratinocytes (NHOK) exposed to ripe BNE also showed significant decrease in population doubling, increase in senescence, cell cycle arrest at G1/S phase and decrease in cell proliferation [76]. It has been reported that BQ may accelerate tumor migration by stimulating MMP-8 expression through MEK pathway in at least some carcinomas of the upper aerodigestive tract. Furthermore, arecoline may be one of the positive MMP-8 regulators among BQ ingredients [77]. Investigation of prostaglandin endoperoxide synthase (PHS) action on the growth of OC in response to BNE exposure of two human oral carcinoma cell lines OEC-M1, and KB, and one normal fibroblast cell line, NF, revealed that BNE significantly inhibited the cell growth of OEC-M1, KB and NF. PHS activity in OEC-M1 and NF was significantly increased by low BNE concentrations but significantly reduced at higher concentrations. The PHS activity in KB, on the other hand, was significantly inhibited by BNE and this effect was intensified as concentration increased [78]. Treatment of human oral mucosal fibroblasts (OMF) with BNE or arecoline induced about 3-fold increase in mRNA levels of the proto-oncogene *c-jun* independent of GSH depletion [79].

The BNE and inflorescence of *Piper betle* (IPB) also induced DNA strand breaks. In addition, BNE, IPB, the BN polyphenol, catechin as well as arecoline decreased cell survival and proliferation. In contrast, another component of BQ, the aqueous extract of lime, was found to increase cell proliferation [80]. AEBN was found to reduce endogenous glutathione (GSH) level, induce

CA and delay cell kinetics in mouse BMC with the induction of SCE probably involving TP53 dependent changes in cell proliferation [81]. Ethyl acetate and *n*-butanol extracts of BN as well as betel leaf are reported to induce CA in human lymphocytes and Chinese hamster ovary (CHO) cells [4]. All components of BQ have been shown to individually enhance chromatid breaks and exchanges in the range of 12–37% in human cells *in vitro*. AEBN also induced DNA strand breaks and enhanced cell proliferation in mouse kidney T1 cells *in vitro* [73]. BNE exposure to CHO-K1 cells caused increased MN frequency, G2/M arrest, cytokinesis failure and an accumulation of hyperploid/aneuploid cells. These events are associated with an increase in intracellular H<sub>2</sub>O<sub>2</sub> level and actin filament disorganization [82]. BNE also elicited actin reorganization resulting in fibroblastoid morphological change, genesis of lamellipodia, loss of subcortical actin and stress fiber formation in cultivated NHOK cells [83].

Arecoline alone has been reported to inhibit cell attachment, cell spreading and cell migration in a dose dependent manner in cultured human gingival fibroblasts (HGF) [84]. In fact, GSH depletion and reduction of glutathione *S*-transferase (GST) activity have been demonstrated in cultured human oral keratinocytes and in fibroblasts treated with arecoline [9]. Arecoline was also reported to be cytotoxic to human buccal fibroblasts in a dose dependent manner wherein the cellular GST activity was downregulated in a dose dependent manner without increase in lipid peroxidation. Addition of extracellular nicotine acted synergistically on the arecoline induced cytotoxicity, indicating that arecoline may render human OMF more vulnerable to other reactive agents in cigarettes via GST reduction. These observations could explain why patients who practice the combined habit of BQ chewing and cigarette smoking are at greater risk of contracting OC [85]. Arecoline inhibited growth of human KB epithelial cells in dose- and time-dependent manners by causing cell cycle arrest in late S and G2/M phases due to induction of cyclin B1, Wee 1, and phosphorylated cdc2 proteins and inhibition of p21 protein expression in KB cancer cells. In primary human gingival keratinocyte (HGK) cell line, arecoline effect appeared to be mediated differently. In this case, arecoline induced p21 but inhibited cdc2 and cyclin B1 proteins. This clearly suggests that differential regulation of S and/or G2/M cell cycle related proteins in the HGK and KB cells play crucial roles in different stages of BQ mediated carcinogenesis [86]. Arecoline could also induce  $\gamma$ -H2AX phosphorylation, a sensitive DNA damage marker, in KB, HEP-2, and 293 cells, suggesting that DNA damage was elicited by arecoline. Moreover, the expression of p53 regulated p21 (WAF1) and p53 activated DNA repair were repressed by arecoline [87]. Arecoline was cytotoxic to HGF cells due to depletion of intracellular thiols and inhibition of mitochondrial activity and induced cell cycle arrest in HGF cells at G2/M phase in a dose dependent manner [88]. Global gene expression profiling in HGF exposed to arecoline revealed that four genes related to maintenance of genome stability and DNA repair were repressed by arecoline [89]. They are *FANCG*, also known as *XRCC9* (tumor suppressor capable of correcting CA), *CHAF1* and *CHAF2* (encoding chromatin assembly factor I or CAF1) and *BRCA1* (breast cancer susceptibility gene implicated in DNA damage response and DNA repair). Among them, at least the *BRCA1* response was dose dependent. *COX-2* and *PTGS2*, which are involved in cancer initiation and progression, were over expressed in HGF cells. *HSP4A1* and *DNAJA1*, which belong to the *HSP70* family of stress-induced proteins and *GDF15/MIC-1*, were also upregulated by arecoline in dose dependent manner [89]. Chen *et al.* established two oral cancer sublines chronically treated with BNE and used methods such as microarray and

immunohistochemistry to screen and validate the genes exhibiting altered expressions in BNE sublines or in cancer tissues. They found that a total of 35 genes were differentially expressed in both sublines. Several functional pathways were significantly altered. Six genes were confirmed over 2-fold of changes, including *Ches1*. Functional analyses showed that overexpression of *Ches1* suppressed cell growth and arrested cells in the G2/M phase. They thus concluded that loss of *Ches1* may be attributed to BNE-induced oral carcinogenesis [90].

Treatment of normal human oral fibroblasts with BNE was also reported to alter miRNA expression profile. BNE-induced overexpression of miR-23a was found to be correlated with an increase of  $\gamma$ -H2AX, a DNA damage marker. *FANCG* was confirmed to be a target of miR-23a by ectopic overexpression or knockdown of miR-23a. The correlation between miR-23a overexpression and BN-chewing habit was also reported in oral cancer patients. Thus, BNE-induced miR-23a was correlated with a reduced *FANCG* expression and DNA double strand break (DSB) repair, which might contribute to BNE-associated human malignancies [91]. Oral fibroblasts with chronic subtoxic BNE treatment were found to exhibit growth arrest and MMP-2 activation. The supernatant of these arrested oral fibroblasts activated the AKT signaling pathway in oral carcinoma cells. Moreover, subcutaneous co-injection of arrested oral fibroblasts into nude mice significantly enhanced the tumorigenicity of xenographic oral carcinoma cells. The investigators therefore concluded that BNE may impair oral fibroblasts and then modulate the progression of oral epithelial oncogenesis *via* their secreted molecules [92]. Various studies have clearly established the mutagenicity of BN and its components. The major metabolite of arecoline, arecoline N-oxide, is reported to be moderately mutagenic to *Salmonella typhimurium* tester strains TA 100 and TA 98. But this mutagenicity was potently inhibited by glutathione, N-acetylcysteine, and cysteine [93]. Aqueous extracts of BQ without tobacco induced mutations in *Salmonella typhimurium* but not in Chinese hamster V79 cells. AEBN, on the other hand, induced mutations in *Salmonella typhimurium* and in Chinese hamster V79 cells besides inducing gene conversion in *Saccharomyces cerevisiae* as well as CA in CHO cells. BN tannin fraction induced gene conversion in *Saccharomyces cerevisiae* [4]. Ames test using *Salmonella typhimurium* strain TA 1535 revealed that arecoline, AEBN and HEBN were weak mutagens while AAEBN and EEEN were strong mutagens suggesting that the mutagenic potential of arecoline could be significantly enhanced by other constituents of BN [5,94,95]. Exposure to BNE was also found to induce mutation at the *hypoxanthine phosphoribosyltransferase* (HPRT) locus in human keratinocytes, which also increased frequency of appearance of MN, intracellular levels of reactive oxygen species (ROS) and 8-hydroxyguanosine in the cells suggesting that stress caused by long term BNE exposure enhanced oxidative stress and genetic damage in human keratinocytes [96].

**(c) Human studies.** Among BN chewers, the possible genomic damage caused by BN without tobacco was confirmed in cytogenetic studies. BNE has been shown to be cytotoxic and genotoxic to human buccal epithelial cells [74]. This may be correlated to its ability to increase DNA strand breaks, MNC formation, gene mutation and CA [16,67]. A study aimed to evaluate the genotoxic effect of BN and tobacco on human peripheral blood lymphocytes revealed anomalies. Binucleated cells with MN, total MN, nucleoplasmic bridge and nuclear buds were higher in chewers whereas elevation in binucleated MN and total MN were significant among subjects with oral submucous fibrosis than nonchewers. Significant positive correlation was also observed between induction of cytokinesis-blocked micronucleus

(CBMN) and consumption of BQ per day [97]. However, there is still a void in the complete understanding of the molecular mechanism by which BN affects DNA repair and genome stability genes. These two are hallmarks of genome fidelity. Arecoline inhibited both expression and transactivation functions of p53. This inhibition is proposed to play an important role in arecoline mediated suppression of DNA repair. It was shown that the expression of p53 mRNA was frequently downregulated in BQ associated OC [87]. Arecoline also arrested cells at prometaphase with large amounts of misaligned chromosomes by stabilizing mitotic spindle assembly, which led to distorted organization of mitotic spindles, misalignment of chromosomes and upregulation of spindle assembly checkpoint (SAC) genes [98]. A chromosomal analysis of patients with OC primarily associated with BN consumption using comparative genomic hybridization revealed that the most common gains of chromosome arms were 8q, 9q and 11q, and the most frequent losses were of chromosome arms 3p and 4q [99].

A study revealed that OSF was largely associated with BN and the exfoliated oral mucosal cells of such patients had significantly higher numbers of MNC. The patients also exhibited increased SCE in circulating blood lymphocytes indicating that the carcinogenic agents in BN produce damage not only in target tissue but also in other tissues [100]. Rooban reviewed the effects of different ways of taking arecoline on salivary flow rates (SRF) and pH of saliva [101]. With an increase in frequency and exposure time of chewing raw BN, both SFR and pH increased. In processed BN chewers, increase in duration and frequency of consumption increased the SFR and decreased the pH, respectively. For chewers taking BN with tobacco, increase in duration was significantly associated with decrease in salivary pH. Similarly, IBP, which contains safrole (4-allyl-1,2-methylenedioxybenzene), a unique ingredient of BQ in Taiwan, forms Safrole–DNA adducts. This has been suggested to play an important role in OC in the population of Taiwan. A high frequency of safrole-like DNA adducts has been reported in BQ associated OSCC and noncancerous matched tissue in contrast to the absence of such adducts in all of non-BQ associated OC. Safrole-DNA adducts are present in oral cancer tissue from patients who have chewed BQ containing high concentration of safrole as well as in peripheral white blood cells. Safrole is classified as a rodent hepatocarcinogen, and chewing BQ may contribute to human exposure to this compound. The saliva of a person chewing BQ contains on average 420  $\mu\text{mol/L}$  of safrole. Interestingly, safrole-DNA adducts were found in liver biopsy specimens of a Taiwanese man suffering from hepatocellular carcinoma, who had chewed BQ for over 32 years. This implies that safrole may be implicated not only in carcinogenesis of the oral cavity of BQ chewers through direct contact, but can also be transported via the oral-digestive tract to distant organs like the liver where it acts a likely cause of liver carcinogenesis [102]. Moreover, individuals with at least one cytochrome P450 - CYP2E1c2 allele had a significantly higher frequency of safrole-DNA adducts formation than those with the CYP2E1c1c1 genotype while chewing less than 20 BQ per day [103]. Hydroxychavicol, a phenolic component of betel leaf, has been found in human saliva at a 4.6 mM concentration after BQ chewing. Hydroxychavicol may induce DNA single strand breaks and 8-hydroxydeoxyguanosine, a marker of oxidative DNA damage, in cultured cells [104].

Hung *et al.* reported the upregulation of Asb6, a coupling protein to the APS adapter protein, which is involved in insulin signaling for glucose transportation, of normal keratinocytes and oral cancer cells under BNE treatment. They also demonstrated a positive correlation between Asb6 upregulation (cancerous tissues

versus adjacent normal tissues) and clinicopathological features such as poor survival status in OSCC patients [105].

In a study pertaining to the contribution of combined BN chewing and cigarette smoking to the risk of OSCC in Taiwan, Wu *et al.* revealed that the alkaline environment created in the oral cavity of BN chewers by lime may enhance nicotine-related oral carcinogenesis through a synergistic effect of nicotine and the alkalinity in inducing higher expression of phosphorylated AKT [106]. AKT/protein kinase B (PKB) is a serine/threonine kinase which is implicated in mediating a variety of biological responses including cell growth, proliferation and survival. AKT is activated by phosphorylation on two critical residues, namely threonine 308 (Thr308) and serine 473 (Ser473), and several studies have found AKT2 to be amplified or overexpressed at the mRNA level in a number of human malignancies [107].

A study involving patients of HNC suggested that BQ chewing may increase mitochondrial DNA (mtDNA) mutation in human oral tissues and that accumulation of mtDNA deletions and subsequent cytoplasmic segregation of these mutations during cell division could be important contributors to the early phase of OC [108]. ZASC1, a zinc finger transcription factor localized on 3q26, is frequently amplified in OSCC. Examination of OSCC patients revealed that increase of ZASC1 gene copy number in recurrent tumors was associated with the consumption of BQ in patients [109]. O(6)-methylguanine-DNA methyltransferase (MGMT) ameliorates mutagenic, carcinogenic and cytotoxic adducts from O(6)-methylguanine in DNA. The absence of MGMT expression associated with promoter hypermethylation has been reported to be related to BQ chewing and, thus, might be a significant event in OC [110]. A high frequency of hypermethylation of p14, p15 and p16 was also detected in the precancerous lesions of BQ chewers in Sri Lanka [111]. Further, it has been proposed that epigenetic silencing of RASSF1A and p16INK4a gene expressions by promoter hypermethylation may play critical roles in BN associated OC [112].

Alphavbeta6 ( $\alpha\text{v}\beta\text{6}$ ) integrin is capable of promoting both tissue fibrosis and carcinoma invasion, and has been reported to be markedly up-regulated in OSF [113]. Moutasim *et al.* investigated the functional role of  $\alpha\text{v}\beta\text{6}$  using oral keratinocyte-derived cells genetically modified to express high  $\alpha\text{v}\beta\text{6}$  (VB6), and also NTERT-immortalized oral keratinocytes, which express low  $\alpha\text{v}\beta\text{6}$  (OKF6/TERT-1). VB6 cells showed significant  $\alpha\text{v}\beta\text{6}$ -dependent activation of TGF- $\beta\text{1}$ , which induced transdifferentiation of oral fibroblasts into myofibroblasts and resulted in up-regulation of genes associated with tissue fibrosis. These experimental *in vitro* findings were confirmed using human clinical samples, which revealed that the stroma of OSF contained myofibroblasts and that TGF- $\beta\text{1}$ -dependent Smad signaling was detectable both in keratinocytes and in myofibroblasts. The investigators also found that arecoline, the major alkaloid of BN up-regulated keratinocyte  $\alpha\text{v}\beta\text{6}$  expression. This was modulated through the  $\text{M}_4$  muscarinic acetylcholine receptor and was suppressed by the  $\text{M}_4$  antagonist, tropicamide. Arecoline-dependent  $\alpha\text{v}\beta\text{6}$  up-regulation promoted keratinocyte migration and induced invasion, raising the possibility that this mechanism may support malignant transformation. This study thus suggests that the pathogenesis of OSF may be epithelial-driven and involve arecoline-dependent up-regulation of  $\alpha\text{v}\beta\text{6}$  integrin [113].

Heat shock protein 47 (HSP47) is a product of CBP2 gene located at chromosome 11q13.5, a region frequently amplified in human cancers. HSP47 expression was reported to be significantly higher in OSCC specimens than normal epithelium, while lower HSP47 expression was associated with lymph node metastasis. No significant difference in HSP47 expression was observed with

respect to age, sex, tumor category, tumor stage and differentiation. Furthermore, arecoline was found to elevate HSP47 expression in a dose- and time-dependent manner in oral epithelial cell line OC2. This study therefore concluded that HSP47 could be used clinically as a marker for lymph node metastasis of oral carcinogenesis [114].

## 7. BETEL NUT AND TUMOR SUPPRESSOR GENES

The *p53* gene is known to be mutated in a variety of human and experimental animal cancers. Similarly, change in cellular level of p53 protein is also known to occur. Accumulation of p53 protein or its stabilization is an important indicator of the presence of mutant p53 protein [115,116]. However, reports pertaining to *p53* mutation status of cancers associated with BN chewing have been widely contradictory. Exposure to BN and/or BQ with or without tobacco has been reported to result in high incidence of *p53* mutations in Taiwanese, Thai, Sri-Lankan and North Indian Population; however, similar exposures resulted in low frequency of *p53* mutations in the populations of India and Papua New Guinea, in other reports. Despite these conflicting observations, a common trend observed among the varying populations was p53 overexpression and nuclear accumulation of p53 protein (see Table 1) [117–127]. A study on northeastern Indian population also found significant influence of p53 codon 72 polymorphism, with interaction between p53 genotype and smoking resulting in a significant risk of OC, while interaction of p53 genotypes with BQ leading to a significant risk of lung cancer [128]. Another study on cancer patients in northeastern India also found that the subjects with family history of cancer were more likely to develop ESCC if they were BQ users and germ line mutations in the DNA repair gene, BRCA2, played a role in this familial aggregation of ESCC [129].

In previously reported transgenerational studies, 6 week old male and female Swiss albino mice were exposed chronically to AEBN in drinking water at a dose of 2 mg ml<sup>-1</sup> for 24 weeks. These mice are referred to as the chronically exposed P generation mice. The F1 generation was raised by inbreeding of P generation

mice exposed to AEBN for 6 weeks. Similarly, the F2 and F3 generations were raised from AEBN exposed F1 and F2 mice, respectively [130,131]. Thus, the transgenerationally exposed F1, F2 and F3 mice received low dose AEBN prior to and during conception and the entire period of development and maturation. In these studies, exposure to AEBN was found to severely impair the ultrastructure of the nucleus, endoplasmic reticulum and mitochondria with a significant reduction in mitochondrial index from the P through F3 generations. This indicates a progressive loss of apoptosis with progression of generation [132]. It was further observed that the exposure to AEBN resulted in an immediate upregulation of p53 protein up to 2–2.5 folds after 6–8 weeks, and Brca1 and Brca2 proteins to 1.4 folds after 2 weeks of exposure. Subsequently, the p53 protein declined to control level and the Brca1 and Brca2 proteins to 70% of the control after 16 weeks of exposure concomitant with the appearance of preneoplastic nodules in the liver. In contrast, in the transgenerationally exposed mice, the level of p53 protein remained largely invariant, and the levels of Brca1 and Brca2 proteins declined rapidly below control level without recording an initial increase. The appearance of pre-neoplastic nodules of the liver was significantly advanced in the transgenerationally exposed mice; developing in 8 weeks in F1, 6 weeks in F2 and 4 weeks in F3 mice. This clearly exhibits progressively increasing genomic instability due to prolonged AEBN exposure and enhanced cancer predisposition.

DNA sequence analyses revealed no mutation in exons 5 and 7 of the *p53* gene and the amplified segment (nucleotides 1–257) of exon 27 of the *Brca2* gene in P, F1, F2 and F3 mice. In contrast, a mis-sense mutation (G→C) was observed in exon 11 of the *Brca1* gene in F1, F2 and F3 mice, but not in P mice. Such a mutation would result in corresponding amino acid replacement Cys→Ser. *In silico* protein modeling revealed that the amino acid substitution was likely to cause structural alterations in the RAD50 binding region of the Brca1 protein, which is crucial for its function in error free repair of DNA single and double strand breaks. These observations clearly indicate that the *p53*, *Brca1* and *Brca2* tumor suppressor genes are intrinsically involved in the process of BN

**Table 1.** p53 associated alterations in betel nut (BN) and/or betel quid (BQ) associated human precancerous lesions/cancers.

#	Exposure condition	Effect(s)	P53 associated alteration(s)	Reference
1	BQ and alcohol	ESCC in Taiwanese population	A:T→G:C transition and G:C→T:A transversion	117
2	BQ	Atrophic oral lichen planus (OLP) in Taiwanese patients	Higher expressions of p53 and proliferating cell nuclear antigen (PCNA)	118
3	BN	Oral cancer in Thailand	Mutations detected in 11.8% (8/68) of betel-related tumors and 7 of 8 mutations were G:C to A:T transitions	119
4	BQ	Leukoplakia and OSCC in North Indian population	p53 missense mutations, p53 antibodies and p53 protein accumulation	120
5	BQ and tobacco	OSCC in Taiwanese population	G:C→A:T transitions	121
6	BQ and tobacco	OSCC in Indian population	Low incidence of p53 mutations	122
7	BQ	OSCC in Sri Lankan population	Point, small deletion and addition type of mutations mainly clustered in exon 5 of the <i>p53</i> gene	123
8	BQ and tobacco	OSCC in Taiwanese population	Mutations in codons 273–282 in exon 8 of p53, nuclear accumulation and positive p53 immunostaining	124
9	BN and tobacco	OSCC in South Indian population	Nuclear p53 staining and p53 expression	125
10	BQ without tobacco	Oral cancers from Papua New Guinea	Low frequency of p53 mutations	126
11	BN	OSCC in Sri Lankan population	Over expressed p53	127
12	BQ and tobacco	Lung cancer and oral cancer in North east Indian population	P53 codon 72 polymorphism	128

doi:10.1371/journal.pone.0042759.t001

induced carcinogenesis in mice as well as in the transgenerational transmission of carcinogenic risk following AEBN exposure. The p53, Brca1 and Brca2 responses were abrogated in the mice exposed transgenerationally to AEBN resulting in significantly increased predisposition to cancer [130,131]. The inactivation of the *p53* gene, which apparently played a crucial role in BN associated cancer in mice, was not achieved through *p53* mutation. The mechanism of p53 inactivation may also involve other routes, which requires to be investigated in the future. One possible alternative mechanism for p53 inactivation in BN induced carcinogenesis may be over expression of MDM2 protein, which has been shown in OSCC [133]. A high prevalence of MDM2 protein was also found in BQ chewing associated OSCC in Taiwan [134]. MDM2 protein has been shown to negatively regulate the function of p53 tumor suppressor protein through two main mechanisms. First, the direct binding of MDM2 to the N-terminal end of p53 inhibits the transcriptional activation function of p53. Second, MDM2 possesses E3 ubiquitin ligase activity that targets p53 for modification and subsequent degradation through the 26S proteasome [135]. Overexpression of MDM2 would therefore lead to carcinogenesis in a p53-dependent manner.

## 8. BETEL NUT POLYPHENOLS AND TANNINS IN CARCINOGENESIS

Toxicity studies relating to BN specific polyphenols and tannins are not conclusive with both carcinogenic and anti-carcinogenic effects being reported in literature. It is reported that ROS produced during autooxidation of BN polyphenols in the BQ chewer's saliva might be crucial in the initiation and promotion of OC [63]. However, the polyphenols are primarily known to be strong antioxidants and, thereby, also considered a food supplement that reduces the risk of degenerative diseases. Huang *et al.* have reported that the antioxidant capacity of the BNE procyanidins increased with the degree of polymerization. Further, they have also demonstrated that BNE which contains catechins based oligomeric and polymeric procyanidins, regulates COX-2 expression *in vitro* and possess anti-inflammatory potential *in vivo* [14]. Similarly, incidence of certain cancers, such as esophageal cancer, has been reported to correlate well with the consumption of tannins-rich food, such as BN, suggesting that tannins might also be carcinogenic. However, other reports indicated that the carcinogenic activity of tannins might be related to components associated with tannins such as "flavolans"- the polymers formed by condensation of flavans, referred to as polyflavonoid tannins or condensed tannins, rather than tannins themselves [136]. More research is required to properly understand this aspect.

## 9. BETEL NUT AND HUMAN GENETIC SUSCEPTIBILITY TO ORAL CANCER

Exposure to BN derived carcinogens, particularly alkaloids, enhance the risk of cancer in BN or BQ chewers in general. However, correlation between prevalence of cancer in human populations in different parts of the world and habit of BN/BQ mastication is not absolute. This suggests that the genetic makeup of the masticator has its own influence on the ultimate manifestation of BN induced cancer. It is becoming obvious that the interplay between the genetic constitution and the environmental factor(s), determine the final risk of human cancers, especially OC, following the exposure to BN or BQ alone or in combination with additives, including tobacco. Mere exposure to BN or BQ may not commit the chewer to cancer. For any given level of exposure to BN carcinogens, only a proportion of exposed individuals will develop cancer, indicating the prevalence of inter-

individual differences in susceptibility [137]. Individual susceptibility to cancer may originate from several factors, including (a) differences in metabolism influencing the metabolic activation of BN derived carcinogens, (b) status of DNA repair pathways and related genes, (c) patterns of expression of proto-oncogenes and tumor suppressor genes and (d) nutritional status of the masticator, etc. Variations in an individual's metabolic phenotype, i.e., phenotypic polymorphism, have also been detected in a variety of enzymes involved in activation and detoxification of chemical carcinogens. It is becoming clearer now that different phenotypic and/or metabolic variations stem from genetic polymorphisms prevalent in different population groups [138]. A number of genetic polymorphisms have been identified, which seem to be associated with the risk of BN induced preneoplastic lesions or pre-cancers, like OL/OE and OSF, as well as with the development of OC in human subpopulations in different regions of the world. These polymorphisms have been mapped to genes with diverse function. However, polymorphisms in a few genes appear to be more significant; these include the DNA repair genes, *XRCC4* in Taiwanese population and *XRCC1* and *XPD* in Indian population, genes encoding detoxifying enzymes such as *NAT2* encoding the most important phase II metabolic enzyme for BQ in Taiwanese population, *GSTT1* and *GSTM1* in Indian and Thai populations, and *CYP2A6* in Sri-Lankan population and genes encoding matrix metalloproteinases, such as *MMP9* in Taiwanese population and *MMP3* in Asian population (see Table 2) [139–165]. This clearly indicates the extremely complex and highly variable influence of the genetic makeup of the population groups on their cancer susceptibility. Further research in this area is also warranted.

## 10. BETEL NUT EXTRACT AND ROLE OF CYCLOOXYGENASE-2

Cyclooxygenase (COX), an inducible enzyme responsible for prostaglandin synthesis, plays an important role in certain inflammatory diseases and carcinogenesis. Tang *et al.* reported that COX-2 protein as well as mRNA expression was significantly enhanced in OSCC as compared to non-cancerous matched tissue (NCMT). Hydroxychavicol, a unique ingredient in BQ, also induced COX-2 overexpression in NHOK, indicating the early involvement of COX-2 in BQ associated OC [166]. Tsai *et al.* also reported that in human BMF, COX2 mRNA increased in a manner dependent upon increase in the dose of arecoline [167]. In addition, pretreatment with the GSH precursor, 2-oxothioazolidine-4-carboxylic acid, led to a decrease in the induction of COX2 mRNA by arecoline and the GSH synthesis inhibitor, buthioninesulfoximine, led to an increase, suggesting that regulation of COX2 expression induced by arecoline is critically dependent on cellular glutathione concentration [167]. Elevated COX2 protein levels have also been detected by immunohistochemistry in human tissues with moderate submucous fibrosis [9]. BNE was also found to induce COX2 mRNA and protein expression and PGE2 and 6-keto-PGF1 $\alpha$  in primary HGK cells [168]. It was suggested that this stimulation of PGE2 production could partly result from the upregulation of COX2 mRNA expression. BN extract also slightly enhanced the activity of COX in the human oral carcinoma cell line, OEC-M1, but inhibited its activity in KB cells at concentrations greater than 50  $\mu$ g/ml after 24 hours of exposure [169]. Upon treatment with BNE, head and neck carcinoma cells showed an increase of vimentin. The activation of extracellular signal-regulated kinase (ERK)/cyclooxygenase (COX)-2/prostaglandin (PGE)-2 cascade underlay the upregulation. These cells also exhibited the enhancement of migration and invasion. By knocking down COX-2 and vimentin expression, the increase of cell mobility was reversed. Tumors exhibiting extensive vimentin

**Table 2.** Genetic polymorphisms and their role(s) in betel nut (BN) and/or betel quid (BQ) associated human precancerous lesions/cancers.

#	Gene/region	Genetic polymorphism(s)	Effect(s)	Reference
1	Nuclear factor-kappa B (NF-κB)	Genetic polymorphisms of <i>NFKB1</i> and <i>NFKBIA</i>	<i>NFKB1</i> -94 ATTG2, <i>NFKBIA</i> -826 T and -881 G alleles are associated with oral carcinogenesis. The combination of <i>NFKB1</i> or <i>NFKBIA</i> gene polymorphisms and tobacco and betel consumption appears related to an increased risk of oral cancer. The genetic polymorphism of <i>NFKBIA</i> -519 might be a predictive factor for the distal metastasis of OSCC in Taiwanese	139
2	Survivin gene	Genetic polymorphisms of survivin gene	The survivin -31GG, +9194 GG, and +9809 TT homozygotes exhibited higher risk for oral cancer compared with the corresponding ancestral genotype, and +9809 SNPs combined with betel quid chewing and/or tobacco consumption could robustly elevate susceptibility to oral cancer. The distribution frequency of the -31 G: +9194 A: +9809 T haplotype was significantly higher in oral cancer patients than in control participants, in Taiwanese men	140
3	Epidermal growth factor receptor (EGFR) gene	Q787Q silent mutation	A high frequency of Q787Q mutation in BN chewing associated Taiwanese OSCC patients	141
4	Chemokine (C-C motif) receptor 2 gene CCR2	V64I CCR2 gene polymorphism	Individuals with GA or at least one A allele had a higher risk for oral cancer, compared to GG genotypes. Moreover, for subjects with GA or at least one A allele of V64I CCR2 gene polymorphism, those exposed to environmental risk factors including alcohol, tobacco and <i>Areca</i> consumptions possessed a significantly higher risk for oral cancer than those unexposed subjects in Chinese population	142
5	Cytochrome gene, <i>CYP26B1</i>	Genetic polymorphism of <i>CYP26B1</i>	Genetic polymorphism AA of <i>CYP26B1</i> appeared to correlate with the risk of oral squamous cell carcinoma (OSCC), and chewing BQ multiplicatively interacted with <i>CYP26B1</i> AA to increase the OSCC risk in Taiwanese population	143
6	Urokinase plasminogen activator (uPA) gene and plasminogen activator inhibitor (PAI)-1	Genetic polymorphisms of uPA gene. At least one 5G allele or 4G/4G genotype of PAI-1	Combination of uPA system gene polymorphisms and betel nut and tobacco consumption was related to the risk of oral cancer, while patients suffering from oral cancer with at least one 5G allele of PAI-1 gene had a low risk for the development of clinical stage III or IV and lymph node metastasis compared with those with 4G/4G homozygotes in Taiwanese population	144
6	Tumor necrosis factor-α (TNF-α)	TNFA genetic variants (-308G>A and -238G>A) with the risk and prognosis of BQ-related	G allele and G/G genotype at TNFA -308 were associated with increased risk of cancer as compared to those with A allele or A/A+A/G genotypes. In addition, G allele and G/G genotype at TNF-α -238 were associated with a borderline but statistically significant increased risk oral and pharyngeal squamous cell carcinoma (OPSCC) in Taiwanese population. Interactions between combined genotypes and smoking status were also found to contribute to risk of BQ-related OPSCC	145
7	Metallothionein 1 (MT-1)	rs8052394, rs11076161, rs8052334, rs964372, rs7191779 and rs708274 genotypes of MT-1	Individuals within Taiwanese population who inherited the MT-1 rs11076161 AA, rs964372 CC, and rs7191779 GC genotypes experienced significant protection against OSCC, whereas individuals carrying the MT-1 rs8052394 An allele seemed exposed to higher risk	146
8	N-acetyl transferase 2 (NAT-2)	Genetic polymorphism in NAT-2 resulting in slow NAT-2 acetylation haplotypes	The genotypic and allelic type of T341C and C481T in NAT-2 are associated with the risk of OPSCC in Taiwanese population	147
9	Glutathione-S-transferase (GST) genes	Polymorphism of GSTT1 gene	GSTT1 null genotype was found to be a significant risk factor for oral as well as gastric cancer in tobacco and BN associated cancer patients from Assam region of NE India	148
10	Microsomal epoxide hydrolase 1 (EPHX1)	139His/Arg genotype and 139Arg/Arg genotype	The 139His/Arg genotype was a significant risk factor for esophageal cancer in tobacco chewers and BQ chewers, while patients with the 139Arg/Arg genotype were at significantly higher risk for developing a well differentiated and moderately differentiated grade of tumor in India	149
11	hOGG1	Single nucleotide polymorphism (SNP) of hOGG1, codon 326	C allele of hOGG1 codon 326 may have a joint effect with BQ chewing on the development of oral cancer in Taiwanese population	150
12	Lysyl oxidase gene, LOX	G to A polymorphism at nucleotide 473 causing a non-conservative Arg158Gln change in the LOX amino acid sequence	The South Asian male patients of OSF older than 50 years had increased Arg158Gln in LOX	151
13	MDM2	Single nucleotide polymorphism in the MDM2 promoter (SNP 309)	The MDM2 SNP 309 GG genotype with mutated p53 contribute to early onset of both sporadic and hereditary malignancies in Taiwanese patients of BN associated OSCC	152

Table 2. Cont.

#	Gene/region	Genetic polymorphism(s)	Effect(s)	Reference
14	XRCC 4 intron 3	Ins/del variant	In smoker and BQ chewer groups, the XRCC4 intron 3 deletion variants exhibited 2.57- and 3.03-fold higher risks than the insertion genotype, respectively, in Taiwanese population	153
15	Cyclooxygenase (COX)	Polymorphisms of COX-2 –765G>C	COX-2 –765C allele vs. –765G/G genotype was a protective factor against OSCC development but was a risk factor for malignant potential of OSF in Taiwanese population	154
16	Matrix metallo-proteinase-9 (MMP-9) promoter	1562 C-to-T polymorphism	Enhanced OSCC risk in young Taiwanese male BN chewers	155
17	Matrix metallo-proteinase-3 (MMP-3) promoter	Insertion/deletion (–1171 5A->6A) polymorphisms	5A genotype polymorphism - enhanced risk of OSF but not OSCC among male Asian BN chewers	156
18	NFκB1 promoter	Insertion/deletion polymorphism (–94 ins/del ATTG) in NFκB1 promoter	NFκB1 insertion and HO-1 L allelotypes – significantly enhanced risks for different subsets of OSCC in male Asian BN chewers	157
19	DNA repair genes <i>XRCC1</i> and <i>XPB</i>	Polymorphisms Arg194Trp, Arg280His, and Arg399Gln of the <i>XRCC1</i> gene and Lys751Gln of the <i>XPB</i> gene	Variant allele of <i>XRCC1</i> 399 codon and <i>XPB</i> – enhanced risk of OC among South Indian BQ chewers and smokers	158
20	Heme oxygenase-1 ( <i>HO-1</i> )	Polymorphisms in a (GT) <sub>n</sub> microsatellite repeat in <i>HO-1</i> promoter in short (S), medium (M) and long (L) alleles	Longer (GT) <sub>n</sub> repeat allele L – higher risk of BN related OSCC; (GT) <sub>n</sub> repeat allele S - may be protective for OSCC in Asian population	159
21	Cytochrome gene <i>CYP2A6</i>	<i>CYP2A6</i> *4C mutation-gene deletion type of polymorphism	Deficient <i>CYP2A6</i> activity due to deletion – reduced risk of oral cancer risk in Sri Lankan BQ chewers	160
22	Cytochrome gene <i>CYP1A1</i>	<i>CYP1A1</i> A/G genotype (Ile/Val) and G/G genotype (Val/Val) in exon 7	<i>CYP1A1</i> exon 7 containing G allele - enhanced risk for OSCC and oral precancerous lesion in Chinese BN chewer and smoker	161
23	Collagen related genes: Collagen 1A1 and 1A2 (COL1A1 and COL1A2), Collagenase-1 (COLase), transforming growth factor β1 (TGF-β1), Lysyl oxidase (LYOXase), and Cystatin C (CST3)	Polymorphisms of six collagen related genes, COL1A1, COL1A2, COLase, TGF-β1, LYOXase and CST3	Multigenic mechanisms involving the collagen related genes enhance susceptibility to OSF among Taiwanese BQ chewers	162
24	Tumor necrosis factor-α (TNF-α)	Bi-allelic promoter region (–308) polymorphism on the TNFα gene	The high production allele, TNF2 - significantly lower among individuals with OSF in Taiwanese population	163
25	Glutathione-S-transferase genes <i>GSTM1</i> and <i>GSTT1</i>	<i>GSTM1</i> and <i>GSTT1</i> null genotypes ( <i>GSTM1</i> *2 and <i>GSTT1</i> *2)	Null genotypes of either or both <i>GSTM1</i> and <i>GSTT1</i> - enhanced risk of development of leukoplakia following exposure to tobacco with or without BQ in South Indian population	164
26	Glutathione-S-transferase genes <i>GSTM1</i> and <i>GSTT1</i>	Genetic polymorphism of <i>GSTM1</i> and <i>GSTT1</i>	Homozygous deletion of <i>GSTM1</i> gene – enhanced risk for oral cancer, which is further compounded by exposure to cigarette smoke, alcohol, and BQ in Thai population	165

doi:10.1371/journal.pone.0042759.t002

and/or COX-2 expression displayed a significantly worse disease-associated survival than contrast groups. The study thus revealed that BN-modulated vimentin expression enhanced the progression of head and neck carcinoma [170]. In another study, OECM-1 and Fadu cells developed a fibroblastoid morphology and there exhibited an increase in vimentin expression after BNE treatment. The treatment also induced the phosphorylation of AKT and glycogen synthase kinase 3β in OECM-1 cells. Blockage of phosphatidylinositol 3-kinase (PI3K)/AKT signaling attenuated vimentin expression when it was induced by BNE. However, it did not affect BNE-mediated extracellular signal-regulated kinase (ERK) activation or cyclooxygenase 2 (COX-2) upregulation. Oral carcinoma tissue samples were found to have significantly higher levels of vimentin and pAKT expression than their controls. Tumors exhibiting no vimentin expression and weak AKT phosphorylation were found to be associated with better survival than groups with higher levels of expression. These results imply that PI3K/AKT activation and vimentin expression are

important pathogenic cascades in BN associated OC [171]. It thus appears that there is conclusive evidence to support a role of increased expression of vimentin in BN associated carcinogenesis [170,171]. Involucrin is a key component of the cornified envelope and a differentiation marker of keratinocytes. Non-toxic BNE treatment of normal human oral keratinocyte (NHOK) was reported to induce a downregulation of involucrin and disruption of involucrin distribution and activation of AKT as well as upregulation of COX-2. The BNE associated downregulation of involucrin through AKT pathway could underlie the BN-associated epithelial pathogenesis [172].

## 11. BETEL NUT IN APOPTOSIS AND AUTOPHAGY

Autophagy is a regulated self cannibalism, classified as type II programmed cell death, and is preceded by the inhibition of the mammalian target of rapamycin (mTOR). The hallmarks of autophagy are the cleavage of the precursor form of microtubule associated protein 1 light chain 3 (LC3-I) (molecular

weight = 18 kDa) to the active form LC3-II (molecular weight = 16 kDa) and the emergence of autophagic vacuoles (AV) and acidic vesicles [173]. Liu *et al.* reported that BNE induced (a) rounding cell morphology and nuclear shrinkage in different types of carcinoma cells, (b) the cleavage of LC3-I, and (c) the emergence of AV and acidic vesicles [173]. On the other hand, arecoline triggered (a) caspase-3 activation, (b) perinuclear chromatin condensation and (c) micronucleation, thus inducing atypical apoptosis. This difference is thought to be due to the ability of BNE, but not arecoline, to inhibit the phosphorylation of the mTOR-Ser2448. Lu *et al.* reported that BNE treatment induced autophagy among oral cancer cells characterized by LC3-II accumulation, genesis of autophagosomes and the appearance of EGFP-LC3 puncta [173]. Significantly, the blockage of BNE induced autophagy increased the proportion of oral cancer cells undergoing apoptotic death indicating that the eventual induction of autophagy was beneficial to cell survival from BNE induced apoptosis [174]. Transmission electron microscopy (TEM) of liver precancerous nodules induced in Swiss Albino mice upon transgenerational exposure to AEBN revealed enhanced cristolysis of mitochondria and formation of AV [132]. Cristolysis would induce deficiency of oxidative ATP production and inhibition of apoptosis. Moreover, the cells would have to meet their nutritional requirements through autophagy, thus, surviving and proliferating in the face of metabolic stress.

## 12. BETEL NUT AND MALIGNANT LESIONS

Studies and follow-up programmes conducted over the past three decades indicate that the rate of malignant transformation of pre-cancer lesions shows a population dependent variability. A study conducted in Karachi, Pakistan (1996–98) on 79 cases of OSCC and 149 hospital controls showed that the risk for developing OC was 19 times higher (95% CI, 4.2–87.7) among cases of OSF than among subjects with no precancerous condition [175]. In a study by Shiu *et al.* conducted in Taiwan, 60 cases of oral and pharyngeal cancers, including lip, tongue, gum, mouth floor, buccal palate, oropharynx and hypopharynx cancers were evaluated by linking a retrospective leukoplakia cohort consisting of 435 patients recruited from hospitals between 1988 and 1998 to a population cancer registry [176]. The risk for malignant transformation increased with time, particularly for BN chewers. Using a Weibull survival model, the adjusted hazard ratio for chewing BN without tobacco was 4.6 (95% CI, 1.3–16.9) after matching for age and sex.

Thus, the presence of pre-cancer lesions appears to correlate well with malignant transformation and eventual development of OC (Figure 5C). Besides OC, individuals who chewed BN or BQ without or with tobacco also reported other aerodigestive cancers. From 1997 to 1998, a hospital based case–control study on oesophageal cancer in Assam, India, included 502 cases (358 men, 144 women) and 994 controls (706 men, 288 women) who were attendants to cancer patients. The risk for esophageal cancer increased with increasing frequency of chewing BQ without or with tobacco and increased substantially when the chewing habit had lasted 20 years or more. A dose–response relationship was also observed for age at starting the habit, with a higher risk for starting at a younger age [177].

## 13. EFFORTS TO CONTROL USE OF BETEL NUT, BETEL QUID AND ADDITIVES, INCLUDING PAAN MASAL AND GUTKHA

It is a scientifically established fact that BN or BQ without or with various additives, including tobacco products, are clearly

associated with increased risk of HNC, especially OC. In spite of this fact, it has remained a challenge for governments and policy makers to contain its usage and reduce the deleterious effects of BN or BQ on population due to its socio-cultural heritage and addictive nature. Global data on efforts to control usage of BN or BQ and its additives strongly suggest that no single strategy is likely to succeed in achieving the desired level of control on BN or BQ consumption. A concerted effort involving radical measures is required in this direction. It may involve health care professionals, media, policy makers, law enforcing agencies and the community at large. Even though no unified guideline exists due to variation in the socio-cultural groups, it appears practical to make sustained behavioral interventions, especially adolescents and young adults, for reducing the use of BN or BQ without or with additives. Lack of scientific information on its deleterious effects on health is conspicuous by its absence. In contrast, various advertisements try to glorify the opposite in order to allure most vulnerable section of the society, the adolescents and young adults. Once addicted, they become a long term consumers of BN or BQ. Media may play a very important role in this effort due to its widespread reach in all strata of the society, particularly the lower strata, which appears to be deeply affected by this practice. Governmental and non-governmental health care professionals may routinely assess and record the usage pattern in the society and rigorously inform the individuals, groups of people or patients about its potential hazard. Chang *et al.* reported that an oral screening program conducted in a tertiary medical centre for detection of oral lesions and oral cancer was effective. The group suggests that individuals aged  $\geq 40$  years or who are habitual cigarette smokers, alcohol consumers, and BQ chewers should receive oral screening regularly so that potential oral cancer can be detected as early as possible [178].

Unnecessary glorification of the BN or BQ products in media must be stopped by enacting appropriate laws. Visual representation of the diseases, particularly OC, caused by the use of these products and appropriate warning signs may be effective in this campaign. The industrial policies, including taxation, need revisit in order to limit such industrial activities to discourage marketing of such products. There is need to have strong political commitment towards this aim. Many countries have made some headway in this direction. A brief summary of such efforts is listed below:

**(a) India.** On 1 August 2002, the Commissioner for Food and Drug Administration and Food (Health) Authority, Maharashtra State, issued a gazette notification banning the manufacture, sale and storage of *gutkha* and *paan masala* or any similar product containing or not containing tobacco. In India, a warning label is now mandatory on packets of commercial BN and tobacco products, but there are no regulations about the size of the letters. Several states are at various stages of passing laws to ban *gutkha* or are in court after being challenged by the industry. A recommendation that *gutkha* should be banned nationwide has been made to the Government of India by the Central Committee on Food Safety [9]. Recently, a ban on using plastic packing was passed by the Supreme Court of India with effect from March 2011 and is being implemented at the time of writing this review ([http://www.tobaccojournal.com/Chewing\\_tobacco\\_plastic\\_pouches\\_banned.50432.0.html](http://www.tobaccojournal.com/Chewing_tobacco_plastic_pouches_banned.50432.0.html)). On the occasion of World No-Tobacco day on the 31<sup>st</sup> May 2012, the state of Bihar joined two other states (Kerala and Madhya Pradesh) in banning manufacture, storage, distribution and sale of any form of tobacco and nicotine containing *paan masala* and *gutkha* for a period of 1 year to begin with (<http://indiatoday.intoday.in/story/bihar-bans-sale-of-gutkha-for-a-year/1/198280.html>).



**(b) North America.** BN figures on the list of herbs that are unacceptable as a non-medicinal ingredient in oral use products. The sale of BN products has been banned in Canada as a result of the link between arecoline and mutagenic effects. The US FDA maintains an import alert within the USA, the main concerns being adulteration and addition of unsafe food additives. In 1976, the US Government announced a ban on interstate traffic of BN [9].

**(c) European Union.** Within the European Union, excluding Sweden, there is legislation banning the sale of tobacco products for oral use. However, there are no specific laws regulating or banning the sale of BN products, even when mixed with smokeless tobacco, as chewing tobacco is excluded from the directive [9].

**(d) United Kingdom.** In the United Kingdom, there is no law to regulate the import or sale of products containing BN. Presently numerous BN preparations, with or without tobacco, are commercially available in shops. The Department of Trade and Industry classifies these products as sweets. Labelling and a list of ingredients on the packaging are sometimes non-existent. Several studies have shown that in most outlets the sales are unrestricted to minors and children under the age of 16 were able to purchase *gukha* easily. Only a few BN products give specific health warnings on the dangers of chewing BN, although most carry the statutory health warning regarding added tobacco. Among 20 commercially processed and packaged BN products on sale in the United Kingdom, only three carried a health warning related to OC; none warned about OSF or potential addiction [9].

**(e) Other Countries.** In the late 1970s, the Public Services of Papua New Guinea issued a ban on BQ chewing in government offices. Possession of BN in the California public school system is grounds for suspension. In Singapore, spitting in public places can lead to a fine, indirectly discouraging the practice of BQ and BN chewing [9].

## Conclusions

BN and products derived from it are widely used as a masticatory among various communities, and in several countries across the world, as a socially endorsed habit [1–11]. Over a long period, several additives got added to a simple BN preparation, thus, creating the BQ and encompassing chewing tobacco in the preparation. The addictive nature of BN and/or the additives that make it BQ, are essentially responsible for its rampant usage among individuals [19–22]. The popularity has led to industrial preparation of convenient substitutes of the BN/BQ in the form of

*paan masala*, *gukha* and the likes in several countries. This review reveals that BN has far reaching consequences on the general health of the masticator, especially in context with the oral health of the users [23–56]. Extensive studies by several workers over the years conclusively prove the role of BN, and its components, primarily the alkaloid arecoline, as a carcinogen [55–61]. These substances not only have general mutagenic, cytotoxic and genotoxic properties, but are also intricately involved in enzymatic, molecular and genetic mechanisms that result in the development of carcinogenesis at various sites, specifically in the oral cavity [58–150]. More research is clearly required to fill many existing gaps in the understanding of the seemingly highly complex interactions of BN with the life process and its manifestation in HNC, particularly OC. Control over human consumption of BN and BQ without or with additives, including tobacco, or its convenient commercial substitutes, such as *gukha* and *paan masala*, is proving to be difficult because the habit is not associated with any social stigma and taboo. In fact, it is other way round in which this practice has largely been given social, religious or other sanctions in different regions of the world. Hence, strong multifaceted intervention is required to discourage or control the habit of BN/BQ mastication. Firstly, legislation against open sale and use of such products should be stricter and more states and countries should bring out such legislations sooner than later. Secondly, public awareness should be created regarding the harmful effects of these products among all sections of society, particularly among children, since the habit starts early in the majority of cases. Lastly, attempts need to be made for harm-reduction of BN/BQ/commercial substitutes. Multi-institutional transnational case controlled studies are also required in order to establish the exact etiopathogenesis and molecular changes of diseases caused by BN/BQ and/or its constituents.

## Acknowledgments

RNS thanks his past students and coworkers whose works may have been used in the review. The authors also thank Dr. Vedant Pahlajani for help in acquisition of clinical pictures and Mr. Pinkhupborlang Malngiang for excellent artwork and photography.

## Author Contributions

Conceived and designed the experiments: RNS RM YC. Performed the experiments: RNS RM YC. Analyzed the data: RNS RM KA. Contributed reagents/materials/analysis tools: RNS RM YC KA. Wrote the paper: RNS YC RM KA.

## References

- Burnell AC, Yule HA (1903) *Hobson-Jobson: A Glossary of Colloquial Anglo Indian Words*. (Ed.) William Crooke.
- Sharan RN, Choudhury Y (2010) Betel nut and susceptibility to cancer. In: Roy D, Dorak MT, editors. *Environment Factors, Genes, and the Development of Human Cancers*. Springer Science+Business Media. Part 3, 401–428.
- Warnakulasuriya S (2002) Areca nut use: an independent risk factor for oral cancer. *Br Med J* 324: 799–800.
- IARC (1985) International Agency for Research on Cancer-Tobacco habits other than Smoking; Betel quid and Areca-nut chewing; and some related nitrosamines. In: IARC Monograph Evaluating Carcinogenic Risk from Chemicals to Humans. Lyon: IARC, 37: p263
- Sharan RN (1996) Association of betel nut with carcinogenesis – A review. *Cancer J* 9: 13–19.
- Nair U, Bartsch H, Nair J (2004) Alert for an epidemic of oral cancer due to use of the betel quid substitutes *gukha* and *paan masala*: a review of agents and causative mechanisms. *Mutagenesis* 19: 251–262.
- Changrai J, Gany F (2005) *Paan* and *Gutka* in the United States: An Emerging Threat. *J Imm Health* 7: DOI: 10.1007/s10903-005-2643-7.
- Reichert PA, Zhang X (2007) Misconceptions related to the areca nut chewing habits of mainland China. *Oral Oncol* 43: 958–959.
- IARC (2004) International Agency for Research on Cancer (IARC) – Summaries & Evaluations: Betel-quid and areca-nut chewing. IARC Monograph Evaluating Carcinogenic Risk from Chemicals to Humans. Lyon: IARC, 85: p39.
- Reichert PA, Creutz U, Scheifele C (2006) The ‘Skull from Bangkok’: a skull of a betel quid chewer in the anthropological collection of Rudolf Virchow (Berlin). *J Oral Pathol Med* 35:410–412.
- Norton SA (1998) Betel: consumption and consequences. *J Am Acad Dermatol* 38: 81–88.
- Lee C-H, Ko AM, Warnakulasuriya S, Yin B-L, Sunarjo, et al. (2011) Intercountry prevalence and practices of betel-quid use in south, southeast and eastern Asia regions and associated oral preneoplastic disorders: an international collaborative study by Asian betel-quid consortium of south and east Asia. *Int J Cancer*. DOI: 10.1002/ijc.25809.
- Nair J, Ohshima H, Friesen M, Croisy A, Bhide SV, et al. (1985) Tobacco-specific and betel nut-specific N-nitroso compounds: occurrence in saliva and urine of betel quid chewers and formation *in vitro* by nitrosation of betel quid. *Carcinogen* 6: 295–303.
- Huang P-L, Chi C-W, Liu T-Y (2010) Effects of *Areca catechu* L. containing procyanidins on cyclooxygenase-2 expression *in vitro* and *in vivo*. *Food Chem Toxicol* 48: 306–313.

15. Nair U, Floyd RA, Nair J, Bussachini V, Friesen M, et al. (1987) Formation of reactive oxygen species and of 8-hydroxydeoxyguanosine in DNA *in vitro* with betel quid ingredients. *Chem Biol Interact* 63: 157–169.
16. Stich HF, Anders F (1989) The involvement of reactive oxygen species in oral cancers of betel quid/tobacco chewing. *Mutat Res* 214: 47–61.
17. Nair UJ, Obe G, Friesen M, Goldberg MT, Bartsch H (1992) Role of lime in the generation of reactive oxygen species from betel-quid ingredients. *Environ Health Perspect* 98: 203–205.
18. Winstock A (2002) Areca nut-abuse liability, dependence and public health. *Addict Biol* 7: 133–138.
19. Chu NS (2002) Neurological aspects of areca and betel chewing. *Addict Biol* 7: 111–114.
20. Chandra PS, Carey MP, Carey KB, Jairam KR (2002) Prevalence and correlates of BN use among psychiatric patients in India. *Drug Alcohol Depend* 69: 1–6.
21. Pickwell SM, Schimelpfening S, Palinkas LA (1994) 'Betelmania'. Betel quid chewing by Cambodian women in the United States and its potential health effects. *West J Med* 160: 326–330.
22. Winstock AR, Trivedy CR, Warnakulasuriya KAAS, Peters TJ (2000) A dependency syndrome related to areca nut use: some medical and psychological aspects among areca nut users in the UK. *Addict Biol* 5: 173–179.
23. Benegal V, Rajkumar RP, Muralidharan K (2008) Does areca nut use lead to dependence? *Drug Alcohol Depend* 97: 114–121.
24. Trivedy C R, Craig G, Warnakulasuriya S (2002) The oral health consequences of chewing areca nut. *Addict Biol* 7: 115–125.
25. Walker DM, Boey G, McDonald LA (2003) The pathology of oral cancer. *Pathol* 35: 376–383.
26. Yen AM, Chen SC, Chang SH, Chen TH (2008) The effect of betel quid and cigarette on multistate progression of oral pre-malignancy. *J Oral Pathol Med* 37: 417–22.
27. Angadi PV, Rao SS (2010) Areca nut in pathogenesis of oral submucous fibrosis: revisited. *Oral Maxillofac Surg*. DOI 10.1007/s10006-010-0219-8.
28. Khirime RD, Mehra YN, Mann SB, Mehta SK, Chakraborti RN (1991) Effect of instant preparation of betel nut (*pan masala*) on the oral mucosa of albino rats. *Indian J Med Res* 94: 119–124.
29. Amarasinghe HK, Usgodaarachchi US, Johnson NW, Laloo R, Warnakulasuriya S (2010) Betel-quid chewing with or without tobacco is a major risk factor for oral potentially malignant disorders in Sri Lanka: a case-control study. *Oral Oncol* 46: 297–301.
30. Tilakarathne WM, Klimkowski MF, Saku T, Peters TJ, Warnakulasuriya S (2006) Oral submucous fibrosis: review on aetiology and pathogenesis. *Oral Oncol* 42: 561–568.
31. Pandya S, Chaudhary AK, M Singh, Singh M, Mehrotra R (2009) Correlation of histopathological diagnosis with habits and clinical findings in oral submucous fibrosis. *Head Neck Oncol* 1:10.
32. Mehrotra R, Pandya S, Chaudhary AK, Kumar M, Singh M (2008) Prevalence of oral premalignant and malignant lesions at a tertiary level hospital in Allahabad, India. *Asia Pacific J Cancer Prev* 9: 263–266.
33. Gupta PC, Ray CS (2004) Epidemiology of betel quid usage. *Ann Acad Med Singapore* 33:31–36.
34. Jacob BJ, Straif K, Thomas G, Ramadas K, Mathew B, et al. (2004) Betel quid without tobacco as a risk factor for oral precancers. *Oral Oncol* 40: 697–704.
35. Ariyawardana A, Athukorala AD, Arulanandam A (2006) Effect of chewing, tobacco smoking and alcohol consumption on oral submucous fibrosis: a case-control study in Sri Lanka. *J Oral Pathol Med* 35: 197–201.
36. Ranganathan K, Devi MU, Joshua E, Kirankumar K, Saraswathi TR (2004) Oral submucous fibrosis: a case-control study in Chennai, South India. *J Oral Pathol Med* 33: 274–277.
37. Chung-Hung T, Shun-Fa Y, Yu-Chao C (2007) The upregulation of cystatin C in oral submucous fibrosis. *Oral Oncol* 43: 680–685.
38. Deng YT, Chnag JZ, Yeh CC, Cheng SJ, Kuo MY (2011) Arecoline stimulated Cyr61 production in human gingival epithelial cells: inhibition by lovastatin. *Oral Oncol* 47: 256–261.
39. Lin PC, Chang WH, Chen YH, Lee CC, Lin YH (2011) Cytotoxic effects produced by arecoline correlated to epigenetic regulation in human K-562 cells. *J Toxicol Environ Health A* 74: 737–745.
40. Tsai CH, Yang SF, Chen YJ, Chu SC, Hsieh YS, et al. (2004) Regulation of interleukin-6 expression by arecoline in human buccal mucosal fibroblasts is related to intracellular glutathione levels. *Oral Dis* 10: 360–364.
41. Tsai CH, Yang SF, Chen YJ, Chou MY, Chang YC (2005) Raised keratinocyte growth factor-1 expression in oral submucous fibrosis *in vivo* and upregulated by arecoline in human buccal mucosal fibroblasts *in vitro*. *J Oral Pathol* 34: 100–105.
42. Tsai CH, Yang SF, Chen YJ, Chou MY, Chang YC (2005) The upregulation of insulin-like growth factor-1 in oral submucous fibrosis. *Oral Oncol* 41: 940–946.
43. Jeng JH, Wang YJ, Chiang BL, Lee PH, Chan CP, et al. (2003) Roles of keratinocyte inflammation in oral cancer: regulating the prostaglandin E2, interleukin-6 and TNF-alpha production of oral epithelial cells by areca nut extract and arecoline. *Carcinogen* 24: 1301–1315.
44. Harvey W, Scutt A, Meghji S, Canniff JP (1986) Stimulation of human buccal mucosa fibroblasts *in vitro* by betel-nut alkaloids. *Arch Oral Biol* 31: 4509.
45. Aziz SR (1997) Oral submucous fibrosis: An unusual disease. *JNJ Dent Assoc* 68: 17–19.
46. van Wyk CW, Stander I, Padayachee A, Grobler-Rabie AF (1993) The areca nut chewing habit and oral squamous cell carcinoma in South African Indians. A retrospective study. *S Africa Med J* 83: 425–429.
47. Meghji S, Scutt A, Harvey W, Canniff JP (1987) An *in vitro* comparison of human fibroblasts from normal and oral submucous fibrosis tissue. *Arch Oral Biol* 32: 213–215.
48. Ni WF, Tsai CH, Yang SF, Chang YC (2007) Elevated expression of NF-kappaB in oral submucous fibrosis - evidence for NF-kappaB induction by safole in human buccal mucosal fibroblasts. *Oral Oncol* 43: 557–562.
49. Lin SC, Lu SY, Lee SY, Lin CY, Chen CH, et al. (2005) Areca (betel) nut extract activates mitogen-activated protein kinases and NF-kappaB in oral keratinocytes. *Int J Cancer* 116: 526–535.
50. Shieh DH, Chiang LC, Shieh TY (2003) Augmented mRNA expression of tissue inhibitor of metalloproteinase-1 in buccal mucosal fibroblasts by arecoline and safole as a possible pathogenesis for oral submucous fibrosis. *Oral Oncol* 39: 728–735.
51. Kato K, Hara A, Kuno T, Kitaori N, Huilan Z, et al. (2005) Matrix metalloproteinases 2 and 9 in oral squamous cell carcinomas: manifestation and localization of their activity. *J Cancer Res Clin Oncol* 131: 340–346.
52. Liu SY, Lin MH, Yang SC, Huang GC, Chang L, et al. (2005) Areca quid chewing enhances the expression of salivary matrix metalloproteinase-9. *J Formos Med Assoc* 102: 113–119.
53. Trivedy CR, Warnakulasuriya KA, Peters TJ, Senkus R, Hazarey VK, et al. (2000) Raised tissue copper levels in oral submucous fibrosis. *J Oral Pathol Med* 29: 241–248.
54. Trivedy C, Meghji S, Warnakulasuriya KA, Johnson NW, Harris M (2001) Copper stimulates human oral fibroblasts *in vitro*: a role in the pathogenesis of oral submucous fibrosis. *J Oral Pathol Med* 30: 465–470.
55. Khanna SS, Karjodkar FR (2006) Circulating Immune Complexes and trace elements (Copper, Iron and Selenium) as markers in oral precancer and cancer: A randomised, controlled clinical trial. *Head Face Med* 2: 33.
56. Wen CP, Tsai MK, Chung WS, Hsu HL, Chang YC, et al. (2010) Cancer risks from betel quid chewing beyond oral cancer: a multiple-site carcinogen when acting with smoking. *Cancer Causes Control* 21: 1427–1435.
57. Wu MT, Lee YC, Chen CJ, Yang PW, Lee CJ (2001) Risk of betel chewing for oesophageal cancer in Taiwan. *Br J Cancer* 85: 658–660.
58. Shiu MN, Chen THH, Chang SH, Hahn LJ (2000) Risk factors for leukoplakia and malignant transformation to oral carcinoma: a leukoplakia cohort in Taiwan. *Br J Cancer* 82: 1871–1874.
59. Lin Y, Jen Y, Wang B, Lee J, Kang B (2005) Epidemiology of oral cavity cancer in Taiwan with emphasis on the role of betel nut chewing. *ORL J Otorhinolaryngol Relat Spec* 67: 230–236.
60. Chen YJ, Chang JT, Liao C, Wang H, Yen T, et al. (2008) Head and neck cancer in the betel quid chewing area: recent advances in molecular carcinogenesis. *Cancer Sci* 99: 1507–1514.
61. Merchant A, Husain SS, Hosain M, Fikree FF, Pitiphat W, et al. (2000) *Paan* without tobacco: an independent risk factor for oral cancer. *Int J Cancer* 86: 128–131.
62. Jeng JH, Chang MC, Hahn LJ (2001) Role of areca nut in betel quid-associated chemical carcinogenesis: current awareness and future perspectives. *Oral Oncol* 37: 477–492.
63. Wary KK, Sharan RN (1991) Cytotoxic and cytostatic effects of arecoline and sodium nitrite on human cells *in vitro*. *Int J Cancer* 47: 396–400.
64. Miyazaki M, Sugawara E, Yoshimura T, Yamazaki H, Kamataki T (2005) Mutagenic activation of betel quid-specific N-nitrosamines catalyzed by human cytochrome P450 coexpressed with NADPH-cytochrome P450 reductase in *Salmonella typhimurium* YG7108. *Mutat Res* 581: 165–171.
65. Saikia JR, Schneeweiss FHA, Sharan RN (1999) Arecoline-induced changes of poly-ADP-ribosylation of cellular proteins and its influence on chromatin organization. *Cancer Lett* 139: 59–65.
66. Deb S, Chatterjee A (1998) Influence of buthionine sulfoximine and reduced glutathione on arecoline-induced chromosomal damage and sister chromatid exchange in mouse bone marrow cells *in vivo*. *Mutagen* 13: 243–248.
67. Panigrahi GB, Rao AR (1989) Study of the genotoxicity of the total aqueous extract of betel nut and its tannin. *Carcinogenesis* 7: 37–39.
68. Chiang C, Chang M, Lee J, Chang J Y, Lee P et al. (2004) Hamsters chewing betel quid or areca nut directly show a decrease in body weight and survival rates with concomitant epithelial hyperplasia of cheek pouch. *Oral Oncol* 40: 720–727.
69. Perera MWS, Gunasinghe D, Perera PAJ, Ranasinghe A, Amaratunga P (2007) Development of an *in vivo* mouse model to study oral submucous fibrosis. *J Oral Pathol Med* 36: 273–280.
70. Jaju RJ, Patel RK, Bakshi SR, Trivedi AH, Dave BJ, et al. (1992) Chromosome damaging effects of *pan masala*. *Cancer Lett* 65: 221–226.
71. Ramchandani AG, D'Souza AV, Borges AM, Bhisey RA (1998) Evaluation of carcinogenic/co-carcinogenic activity of a common chewing product, *pan masala*, in mouse skin, stomach and esophagus. *Int J Cancer* 75: 225–232.
72. Bhisey RA, Ramchandani AG, D'Souza AV, Borges AM, Notani PN (1999) Long-term carcinogenicity of *pan masala* in Swiss mice. *Int J Cancer* 83: 679–84.
73. Wary KK, Sharan RN (1988) Aqueous extract of betel-nut of Northeastern India induces DNA strand breaks and enhances rate of cell proliferation *in vitro*. *J Cancer Res Clin Oncol* 114: 579–582.

74. Sundqvist K, Liu Y, Nair J, Bartsch H, Arvidson K, et al. (1989) Cytotoxic and genotoxic effects of areca nut-related compounds in cultured human buccal epithelial cells. *Cancer Res* 49: 5294–5298.
75. Sharan RN, Wary KK (1992) Study of unscheduled DNA synthesis following exposure of human cells to arecoline and extracts of betel nut *in vitro*. *Mutat Res* 278: 271–276.
76. Lu S, Chang K, Liu C, Tseng Y, Lu H, et al. (2006) Ripe areca nut extract induces G<sub>1</sub> phase arrests and senescence-associated phenotypes in normal human oral keratinocyte. *Carcinogenesis* 27: 1273–1284.
77. Liu SY, Liu YC, Huang WT, Huang GC, Chen TC, et al. (2007) Up-regulation of matrix metalloproteinase-8 by betel quid extract and arecoline and its role in 2D motility. *Oral Oncol* 43: 1026–33.
78. Yang CY, Meng CL, van der Bijl P, Lee HK (2002) The effect of betel nut extract on cell growth and prostaglandin endoperoxide synthase in human epidermoid carcinoma cells. *Prostaglandin Other Lipid Mediat* 67: 181–195.
79. Ho TJ, Chiang CP, Hong CY, Kok SH, Kuo YS, et al. (2000) Induction of the c-jun protooncogene expression by areca nut extract and arecoline on oral mucosal fibroblasts. *Oral Oncol* 36: 432–436.
80. Jeng JH, Kuo ML, Hahn LJ, Kuo MY (1994) Genotoxic and non-genotoxic effects of betel quid ingredients on oral mucosal fibroblasts *in vitro*. *J Dental Res* 73: 1043–1049.
81. Kumpawat K, Deb S, Ray S, Chatterjee A (2003) Genotoxic effect of raw betel-nut extract in relation to endogenous glutathione levels and its mechanism of action in mammalian cells. *Mutat Res* 538: 1–12.
82. Lin CC, Chang MC, Chang HH, Wang TM, Tseng WY et al. (2009) Areca nut-induced micronuclei and cytokinesis failure in Chinese hamster ovary cells is related to reactive oxygen species production and actin filament deregulation. *Environ Mol Mutagen* 50: 367–374.
83. Yang SC, Lin SC, Chiang WF, Yen CY, Lin CH, et al. (2003) Areca nut extract treatment elicits the fibroblastoid morphological changes, actin reorganization and signaling activation in oral keratinocytes. *J Oral Pathol Med* 32: 600–605.
84. Jeng JH, Lan WH, Hahn LJ, Hsieh CC, Kuo MY (1996) Inhibition of the migration, attachment, spreading, growth and collagen synthesis of human gingival fibroblasts by arecoline, a major areca alkaloid, *in vitro*. *J Oral Pathol Med* 25: 371–375.
85. Chang YC, Hu CC, Tseng TH, Tai KW, Lii CK, et al. (2001) Synergistic effects of nicotine on arecoline-induced cytotoxicity in human buccal mucosal fibroblasts. *J Oral Pathol Med* 30: 458–464.
86. Lee P, Chang M, Chang W, Wang T, Wang Y (2006) Prolonged exposure to arecoline arrested human KB epithelial cell growth: Regulatory mechanisms of cell cycle and apoptosis. *Toxicol* 220: 81–89.
87. Tsai YS, Lee KW, Huang JL, Liu YS, Juo SH et al. (2008) Arecoline, a major alkaloid of areca nut, inhibits p53, represses DNA repair, and triggers DNA damage response in human epithelial cells. *Toxicol* 249: 230–247.
88. Chang YC, Hu CC, Lii CK, Tai KW, Yang SH, et al. (2001) Cytotoxicity and arecoline mechanisms in human gingival fibroblasts *in vitro*. *Clin Oral Investig* 5: 51–56.
89. Chiang S, Jiang S, Wang Y, Chiang H, Chen P, et al. (2007) Characterization of arecoline-induced effects on cytotoxicity in normal human gingival fibroblasts by global gene expression profiling. *Toxicol Sci* 100: 66–74.
90. Chen YJ, Liao CT, Chen PJ, Lee LY, Li YC et al (2011) Downregulation of *Ches1* and other novel genes in oral cancer cells chronically exposed to areca nut extract. *Head Neck* 33: 257–266.
91. Tsai YS, Lin CS, Chiang SL, Lee CH, Lee KW, et al. (2011) Areca nut induces miR-23a and inhibits repair of DNA double-strand breaks by targeting FANCG. *Toxicol Sci* 123: 480–490.
92. Lu HH, Liu CJ, Liu TY, Kao SY, Lin SC, et al. (2008) Areca-treated fibroblasts enhance tumorigenesis of oral epithelial cells. *J Dent Res* 87: 1069–1074.
93. Lin KH, Lin CY, Liu CC, Chou MY, Lin JK (2011) Arecoline N-oxide: its mutagenicity and possible role as ultimate carcinogen in areca oral carcinogenesis. *J Agri Food Chem* 59: 3420–3428.
94. Sharan RN (1994) Biochemical investigation of carcinogenic potency of betel nut (Kwai) of north-east India. In: *Oral Oncology*, vol. III, New Delhi: Macmillan India Ltd. pp. 190–193.
95. Balachandran B, Sharan RN (1995) Induction of mutations by different extracts of betel nut and radiation: Their implication in carcinogenesis. In: *Radiation Research*, Editors U. Hagen, H. Jung, and C. Streffler, vol. 1, Würzburg: Universitätsdruckerei H. Stürtz AG. pp 165.
96. Lai K, Lee T (2006) Genetic damage in cultured human keratinocytes stressed by long-term exposure to areca nut extracts. *Mutat Res Fundam Mol Mech Mutag* 599: 66–75.
97. Joshi MS, Verma Y, Gautam AK, Shivgotra VK, Parmar G, et al. (2011) Assessment of genetic damage among chewers of mixture containing mainly areca nut and tobacco. *Asia Pacific J Public Health* 23: 852–860.
98. Wang YC, Tsai YS, Huang JL, Lee KW, Kuo CC et al. (2010) Arecoline arrests cells at prometaphase by deregulating mitotic spindle assembly and spindle assembly checkpoint: implication for carcinogenesis. *Oral Oncol* 46: 255–262.
99. Lin SC, Chen YJ, Kao SY, Hsu MT, Lin CH, et al. (2002) Chromosomal changes in betel-associated oral squamous cell carcinomas and their relationship to clinical parameters. *Oral Oncol* 38: 266–273.
100. Desai SS, Ghaisas SD, Jakhi SD, Bhide SV (1996) Cytogenetic damage in exfoliated oral mucosal cells and circulating lymphocytes of patients suffering from precancerous oral lesions. *Cancer Lett* 109: 9–14.
101. Rooban T, Mishra G, Elizabeth J, Ranganathan K, Saraswathi TR (2006) Effect of habitual areca nut chewing on resting whole mouth salivary flow rate and PH. *Indian J Med Sci* 60: 95–105.
102. Liu CJ, Chen CL, Chang KW, Chu CH, Liu TY (2000) Safole in betel quid may be a risk factor for hepatocellular carcinoma: case report. *CMAJ* 162: 359–360.
103. Liu TY, Chung YT, Wang PF, Chi CW, Hsieh LL (2004) Safole-DNA adducts in human peripheral blood—an association with areca quid chewing and CYP2E1 polymorphisms. *Mutat Res* 559: 59–66.
104. Chen YJ, Chang J, Liao CT, Wang HM, Yen TC, et al. (2008) Hydroxychavicol, a phenolic component of betel leaf, has been found in human saliva at a 4.6 mM concentration after betel quid chewing. Hydroxychavicol may induce the formation of single-strand DNA breaks and 8-hydroxydeoxyguanosine – a marker of oxidative DNA damage – in cultured cells. *Cancer Sci* 99: 1507–1514.
105. Hung KF, Lai KC, Liu TY, Liu CJ, Lee TC et al (2009) Asb6 upregulation by Areca nut extracts is associated with betel quid-induced oral carcinogenesis. *Oral Oncol* 45: 543–548.
106. Wu HT, Ko SY, Fong JH, Chang KW, Liu TY, et al. (2009) Expression of phosphorylated Akt in oral carcinogenesis and its induction by nicotine and alkaline stimulation. *J Oral Pathol Med* 38: 206–213.
107. Ciceas J (2008) The potential role of Akt phosphorylation in human cancers. *Int J Biol Markers* 23: 1–9.
108. Lee H, Yin P, Yu T, Chang Y, Hsu W, et al. (2001) Accumulation of mitochondrial DNA deletions in human oral tissues — effects of betel quid chewing and oral cancer. *Mutat Res Genetic Toxicol Environ Mutagen* 493: 67–74.
109. Chiang WF, Hung PS, Liu SY, Yuan TC, Chang KW, et al. (2011) Increase of ZASC1 gene copy number in recurrent oral carcinoma. *Oral Dis* 17: 53–59.
110. Huang SH, Lee HS, Mar K, Ji DD, Huang MS, et al. (2010) Loss expression of O6-methylguanine DNA methyltransferase by promoter hypermethylation and its relationship to betel quid chewing in oral squamous cell carcinoma. *Oral Surg Oral Med Oral Pathol Oral Radiol Endo* 109: 883–889.
111. Takeshima M, Saitoh M, Kusano K, Nagayasu H, Kurashige Y, et al. (2008) High frequency of hypermethylation of p14, p15 and p16 in oral pre-cancerous lesions associated with betel-quid chewing in Sri Lanka. *J Oral Pathol Med* 37: 475–479.
112. Tran TN, Liu Y, Takagi M, Yamaguchi A, Fujii H (2005) Frequent promoter hypermethylation of RASSF1A and p16INK4a and infrequent allelic loss other than 9p21 in betel-associated oral carcinoma in a Vietnamese non-smoking/non-drinking female population. *J Oral Pathol Med* 34: 150–156.
113. Moutasim KA, Jenei V, Sapienza K, Marsh D, Weinreb PH, et al. (2011) Betel-derived alkaloid up-regulates keratinocyte alpha6beta6 integrin expression and promotes oral submucous fibrosis. *J Pathol* 223: 366–377.
114. Lee SS, Tseng LH, Li YC, Tsai CH, Chang YC (2011) Heat shock protein 47 expression in oral squamous cell carcinomas and upregulated by arecoline in human oral epithelial cells. *J Oral Pathol Med* 40: 390–396.
115. Hollstein M, Sidransky D, Vogelstein B, Harris CC (1991) P53 mutation in human cancer. *Science* 253: 49–53.
116. Harris CC, Hollstein M (1993) Clinical implications of the p53 tumor-suppressor gene. *N Engl J Med* 329: 1318–1327.
117. Goan YG, Chang HC, Hsu HK, Chou YP, Cheng JT (2005) Risk of p53 gene mutation in esophageal squamous cell carcinoma and habit of betel quid chewing in Taiwanese. *Cancer Sci* 96: 758–765.
118. Lee JJ, Kuo MY, Cheng SJ, Chiang CP, Jeng JH, et al. (2005) Higher expressions of p53 and proliferating cell nuclear antigen (PCNA) in atrophic oral lichen planus and patients with areca quid chewing. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 99: 471–478.
119. Thongsuksai P, Boonyaphiphat P, Sriplung H, Sudhikaran W (2003) p53 mutations in betel-associated oral cancer from Thailand. *Cancer Lett* 201: 1–7.
120. Ralhan R, Agarwal S, Nath N, Mathur M, Waslyk B, et al. (2001) Correlation between p53 gene mutations and circulating antibodies in betel- and tobacco-consuming North Indian population. *Oral Oncol* 37: 243–250.
121. Hsieh LL, Wang PF, Chen Ich, Liao CT, Wang HM, et al. (2001) Characteristics of mutations of the p53 gene in oral squamous cell carcinoma associated with betel quid chewing and cigarette smoking in Taiwanese. *Carcinogenesis* 22: 1497–1503.
122. Kannan K, Munirajan AK, Krishnamurthy J (1999) Low incidence of p53 mutation in betel quid and tobacco chewing-associated oral squamous carcinoma from India. *Int J Oncol* 15: 1133–1136.
123. Chiba I, Muthumala M, Yamazaki Y, Zaman AU, Iizuka T, et al. (1998) Characteristics of mutations in the p53 gene of oral squamous-cell carcinomas associated with betel-quid chewing in Sri Lanka. *Int J Cancer* 77: 839–842.
124. Wong YK, Liu TY, Chang KW, Lin SC, Chao TW, et al. (1998) p53 alterations in betel quid- and tobacco-associated oral squamous cell carcinomas from Taiwan. *J Oral Pathol Med* 27: 243–248.
125. Kuttan NAA, Rosin MP, Ambika K, Priddy RW, Bhakthan NMG, et al. (1995) High prevalence of expression of p53 oncoprotein in oral carcinomas from India associated with betel and tobacco chewing. *Eur J Cancer* 31: 169–173.

126. Thomas S, Brennan J, Martel G (1994) Mutations in the conserved regions of p53 are infrequent in betel-associated oral cancers from Papua New Guinea. *Cancer Res* 54: 3588–3593.
127. Ranasinghe AW, Warnakulasuriya KAAS, Johnson NW (1993) Low prevalence of expression of p53 oncoprotein in oral carcinomas from Sri Lanka associated with betel and tobacco chewing. *Eur J Cancer B Oral Oncol* 29: 147–150.
128. Ihsan R, Devi TR, Yadav DS, Mishra AK, Sharma J, et al. (2011) Investigation on the role of p53 codon 72 polymorphism and interactions with tobacco, betel quid, and alcohol in susceptibility to cancers in a high-risk population from North East India. *DNA Cell Biol* 30: 163–171.
129. Kaushal M, Chattopadhyay I, Phukan R, Purkayastha J, Mahanta J et al. (2010) Contribution of germ line BRCA2 sequence alterations to risk of familial esophageal cancer in a high-risk area of India. *Dis Esophagus* 23: 71–75.
130. Choudhury Y, Sharan RN (2009) Altered p53 response and enhanced transgenerational transmission of carcinogenic risk upon exposure of mice to betel nut. *Environ Toxicol Pharmacol* 31: 57–69.
131. Choudhury Y, Sharan RN (2010a) Altered BRCA1 and BRCA2 responses and mutation of BRCA1 gene in mice exposed chronically and transgenerationally to aqueous extract of betel nut (AEBN). *Environ Toxicol Pharmacol* 27: 127–138.
132. Choudhury Y, Sharan RN (2010b) Ultrastructural alterations in liver of mice exposed chronically and transgenerationally to aqueous extract of betel nut: Implications in betel-nut induced carcinogenesis. *Microsc Res Tech* 73: 530–539.
133. Shwe M, Chiguchi G, Yamada S, Nakajima T, Maung KK, et al. (2001) P53 and MDM2 co-expression in tobacco and betel-chewing associated oral squamous cell carcinomas. *J Med Dent Sci* 48: 113–119.
134. Huang JS, Ho TJ, Chiang CP, Kok SH, Kuo YS, et al. (2001) MDM2 expression in areca quid chewing-associated oral squamous cell carcinomas in Taiwan. *J Oral Pathol Med* 30: 53–58.
135. Manfredi JJ (2010) The Mdm2-p53 relationship evolves: Mdm2 swings both ways as an oncogene and a tumor suppressor. *Gene Dev* 24: 1580–1589.
136. Chung KT, Wong TY, Wei CI, Huang YW, Lin Y (1998) Tannins and human health: a review. *Crit Rev Food Sci Nutr* 38: 421–464.
137. Spitz MR, Bondy ML (1993) Genetic susceptibility to cancer. *Cancer* 72: 991–995.
138. Bartsch H, Hietanen E (1996) The role of individual susceptibility in cancer burden related to environmental exposure. *Environ Health Perspect* 104: 569–577.
139. Hsieh CH, Chang JW, Hsieh JJ, Hsu T, Huang SF, et al. (2011) Epidermal growth factor receptor mutations in patients with oral cavity cancer in a betel nut chewing-prevalent area. *Head Neck*, doi: 10.1002/hed.21665.
140. Lin C-W, Hsieh Y-S, Hsin C-H, Su C-W, Lin C-H, et al. (2012) Effects of *NFKB1* and *NFKBIA* Gene Polymorphisms on Susceptibility to Environmental Factors and the Clinicopathologic Development of Oral Cancer. *PLoS ONE* 7(4): e35078. doi:10.1371/journal.pone.0035078.
141. Weng CJ, Hsieh YH, Chen MK, Tsai CM, Lin CW, et al. (2012) Survivin SNP-carcinogen interactions in oral cancer. *J Dent Res* 91: 358–363.
142. Chen MK, Yah KT, Chiou HL, Lin CW, Chung TT, et al. (2011) CCR2-641 gene polymorphism increase susceptibility to oral cancer. *Oral Oncol* 47: 577–582.
143. Chen PH, Lee KW, Chen CH, Shieh TY, Ho PS, et al. (2011) CYP26B1 is a novel candidate gene for betel quid-related oral squamous cell carcinoma. *Oral Oncol* 47: 594–600.
144. Weng CJ, Lin CW, Chung TT, Tsai CM, Chen MK (2011) Impact of uPA system gene polymorphisms on the susceptibility of environmental factors to carcinogenesis and the development of clinicopathology of oral cancer. *Ann Surg Oncol* 18: 805–812.
145. Yang CM, Hou YY, Chiu YT, Chen HC, Chu ST, et al. (2011) Interaction between tumour necrosis factor- $\alpha$  gene polymorphisms and substance use on risk of betel quid-related oral and pharyngeal squamous cell carcinoma in Taiwan. *Arch Oral Biol* 56: 1162–1169.
146. Zavras A, Yoon AJ, Chen MK, Lin CW, Yang SF (2011) Metallothionein-I genotypes in the risk of oral squamous cell carcinoma. *Ann Surg Oncol* 18: 1478–1483.
147. Hou YY, Ou HL, Chu ST, Wu PC, Lu PJ, et al. (2011) NAT2 slow acetylation haplotypes are associated with the increased risk of betel quid-related oral and pharyngeal squamous cell carcinoma. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 112: 484–492.
148. Yadav DS, Devi TR, Ihsan R, Mishra AK, Kaushal M, et al. (2010) Polymorphisms of glutathione-S-transferase genes and the risk of aerodigestive tract cancers in the Northeast Indian population. *Genet Test Mol Biomarkers* 14: 715–723.
149. Ihsan R, Chattopadhyay I, Phukan R, Mishra AK, Purkayastha J, et al. (2010) Role of epoxide hydrolase 1 gene polymorphisms in esophageal cancer in a high-risk area in India. *J Gastroenterol Hepatol* 25: 1456–1462.
150. Tsou YA, Hua CH, Tseng HC, Hsu CF, Tsai CW, et al. (2010) The joint effect of hOGG1 single nucleotide polymorphism and betel quid chewing on oral cancer in Taiwan. *Anticancer Res* 30: 4205–4208.
151. Shieh TM, Tu HF, Ku TH, Chang SS, Chang KW, et al. (2009) Association between lysyl oxidase polymorphisms and oral submucous fibrosis in older male areca chewers. *J Oral Pathol Med* 38: 109–113.
152. Huang SF, Chen IH, Liao CT, Wang HM, Liou SH, et al. (2009) Combined effects of MDM2 SNP 309 and p53 mutation on oral squamous cell carcinomas associated with areca quid chewing. *Oral Oncol* 45: 16–22.
153. Chiu CF, Tsai MH, Tseng HC, Wang CL, Wang CH, et al. (2008) A novel single nucleotide polymorphism in XRCC4 gene is associated with oral cancer susceptibility in Taiwanese patients. *Oral Oncol* 44: 898–902.
154. Lin YC, Huang HI, Wang LH, Tsai CC, Lung O, et al. (2008) Polymorphisms of COX-2 -765G>C and p53 codon 72 and risks of oral squamous cell carcinoma in a Taiwan population. *Oral Oncol* 44: 798–804.
155. Tu HF, Wu CH, Kao SY, Liu CJ, Liu TY, et al. (2007) Functional -1562 C-to-T polymorphism in matrix metalloproteinase-9 (MMP-9) promoter is associated with the risk for oral squamous cell carcinoma in younger male areca users. *J Oral Pathol Med* 36: 409–414.
156. Tu HF, Liu CJ, Chang CS, Lui MT, Kao SY, et al. (2006) The functional (-1171 5A->6A) polymorphisms of matrix metalloproteinase 3 gene as a risk factor for oral submucous fibrosis among male areca users. *J Oral Pathol Med* 35: 99–103.
157. Lin SC, Liu CJ, Yeh WI, Lui MT, Chang KW, et al. (2006) Functional polymorphism in NFKB1 promoter is related to risks of oral squamous cell carcinoma occurring on older male areca (betel) chewers. *Cancer Lett* 243: 47–54.
158. Ramachandran S, Ramadas K, Hariharan R, Rejnish KR, Radhakrishna PM (2006) Single nucleotide polymorphisms of DNA repair genes XRCC1 and XPD and its molecular mapping in Indian oral cancer. *Oral Oncol* 42: 350–362.
159. Chang KW, Lee TC, Yeh WI, Chung MY, Liu CJ, et al. (2004) Polymorphism in heme oxygenase-1 (HO-1) promoter is related to the risk of oral squamous cell carcinoma occurring on male areca chewers. *Br J Cancer* 18: 1551–1555.
160. Topcu Z, Chiba I, Fujieda M, Shibata T, Ariyoshi N, et al. (2002) CYP2A6 gene deletion reduces oral cancer risk in betel quid chewers in Sri Lanka. *Carcinogenesis* 23: 595–598.
161. Kao SY, Wu HC, Lin SC, Yap SK, Chang CS, et al. (2002) Genetic polymorphism of cytochrome P4501A1 and susceptibility to oral squamous cell carcinoma and oral precancer lesions associated with smoking/betel use. *J Oral Pathol Med* 31: 505–511.
162. Chiu CJ, Chang CP, Hahn LJ, Hsieh LL, Chen CJ (2002) Interaction of collagen-related genes and susceptibility to betel quid-induced oral submucous fibrosis. *Cancer Epidemiol Biomarkers Prev* 11: 646–653.
163. Chiu CJ, Chiang CP, Chang ML, Chen HM, Hahn LJ, et al. (2001) Association between genetic polymorphism of tumor necrosis factor- $\alpha$  and risk of oral submucous fibrosis, a pre-cancerous condition of oral cancer. *J Dent Res* 80: 2055–2059.
164. Nair UJ, Nair J, Mathew B, Bartsch H (1999) Glutathione S-transferase M1 and T1 null genotypes as risk factors for oral leukoplakia in ethnic Indian betel quid/tobacco chewers. *Carcinogenesis* 20: 743–748.
165. Kietthubthwe S, Sriplung H, Au WW (2001) Genetic and environmental interactions on oral cancer in Southern Thailand. *Environ Mol Mutagen* 37: 111–116.
166. Tang DW, Lin SC, Chang KW, Chi CW, Chang CS, et al. (2003) Elevated expression of cyclooxygenase (COX)-2 in oral squamous cell carcinoma—evidence for COX-2 induction by areca quid ingredients in oral keratinocytes. *J Oral Pathol Med* 32: 522–529.
167. Tsai CH, Chou MY, Chang YC (2003) The upregulation of cyclooxygenase-2 expression in human buccal mucosal fibroblasts by arecoline: A possible role in the pathogenesis of oral submucous fibrosis. *J Oral Pathol Med* 32: 146–153.
168. Jeng JH, Ho YS, Chan CP, Wang YJ, Hahn LJ, et al. (2000) Areca nut extract up-regulates prostaglandin production, cyclooxygenase-2 mRNA and protein expression of human oral keratinocytes. *Carcinogenesis* 21: 1365–1370.
169. Yang CY, Meng CL, van der Bijl P, Lee HK (2002) The effect of betel nut extract on cell growth and prostaglandin endoperoxide synthase in human epidermoid carcinoma cells. *Prostaglandin Other Lipid Mediat* 67: 181–195.
170. Tseng YH, Yang CC, Lin SC, Cheng CC, Lin SH, et al. (2011) Areca nut extract upregulates vimentin by activating PI3K/AKT signaling in oral carcinoma. *J Oral Pathol Med* 40: 160–166.
171. Tseng YH, Chang KW, Yang CC, Liu CJ, Kao SY, et al. (2012) Association between areca-stimulated vimentin expression and the progression of head and neck cancers. *Head Neck* 34: 245–253.
172. Tseng YH, Chang CS, Liu TY, Kao SY, Chang KW, et al. (2007) Areca nut extract treatment down-regulates involucrin in normal human oral keratinocyte through PI3K/AKT activation. *Oral Oncol* 43: 670–679.
173. Liu SY, Lin MH, Hsu YR, Shih YY, Chiang WF et al. (2008) Arecoline and the 30–100 kDa fraction of areca nut extract differentially regulate mTOR and respectively induce apoptosis and autophagy: a pilot study. *J Biomed Sci* 15: 823–831.
174. Lu HH, Kao SY, Liu TY, Liu ST, Huang WP, et al. (2010) Areca nut extract induced oxidative stress and upregulated hypoxia inducing factor leading to autophagy in oral cancer cells. *Autophagy* 6: 725–737.
175. Merchant A, Husain SSM, Hosain M, Fikree FF, Pritiphat W, et al. (2000) *Paan* without tobacco: An independent risk factor for oral cancer. *Int J Cancer* 86: 128–131.
176. Shiu MN, Chen THH, Chang SH, Hahn LJ (2000) Risk factors for leukoplakia and malignant transformation to oral carcinoma: A leukoplakia cohort in Taiwan. *Br J Cancer* 82: 1871–1874.

177. Phukan RK, Ali MS, Chetia CK, Mahanta J (2001) Betel nut and tobacco chewing potential risk factors of cancer of oesophagus in Assam, India. *Br J Cancer* 85: 661–667.
178. Chang IH, Jiang RS, Wong YK, Wu SH, Chen FJ (2011) Visual screening of oral cavity cancer in male population: experience from a medical center. *J Chin Med Assoc* 74: 561–566.