

Biology 4361

# Early Development and Axis Formation in Amphibians

July 9, 2009

# The Amphibian Model

Common vertebrate (e.g. *Rana*, *Xenopus*)

Manipulable, observable (i.e. large eggs and embryos)

- 1) How are the body axes established?
  - How do relatively homogeneous cells (i.e. zygotes) establish polarity (complexity from simplicity)?
- 2) How are the germ layers determined?
  - How do zygotes send their cellular progeny (i.e. blastomeres) down pathways toward different fates?
- 3) How is development organized in a regulative system?

# Amphibian Development - Overview

Fertilization, cortical rotation

Cleavage

Gastrulation

Axis and germ layer determination

The “Organizer”

Inductions:

Mesoderm

Dorsal/organizer

Ectoderm

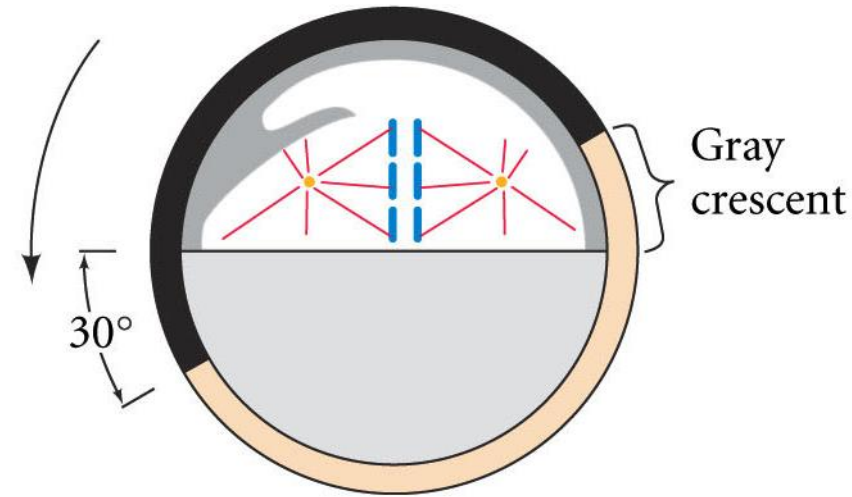
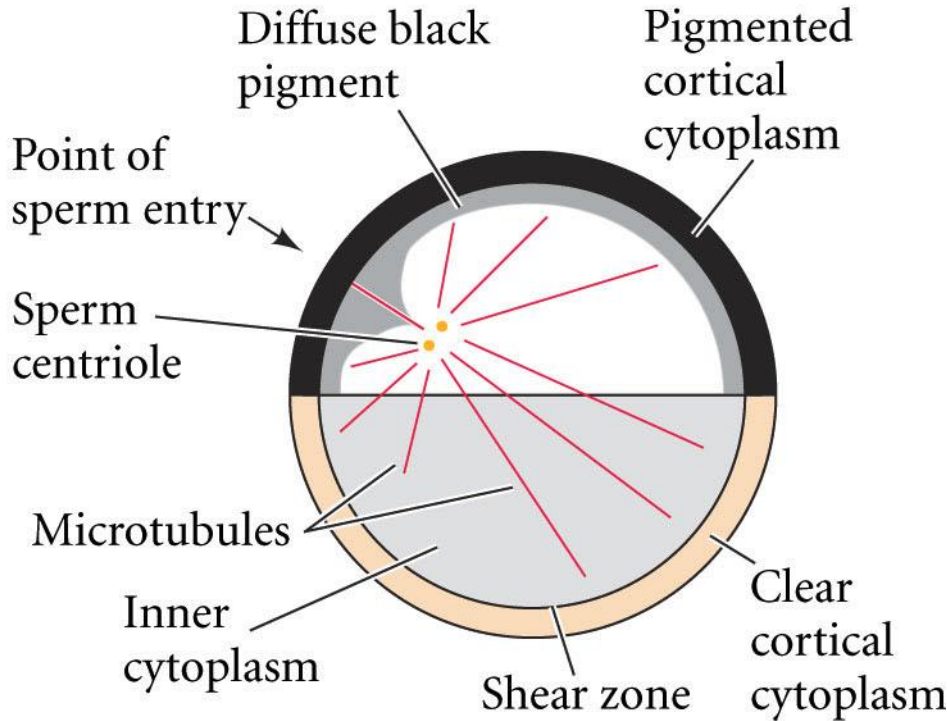
Axes Summary

Left – Right Asymmetry



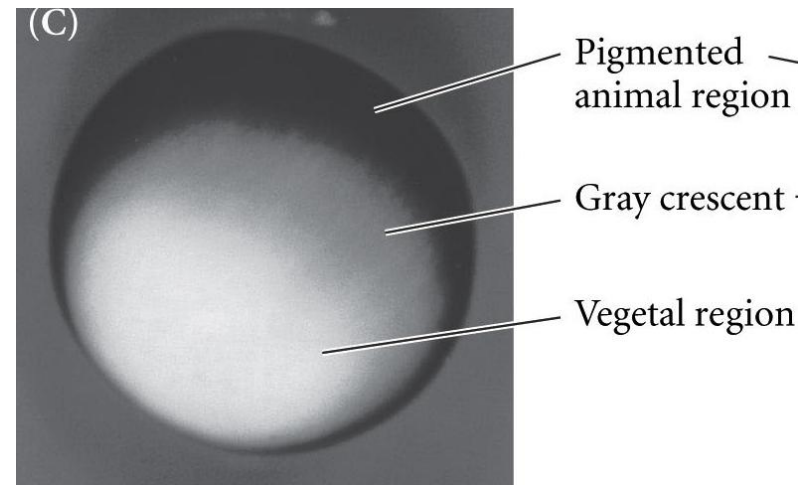
*(Photo by Harland lab/UC Berkeley)*

# Cortical Rotation



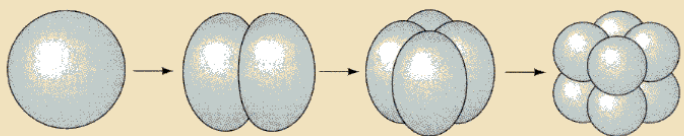
Fertilization – animal hemisphere

Microtubular network originates at fertilization point



## Holoblastic (complete cleavage)

(VAGE)

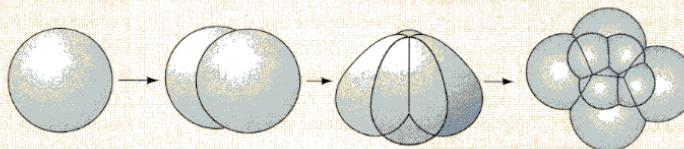


### Species

echinoderms,  
amphioxus

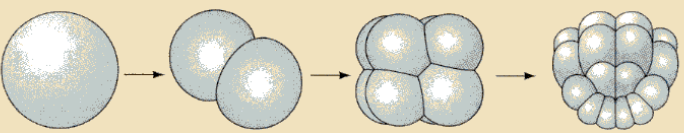
### Cleavage

Radial



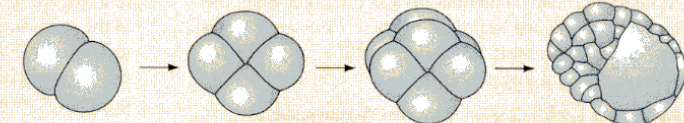
annelids, molluscs,  
flatworms

Spiral



tunicates

Bilateral



mammals, nematode

Rotational

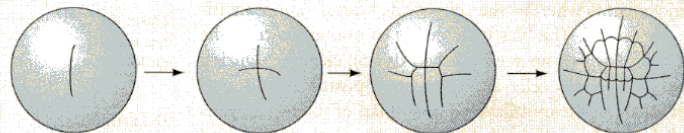
Yolk classification

Isolecithal

Mesolecithal

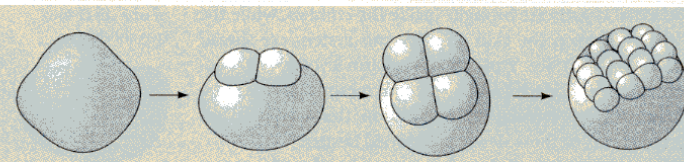
## Meroblastic (incomplete cleavage)

ii)



cephalopod molluscs

Bilateral

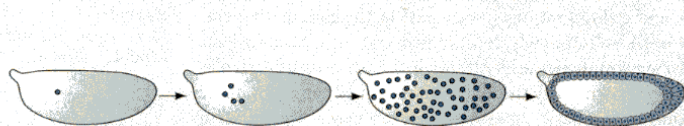


fish, reptiles, birds

Discoidal

Telolecithal

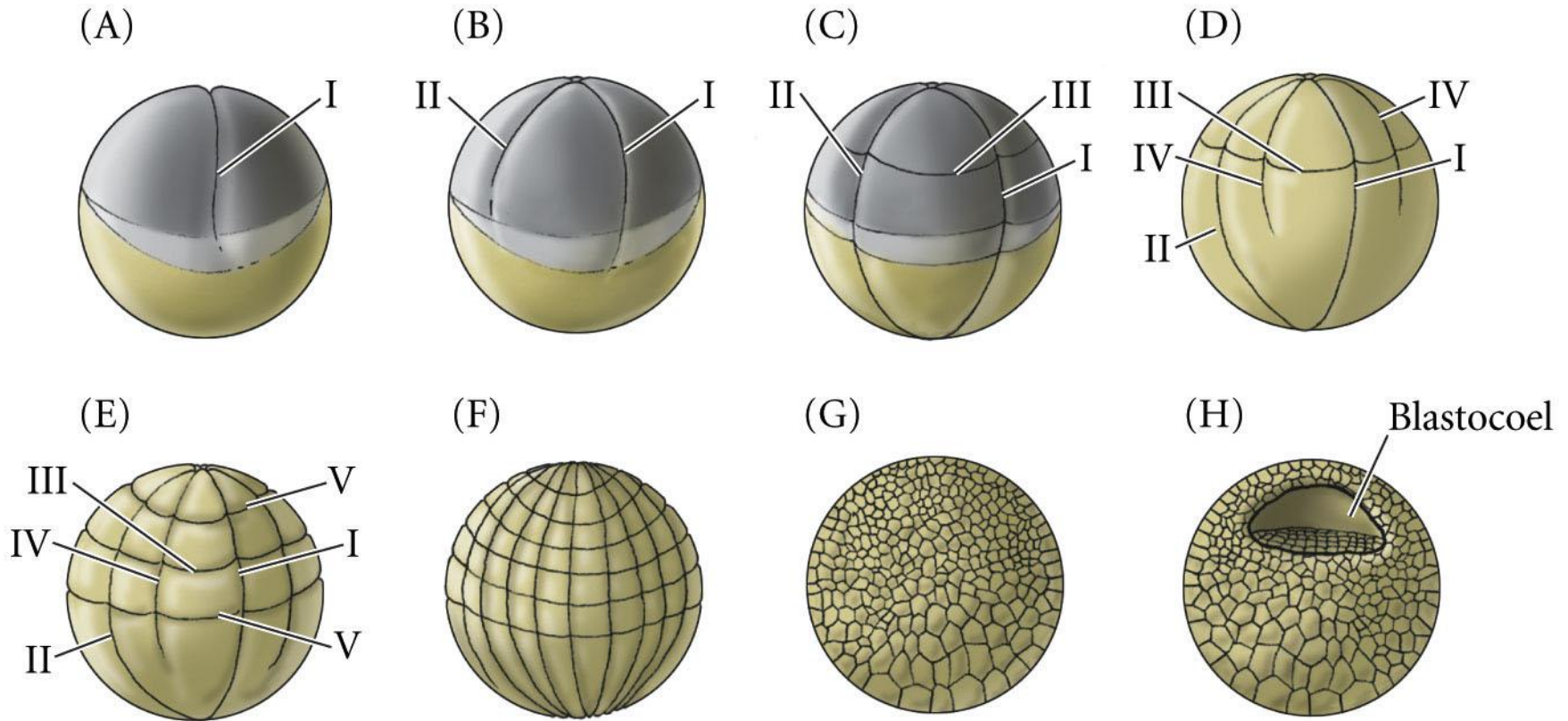
Centrolethical



most insects

Superficial

# Unequal Radial Holoblastic Cleavage



Cell cycles regulated by mitosis-promoting factor (MPF)

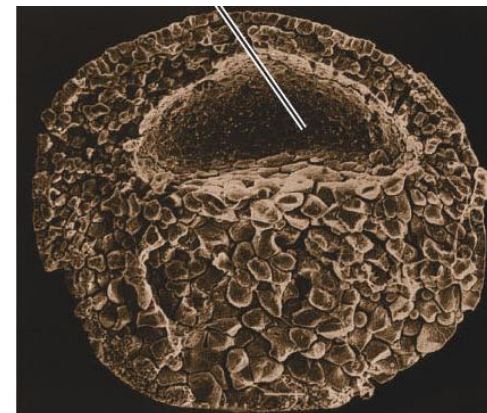
- no G phases

... until **mid-blastula transition**

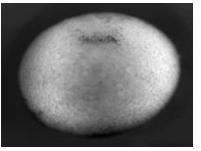
- promoters demethylated (i.e. de-repressed)

- transcription factors formed in vegetal cytoplasm

- embryonic control of development

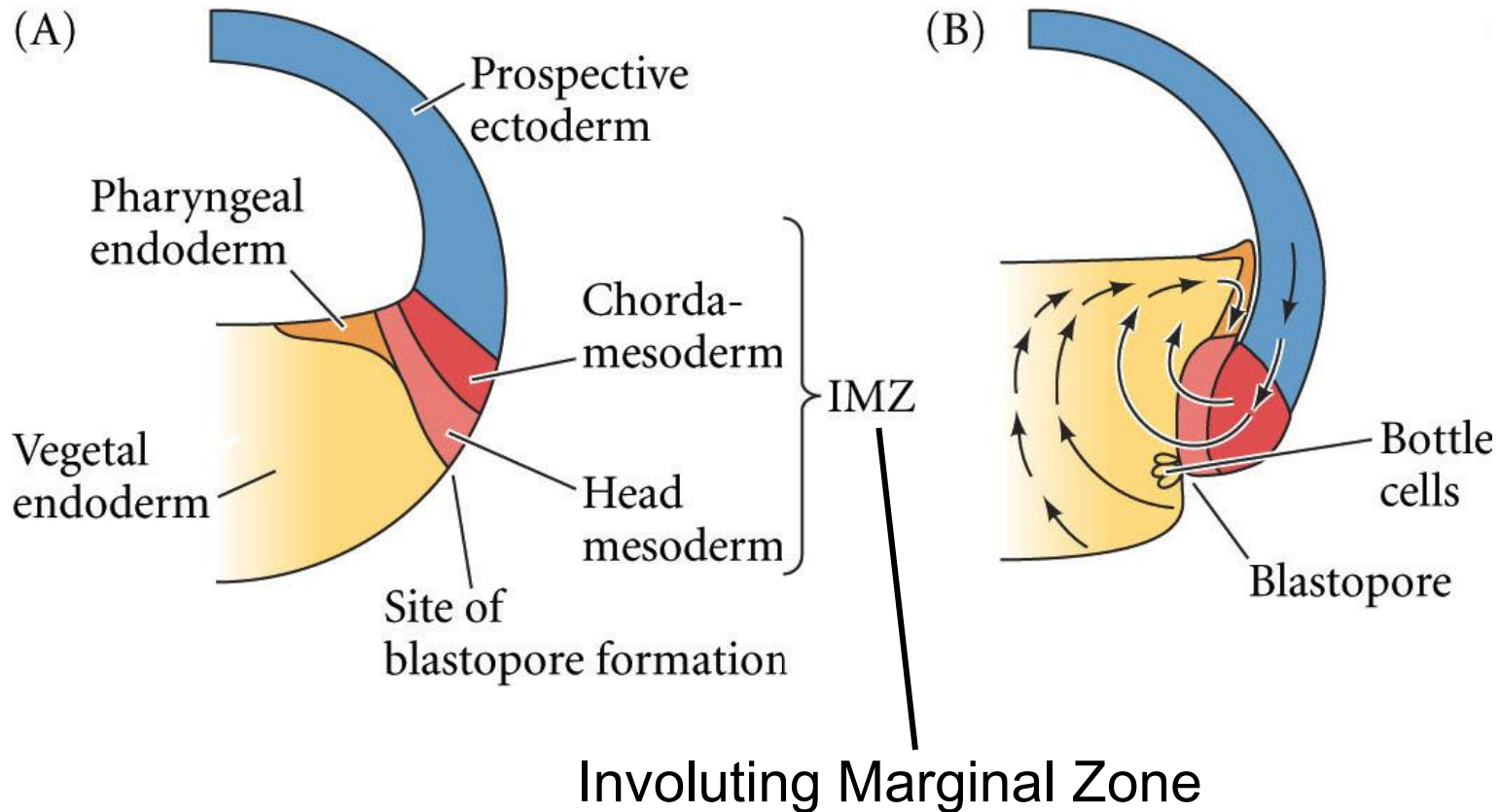


# Mechanics of Gastrulation

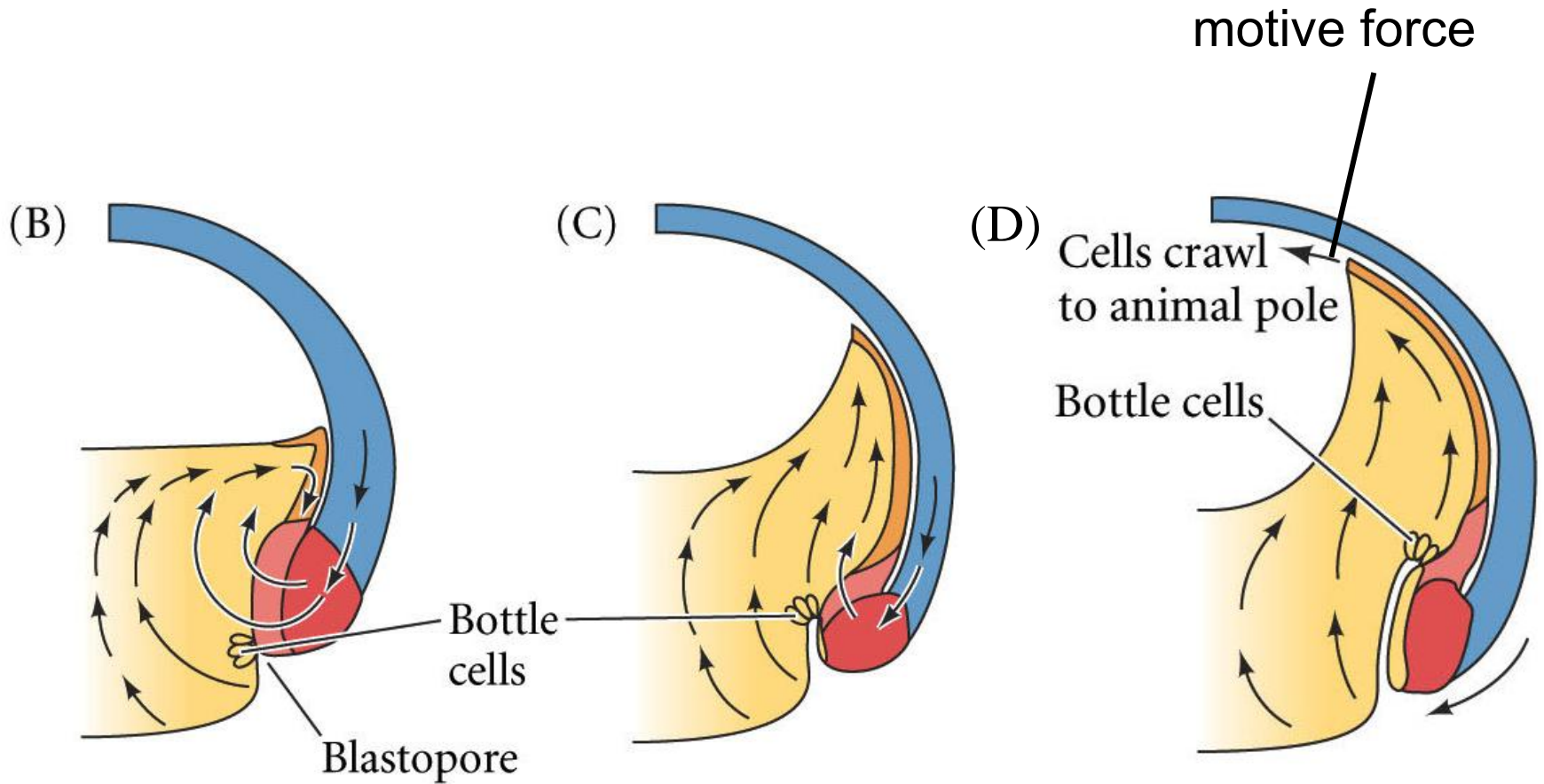
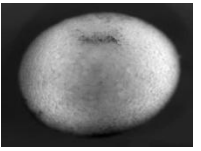


## Formation of the dorsal lip

- vegetal rotation
- invagination of bottle cells
- involution of marginal zone cells

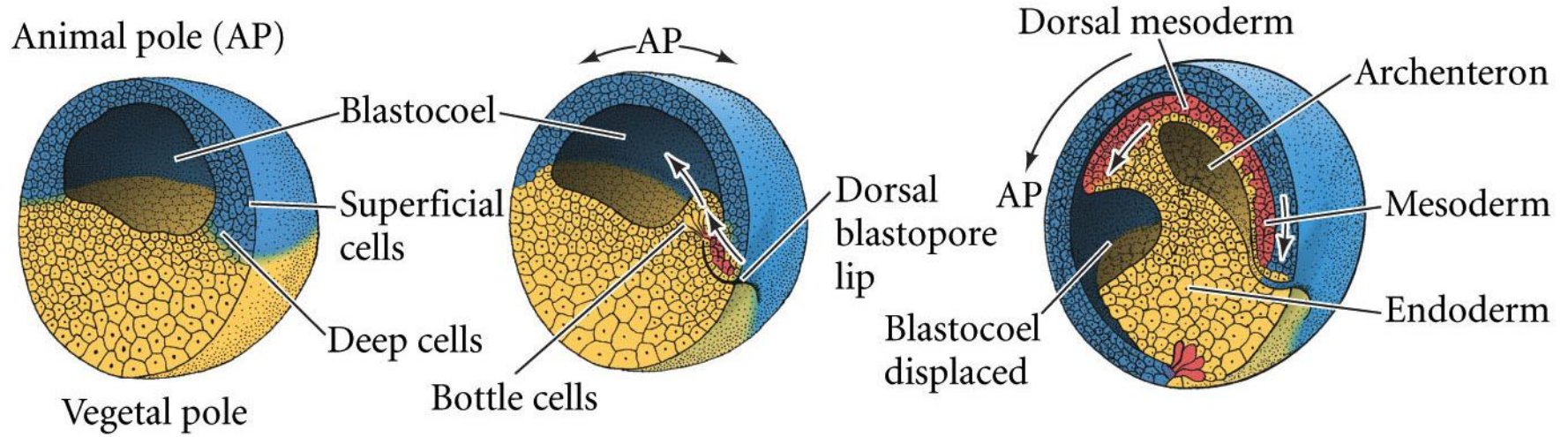


# Mechanics of Gastrulation - 2

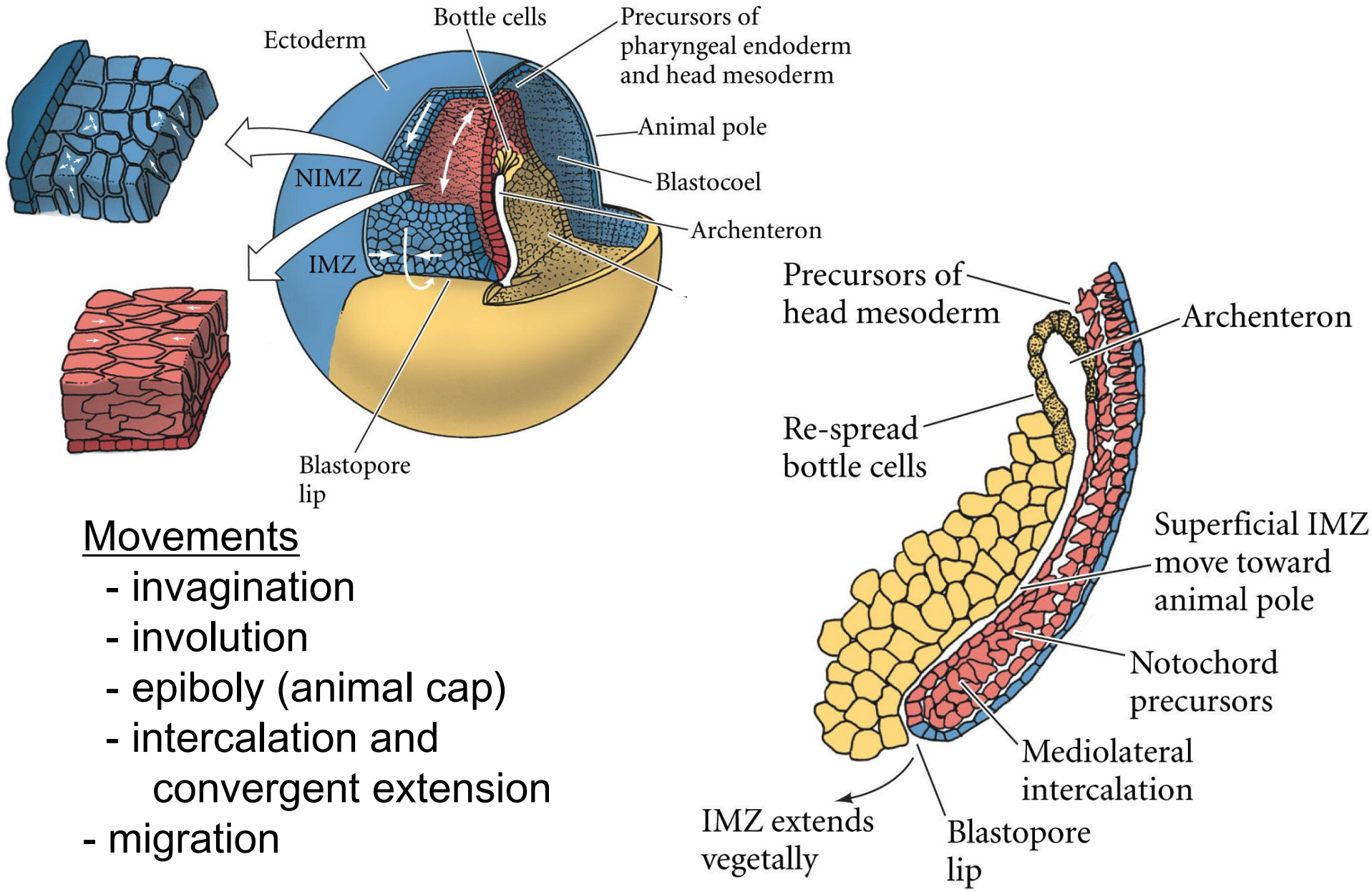




# Xenopus Gastrulation



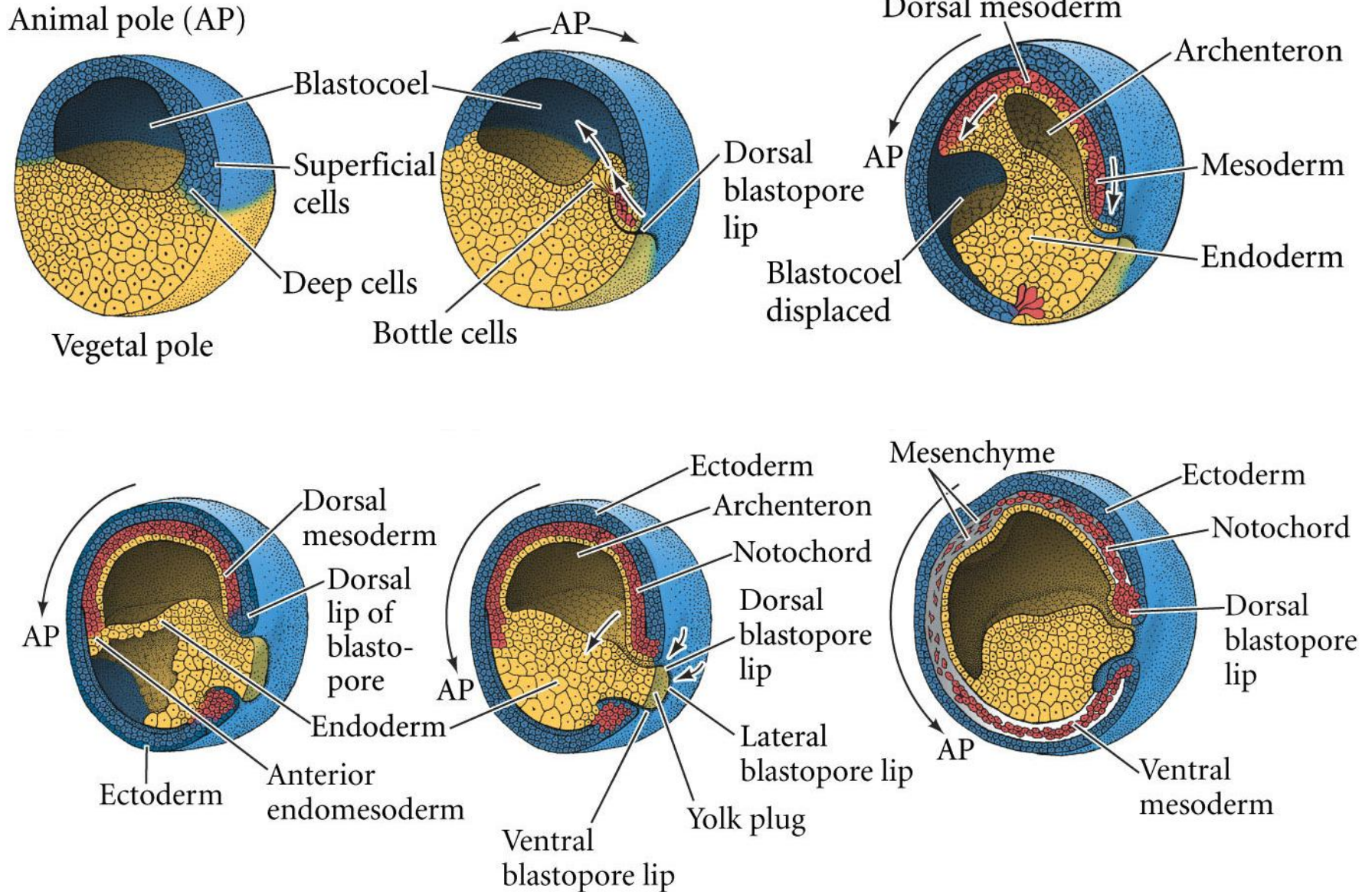
# Cell Movements during Gastrulation



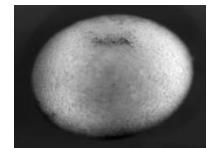
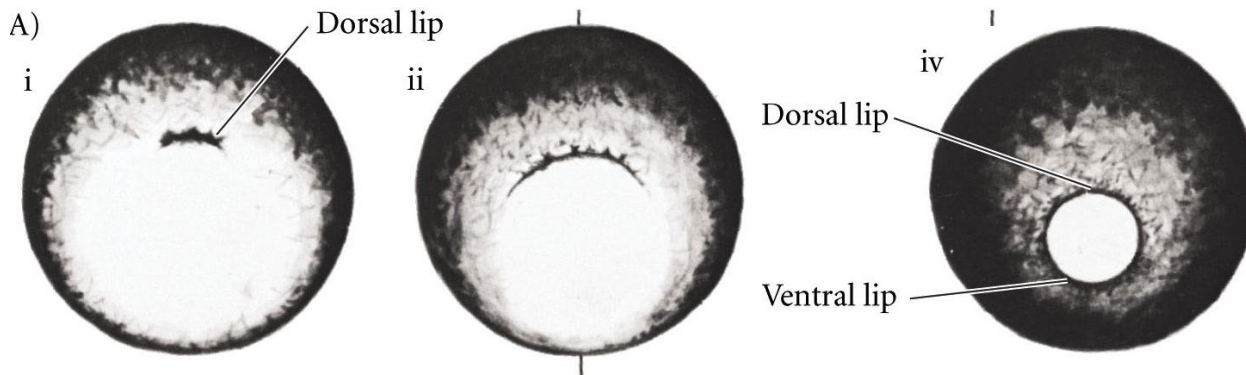
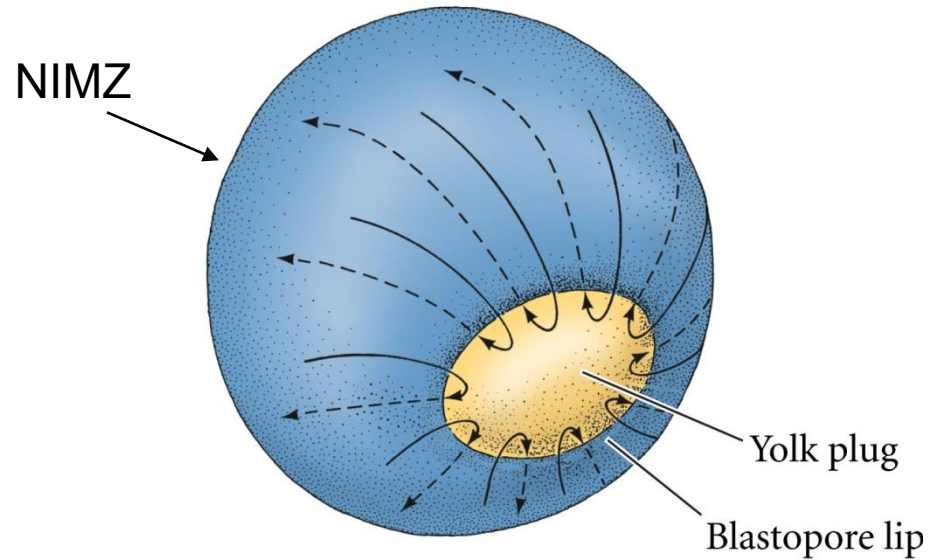
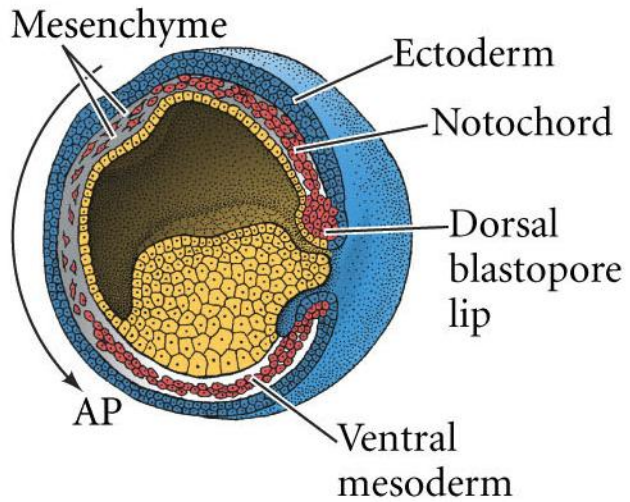
## Movements

- invagination
- involution
- epiboly (animal cap)
- intercalation and convergent extension
- migration

# Xenopus Gastrulation



# Xenopus Gastrulation - Blastopore



# Amphibian Development - Overview

Fertilization, cortical rotation

Cleavage

Gastrulation

Axis and germ layer determination

The “Organizer”

Inductions:

Mesoderm

Dorsal/organizer

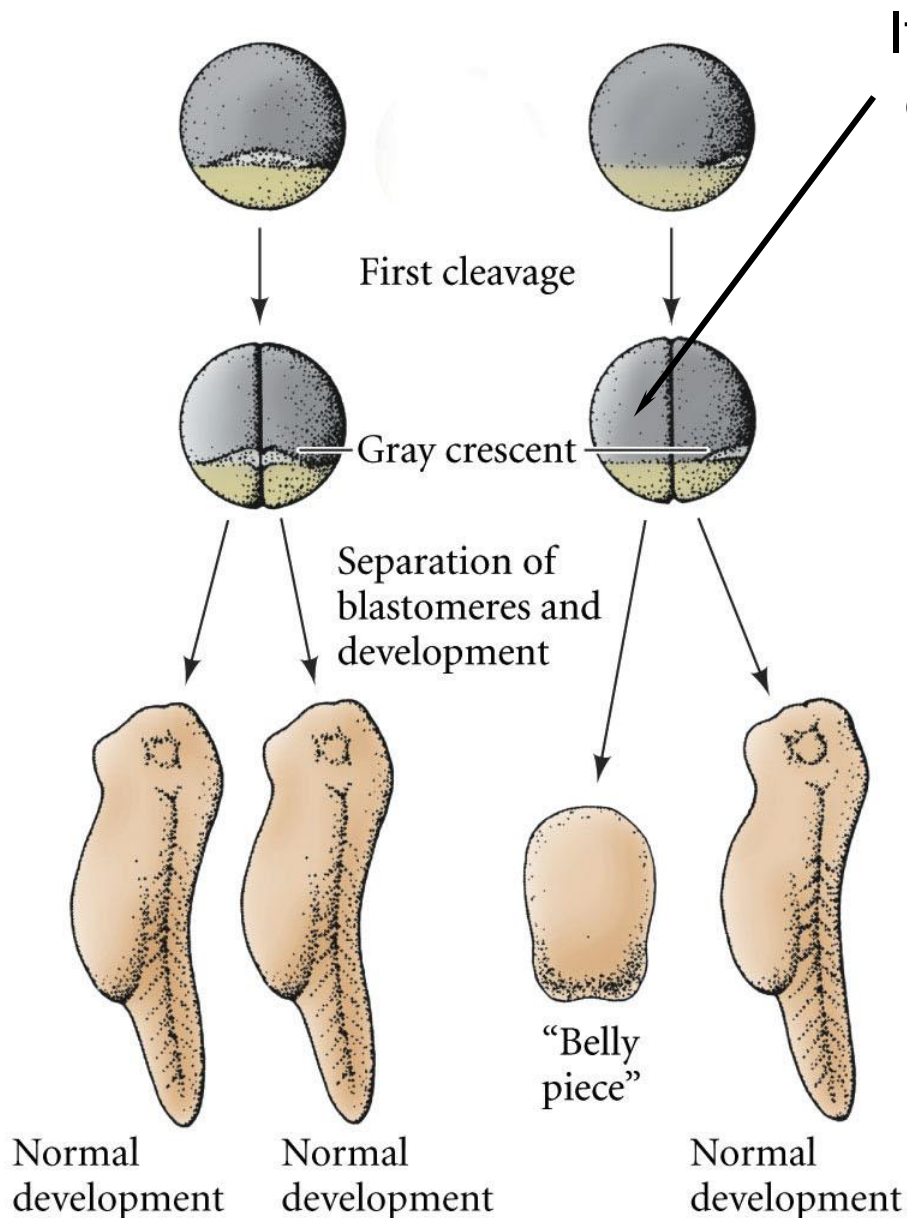
Ectoderm

Axes Summary



*(Photo by Harland lab/UC Berkeley)*

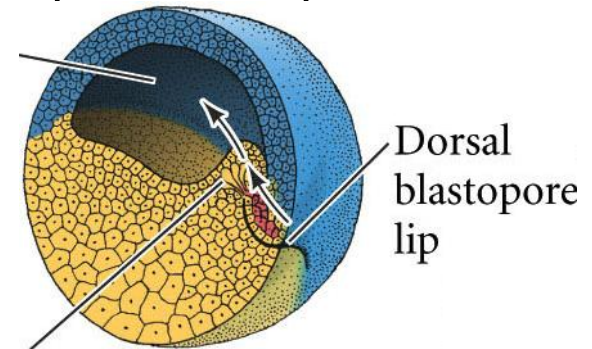
# Determination of Amphibian Axes



If one blastomere received no gray crescent material, resulted in

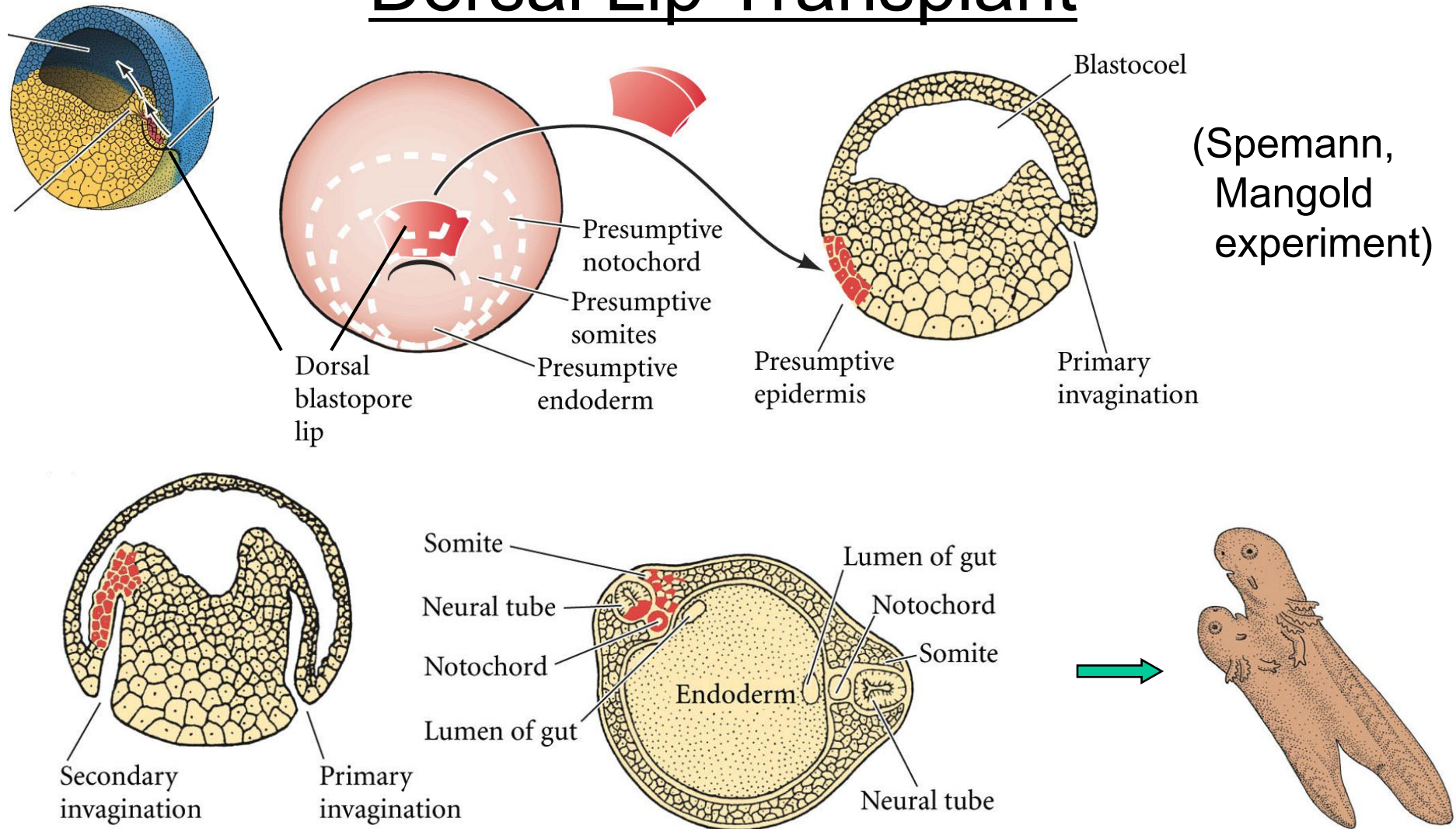
- "belly piece" – blood, mesenchyme, gut cells
- *no dorsal structures* (e.g. notochord, somites)

The gray crescent area is critical for proper development



Gray crescent = future dorsal lip of the blastopore

# Dorsal Lip Transplant



**Dorsal lip = "Organizer"**

- organizes secondary D-V axis
- induced ventral cells to change fates

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*(Photo by Harland lab/UC Berkeley)*



# The Organizer

Transplantation experiments established the *organizing properties* of the **dorsal blastopore lip**, which ...

- 1) Self-differentiates (all other tissues conditionally specified)
- 2) Establishes the dorsal-ventral axis
- 3) Specifies multiple tissues, including...
  - dorsal mesoderm, which includes...
    - head mesoderm (prechordal plate)
    - chordamesoderm (notochord)
- 4) Dorsalizes surrounding mesoderm into paraxial mesoderm
- 5) Induces the neural tube
- 6) Initiates the movements of gastrulation

**How is the dorsal lip specified?**

# Dorsal Signal: $\beta$ -Catenin

## $\beta$ -catenin

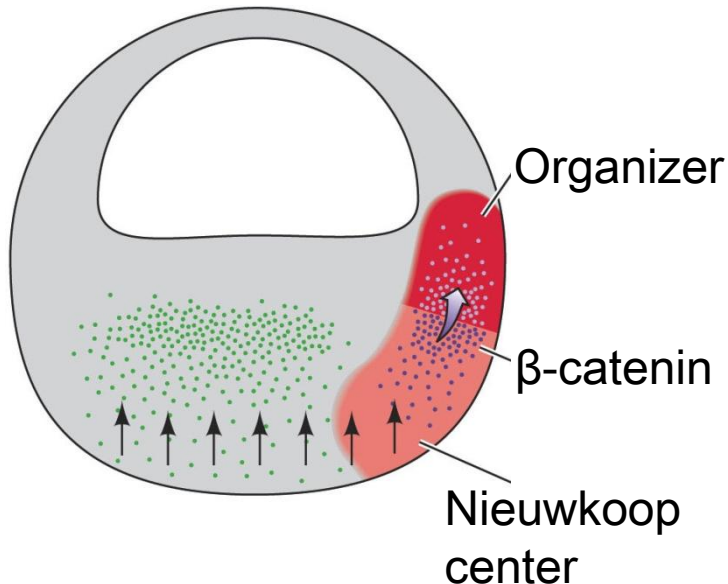
nuclear transcription factor (in Wnt pathway)

(in sea urchins, specifies micromeres, endomesoderm)

(in *Xenopus*, specifies dorsal structures; e.g. organizer)

$\beta$ -catenin is initially distributed throughout the embryo,

- accumulates only in prospective dorsal cells.
- concentrated in the Nieuwkoop center and organizer



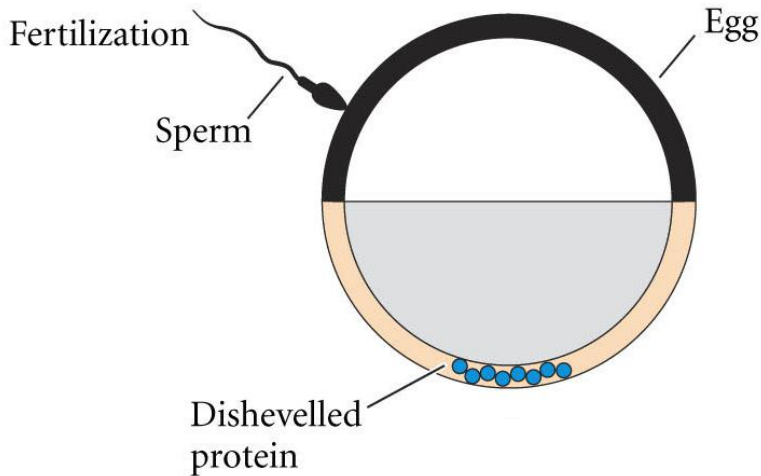
Dorsalization of  $\beta$ -catenin:

- a) protect  $\beta$ -catenin in dorsal area,
- b) degrade  $\beta$ -catenin everywhere else.

Mechanism - cortical rotation ...

# Disheveled/ $\beta$ -Catenin/Cortical Rotation

*$\beta$ -catenin induces cells to dorsal fates*



$\beta$ -catenin is initially distributed throughout the oocyte

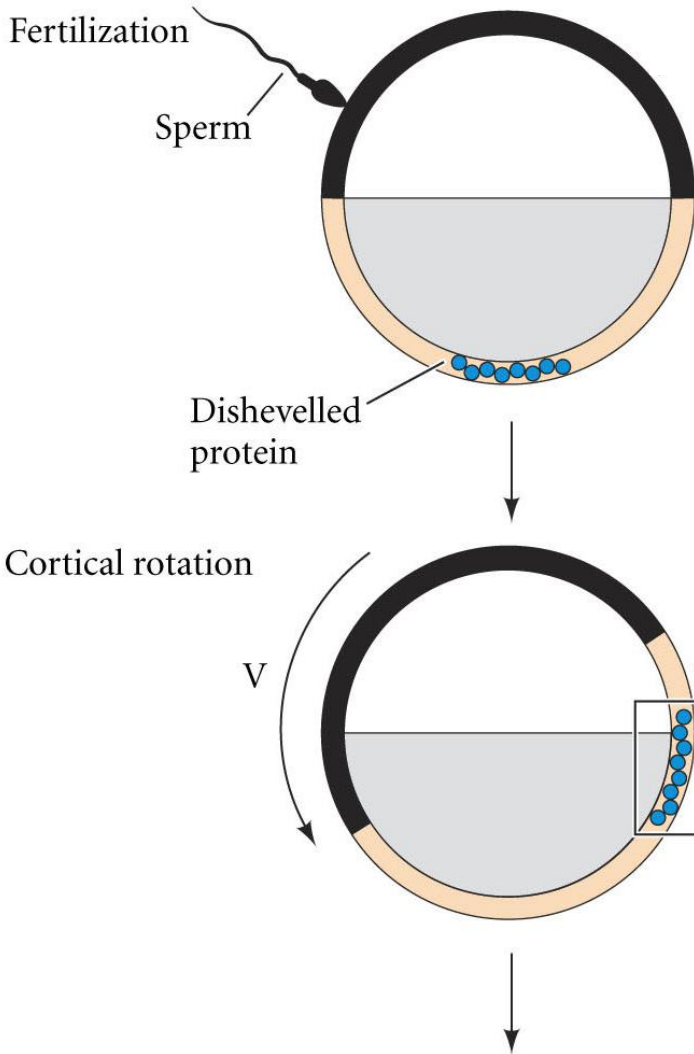
**Glycogen synthase kinase 3 (GSK3)** is also distributed throughout oocyte

GSK3 marks  $\beta$ -catenin for degradation

**Dishevelled (Dsh)** blocks GSK3 activity

Dsh localizes in the cytoplasmic cortex at the vegetal pole

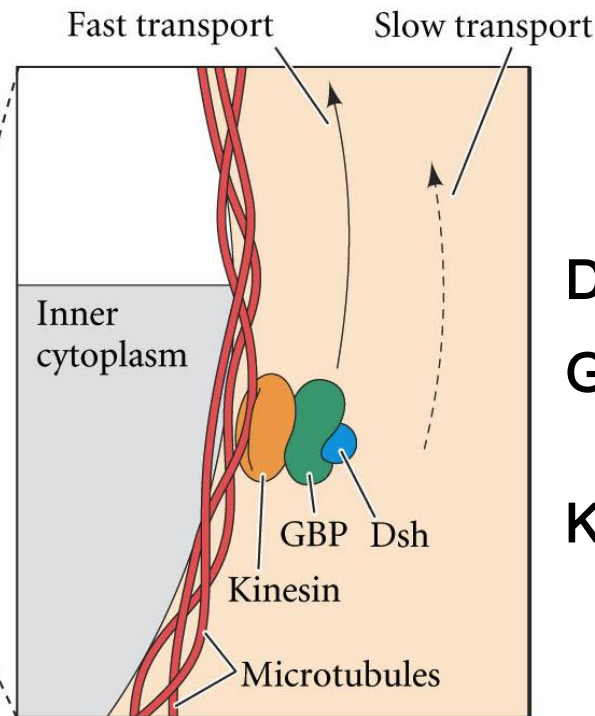
# Dishevelled/ $\beta$ -Catenin/Cortical Rotation



$\beta$ -catenin, GSK3 distributed throughout

**Dishevelled** - blocks GSK3-mediated  $\beta$ -catenin degradation

- anywhere Dsh exists,  $\beta$ -catenin survives



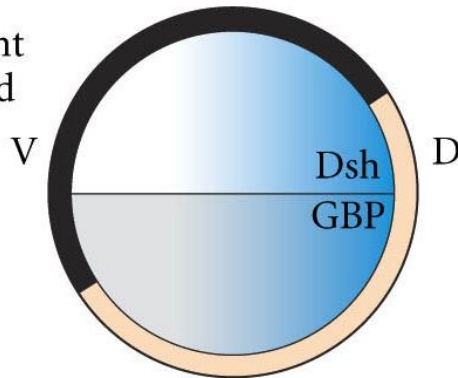
**Dsh** - Dishevelled

**GBP** - GSK3 binding protein

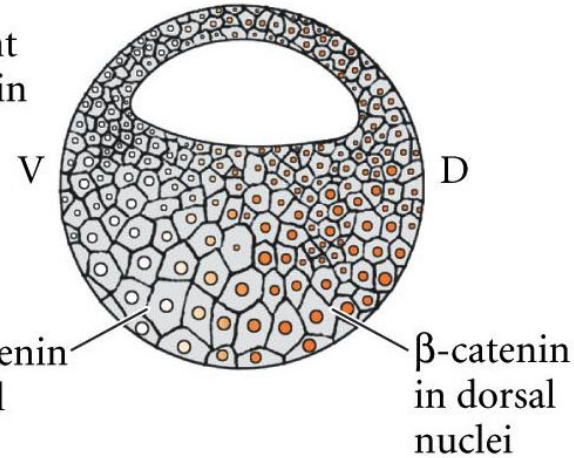
**Kinesin** - motor protein

# Disheveled/GSK3/ $\beta$ -Catenin

Dorsal enrichment of Dsh and GBP

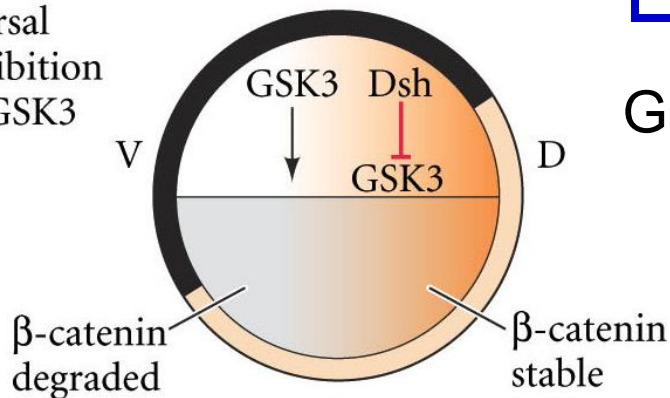


Dorsal enrichment of  $\beta$ -catenin



At the blastula stage,  $\beta$ -catenin is located exclusively in the future dorsal region

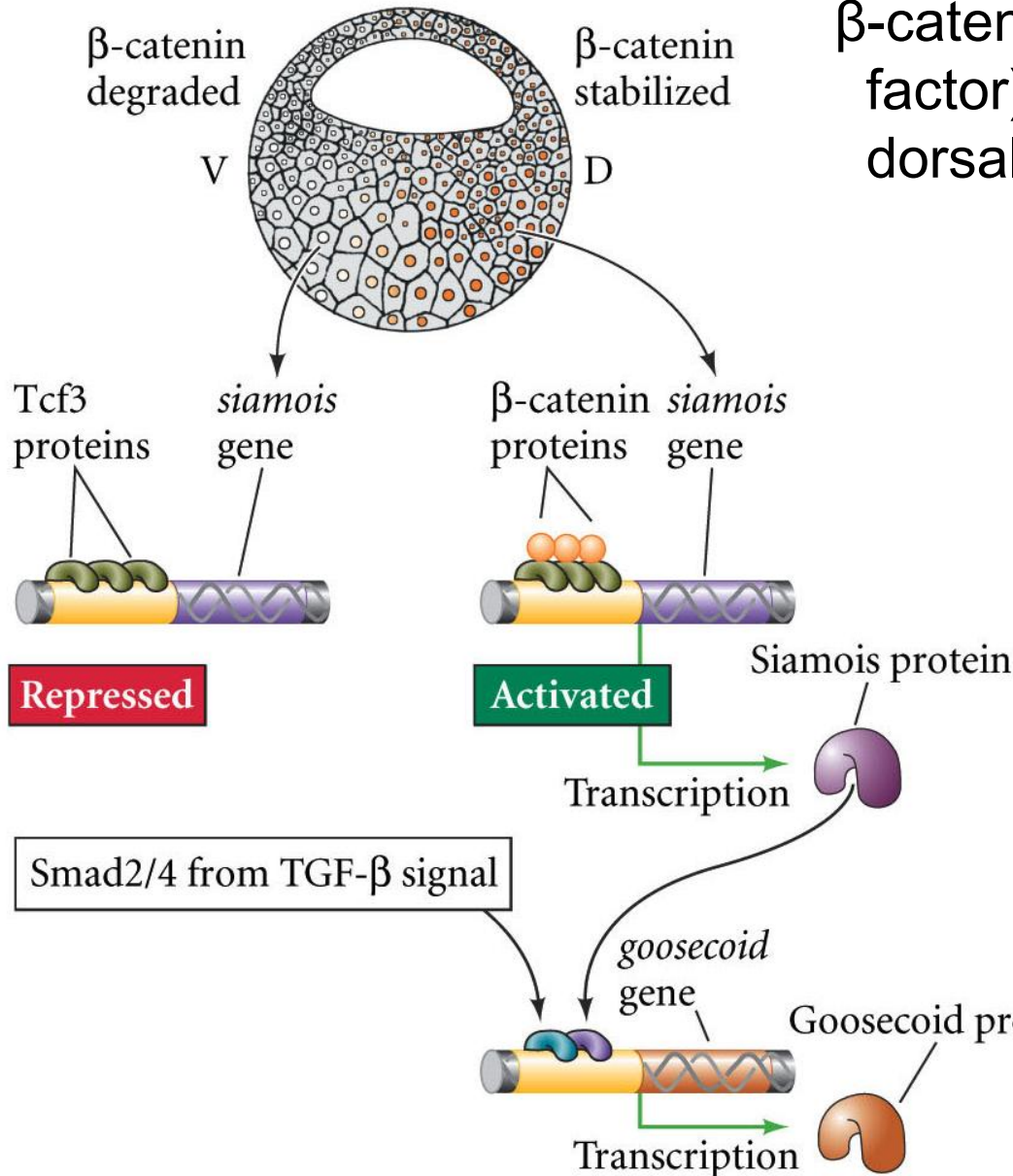
Dorsal inhibition of GSK3



GSK3 mediates  $\beta$ -catenin destruction .....  
but Disheveled and GBP block GSK3;

- resulting in  $\beta$ -catenin present *only* in the marginal area opposite the point of sperm entry (i.e. future dorsal lip)

# Organizer Induction



$\beta$ -catenin acts with Tcf3 (transcription factor); stimulates expression of dorsalizing genes:

**Siamois** – TF; activates *Xlim*, *goosecoid* (dorsal determinants)

**Goosecoid** protein – TF responsible for organizer properties

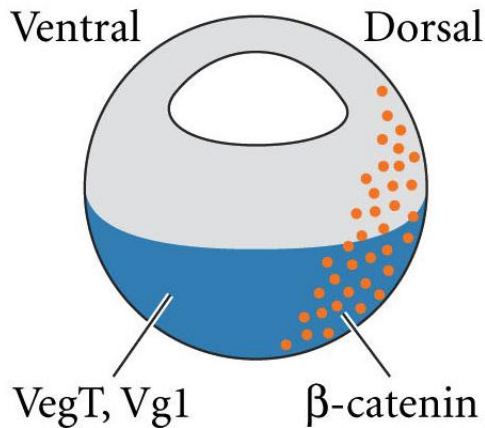
*Goosecoid* also plays a part in specifying dorsal mesoderm; however, additional vegetal factors are needed: e.g.

**Vegetal TGF- $\beta$  signals**

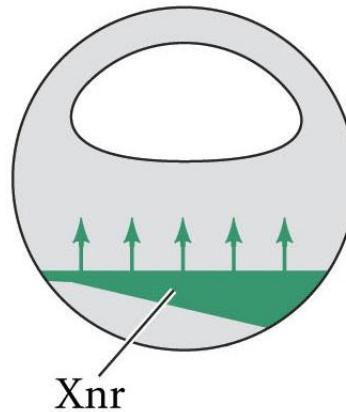
# Mesoderm/Organizer Induction

VegT, Vg1, Xnr(s) – *Xenopus* nodal related genes; TGF- $\beta$  family

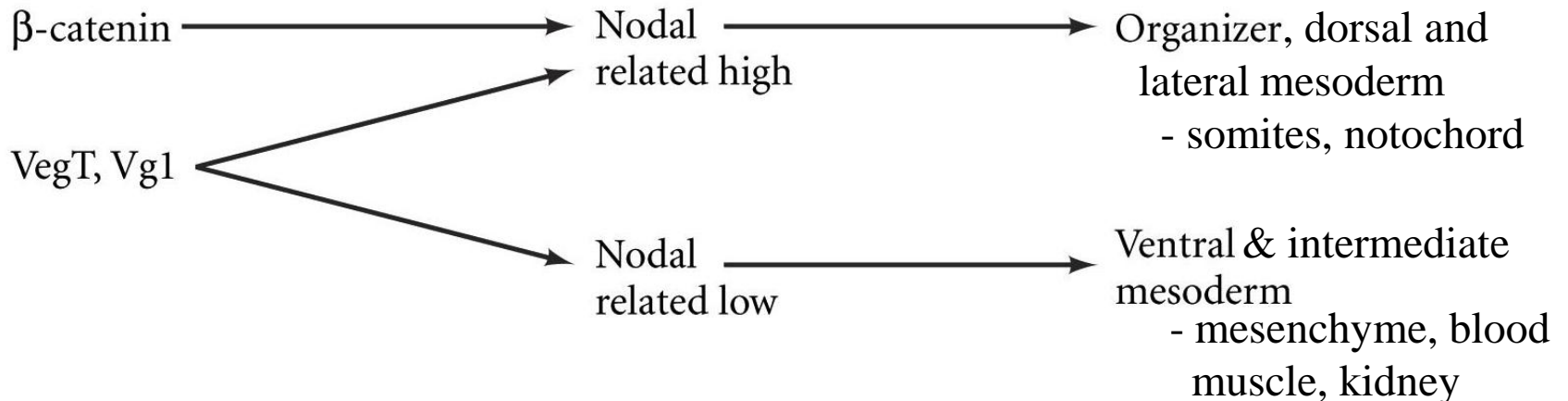
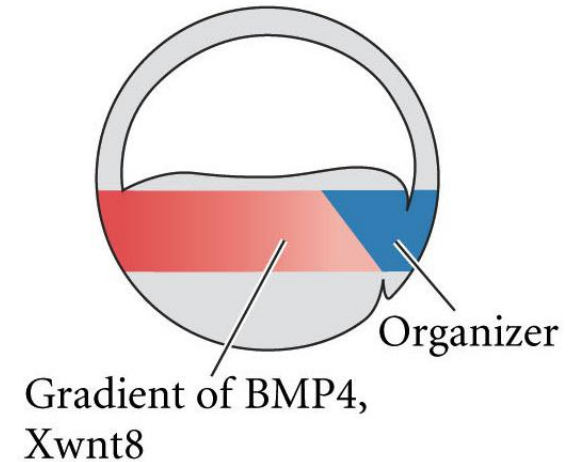
(A) Stage 8 (mid-blastula)



(B) Stage 9 (late blastula)

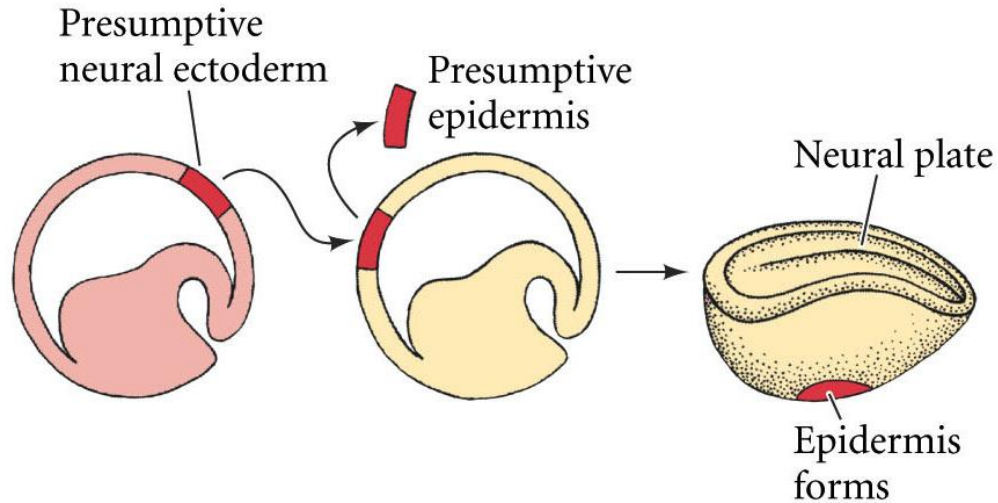


(C) Stage 10 (early gastrulation)



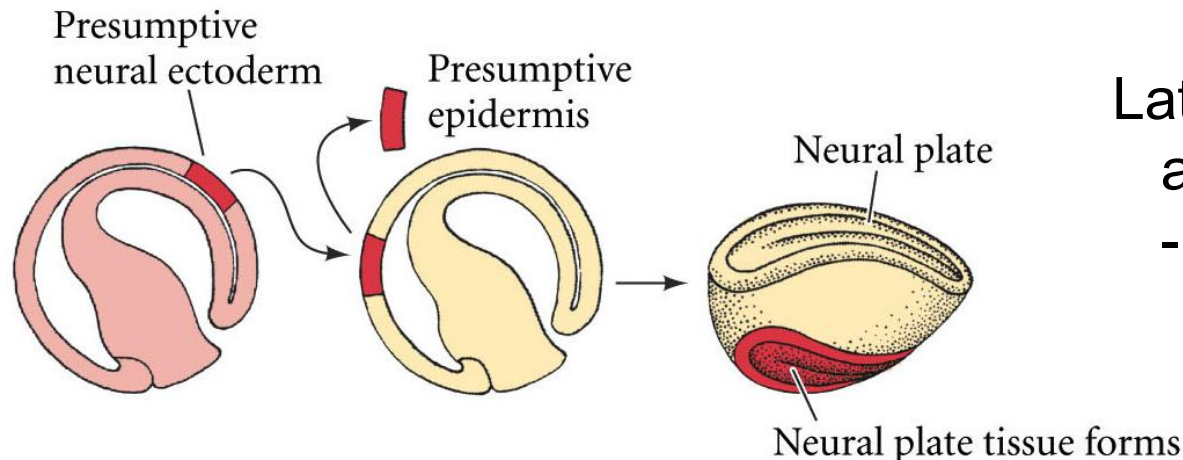
# Determination of Ectoderm

## (A) TRANSPLANTATION IN EARLY GASTRULA



Early gastrulation stage cells are uncommitted  
- exhibit regulative development

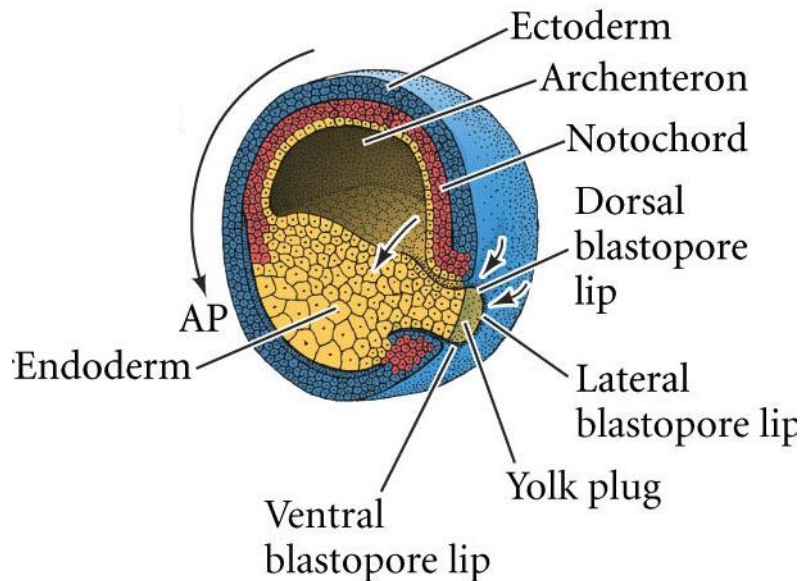
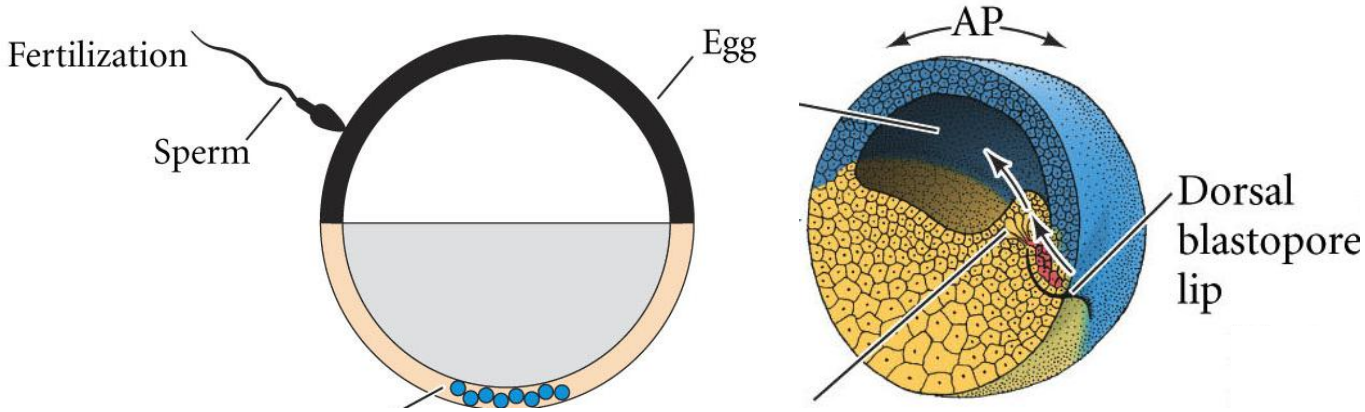
## (B) TRANSPLANTATION IN LATE GASTRULA



Later gastrula cells are determined  
- exhibit autonomous development



# Establishment of Axes - Summary



**D-V axis** – set up at fertilization

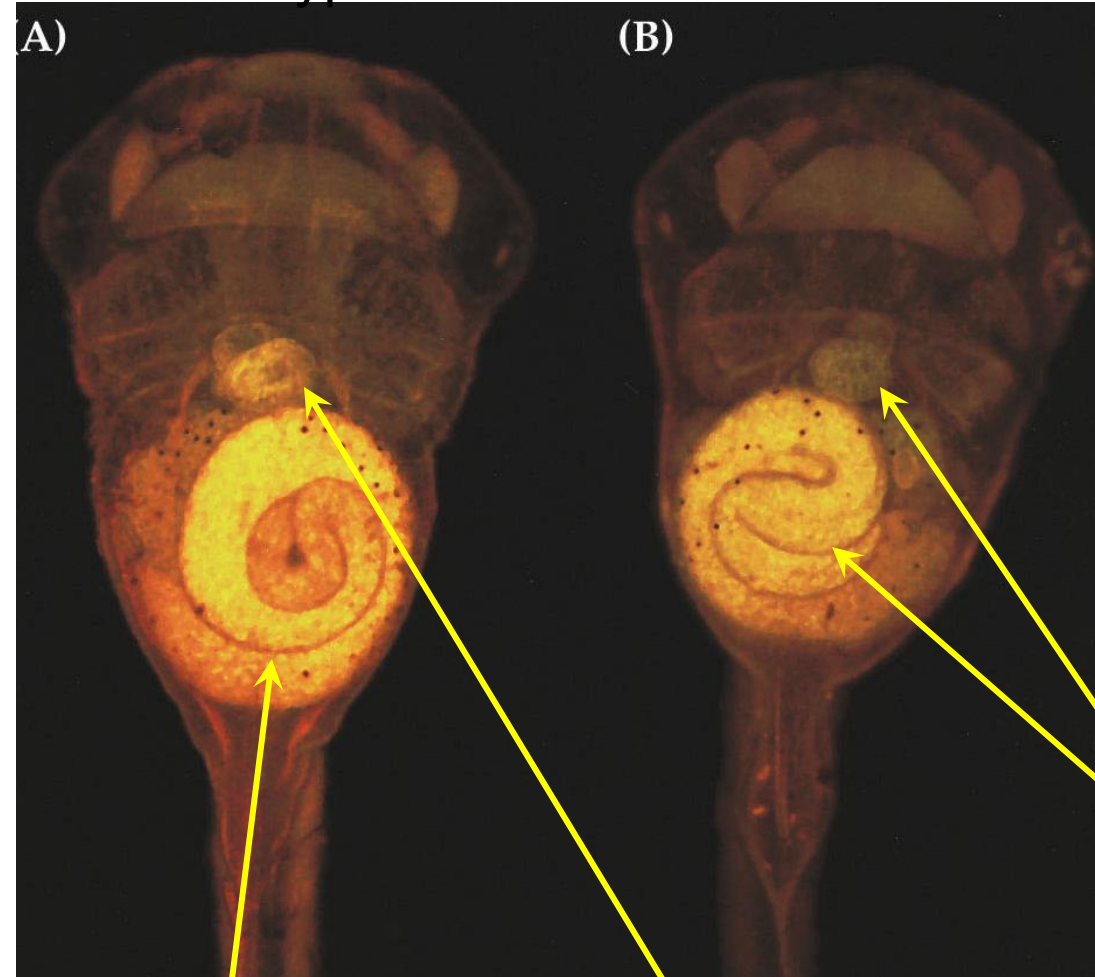
**A-P axis** – established by gastrulation movements across the dorsal lip of the blastopore

**L-R axis** – Nodal expression on Left, not Right

# Left – Right Asymmetry

wild-type

*Xnr1*<sup>-/-</sup>



gut coiling – counter-clockwise

heart loops to the left

Left–Right axis established by *Xenopus nodal-related* (Xnr1)

**Nodal** expression:

- common to all vertebrates
- expressed on the **left side!**

Xnr1 expression is limited to the left side in a process involving cortical rotation and Vg1

Block Xnr1 expression = random gut coiling, heart looping