



Sterculic Acid and Its Analogues Are Potent Inhibitors of *Toxoplasma gondii*

Pan Hao¹, Intisar Q. M. Alaraj², Juma'a R. Al Dulayymi², Mark S. Baird², Jing Liu¹, Qun Liu^{1*}

¹National Animal Protozoa Laboratory, College of Veterinary Medicine, China Agricultural University, Beijing, China; ²School of Chemistry, Bangor University, Bangor, UK

Abstract: Toxoplasmosis is a serious disease caused by *Toxoplasma gondii*, one of the most widespread parasites in the world. Lipid metabolism is important in the intracellular stage of *T. gondii*. Stearoyl-CoA desaturase (SCD), a key enzyme for the synthesis of unsaturated fatty acid is predicted to exist in *T. gondii*. Sterculic acid has been shown to specifically inhibit SCD activity. Here, we examined whether sterculic acid and its methyl ester analogues exhibit anti-*T. gondii* effects in vitro. *T. gondii*-infected Vero cells were disintegrated at 36 hr because of the propagation and egress of intracellular tachyzoites. All test compounds inhibited tachyzoite propagation and egress, reducing the number of ruptured Vero cells by the parasites. Sterculic acid and the methyl esters also inhibited replication of intracellular tachyzoites in HFF cells. Among the test compounds, sterculic acid showed the most potent activity against *T. gondii*, with an EC₅₀ value of 36.2 μM, compared with EC₅₀ values of 248-428 μM for the methyl esters. Our study demonstrated that sterculic acid and its analogues are effective in inhibition of *T. gondii* growth in vitro, suggesting that these compounds or analogues targeting SCD could be effective agents for the treatment of toxoplasmosis.

Key words: *Toxoplasma gondii*, sterculic acid, anti-*Toxoplasma gondii* effect, stearoyl-CoA desaturase

INTRODUCTION

Toxoplasma gondii is a zoonotic pathogen and one of the most widespread parasites, and chronically infects approximately 30% of the global human population, as well as animal species [1]. *T. gondii* is the etiological agent of toxoplasmosis, causing severe neurological deficits in immunosuppressed individuals and causing hydrocephalus, chorioretinitis, and blindness in congenitally infected newborns [2]. The main effect of *T. gondii* infection in farm animals is abortion, which causes significant reproductive losses, and as a result, economic losses [3]. The most frequently used and successful chemotherapy of *T. gondii* infection is a combination of antifolates, such as pyrimethamine and sulfadiazine. However, other options are desirable because of the emergence of resistant strains and side effects caused by the available anti-parasitic agents [4].

Sterculic acid is naturally found in plants such as *Sterculia*

foetida, which is used as an anti-parasite drug in traditional Chinese medicine. Sterculic acid specifically inhibits the conversion of stearic acid to oleic acid by Δ⁹-desaturation [5]. Methyl esters of sterculic acid and analogues were shown to have parasitocidal activities against the asexual blood stage of *Plasmodium falciparum*, probably by targeting stearoyl-CoA desaturase (SCD) [6]. We investigated the anti-parasitic activity of sterculic acid and methyl esters against *T. gondii* in vitro. Herein, we show that sterculic acid and its analogue methyl esters also inhibit the growth of *T. gondii* tachyzoites in vitro.

MATERIALS AND METHODS

Candidate compounds

Sterculic acid is a cyclopropene fatty acid. The analogues used in our experiments were methyl esters of cyclopropene fatty acids that had similar structures to sterculic acid (Fig. 1). INT15 is the corresponding methyl ester of sterculic acid while INT13 and INT14 has 1 less or 1 more carbon atom than INT15 in the chains, respectively. INT21 is composed of the backbone of INT15 and a methoxyl. The compounds were prepared as previously described [6-8]. DMSO was used to dissolve these compounds at 1 M as stock solutions.

•Received 2 December 2015, revised 10 March 2016, accepted 13 March 2016.

*Corresponding author (qunliu@cau.edu.cn)

© 2016, Korean Society for Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

MTS assay

This experiment was performed as previously described with minor modifications [4]. Vero cells of exponential growth stage were digested and cultured in 96-well plates at 6×10^3 cells/well. The adherent cells were infected with *T. gondii* tachyzoites resuspended in RPMI 1640 containing 10% FBS (3×10^4 tachyzoites in 100 μ l medium per well). The infected cells were washed 6 hr after inoculation, and RPMI 1640 medium without FBS was added. After 18 hr, the medium in each well was changed into RPMI 1640 supplemented with 2% FBS along with different concentrations of sterculic acid or the methyl esters (final concentrations 15.6-1,000 μ M). After treatment for 24 hr, anti-*T. gondii* activity and host cell cytotoxicity of the test compounds were evaluated by the 3-(4,5-dimethylthiazolyl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium, inner salt (MTS) assay using the CellTiter 96® Aqueous One Solution Cell Proliferation Assay kit (Promega Biotech, Beijing, China) according to the manufacturer's instruction. The OD value of each well was assayed at the wavelength of 450 nm on a microplate reader (Model 680, Bio-Rad, Richmond, California, USA). Cell viability was expressed as percentage of the control value. The 50% cytotoxicity concentration (CC_{50}) was expressed as the concentration reducing Vero cell growth by 50%, and the 50% effective concentration (EC_{50}) was the concentration inhibiting *T. gondii*-induced cytopathic effect by 50%. Anti-*T. gondii* activity was expressed as the selectivity index calculated from the CC_{50} and EC_{50} values (CC_{50}/EC_{50}).

Statistical analysis

All experiments and measurements were performed in triplicate. The number of extracellular tachyzoites was compared using the Student's *t*-test. The proliferation results were statistically analyzed using the chi-square test (PASW Statistics 18). EC_{50} and CC_{50} were calculated in GraphPad Prism 5 using non-linear regression.

RESULTS

Inhibitory activity against tachyzoite egress from host cells

Normal growth and proliferation of intracellular tachyzoites result in host cell rupture and thus release of tachyzoites from the host cell. Prevention of tachyzoite egress from Vero cells was used as a marker for antiparasitic activity of the test compounds. The high concentration of sterculic acid (1 mM)

caused cell death within 12 hr (data not shown), while methyl ester analogues were not cytotoxic at 1 mM. Thus, concentrations of 0.1 mM sterculic acid and 1 mM methyl esters were tested for antiparasitic activity in the Vero cell assay. Vero cells were unaffected at 12 hr and were of the same appearance in the treated and control groups. In the control group, tachyzoites started to egress at 24 hr, and by 36 hr most of the tachyzoites were released from the host cells with only a small number of intact cells remaining. Most host cells of all the treated groups remained intact at 36 hr (Fig. 2A-E), with a smaller number of tachyzoites released (Fig. 2G). The lowest number of extracellular tachyzoites was seen in the INT21-treated group, and the intracellular tachyzoites seemed to cease developing at 36 hr.

Inhibitory activity against tachyzoite proliferation

The decreased tachyzoite egress indicated that the test compounds inhibited the growth of *T. gondii* tachyzoites. We then examined the anti-proliferation activity of the test compounds using the indirect fluorescent antibody assay. The number of parasitophorous vacuoles containing 1-8 intracellular tachyzoites was counted after incubation in HFF for 18 hr. All test compounds inhibited tachyzoite proliferation. Significantly more parasitophorous vacuoles of all treated groups remained in the 1 or 2 tachyzoites per vacuole stage than the DMSO control (Fig. 3). Of the test compounds, INT21 was most effective, with over 90% parasitophorous vacuoles arresting at 2 or 4 tachyzoites stage and only 3% grown into 8 tachyzoites. In other treated groups, about 30% of parasitophorous vacuoles contained 8 tachyzoites. In comparison, approximately 50% parasites developed to 4 tachyzoites and 40% to 8 per vacuole in the DMSO culture.

Anti-*T. gondii* activity by MTS assay

Anti-*T. gondii* activity was also tested using the MTS assay. Vero cells grew normally in culture medium with methyl esters (INT13, INT14, and INT15) no matter whether the test concentration was low or high. INT21 seemed to promote the cell growth, especially in the low concentration. In contrast, sterculic acid was cytotoxic to Vero cells (Fig. 4A), with a mean CC_{50} of 215 μ M. The viability of Vero cells infected with *T. gondii* was increased with increasing compound concentrations, indicating anti-*T. gondii* activity of the test compounds (Fig. 4B). Sterculic acid had the lowest EC_{50} value (36.2 μ M). The mean EC_{50} values for the methyl esters ranged from 248.0 μ M

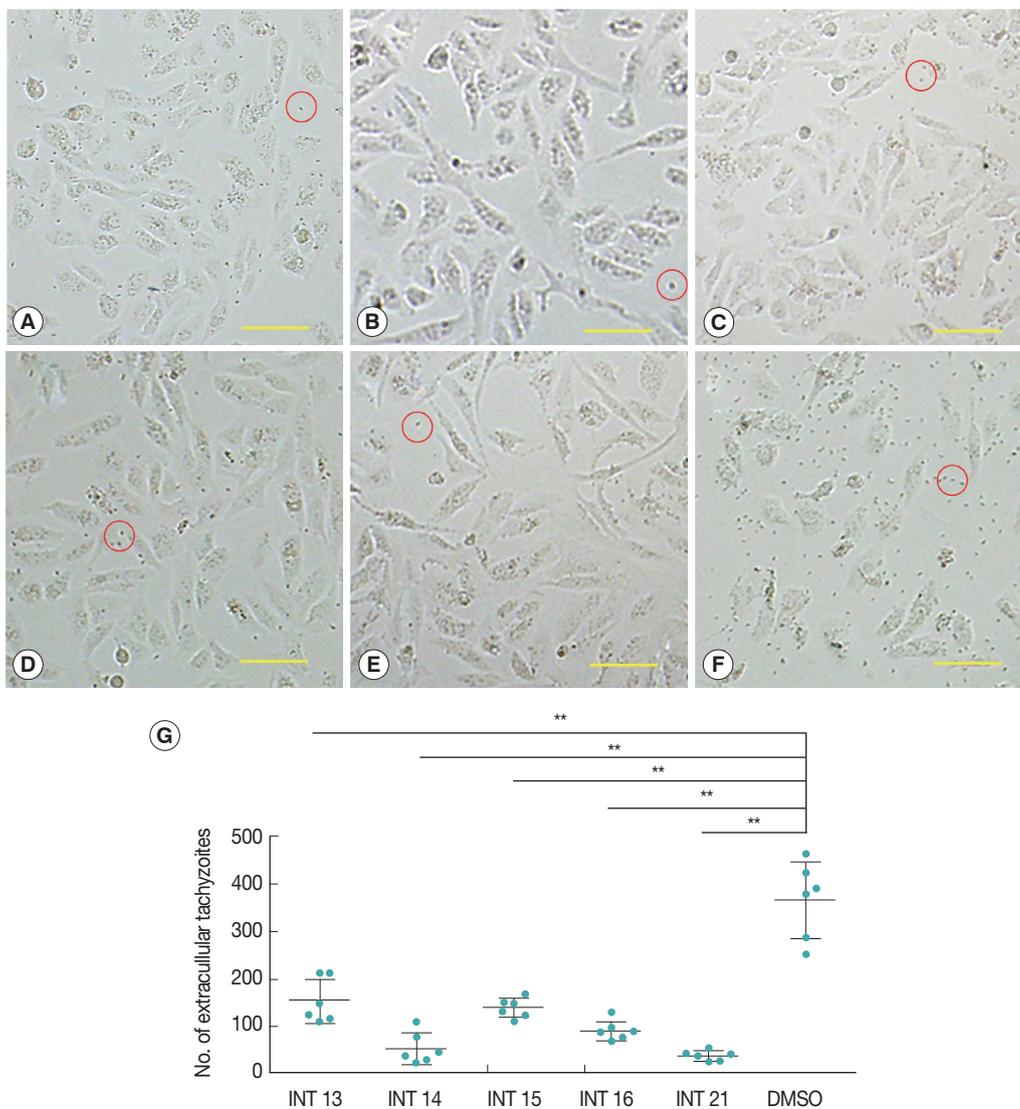


Fig. 2. Sterculic acid and methyl esters inhibited the egress of tachyzoites from Vero cells. Representative pictures were presented to show that most of host cells remained intact after incubation with INT13 (A), INT14 (B), INT15 (C), INT16 (D), or INT21 (E) for 36 hr, with a small number of released tachyzoites, compared with numerous tachyzoites and a smaller number of intact host cells in the negative control group (F). Scale bar = 100 μ m. Extracellular tachyzoites are marked in red circles. (G) The number of released tachyzoites in the culture incubated with the test compounds was decreased relative to the DMSO control. Extracellular tachyzoites were counted in 6 random fields under the microscope per group. Data are presented as mean \pm SD. ** $P < 0.01$.

(INT21) to 428.4 μ M (INT14). All the test compounds have a selectivity index of > 2 (Table 1). The antiparasitic activity in MTS assay was consistent with the release assay and proliferation assay results, with INT21 being the most effective moiety of the methyl esters.

DISCUSSION

Our study showed that sterculic acid and its analogues all

exhibited anti-*T. gondii* activities in vitro in 3 different assays – egress, proliferation, and MTS. Among the compounds examined, sterculic acid exhibited the lowest EC_{50} value against *T. gondii* by the MTS assay while it was also most toxic to Vero cells, hindering us from testing its effect in the proliferation assay with a higher concentration. The effect of sterculic acid on Vero cells or HFFs has not been reported previously. Our results showed that sterculic acid is cytotoxic at a high concentration, which was similar to a report that the viability of differ-

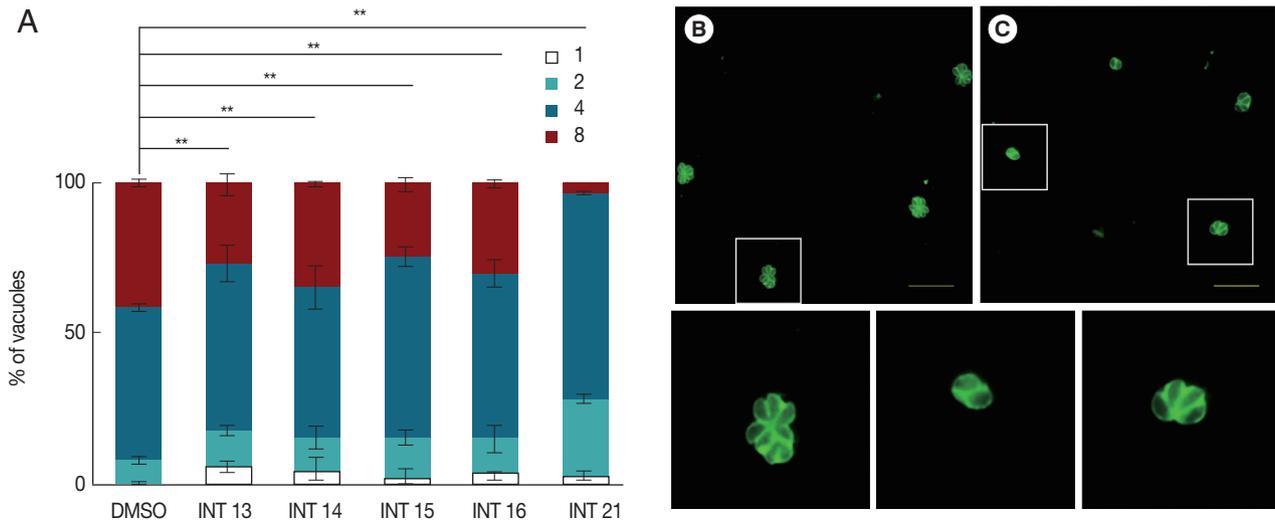


Fig. 3. Sterculic acid and all methyl ester analogues inhibit tachyzoite proliferation. Freshly released parasites were used to infect HFF cell monolayers. (A) Invaded parasites were allowed to replicate for 18 hr in the culture medium containing the test compounds, and the number of vacuoles containing 1, 2, 4, or 8 parasites was counted under a fluorescence microscope after anti-*T. gondii* staining. (B) Representative fluorescent microscopic images were presented to show that many vacuoles contained 8 tachyzoites in DMSO control groups, (C) while 4 or 2 tachyzoites were often observed in a vacuole in INT21-treated groups after culture for 18 hr. The magnified images of the areas marked with white boxes in (B) and (C) were presented at the bottom in order. Data are presented as mean \pm SD from 3 independent experiments each in triplicate. ****** $P < 0.01$.

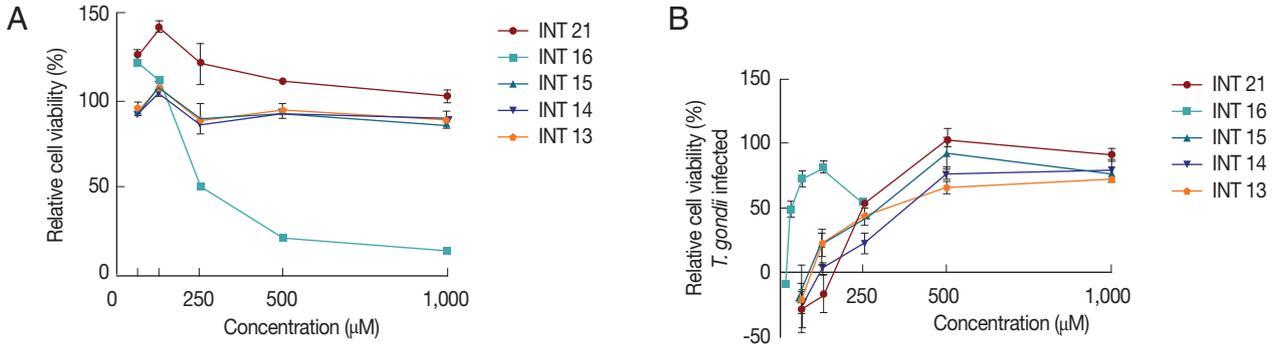


Fig. 4. Relative cell viability was calculated from the MTS assay. The cell viability of 0.1% DMSO was regarded as 100%. (A) Sterculic acid was cytotoxic to Vero cells at high concentrations while the methyl esters displayed no cytotoxicity. (B) All test compounds inhibited *T. gondii* growth as shown by increased viability of Vero cells infected with *T. gondii*.

Table 1. In vitro anti-*T. gondii* activity of sterculic acid and its methyl ester analogues

Compound code	EC ₅₀ ^a (μM)	CC ₅₀ ^b (μM)	SI ^c
Sterculic acid (INT16)	36.2 \pm 5.1	215.2 \pm 15.8	5.9
INT13	356.9 \pm 14.1	> 1,000	> 2.8
INT14	428.4 \pm 15.7	> 1,000	> 2.3
INT15	306.4 \pm 18.8	> 1,000	> 3.3
INT21	248.0 \pm 17.9	> 1,000	> 4.0

Results are presented as mean \pm SD from 3 independent experiments with triplicate wells in each experiment.

^aEC₅₀, half maximal effective concentration. Concentration required to inhibit *T. gondii*-induced cytopathic effect by 50%.

^bCC₅₀, half maximal cytotoxic concentration. Concentration required to reduce Vero cell growth by 50%.

^cSelectivity index = CC₅₀/EC₅₀

entiated adipocyte cultures was decreased at higher levels of sterculic acid (100 μ M or 200 μ M) [10]. In contrast, the methyl esters did not exhibit negative effects on Vero cells and even increased the cell viability in low concentrations. Increased cell proliferation under methyl sterculate treatment was also observed in early reports, and methyl ester of sterculic acid has been identified as one of the mitogenic principles [11,12]. INT21, the only methoxy analogue, was found to have little cytotoxicity and the most active in the inhibition of *T. gondii* growth among the methyl esters, which consisted with a previous report that the compound was the most active against *P. falciparum* among 7 methyl esters tested [6].

Sterculic acid directly inhibits stearoyl-coenzyme A desaturase (SCD) activity without affecting the processes required for adipocyte differentiation, *scd* gene expression or SCD protein translation in 3T3-L1 adipocytes [13]. Sterculic acid methyl ester also inhibits $\Delta 9$ -desaturase of the bacteria *Rhodococcus opacus*, leading to an almost complete suppression of growth at 0.25 mM and even cell lysis at 0.75 mM [14]. Methyl esters of sterculic acid have also been shown to inhibit SCD of *P. falciparum* and exhibited antimalarial activities in vitro [6]. By sequence alignment with *P. falciparum* SCD in the ToxoDB database, a putative TgSCD (TGGT1_238950) was predicted to exist in *T. gondii*, retaining classic $\Delta 9$ -desaturase amino acid sequences, such as 3 Histidine-Boxes and 8 conserved His residues. Neither the transcription nor the translation of *T. gondii* SCD was affected in the presence of the test compounds in our research, determined by qPCR and western blot (data not shown). We implied that sterculic acid and its analogues might only inhibit the SCD activity in *T. gondii*.

SCD is the endoplasmic reticulum-resident enzyme that catalyzes the introduction of the first double bond in the cis- $\Delta 9$ position of several saturated fatty acyl-CoAs, principally palmitic acid (C16:0) and stearic acid (C18:0), to render palmitoleic acid (C16:1) and oleic acid (C18:1). The presence of SCD has been described in a wide variety of organisms [15]. SCD1, an isoform of SCD in mammalian organisms, apart from its role in monounsaturated fatty acids synthesis, has been recently revealed a novel function in the modulation of metabolic and signaling processes related to cell proliferation, survival, and malignant transformation to cancer [16]. In apicomplexan parasites, SCD has not been deeply studied but only identified in *P. falciparum*. It converted C18:0 to C18:1 oleic acid and was important for the blood-stage survival [6]. However, C18:1 in *T. gondii* was presumably synthesized from

the elongation of C16:1 palmitoleic acid by ELO-A (1 of 3 elongases in the fatty acid elongation system) [17]. Besides de novo synthesis of the fatty acids, *T. gondii* can also salvage lipids from the host cells and the environment [18,19]. It is possible that *T. gondii* desaturates salvaged C18:0 to C18:1 by SCD. Moreover, SCD can also catalyze the synthesis of C16:1 using C16:0 as the substrate [20], suggesting that the synthesis of C16:1 palmitoleic acid may be mediated by SCD in *T. gondii*. Therefore, TgSCD might be an essential enzyme in the mono-unsaturated fatty acid synthesis in this parasite. We inferred that the anti-*T. gondii* activity of sterculic acid and its analogues is most likely mediated by inhibiting TgSCD.

In conclusion, sterculic acid and its analogues inhibited *T. gondii* growth in vitro, most likely by targeting the synthesis of unsaturated fatty acid, which is essential for the development and proliferation of *T. gondii*. In vivo anti-*T. gondii* effects of sterculic acid analogues and other $\Delta 9$ -desaturase inhibitors warrant further research. The function of TgSCD also requires further investigation.

ACKNOWLEDGMENTS

This work was financed through the National Natural Science Foundation of China (No. 31372424).

CONFLICT OF INTEREST

We have no conflict of interest related to this work.

REFERENCES

- Lüder CG, Bohne W, Soldati D. Toxoplasmosis: a persisting challenge. *Trends Parasitol* 2001; 17: 460-463.
- Martins-Duarte ES, Urbina JA, de Souza W, Vommaro RC. Antiproliferative activities of two novel quinuclidine inhibitors against *Toxoplasma gondii* tachyzoites in vitro. *J Antimicrob Chemother* 2006; 58: 59-65.
- Ferra B, Holec-Gasior L, Kur J. Serodiagnosis of *Toxoplasma gondii* infection in farm animals (horses, swine, and sheep) by enzyme-linked immunosorbent assay using chimeric antigens. *Parasitol Int* 2015; 64: 288-294.
- Kavitha N, Noordin R, Chan KL, Sasidharan S. In vitro anti-*Toxoplasma gondii* activity of root extract/fractions of *Eurycoma longifolia* Jack. *BMC Complement Altern Med* 2012; 12: 91.
- Jeffcoat R, Pollard MR. Studies on the inhibition of the desaturases by cyclopropanoid fatty acids. *Lipids* 1977; 12: 480-485.
- Gratraud P, Huws E, Falkard B, Adjalley S, Fidock DA, Berry L, Jacobs WR Jr, Baird MS, Vial H, Kremer L. Oleic acid biosynthe-

- sis in *Plasmodium falciparum*: characterization of the stearoyl-CoA desaturase and investigation as a potential therapeutic target. PLoS One 2009; 4: e6889.
7. Baird MS, Dale CM, Lytollis W, Simpson MJ. A new approach to cyclopropene fatty acids. Tetrahedron Lett 1992; 33: 1521-1522.
 8. Baird MS, Grehan B. A new approach to cyclopropene fatty acids involving 1,2-deiodination. J Chem Soc Perkin Trans 1993; 1: 1547-1548.
 9. Weiss LM, Kim K. *Toxoplasma gondii* - the Model Apicomplexan: Perspectives and Methods. 2nd ed. Salt Lake City, USA. Academic Press. 2014, pp 594-595.
 10. Kadegowda AK, Burns TA, Pratt SL, Duckett SK. Inhibition of stearoyl-CoA desaturase 1 reduces lipogenesis in primary bovine adipocytes. Lipids 2013; 48: 967-976.
 11. Scarpelli DG. Mitogenic activity of sterculic acid, a cyclopropenoid fatty acid. Science 1974; 185: 958-960.
 12. Leveille AS, Fischer-Dzoga K. The mitogenic effects of methyl sterculate on aortic smooth muscle cells. Artery 1982; 11: 207-224.
 13. Gomez FE, Bauman DE, Ntambi JM, Fox BG. Effects of sterculic acid on stearoyl-CoA desaturase in differentiating 3T3-L1 adipocytes. Biochem Biophys Res Commun 2003; 300: 316-326.
 14. Waltermann M, Steinbuchel A. In vitro effects of sterculic acid on lipid biosynthesis in *Rhodococcus opacus* strain PD630 and isolation of mutants defective in fatty acid desaturation. FEMS Microbiol Lett 2000; 190: 45-50.
 15. Scaglia N, Igal RA. Stearoyl-CoA desaturase is involved in the control of proliferation, anchorage-independent growth, and survival in human transformed cells. J Biol Chem 2005; 280: 25339-25349.
 16. Igal RA. Stearoyl-CoA desaturase-1: a novel key player in the mechanisms of cell proliferation, programmed cell death and transformation to cancer. Carcinogenesis 2010; 31: 1509-1515.
 17. Ramakrishnan S, Docampo MD, Macrae JI, Pujol FM, Brooks CF, van Dooren GG, Hiltunen JK, Kastaniotis AJ, McConville MJ, Striepen B. Apicoplast and endoplasmic reticulum cooperate in fatty acid biosynthesis in apicomplexan parasite *Toxoplasma gondii*. J Biol Chem 2012; 287: 4957-4971.
 18. Charron AJ, Sibley LD. Host cells: mobilizable lipid resources for the intracellular parasite *Toxoplasma gondii*. J Cell Sci 2002; 115: 3049-3059.
 19. Mazumdar J, Wilson EH, Masek K, Hunter CA, Striepen B. Apicoplast fatty acid synthesis is essential for organelle biogenesis and parasite survival in *Toxoplasma gondii*. Proc Natl Acad Sci USA 2006; 103: 13192-13197.
 20. Poloni S, Blom HJ, Schwartz IV. Stearoyl-CoA Desaturase-1: is it the link between sulfur amino acids and lipid metabolism? Biology 2015; 4: 383-396.