

# Gene Therapy for Head and Neck Squamous Cell Carcinoma Using KITENIN (KAI1 COOH-Terminal Interacting Tetraspanin)-Antisense Therapy

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**Purpose:** KAI1 COOH-terminal interacting tetraspanin (KITENIN) has been found to act as a promoter of metastasis in murine models of colon cancer and squamous cell carcinoma (SCC). The suppression of tumor progression and metastasis of established colon cancer in mice was observed after intravenous delivery of small interfering RNA (siRNA) targeting KITENIN. The purpose of this study was to investigate the efficacy of gene therapy targeting KITENIN in human head and neck SCC. **Materials and Methods:** SNU-1041, a well-established human hypopharyngeal SCC cell line, was used. KITENIN expression in SNU-1041 was measured by Western blot analysis. The cells were prepared, maintained in culture dishes with media, and divided into two groups: the si-KITENIN group and the scrambled group (control). The siRNA targeting KITENIN (si-KITENIN) and scrambled DNA were transfected into the SNU-1041 cells in each group. The effect of gene therapy was compared by in vitro experiments to evaluate invasion, migration, and proliferation. **Results:** KITENIN was strongly expressed in the SNU-1041 cells, and the number of invaded cells was reduced more in the si-KITENIN group than in the scrambled group ( $p < 0.001$ ). The speed for the narrowing gap, made through adherent cells, was lower in the si-KITENIN group ( $p < 0.001$ ), and the number of viable proliferating cells was reduced in the si-KITENIN group compared to the scrambled group ( $p < 0.001$ , the third day). KITENIN protein expression was no longer identified in the si-KITENIN group. **Conclusion:** Gene therapy using an anti-KITENIN strategy might be effective for head and neck squamous carcinoma.

**Key Words:** Gene therapy, head and neck cancer, squamous cell carcinoma, KITENIN protein, small interfering RNA (siRNA)

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## INTRODUCTION

Reducing the progression and metastasis of head and neck squamous cell carcinoma (SCC) requires an understanding of molecular mechanisms involved in inva-

sion and dissemination.<sup>1</sup> Gene therapy for tumor metastasis is a promising because it inhibits the spread of cancer regardless of the tumorigenicity.<sup>2,3</sup>

A large body of evidence suggests that down-regulation of KAI1/CD82 expression occurs with invasive and metastatic disease, and that it is an important step in the progression of many human malignancies.<sup>4-8</sup> KAI1, a tumor metastasis suppressor gene, is a transmembrane glycoprotein that is a member of the tetraspanin superfamily. The metastasis suppressor function was recently reported to be decreased in a spliced variant of KAI1 at the COOH-terminal region, suggesting that the COOH-terminal region of KAI1 is important for the effects of KAI1 on cell motility.<sup>9</sup> Subsequently, a protein that interacted with the COOH-terminal cytoplasm domain of KAI1 was identified via a two-hybrid yeast system.<sup>10</sup> The function of this protein Vang-like 1 (VANGL1<sup>11</sup>) during carcinogenesis has not been fully described, and it has been renamed as KAI1 COOH-terminal interacting tetraspanin (KITENIN).

KITENIN-overexpressing CT-26 mouse colon cancer cells have shown increased tumorigenicity and early hepatic metastasis *in vivo*, as well as increased invasiveness and adhesiveness to fibronectin *in vitro*, compared to parental cells.<sup>10</sup> Moreover, the suppression of progression and metastasis of established colon cancer has been observed in mice after intravenous delivery of small interfering RNA (siRNA) targeting KITENIN.<sup>12</sup>

We recently reported that KITENIN promoted pulmonary metastasis in a murine model of SCC by increasing cancer cell invasion and adhesion to fibronectin,<sup>13</sup> and that KITENIN represented a more aggressive phenotype in a murine model of oral cavity squamous carcinoma.<sup>14</sup> Furthermore, we investigated the expression of KITENIN in human laryngeal SCC, and found that KITENIN expression was significantly increased in laryngeal cancer tissues, compared to the adjacent normal tissue mucosa, as well as in metastatic lymph nodes compared to non-metastatic lymph nodes. High KITENIN expression was associated significantly with an advanced disease stage, extent of the tumor, and lymph node metastasis.<sup>15</sup>

siRNA is a small double-stranded, non-protein coding RNA (21-31 nucleotides) involved in gene silencing functions, especially RNA interference (RNAi). This ability of siRNA has provided researchers with a novel tool to block the expression of disease-causing genes, provided that their mRNA sequences are known. siRNAs can be delivered to cells either exogenously as synthetic agents or endogenously

as gene-coding siRNAs. Recent studies have demonstrated the general application of siRNAs to silence gene expression in a range of cell types and in mammalian models.<sup>16</sup>

The purpose of this study was to investigate the efficacy of gene therapy (siRNA targeting KITENIN) in human head and neck SCC.

## MATERIALS AND METHODS

### Cell culture and transfection

SNU-1041 cells,<sup>17-19</sup> a well-established human head and neck SCC cell line, were grown in Dulbecco's modified Eagle's medium (DMEM, Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum (Hyclone, Logan, UT, USA) in a humidified atmosphere of 5% CO<sub>2</sub> at 37°C. The cells were prepared and maintained in culture dishes with media, and divided into two groups: the si-KITENIN group and the scrambled group (control). The siRNA targeting KITENIN (si-KITENIN) and the scrambled DNA were transfected into the SNU-1041 cells in each group. The sequence of siRNA was as follows: human KITENIN 5'-GCUUG-GACUUCAGCCUCGUAGUCAA-3'. The transfections were performed using the Lipofectamine RNAiMAX (Invitrogen) according to the manufacturer's instructions. The cells were maintained and used for further analysis.

### Western blot analysis

Western blot analysis was used to investigate KITENIN expression before and after transfection of siRNA (si-KITENIN). The tissues were solubilized in NP40 lysis buffer containing a protease inhibitor mixture (Roche, Indianapolis, IN) and 1 mM phenylmethylsulfonyl fluoride. The resolved proteins (50 µg) were transferred to a nitrocellulose membrane and blotted with the KITENIN antibody and anti-rabbit immunoglobulin-horseradish peroxidase (Amer sham, Arlington Heights, IL, USA). The blot was reprobed with anti-actin antibody (I-19; Santa Cruz Biotechnology, Santa Cruz, CA, USA) to control loading variations.

### Cell invasion assay

Cell migration was measured using a Transwell migration apparatus (Costar Inc., Cambridge, UK) with minor modification as described previously.<sup>9</sup> The filters (8-µm pore size) were coated with 1% gelatin solution on both the top and bottom surfaces. The cells from each group (si-KITENIN group, scrambled group) were harvested, washed once in serum-free

DMEM/0.2% bovine serum albumin (BSA), and resuspended at  $2 \times 10^6$  cells/ $\mu\text{L}$  DMEM/0.2% BSA. To start the assay, 120  $\mu\text{L}$  ( $2.4 \times 10^5$  cells) was loaded onto the upper chamber of the Transwell, whereas 400  $\mu\text{L}$  DMEM/0.2% BSA containing 20  $\mu\text{g}/\text{mL}$  human plasma fibronectin (Calbiochem, La Jolla, CA, USA), a chemotactic factor, was loaded onto the lower chamber. The Transwell apparatus was incubated for 24 hours at 37°C. At the end of incubation, the cells attached to the membranes were fixed and stained with Diff-Quick (International Reagents, Kobe, Japan) by following the manufacturer's protocol. The cells on the top surface of the filters were wiped off with cotton balls, and the cells that migrated to the bottom surface were counted in ten random squares of  $0.5 \times 0.5 \text{ mm}^2$  by microscopic field of view. The results were expressed as mean $\pm$ SE of the number of cells/field.

#### Cell migration assay

si-KITENIN-transfected and scrambled DNA-transfected SNU-1041 cells were seeded in a 6-well plate ( $1 \times 10^5$  cells/well). Twenty-four hours later, the media were changed to serum-free DMEM and incubated for 12 hours. After the media were removed from the wells, a straight transverse line through the adherent cells was drawn using a ruler and a 1000  $\mu\text{L}$ -tip, resulting in a uniform gap. The media were changed to DMEM supplemented with 2% fetal bovine serum. At 0, 20, 28, and 44 hours later; the distances between the gaps were measured in centimeters after capture of six random sites by the microscopic field of view.

#### Cell proliferation assay

The proliferation and viability of the cells were measured using an enhanced cell viability assay kit, EZ CyTox (Daeil Lab Service Co., Seoul, Korea). The cells were seeded at  $2 \times 10^4$  cells/well in 48-well plates, transfected with si-KITENIN and scrambled DNA, and grown overnight in the presence of serum. Next, EZ CyTox was added according to the manufacturer's instructions for 2 hours, after which the discolored cells were moved into 96-well plates for analysis. Absorbance at 450 nm was determined using a microplate reader with SOFTmax PRO software (Molecular Devices, Sunnyvale, CA, USA).

#### Statistical analysis

Experimental differences were tested for statistical significance with SPSS 16.0 (SPSS Inc., Chicago, IL, USA). The Student's t-test was used, and  $p$  values of less than 0.05 were considered statistically significant.

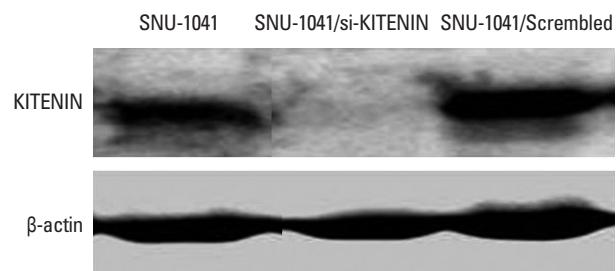
## RESULTS

#### Western blot analysis

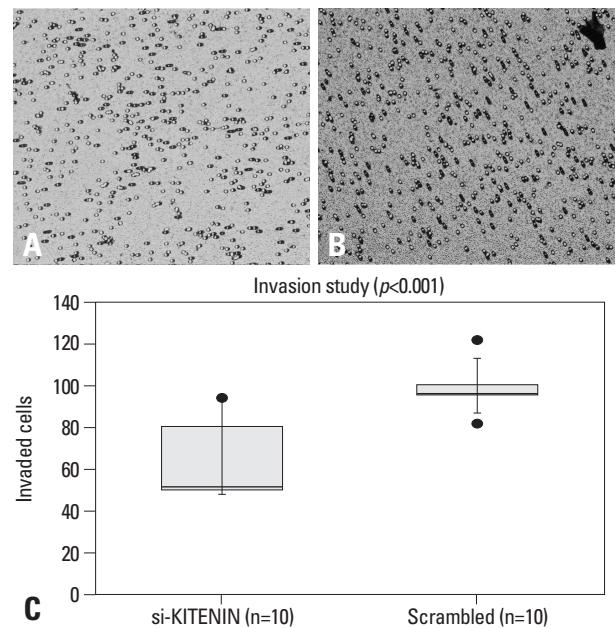
Fig. 1 shows that KITENIN was strongly expressed in the SNU-1041 cells and scrambled DNA-transfected SNU-1041 cells (SNU-1041/Scrambled). However, the expression of KITENIN protein was not observed in the si-KITENIN-transfected SNU-1041 cells (SNU-1041/si-KITENIN), demonstrating that KITENIN was effectively blocked by siRNA.

#### Cell invasion assay

As shown in Fig. 2, the number of invading si-KITENIN-transfected SNU-1041 cells was  $64.2 \pm 19.2$ , whereas the number of the scrambled DNA-transfected SNU-1041 cells



**Fig. 1.** KITENIN expression in SNU-1041 cells: KITENIN was strongly expressed in SNU-1041 cells and scrambled DNA-transfected SNU-1041 cells (SNU-1041/Scrambled). The expression of KITENIN protein was not observed in si-KITENIN-transfected SNU-1041 cells (SNU-1041/si-KITENIN).

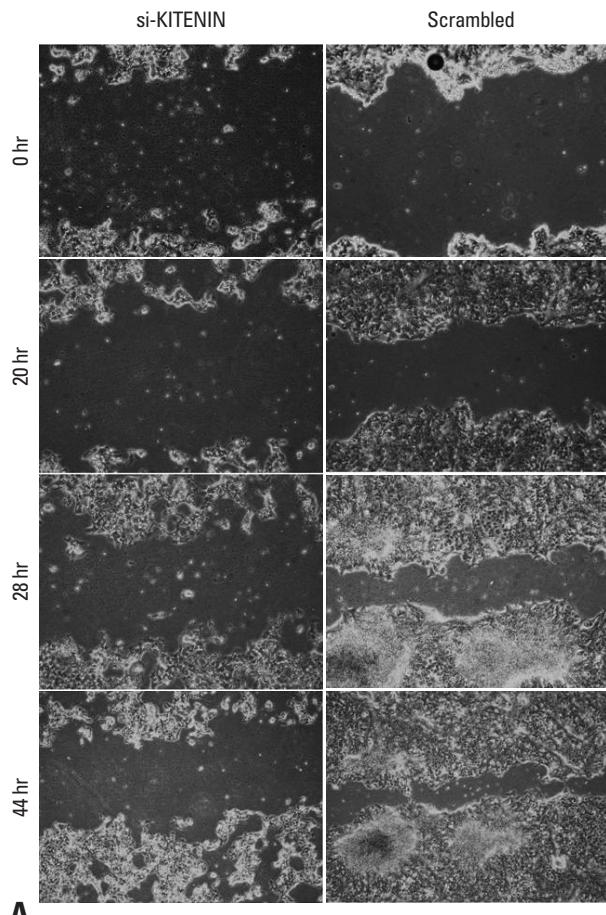
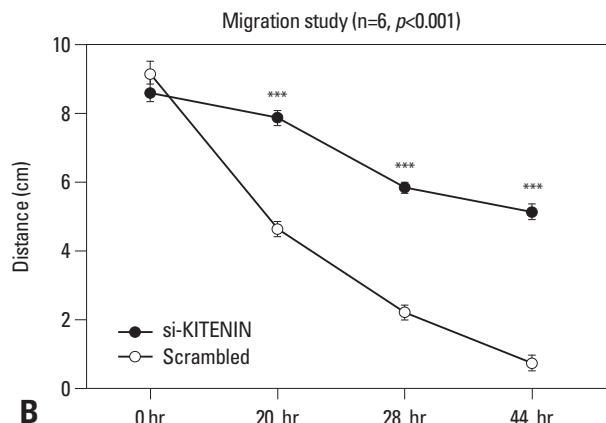


**Fig. 2.** Cell invasion assay: the number of invading si-KITENIN-transfected SNU-1041 cells (A) was  $64.2 \pm 19.2$ , whereas the number was  $98.6 \pm 10.2$  for the scrambled DNA-transfected SNU-1041 cells (B), as measured by 10 random squares of  $0.5 \times 0.5 \text{ mm}^2$  on the microscopic field of view under conditions with 20  $\mu\text{g}/\text{mL}$  of fibronectin; the difference between the two groups was statistically significant ( $p < 0.001$ )(C).

was  $98.6 \pm 10.2$ , as measured by 10 random squares of  $0.5 \times 0.5 \text{ mm}^2$  by the microscopic field of view under conditions with  $20 \mu\text{g/mL}$  of fibronectin; the difference between the two groups was statistically significant ( $p < 0.001$ ).

### Cell migration assay

The artificial wound gap became significantly narrower in

**A**

**Fig. 3.** Cell migration assay: the artificial wound gap became significantly narrower in the control group as time passed by 20, 28, 44 hours, compared to the si-KITENIN group ( $p < 0.001$ ) (B). In the scrambled group, the gap was nearly filled within 44 hours. However, it remained wide open in the si-KITENIN group (A).

the control group as time passed by 20, 28, 44 hours, compared to the si-KITENIN group ( $p < 0.001$ ) (Fig. 3). For the scrambled group, the gap was nearly filled within 44 hours, however, it remained wide open in the si-KITENIN group.

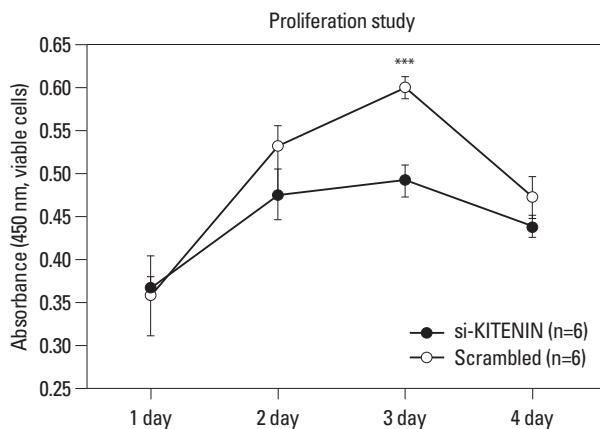
### Cell proliferation assay

The number of proliferating cells, as determined by absorbance, was significantly decreased in the si-KITENIN group compared to the scrambled group (Fig. 4). On the third day, absorbance was  $0.4933 \pm 0.0484$  in the si-KITENIN group, whereas  $0.6023 \pm 0.0319$  in the scrambled group ( $p < 0.001$ ).

## DISCUSSION

siRNA, known as RNAi, is a powerful tool for strong and specific suppression of gene expression.<sup>16</sup> The use of these molecules can be applied to a wide range of cancers and other proliferative disorders where aberrant gene expression occurs. Therefore, oncogenic and mutant tumor suppressor genes might represent potential targets for the RNAi approach. It has been shown that mutated p53 protein, involved in almost half of human malignancies, was eliminated by siRNA and the function of wild-type p53 restored.<sup>20</sup> Other examples include: elimination of the Ras oncogene using siRNAs,<sup>21</sup> transfection of leukemic cells with siRNAs targeting a BCR-ABL fusion transcript, thereby inducing apoptosis,<sup>22,23</sup> and targeting of VEGF by siRNAs.<sup>24</sup>

In this study, siRNA targeting KITENIN was effective. SNU-1041, an established human head and neck squamous carcinoma cell line,<sup>17-19</sup> expressed very high levels of KIT-



**Fig. 4.** Cell proliferation assay: the proliferating cells, as determined by absorbance, were significantly decreased in the si-KITENIN group compared to the scrambled group. On the third day, absorbance was  $0.4933 \pm 0.0484$  in the si-KITENIN group and  $0.6023 \pm 0.0319$  in the scrambled group ( $p < 0.001$ ).

ENIN, which functions as a metastasis-promoting gene. Our previous data demonstrated that many human head and neck squamous cancer tissues expressed KITENIN, and that the level of KITENIN correlated with the disease stage. After transfection of si-KITENIN into SNU-1041 cells, KITENIN was no longer identified in the cells. siRNA-transfected SNU-1041 cells showed reduced *in vitro* invasion, migration, and proliferative characteristics compared to the cells in the control group.

siRNAs can be delivered in two ways: exogenous administration of synthetic siRNAs and endogenous expression of siRNA via plasmid or viral vectors.

Generally, synthetic siRNAs are delivered to cells in culture via liposome-based transfection reagents. In this study, we used the lipofectamine trasfection reagent (RNAiMAX, Invitrogen) and the experiments were successful. However, this delivery offers only immediate and/or short-term effects.<sup>25</sup> Therefore, siRNA delivery systems which can provide long-term biological effects are needed.

To overcome these problems, several research groups have shown that short hairpin siRNA can be produced from expression plasmids that contain promoters that are dependent on either RNA polymerase (pol) II or pol III.<sup>26-29</sup> Brummelkamp, et al.<sup>26</sup> developed a new expression vector system, pSUPER (suppression of endogenous RNA) that directs the synthesis of siRNA-like transcripts in mammalian cells, and Lee, et al.<sup>12</sup> used the pSUPER vector system to deliver KITENIN siRNA for efficient and stable suppression of KITENIN expression in established tumors of syngeneic mice. The latter observed marked inhibition of the growth of established tumors as well as suppression of distant metastases of colon cancer, following four weekly or semiweekly intravenous injections of pSUPER-KITENIN. In addition to plasmid vectors, adenoviral and lentiviral-based vectors have also been developed and used successfully.<sup>30,31</sup> For our next *in vivo* study with animals on anti-KITENIN delivery, plasmid or viral vector systems will be studied.

In conclusion, gene therapy using anti-KITENIN strategies may help or delay the progression (invasion, migration and proliferation) of head and neck squamous carcinoma.

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