

## Evaluation of a Rapid Lateral Flow Point-of-Care Test for Detection of *Cryptosporidium*

Molly E. Fleece,<sup>1</sup> Jack Heptinstall,<sup>2</sup> Shaila S. Khan,<sup>3</sup> Mamum Kabir,<sup>3</sup> Joel Herbein,<sup>2</sup>  
Rashidul Haque,<sup>3</sup> and William A. Petri Jr.<sup>1\*</sup>

<sup>1</sup>Department of Medicine, University of Virginia, Charlottesville, Virginia; <sup>2</sup>TechLab, Inc., Blacksburg, Virginia;  
<sup>3</sup>International Centre for Diarrhoeal Disease Research, Bangladesh (icddr), Dhaka, Bangladesh

**Abstract.** A new rapid lateral flow fecal antigen detection test for *Cryptosporidium* was evaluated using diarrheal stool samples from a cohort of children in Bangladesh. The test had a sensitivity of 100% and a specificity of 94% when compared with enzyme-linked immunosorbent assay antigen detection.

Diarrheal diseases are a major cause of morbidity and mortality in the world.<sup>1,2</sup> *Cryptosporidium* is an enteric protozoan parasite that is transmitted through the fecal-oral route, typically by consumption of contaminated food and water.<sup>3,4</sup> The parasite has a low infectious dose resulting in diarrhea and abdominal pain.<sup>4,5</sup> *Cryptosporidium* is a common cause of waterborne diarrheal disease worldwide, both in developing and developed countries as well as urban and rural areas; however, due to poor sanitation and urban crowding, *Cryptosporidium* infections are more prevalent in underdeveloped areas.<sup>1,4</sup> Although infections do occur in immunocompetent hosts, immunocompromised hosts and children tend to have a more severe and prolonged disease course.<sup>4–6</sup> There is need for a practical point-of-care diagnostic test that is rapid, reliable, and feasible for use in the field.

*Cryptosporidium* lateral flow (TechLab, Inc., Blacksburg, VA) is a newly developed immunochromatographic assay that qualitatively detects *Cryptosporidium* antigen in fecal specimens. It is a dipstick that uses a monoclonal antibody sandwich design to detect *Cryptosporidium* oocyst wall antigen. The assay flow begins with a diluted specimen that is drawn up via capillary action, the liquid fraction of which liberates membrane-embedded gold particles conjugated with anti-*Cryptosporidium* antigens. This mixture then flows to the visible reaction window where additional anti-*Cryptosporidium* antibodies are immobilized and capture antigen–gold complexes for a visual positive result.

The data presented here are of the first field test of the *Cryptosporidium* lateral flow focusing on the sensitivity and specificity of this rapid dipstick test. All diarrheal stool samples were collected from a cohort of children living in an urban slum in Bangladesh where *Cryptosporidium* is prevalent.<sup>7</sup> The specimens were tested at the International Centre for Diarrhoeal Disease Research, Bangladesh. The samples were stored on average for 2 years at  $-20^{\circ}\text{C}$  until testing in batches. Real-time polymerase chain reaction (PCR) testing had been performed on all diarrheal stool samples before this study.<sup>8</sup> As a comparison of measurement of the presence/absence of *Cryptosporidium* antigen, enzyme-linked immunosorbent assays (ELISAs) were performed using the *Cryptosporidium II* test (TechLab, Inc.). The lateral flow was tested on 50 diarrheal stool samples known to be *Cryptosporidium* positive by PCR and 50 negative diarrheal stool samples. In addition, 100 randomly

selected diarrheal stool specimens from children 6–12 months of age were tested using *Cryptosporidium* lateral flow and compared with the results of PCR testing.

Fecal samples were brought to room temperature and mixed thoroughly before beginning the test. Fifty microliters of specimen were transferred via pipette into the specimen dilution tube containing diluent (buffered protein solution). The sample end of a test strip was inserted into the specimen dilution tube. Results were read visually after 10 minutes. A sample was interpreted as positive if both test and control lines were present (Figure 1). The color of the lines ranged from dark red to light pink, recognizing that color intensity did not correlate with strength of positivity. A sample was interpreted as negative if only the control line was visible. The test was considered invalid if the control line was absent.

We first tested 50 diarrheal stool samples known to contain *Cryptosporidium* DNA by PCR and 50 negative controls. Using ELISA as the reference standard for antigen detection, the *Cryptosporidium* lateral flow had a sensitivity of 100%, 94% specificity, 89% positive predictive value, and 100%

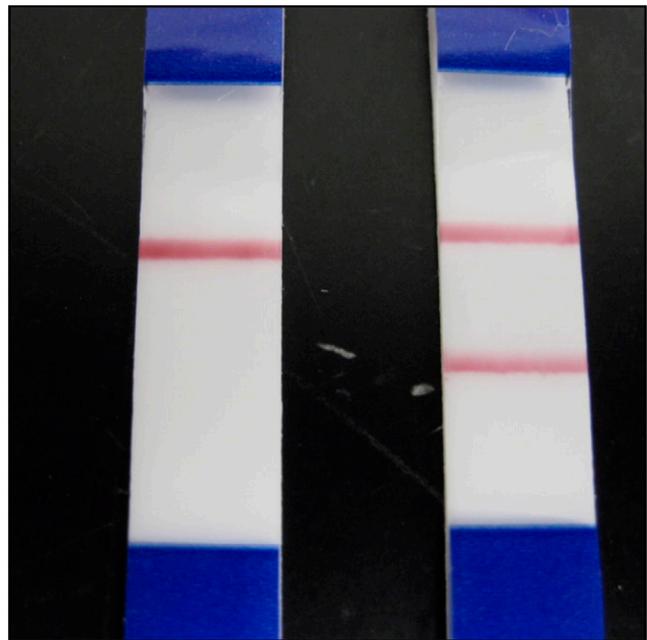


FIGURE 1. Lateral flow test for the detection of *Cryptosporidium* in stool specimens. The lateral flow on the left is an example of a negative test result where only the control line (upper) is positive. A positive test result is shown on the right with both the control and test lines visible.

\*Address correspondence to William A. Petri Jr., Infectious Diseases and International Health, 1709A Carter-Harrison Bldg., University of Virginia, Charlottesville, VA 22908-1340. E-mail: wap3g@virginia.edu

TABLE 1

Comparison of the *Cryptosporidium* lateral flow to the *Cryptosporidium II* ELISA for diarrheal stool samples

Assay type	<i>Cryptosporidium II</i> ELISA (+)	<i>Cryptosporidium II</i> ELISA (-)
Lateral flow (+)	34	4
Lateral flow (-)	0	62

ELISA = enzyme-linked immunosorbent assay.

negative predictive value (Table 1). Three of the four discrepant specimens (i.e., that were positive by *Cryptosporidium* lateral flow and negative by *Cryptosporidium II* test) were confirmed negative via PCR (with the fourth PCR positive).

We also evaluated the field adaptability of the lateral flow by testing 100 randomly selected diarrheal stool samples from the same cohort in Bangladesh, the vast majority of which did not have *Cryptosporidium*. There were no false positives: none of the 96 *Cryptosporidium*-negative samples had a positive lateral flow result. Of the four diarrhea samples with detectable *Cryptosporidium* DNA by PCR, the *Cryptosporidium* lateral flow detected one true positive sample with a  $C_t$  value 31.5. The three PCR (+) samples that were not detected by the lateral flow were most likely true negatives (i.e., *Cryptosporidium* was not the cause of diarrhea), as they had substantially lower amounts of *Cryptosporidium* DNA ( $C_t$  values of 35.2, 36.2, and 38.0). It has previously been shown that the strength of association of PCR (+) samples with diarrhea increases at higher pathogen loads.<sup>9</sup>

Available alternative rapid antigen detection dipstick tests include the Crypto Uni-Strip (Coris BioConcept, Gembloux, Belgium), RIDA QUICK *Cryptosporidium* (R-Biopharm, Darmstadt, Germany), and Crypto + *Giardia* dipstick (CLONIT, Milano, Italy).<sup>10-14</sup> All these tests have comparable time to results and easy visual result interpretation; however, the other available rapid antigen detection tests above involve at least one additional step in comparison to the *Cryptosporidium* lateral flow test. We concluded that the *Cryptosporidium* lateral flow has a comparable sensitivity and specificity to the *Cryptosporidium II* ELISA and is rapid, reliable, and easy to use in the field.

Received February 20, 2016. Accepted for publication May 25, 2016.

Published online August 29, 2016.

Financial support: This work was supported by NIH grant 5R01 AI043596 to William A. Petri Jr. William A. Petri Jr. is a consultant for TechLab, Inc.

Authors' addresses: Molly E. Fleece and William A. Petri Jr., Division of Infectious Diseases and International Health, Department of Medicine, University of Virginia, Charlottesville, VA, E-mails: mef8w@hscmail.mcc.virginia.edu and wap3g@virginia.edu. Jack Heptinstall and Joel Herbein, Research and Development, Techlab, Inc., Blacksburg, VA, E-mails: jheptinstall@techlab.com and jherbein@techlab.com. Shaila S. Khan and Rashidul Haque, International Centre for Diarrhoeal Disease Research, Bangladesh (icddr,b), Dhaka, Bangladesh, E-mails: skhan@icddr.org and rhaque@icddr.org. Mamun Kabir, Parasitology,

International Centre for Diarrhoeal Disease Research, Bangladesh (icddr,b), Dhaka, Bangladesh, E-mail: mamunk@icddr.org.

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## REFERENCES

- Fayer R, Ungar BLP, 1987. *Cryptosporidium* spp. and cryptosporidiosis. *Pediatr Infect Dis J* 6: 879.
- Liu L, Johnson HL, Cousens S, Perin J, Scott S, Lawn JE, Rudan I, Campbell H, Cibulskis R, Li M, Mathers C, Black RE, 2012. Global, regional, and national causes of child mortality: an updated systematic analysis for 2010 with time trends since 2000. *Lancet* 379: 2151–2161.
- Fayer R, Morgan U, Upton SJ, 2000. Epidemiology of *Cryptosporidium*: transmission, detection and identification. *Int J Parasitol* 30: 1305–1322.
- Chappell CL, Okhuysen PC, Langer-Curry R, Widmer G, Akiyoshi DE, Tanriverdi S, Tzipori S, 2006. *Cryptosporidium hominis*: experimental challenge of healthy adults. *Am J Trop Med Hyg* 75: 851–857.
- DuPont HL, Chappell CL, Sterling CR, Okhuysen PC, Rose JB, Jakubowski W, 1995. The infectivity of *Cryptosporidium parvum* in healthy volunteers. *N Engl J Med* 332: 855–859.
- Guerrant DL, Moore SR, Lima AA, Patrick PD, Schorling JB, Guerrant RL, 1999. Association of early childhood diarrhea and cryptosporidiosis with impaired physical fitness and cognitive function four–seven years later in a poor urban community in northeast Brazil. *Am J Trop Med Hyg* 61: 707–713.
- Mondal D, Haque R, Sack RB, Kirkpatrick BD, Petri WA Jr, 2009. Attribution of malnutrition to cause-specific diarrheal illness: evidence from a prospective study of preschool children in Mirpur, Dhaka, Bangladesh. *Am J Trop Med Hyg* 80: 824–826.
- Haque R, Roy S, Siddique A, Mondal U, Rahman SMM, Mondal D, Houpt E, Petri WA Jr, 2007. Multiplex real-time PCR assay for detection of *Entamoeba histolytica*, *Giardia intestinalis*, and *Cryptosporidium* spp. *Am J Trop Med Hyg* 76: 713–717.
- Liu J, Kabir F, Manneh J, Lertsethakarn P, Begum S, Gratz J, Becker SM, Operario DJ, Taniuchi M, Janaki L, Platts-Mills JA, Haverstick DM, Kabir M, Sobuz SU, Nakjarung K, Sakpaisal P, Silapong S, Bodhidatta L, Qureshi S, Kalam A, Saidi Q, Swai N, Mujaga B, Maro A, Kwambana B, Dione M, Antonio M, Kibiki G, Mason CJ, Haque R, Iqbal N, Zaidi AK, Houpt ER, 2014. Development and assessment of molecular diagnostic tests for 15 enteropathogens causing childhood diarrhoea: a multicentre study. *Lancet Infect Dis* 14: 716–724.
- Llorente M, Clavel A, Varea M, Olivera S, Castillo F, Sahagún J, Rubio M, Gómez-Lus R, 2002. Evaluation of an immunochromatographic dip-strip test for the detection of *Cryptosporidium* oocysts in stool specimens. *Eur J Clin Microbiol Infect Dis* 21: 624–625.
- Coris BioConcept Inc., 2012. *Crypto-Strip* (package insert). Gembloux, Belgium: Coris BioConcept, Inc.
- Hawash Y, 2014. Evaluation of an immunoassay-based algorithm for screening and identification of *Giardia* and *Cryptosporidium* antigens in human faecal specimens from Saudi Arabia. *J Parasitol Res* 2014: 213745.
- R-Biopharm Inc., 2010. *RIDA QUICK Cryptosporidium* (package insert). Darmstadt, Germany: R-Biopharm Inc.
- CLONIT, 2012. *CRYPTO + GIARDIA dipstick* (package insert). Milano, Italy: CLONIT Inc.