

The predictive value of stimulation index calculated by modified mixed lymphocyte culture in the detection of GVHD following hematopoietic stem cell transplantation

Hematopoetik kök hücre naklini takiben gelişen GVHD'nin saptanmasında modifiye mikst lenfosit kültür testinde hesaplanan stimülasyon indeksinin belirleyici değeri

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Abstract

Objective: Mixed lymphocyte culture (MLC) is one of the routine tests performed prior to hematopoietic stem cell transplantation (HSC) as a predictive assay for assessing the quality of donor matching and graft-versus-host disease (GVHD). The stimulation index is one of the formulas of the MLC test, and it is used for evaluation of matching between donor and recipient. Modified MLC (mMLC) test is produced by adding various cytokines to the MLC test, and increased sensitivity has been reported with this modification.

Materials and Methods: The importance of the stimulation index values in MLC and mMLC tests was evaluated in 59 patients who received HSCs from human leukocyte antigen-identical sibling donors. In the mMLC test, cytokines were added as interleukin (IL)-2, IL-2 + IL-4 and IL-2 + interferon (IFN)-gamma + tumor necrosis factor (TNF)-alpha. Stimulation index values in mMLC test were compared with stimulation index values in MLC test.

Results: Twenty-three (39%) patients developed GVHD. When evaluated in terms of stimulation index >1 patients, in MLC, 55% of the patients developed GVHD (p=0.229), whereas these values were 75% in the IL-2 added mMLC test (p=0.035), 100% in the IL-2 + IL-4 added mMLC test (p=0.076) and 85.7% in the IL-2 + IFN-gamma + TNF-alpha added mMLC test (p=0.015).

Conclusion: mMLC increased the sensitivity of the test. The relation between the positive results and evidence of GVHD after transplantation was found significant. (*Turk J Hematol 2010; 27: 263-8*)

Key words: Stimulation index, mixed lymphocyte culture, hematopoietic stem cell transplantation, cytokines

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Özet

Amaç: Mikst lenfosit kültür testi hematopoetic kök hücre naklinden önce, donör uyumunu ve graft versus host hastalığını önceden belirlemek amacı ile yapılan bir testtir. Stimülasyon indeksi mikst lenfosit kültür testinde kullanılan formüllerden biridir ve alıcı ile verici arasındaki uyumu belirlemek için yapılır. Modifiye mikst lenfosit kültür testi ise mikst lenfosit kültür testine çeşitli sitokinlerin ilave edilmesiyle yapılır ve yapılan bu değişiklikle duyarlılıkta artış olduğu saptanmıştır.

Yöntem ve Gereçler: HLA uyumlu donörden hematopoetik kök hücre nakli yapılacak 59 hastada Mikst Lenfosit Kültür ve Modifiye Mikst Lenfosit Kültürdeki stimülasyon indeksi değerlerine bakılmıştır. Modifiye mikst lenfosit kültür testinde sitokinler interlökin-2, interlökin-2 + interlökin-4 ve interlökin-2 + interferon-gama + tümör nekrozis faktör-alfa şeklinde eklenmiş ve modifiye mikst lenfosit kültüründeki stimülasyon indeksi değerleri, mikst lenfosit kültüründeki stimülasyon indeksi değerleriyle karşılaştırılmıştır.

Bulgular: Yirmi üç hastada (%39) graft versus host hastalığı oluşmuştur. Mikst lenfosit kültürde stimülasyon indeksi >1 olan hastalara bakıldığında hastaların %55'inde graft versus host hastalığı görülmüştür (p:0,229). Bunun yanı sıra interlökin-2 ilavesi ile yapılan modifiye mikst lenfosit kültür testinde stimülasyon indeksi >1 olan hastaların %75'inde (p=0,035); interlökin-2 + interlökin-4 ilavesi ile yapılan modifiye mikst lenfosit kültür testinde stimülasyon indeksi >1 olan hastaların %100'ünde (p=0,076) ve interlökin-2 + interferon-gama + tümör nekrozis faktör-alfa ilavesi ile yapılan modifiye mikst lenfosit kültür testinde stimülasyon indeksi >1 olan hastaların %85,7'sinde (p=0,015) graft versus host hastalığı görülmüştür.

Sonuç: Modifiye mikst lenfosit kültür, testin duyarlılığını artırmıştır ve pozitif sonuçlar ile transplantasyondan sonra graft versus host hastalığının gelişmesi arasındaki ilişki anlamlı bulunmuştur.

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Anahtar kelimeler: Stimülasyon indeksi, mikst lenfosit kültür, hematopoetik kök hücre nakli, sitokinler

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Introduction

Receiving a hematopoietic stem cell transplantation (HSCT) from a matched donor is a lifesaving treatment modality in some diseases. The degree of human leukocyte antigen (HLA) donor/recipient match has a significant association with graft-versus-host disease (GVHD). To date, donor selection has been based on match for the antigens encoded by the HLA class I (A, B, C) and class II (DR) loci. Unrecognized or undefined mismatch for class II genes between the donor and recipient can exist despite a match for HLA-A, B, C and DR and could contribute to the high risk of GVHD after transplantation. Mixed lymphocyte culture (MLC) test is a method used in determination of class II antigens that evaluates cell proliferation based on compatibility of HLA antigens between the recipient and donor. MLC is a cellular test that exhibits the appropriateness of HLA antigens of the donor/recipient. One of the formulas used for this evaluation is the stimulation index (SI) [1,2]. However, the MLC test fails to detect the minor histocompatibility antigens contributing to GVHD and tissue rejection and also the response stimulated by HLA-DP. On the other hand, it is reported that GVHD and tissue rejection could be predicted by cytokine-modified mixed lymphocyte culture (mMLC) test [3-6]. Cytokines play a critical role after allogeneic recognition in the

MLC. The MLC is not only a clinical method in HSCT but also an important model to show T cell activation and cytokine interaction following alloantigen recognition. The T helper 1 cytokine interferon-gamma (IFN- γ) is known to induce cytotoxic T lymphocytes (CTL) by enhancing the expression of both HLA class I and class II molecules. Synergistic effects of interleukin-2 (IL-2) as well as tumor necrosis factor-alpha (TNF- α) on the production of IFN- γ are supposed, whereas IFN- γ itself also shows a stimulatory influence on the expression of the IL-2 receptor in T lymphocytes [7].

Interleukin-4 (IL-4) is the developmental factor for B-lymphocytes and among the molecules that establishes association with HLA class II products [5,6]. TNF- α activates leukocytes, particularly neutrophils [8]. In both murine and human MLC reactions, TNF- α enhances the proliferative response [9].

In this study, cytokine-spiked mMLC test in addition to the MLC test were administered to patient-donor pairs. In mMLC, cytokines were added to the test individually or in combination. The probability of GVHD was thus determined by stimulating the antigens that were not exhibited in MLC, with an *in vivo* test. The association of SI results obtained from the MLC and mMLC tests with GVHD development was investigated. The relation between mMLC results and grade or organ involvement of GVHD was not considered.

Materials and Methods

MLC and mMLC tests were performed in a total of 59 full match-related, recipient and donor pairs at Istanbul University Medical Faculty, Medical Biology Department. These cases were received from eight different centers. Written informed consents were obtained from all patients or their parents. This study was approved by local ethics committee.

Allogeneic HSCT was performed from full match-related donors, and bone marrow was used as HSC sources for each patient.

The conditioning regimen was changed according to the underlying disease and was mainly with busulfan and cyclophosphamide (BU/CY). In patients with multiple myeloma, the regimen consisted of high-dose melphalan. In patients with hemoglobinopathy and aplastic anemia, antithymocyte globulin (ATG) was incorporated into the regimen.

GVHD prophylaxis consisted of cyclosporin A (CsA) + short course methotrexate (MTX) in all cases.

The diagnostic tests for GVHD usually depended on the symptoms, but could include: gastrointestinal endoscopy, with or without a biopsy, liver function tests (aspartate aminotransferase [AST], alkaline phosphatase [ALP] and bilirubin levels are increased), liver biopsy (if the patient only has liver symptoms), lung X-rays, and skin biopsy [10,11].

The MLC test was administered in one-way and two-stage manner (recipient and donor) in the culture laboratory of the department. In this study, we evaluated donor-directed. The peripheral blood mononuclear cells of the recipient and donor pairs were obtained via Ficoll Hypaque gradient centrifuge method. The lymphocytes obtained were washed three times with RPMI 1640 (with glutamine, Hepes, Sigma-50 u/ml penicillin and 50 µg/ml streptomycin) consequently followed by counting the cells in a medium containing RPMI 1640 and human serum (9:1). Donor lymphocytes were used at a final concentration of 1×10^6 cell/ml. Recipient lymphocytes were used at a final concentration of 2×10^6 cell/ml. The recipient cells were irradiated at 5000 rad (cGy). Donor cells (100 µl) and irradiated

recipient cells (50 µl) were added to the culture plates. All experiments were performed in triplicate. Results were normalized and presented as means. Cytokines were added in the same way in addition to MLC test, by producing cell batch (IL-2: 2.4 U/µl, IL-4: 14.5 U/µl, IFN-γ: 200 U/µl, TNF-α: 100 U/µl) (Table 1).

³H Thymidine was placed in each well after the cells were incubated in 96-well plates for 96 hours (h) at 37°C and 5% CO₂. All the cells were collected after 16-18 h. Cultures were harvested onto glass fiber filters. Glass fiber filters are dried at room temperature. The numeric information is defined in β counter (Packard Tricarb 1000 TR) by placing filter papers in scintillation solution (toluol + 2,5 diphenyl oxazole). Blank values are deducted from the measurements. Sample filters were counted using β counter. SI formula was used for evaluation of the results. On the MLC test, a SI value ≤1 indicates that the donor is not reactive to the recipient.

$$SI = DR^* / DD$$

Statistical analyses were performed using SPSS 10.0 statistical software program. The relationship between development of GVHD and MLC/mMLC was analyzed with Fisher's exact and Independent Samples T Test.

Results

The mean age of the 59 patients who underwent MLC test and HSCT was 19.91 ± 1.96 (1-51) and the mean age of the donors was 20.61 ± 1.91 (1-51). Forty-one percent of the patients were female (n=24) and 59% were male (n=35). The median age was 21.50 ± 2.88 (1-45) for the female patients and 20.51 ± 2.53 (1-51) for the males. Twelve female recipients received stem cells from a female donor and 14 from a male donor, whereas 17 of the male recipients received stem cells from a female donor and 16 from a male donor. Eighteen patients were previously diagnosed with chronic myeloid leukemia (CML) and the rest of the group were as follows: 15 acute myeloid leukemia (AML), 9 aplastic anemia, 7 acute lymphoid

Table 1. Localization of the cells on the culture plate on the MLC and mMLC test

MLC	MLC	mMLC	mMLC
Donor-Donor (DD)	Donor-Recipient* (DR)	DD + IL-2	DR* + IL-2
		DD + IL-2 + IL-4	DR* + IL-2 + IL-4
		DD + IL-2 + IFN-γ + TNF-α	DR* + IL-2 + IFN-γ + TNF-α

* irradiated cells

leukemia (ALL), 3 thalassemia major, 3 multiple myeloma (MM), 2 non-Hodgkin lymphoma (NHL), 1 myelodysplastic syndrome (MDS), and 1 sickle cell anemia (Table 2).

Cytokine-spiked mMLC test was administered to recipient and donor pairs with an adequate number of cells.

GVHD developed in 23 (39%) of the 59 patients who underwent HSCT. Acute and chronic GVHD were not classified or analyzed separately. The evaluation of the MLC test for SI in 59 patients

revealed SI values ≤ 1 in 50 patients, while 9 patients had SI values > 1 . In patients with SI values ≤ 1 , after transplantation, GVHD was detected in 18 patients and not detected in 32 patients. In the group with SI > 1 , 4 had no complications while 5 had GVHD. The statistical evaluation of the association between the SI values and GVHD development revealed no significance ($p=0.229$) (Table 3).

mMLC test was performed by adding IL-2 in 53 of these patients. While 45 patients revealed SI values ≤ 1 , 8 patients had SI values > 1 . While no GVHD was observed in 30 of the patients with SI ≤ 1 , GVHD occurred in 15 patients. Six of the 8 patients with SI > 1 developed GVHD while 2 patients did not. The statistical evaluation of the relation between SI values and GVHD development revealed a statistically significant result ($p=0.035$) (Table 3).

mMLC test was performed by adding IL-2+IL-4 in 24 of these patients. While 22 patients revealed SI values ≤ 1 , 2 patients had SI values > 1 . While no post-transplant GVHD occurred in 17 of those patients with a SI ≤ 1 , 5 patients developed this disease. GVHD could be determined in 2 patients with SI > 1 . In the group spiked with IL-2+IL-4 cytokines, the statistical evaluation of the relation between SI values and GVHD development revealed a non-significant result ($p=0.076$) (Table 3).

mMLC test was performed by adding IL-2+IFN- γ +TNF- α in 51 of the patient-donor pairs. While 44 patients revealed SI values ≤ 1 , 7 patients had SI values > 1 . In 29 of those patients with SI ≤ 1 , no post-transplant GVHD occurred, while 15 patients experienced GVHD. While it was not possible to determine GVHD in 1 patient with SI > 1 , the disease had developed in the remaining 6 of those 7 patients. There was a statistically significant relation between SI values and GVHD development ($p=0.015$) (Table 3).

Table 2. Patient characteristics

	n	%
Age		
≤ 15 (Pediatric Group)		
Male	16	27
Female	11	19
15-51 (Adult Group)		
Male	19	32
Female	13	22
Gender		
Male	35	59
Female	24	41
Malignant Diseases		
CML	18	31
AML	15	25
Aplastic Anemia	9	15
ALL	7	12
Thalassemia Major	3	5
Multiple Myeloma	3	5
NHL	2	3
MDS	1	2
Sickle Cell Anemia	1	2

Table 3. GVHD status and SI values calculated by MLC, MLC+IL-2, MLC+IL-2+ IL-4, and MLC+IL-2+IFN- γ + TNF- α

		MLC			MLC + IL-2			MLC + IL-2 + IL-4			MLC+IL-2+IFN- γ + TNF- α		
		SI ≤ 1	SI > 1	Total	SI ≤ 1	SI > 1	Total	SI ≤ 1	SI > 1	Total	SI ≤ 1	SI > 1	Total
GVHD (-)	n	32	4	36	30	2	32	17	0	17	29	1	30
	%	64.0	44.4	(61.0)	66.7	25	(60.4)	77.3	0	(70.8)	65.9	14.3	(58.8)
GVHD (+)	n	18	5	23	15	6	21	5	2	7	15	6	21
	%	36.0	55.6	(39.0)	33.3	75	(39.6)	22.7	100	(29.2)	34.1	85.7	(41.2)
Total	n	50	9	59	45	8	53	22	2	24	44	7	51
	%	100	100	(100)	100	100	(100)	100	100	(100)	100	100	(100)
Fisher's exact test		p=0.229			p=0.035			p=0.076			p=0.015		

When the mean SI values obtained on the MLC and mMLC tests were evaluated with respect to GVHD development, the mean SI value of the 36 of the 59 patients undergoing MLC test who did not develop GVHD was 0.63 ± 0.14 , while the mean SI value in the 23 patients who developed GVHD was 0.71 ± 0.10 ; the difference between the two groups was not significant ($p=0.956$) (Table 4).

The mean SI value of the 32/53 patients undergoing IL-2-spiked mMLC test who did not develop GVHD was 0.52 ± 0.07 while this value was 1.01 ± 0.25 in the 21/53 patients who did develop GVHD; the difference was statistically significant ($p=0.008$) (Table 4).

The mean SI value of the 17/24 patients undergoing IL-2+IL-4-spiked mMLC test who did not develop GVHD was 0.59 ± 0.05 , while this value was 2.44 ± 1.87 in the 7/24 patients who did develop GVHD; the difference was statistically significant ($p=0.001$) (Table 4).

The mean SI value of the 30/51 patients undergoing IL-2 + IFN- γ + TNF- α -spiked mMLC test who did not develop GVHD was 0.41 ± 0.05 , while this value was 3.99 ± 2.51 in the 21/51 patients who did develop GVHD; the difference was statistically significant ($p=0.005$) (Table 4).

Discussion

Despite the technological and scientific advances in pre-transplant tissue typing tests and the fact that HSCT is performed between HLA-identical pairs, GVHD still represents the most significant

complication occurring after transplantation [12]. The possible reasons for this are thought to be polymorphic HLA determinants and/or minor histocompatibility systems not yet detected [6,13].

In the present study, the results from the MLC and cytokine-spiked mMLC tests were compared with regards to GVHD development in HSCT recipients. The association between GVHD development and SI values, as ≤ 1 or > 1 , was investigated.

In mMLC, cell surface antigens were increased by providing treatment of stimulator cells with cytokines (IL-2, IL-4, TNF- α , IFN- γ) that are known to increase the expression of HLA and non-HLA antigens. By adding exogenous cytokines to MLC cultures, amplification of weak proliferative responses was achieved. Using these amplifications, positive MLC reactions were frequently achieved amongst HLA-identical siblings [6,7,14].

In a study by Bishara et al. [9], pre-determination of GVHD and tissue rejection was targeted by modifying the one-way MLC test. Three separate modifications were used in this study. In the first modification, IL-1 α , IL-2 and IL-4 were added separately and in combination to the MLC test. Addition of IL-2 and IL-4 increased the MLC response in all unpaired controls and in certain HLA-identical pairs. The second modification was made by pre-treatment of the stimulator cells with IFN- γ , TNF- α and IL-4. This modification resulted in positive response in all the cases. The third modification was performed by re-addition of cytokines to the MLC test where stimulator cells were pre-treated with added cytokines. As a result of this combined application, a high rate of positivity was also detected among the HLA-identical pairs. It was concluded that this result could be used in determining undetectable minor antigenic differences. The Bishara study reported that the mMLC study could be beneficial in choosing the most compatible donor in the presence of multiple HLA-identical donors.

In the study by Visentainer et al. [15], exogenous cytokines were added to the one-way MLC test. The investigators, thus intending to increase the sensitivity of the MLC test, stimulated the stimulator cells by IL-4 or IFN- γ and added IL-2 or IL-4 to the responding cells at the start of the culture. By addition of different doses of cytokine in the autologous cultures and compatible recipient-donor cultures, advantageous results were obtained as compared to the MLC test. Their study revealed that pre-treatment of stimulator cells with IL-4 or IFN- γ did not increase allogeneic response in the MLC test; however, IL-2 and IL-4

Table 4. The results for mean MLC, MLC+IL-2, MLC+IL-2+IL-4 and MLC+IL-2+IFN- γ +TNF- α SI values and GVHD

		GVHD (-)	GVHD (+)	p
MLC	n	36	23	0.956
	%	61	39	
	SI	0.63 ± 0.14	0.71 ± 0.10	
MLC + IL-2	n	32	21	0.008
	%	60.4	39.6	
	SI	0.52 ± 0.07	1.01 ± 0.25	
MLC + IL-2 + IL-4	n	17	7	0.001
	%	70.8	29.2	
	SI	0.59 ± 0.05	2.44 ± 1.87	
MLC + IL-2 + IFN- γ + TNF- α	n	30	21	0.005
	%	58.8	41.2	
	SI	0.41 ± 0.05	3.99 ± 2.51	

addition at the start of the culture increased all the responses including the autologous response.

Another trial by Visentainer et al. [15] investigated the association between chronic GVHD occurring after SCT and the one-way MLC test. They found in that study that the MLC test was not adequate for pre-determining acute GVHD; however, it could be the predictor of chronic GVHD when a result above Relative Response Index (RRI)=4.5% was achieved.

As could be understood from the study conducted, the objective of modification of the MLC test is to predict post-transplant GVHD and tissue rejection. In our study, in addition to the MLC test, three different series were formed with addition of different cytokines. In the first series, IL-2 was added separately to the MLC test. In the second series, IL-2+IL-4 were added. In the third series, IL-2+IFN- γ +TNF- α were added. In the mMLC tests, an increase was observed in the SI values. GVHD developed in 39% of the patients undergoing HSCT. GVHD was detected in 55.6% of the group with SI > 1 in the MLC test, while these figures were 75.6%, 100% and 85.7%, respectively, in the series spiked with IL-2, IL-2+IL-4 and IL-2+IFN- γ +TNF- α [15].

When the mean SI values obtained on the MLC and mMLC tests were evaluated with respect to GVHD development, in the cytokine-spike groups, the statistical evaluation of the relation between mean SI values and GVHD development revealed significant results.

In conclusion, the mMLC test sensitized by addition of IL-2 and IL-2+IFN- γ +TNF- α cytokines is important in determining post-transplant GVHD. We believe that studying the mMLC test could be beneficial in choosing the most compatible donor when multiple HLA-identical donors are present.

Conflict of interest

No author of this paper has a conflict of interest, including specific financial interests, relationships, and/or affiliations relevant to the subject matter or materials included in this manuscript.

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