

Combination of COX-2 expression and *PIK3CA* mutation as prognostic and predictive markers for celecoxib treatment in breast cancer

Sandrine Tury¹, Véronique Becette², Franck Assayag³, Sophie Vacher¹, Camille Benoist¹, Maud Kamal⁴, Elisabetta Marangoni³, Ivan Bièche¹, Florence Lerebours⁴, Céline Callens¹

¹Pharmacogenomic Unit, Genetics Laboratory, Institut Curie, Paris, France

²Department of Pathology, Institut Curie, Hôpital René Huguenin, Saint-Cloud, France

³Laboratory of Preclinical Investigations, Translational Research Department, Institut Curie, Paris, France

⁴Department of Medical Oncology, Institut Curie, Paris and Saint-Cloud, France

Correspondence to: Céline Callens, email: celine.callens@curie.fr

Keywords: breast cancer, *PIK3CA*, celecoxib, prognosis, predictive biomarker

Received: April 29, 2016

Accepted: October 26, 2016

Published: November 08, 2016

ABSTRACT

COX-2 expression level and prognostic value are still a matter of debate in breast cancer (BC). We addressed these points in the context of *PIK3CA* mutational status. Based on an interesting study of aspirin efficacy in colorectal cancer, we hypothesized that celecoxib antitumoral activity may be restricted to *PIK3CA* mutated BC.

COX-2 mRNA expression was analyzed in 446 BC samples and in 61 BC patient-derived xenografts (PDX) using quantitative RT-PCR. The prognostic impact of COX-2 expression level was assessed independently and according to *PIK3CA* mutational status in our cohort and in a validation set of 817 BC. The antitumoral activity of celecoxib was tested in two triple-negative (TN) PDX with a *PIK3CA* wild-type (wt) or mutated genotype.

COX-2 mRNA was overexpressed in 2% of BC and significantly associated with TN subtype. Metastasis-free survival (MFS) was significantly better in patients with high COX-2 expression level, the prognosis of whom was similar to patients with *PIK3CA* mutations. TCGA validation cohort confirmed that patients with low COX-2 expression *PIK3CA* wt tumors had the worse disease-free survival (DFS) compared to all other subgroups. Celecoxib had a significant antitumoral effect in *PIK3CA* mutated PDX only. Celecoxib antitumoral activity involved S6 ribosomal protein and AKT phosphorylation.

Low expression of COX-2 has a significant negative impact on the MFS/DFS of BC patients. Antitumoral effect of celecoxib is restricted to *PIK3CA* mutated PDX. These results suggest that *PIK3CA* mutation may be a new predictive biomarker for celecoxib efficacy.

INTRODUCTION

The cyclooxygenase-2 (COX-2) also known as the prostaglandin-endoperoxide synthase-2 (PTGS-2) is an inducible enzyme involved in inflammatory and oncogenic processes. It is responsible for the synthesis of prostaglandins from arachidonic acid [1] and is reported to induce the expression of aromatase in breast tissue [2, 3]. COX-2 expression level in breast carcinomas and normal breast tissue is not well established and

reports are controversial. COX-2 expression levels in ductal carcinoma *in situ* (DCIS) and invasive carcinoma were reported to be similar in a meta-analysis of COX-2 expression levels in breast cancers (BC). No clear conclusion on COX-2 expression levels in normal breast epithelium was however reported in the latter study [4]. A recent study using immunohistochemistry (IHC) assessed COX-2 expression on BC and adjacent normal tissues from 96 premenopausal women. COX-2 expression in the normal breast epithelium fluctuated (more than 40-fold)

among women and was correlated with COX-2 expression levels in DCIS and invasive cancer, independently of known prognostic features. The authors suggested that factors regulating physiological COX-2 expression may be the primary drivers of COX-2 expression in BC. Thus, baseline COX-2 expression level may be an indicator of BC risk, and predict chemo preventive and therapeutic efficacy of COX-2 inhibitors in young women [5]. The prognostic value of COX-2 is still debated. Several studies suggested that COX-2 is implicated in BC progression, where COX-2 overexpression was shown to be associated with poorer outcome. On the other hand, this negative prognostic impact may be counterbalanced by hormonal treatment [6–10].

Giving the putative prognostic role of COX-2, the potential therapeutic benefit of COX-2 inhibitors has been investigated. Celecoxib, a non-steroidal anti-inflammatory drug (NSAID), is a specific COX-2 inhibitor. Celecoxib acts mainly by decreasing the formation of downstream target proteins prostaglandin, prostacyclin or thromboxane involved in cell proliferation and angiogenesis. Celecoxib has thus been examined for its antitumoral properties [11, 12].

In vitro and *in vivo* studies have shown an antitumoral effect of celecoxib in BC. Celecoxib significantly decreased tumor volume by 32% in rats with chemically induced mammary tumor [13]. Celecoxib was also reported to significantly decrease tumor incidence rate and delayed tumor emergence in similar animal models [14].

Although preclinical data were optimistic, the clinical trials results testing the efficacy of celecoxib in BC patients were disappointing. The combination of celecoxib and aromatase inhibitors was tested in clinical trials since celecoxib may enhance aromatase inhibitors' efficacy. In DCIS, two studies have led to conflicting results [15, 16]. In a phase II trial for advanced BC women with progressive disease under tamoxifen, celecoxib in association with exemestane did not improve clinical outcome as compared to exemestane alone [17]. Another multicentric randomized phase II study of neoadjuvant epirubicin/cyclophosphamide followed by docetaxel (EC-D) with or without celecoxib showed that celecoxib is not likely to improve complete pathological response rates in addition to EC-D in patients with HER2-negative tumor [18]. A trial on 90 DCIS postmenopausal patients with ER-positive carcinoma showed that two weeks presurgical treatment with celecoxib alone or in combination with exemestane had no effect on proliferation or apoptose [15]. More recently, a monocentric phase II neoadjuvant trial in postmenopausal women with ER-positive DCIS (n=95) showed that concomitant administration of celecoxib and exemestane during 12 weeks induced a significant reduction in tumor cell proliferation and COX-2 expression. These results suggest that COX-2 high expression levels may be a predictive marker for

early relapse in patients with DCIS and may help in the clinical decision for treatment of DCIS [16]. Despite the different encouraging results, it remains to be established whether BC patients might actually benefit from celecoxib treatment.

In 2012 Liao *et al.* showed that aspirin, a non-selective COX inhibitor, increased overall survival in patients with colorectal cancer harboring an activating mutation in the *PIK3CA* gene. These results substantiate an interaction between the cyclooxygenase activity and the PI3K/AKT pathway [19]. Other studies confirmed the benefit of aspirin treatment on overall survival in *PIK3CA* mutated colorectal cancer [20]. As *PIK3CA* mutations are reported in 10-40% of BCs [21] we hypothesized that mutated-*PIK3CA* breast tumor expressing COX-2 could benefit from treatment with a COX-2 inhibitor such as celecoxib.

In the present study we first evaluated COX-2 expression levels and prognostic value according to the *PIK3CA* mutational status in a large retrospective cohort of BC patients. We then investigated the antitumoral effect of celecoxib depending on *PIK3CA* mutation in triple-negative patients-derived xenograft models (PDX). Finally, we assessed potential predictive biomarkers and secondary resistance mechanisms associated with celecoxib antitumoral properties.

RESULTS

COX-2 overexpression is rare and associated with TNBC

We first quantified the expression level of *COX-2* transcript by quantitative RT-PCR (qRT-PCR) in a cohort of 446 BC samples composed of 68 HR⁺ERBB2⁻, 42 HR⁻ERBB2⁺, 285 HR⁺ERBB2⁻ and 51 HR⁺ERBB2⁺ cases.

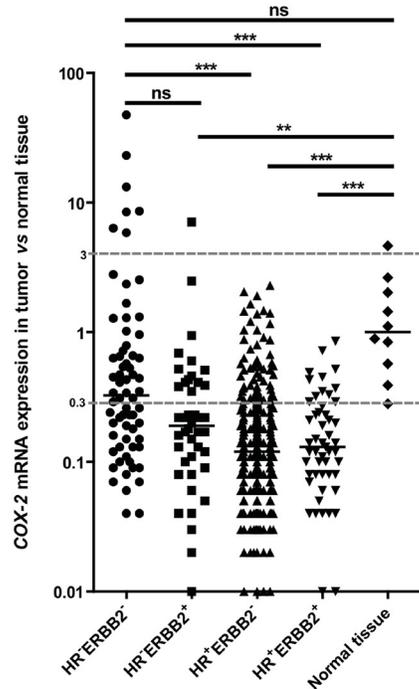
COX-2 transcript was underexpressed (relative expression <0.3 compared to normal tissue as detailed in material and method section) in 74% (332/446) and overexpressed (relative expression >3 compared to normal tissue) in 2% (8/446) of cases. By comparison with normal tissue *COX-2* mRNA relative expression was significantly higher in triple-negative subtype than in HR⁺ERBB2⁻ and in HR⁺ERBB2⁺ subtypes (Figure 1A). This result was identical when evaluating *COX-2* mRNA expression without normal tissue comparison (Supplementary Figure S1A). Considering the relative expression cut-off of 3, *COX-2* overexpression is strongly associated with the triple-negative subtype (10%, 7/68 tumors, p<0.0001)

COX-2 transcript was also quantified by qRT-PCR in 61 tumors collected on PDX (15 HR⁺, 6 ERBB2⁺ and 40 triple negative tumors). In PDX, the strongest expression levels of *COX-2* were found in triple-negative (median 36 [0–1673]) compared to luminal (median 0 [0–202]) (p=0.0006) and ERBB2 positive subtypes (median 6 [0–601]) (p=ns) (Supplementary Figure S1B).

COX-2 IHC staining in 26 primary tumors representative of our cohort revealed no labeling when the expression of *COX-2* transcript was inferior to 2 (relative expression compared to normal tissue) and the staining becomes more intense as the level of expression of transcript increases (Table 1 and examples are shown in Figure 1B).

The same observation was made in 14 PDX where COX-2 IHC staining was more intense when *COX-2* transcript was more expressed (Table 2). These data show a good correlation between the *COX-2* mRNA and the COX-2 protein expression levels except for two cases (2/26, 8%) of primary tumor (3395 and 5015). Technical difficulties

A



B

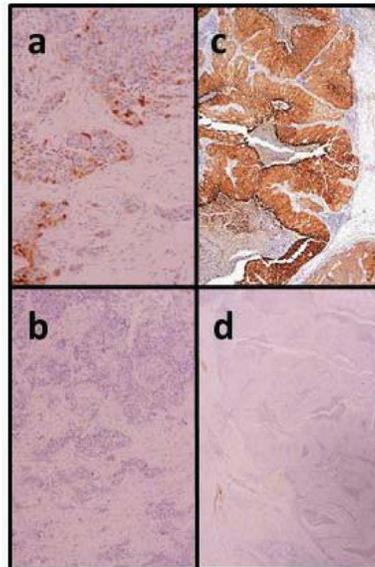


Figure 1: *COX-2* mRNA and protein expression in patient breast tumors. A. *COX-2* mRNA expression levels in 446 breast tumor samples (68 HR-ERBB2-, 42 HR-ERBB2+, 285 HR+ERBB2- and 51 HR+ERBB2+) and in 10 normal breast tissues using qRT-PCR. HR for hormone receptors. *COX-2* mRNA expression in breast tumor samples is expressed compared to *COX-2* mRNA expression in normal tissue. B. COX-2 protein expression in a mRNA *COX-2* expressing tumor, weak (a) and strong (c). b and d: the same tumors stained with the isotypic control antibody.

Table 1: Correlation between *COX-2* mRNA and *COX-2* protein expression level on 26 primary breast tumors

samples ID	<i>COX-2</i> mRNA expression	<i>COX-2</i> protein expression	positive cells (%)	staining intensity
4410	<0.05	negative	0	0
4207	<0.05	negative	0	0
5396	<0.05	negative	60	1
6645	0.03	negative	0	0
5461	0.04	negative	0	0
4393	0.20	negative	0	0
5470	0.20	negative	0	0
6605	0.79	negative	2	2
3395	1	positive (moderate)	25	3
6189	1	negative	80	1
6189 ^a	1			
6601	1	negative	80	1
6891	1	negative	0	0
5708	1.47	negative	0	0
2421	1.67	negative	0	0
2690	1.91	negative	0	0
6602 ^b	2.05			
5295	2.53	positive (moderate)	50	2 to 3
5295	2.53	positive (moderate)	20	2 to 3
5015	5.85	negative	0	0
6874	6.33	positive (moderate)	40	2 to 3
6889	7.07	positive (high)	70	2 to 3
6889	7.07	positive (moderate)	20	2 to 3
5392	8.46	positive (moderate)	<<1	3
2346	8.58	positive (high)	100	2 to 3
6876	47.31	positive (high)	70	2

^aCorresponding to ductal carcinoma *in situ* (DCIS) case.

^bCorresponding to neuroendocrine carcinoma case.

prevented us from accurately determine the percent of positive cells and intensity of the staining in particular histological types of breast carcinomas like ductal carcinoma *in situ* (6189), neuroendocrine carcinoma (6602) and metaplastic carcinoma (HBCx-60).

Relation between *COX-2* mRNA level and clinico-pathological parameters

The characteristics of the 446 breast tumors according to the individual *COX-2* mRNA level are shown in Table 3. Age of patients, SBR histological grade, lymph node status, tumor size and *Ki67* mRNA expression were not statistically different in patients with different *COX-2* expression levels. Hormone receptor status was the only parameter associated with *COX-2* mRNA level (ER α : $p < 10^{-4}$; PR: $p = 0.0016$) (Table 3).

We then tested the relation between *COX-2* mRNA expression levels and both *EGFR* mRNA level and *PIK3CA* mutation status, previously determined in these tumor samples [33, 34]. *PIK3CA* mutations were detected in 33% of patients (148/446). *COX-2* mRNA level was tightly linked to *EGFR* mRNA levels ($p < 10^{-4}$) and to *PIK3CA* mutations ($p < 10^{-4}$) (Table 3).

Prognostic impact of *COX-2* mRNA expression level

SBR grade ($p = 1.5 \cdot 10^{-4}$), lymph node status ($p = 1.9 \cdot 10^{-3}$), tumor size ($p = 1.4 \cdot 10^{-5}$), ER ($p = 8.4 \cdot 10^{-6}$) and PR ($p = 8.6 \cdot 10^{-6}$) status as well as *PIK3CA* mutations ($p = 0.02$) all had prognostic value as measured by the 5-years MFS. A trend towards a worse MFS among patients with low *COX-2* expression (using optimal cut-

Table 2: Correlation between *COX-2* mRNA and *COX-2* protein expression level on 14 PDX samples

samples ID	<i>COX-2</i> mRNA expression	<i>COX-2</i> protein expression	positive cells (%)	staining intensity
HBCx-10	0	negative	0	0
HBCx-51	0	negative	0	0
HBCx-22	1	negative	0	0
HBCx-28	7	negative	0	0
HBCx-43	26	negative	0	0
HBCx-16	38	negative	0	0
HBCx-49	38	positive (weak)	1	3
HBCx-30	39	positive (weak)	<1	2
HBCx-4B^b	218	positive (moderate)	15	2 to 3
HBCx-23	264	positive (moderate)	5	3
HBCx-8	339	positive (high)	50	2 to 3
HBCx-60 ^a	504			
HBCx-52^b	579	positive (high)	60	2 to 3
HBCx-15	658	positive (moderate)	15	2 to 3

^aCorresponding to metaplastic carcinoma case.

^bCorresponding to PDX selected for *in vivo* experiments.

off determined as described in material and methods section) was observed ($p=0.05$) (Table 4). This trend became statistically significant when evaluating the prognostic impact of low *COX-2* expression for the complete follow-up delay of this cohort ($p=0.007$) (Figure 2A). Multivariate analysis (Cox proportional hazards model) was also used to assess the influence of *COX-2* mRNA level on MFS, together with histological grade, lymph-node status, tumor size, estrogen and progesterone receptor status and *PIK3CA* mutations. Lymph node status >3 ($p=0.02$), SBR grade III ($p=0.04$), tumor size $>25\text{mm}$ ($p=0.02$) and low *COX-2* mRNA expression ($p=0.01$) were statistically associated with poor prognosis (Supplementary Table S1).

***COX-2* expression presents a prognostic value in *PIK3CA* wild-type patients**

We then assessed the prognostic impact of *COX-2* expression depending on the *PIK3CA* mutational status in the cohort of 446 patients. Independently of the subtype of BC and adjuvant treatment received (chemotherapy, hormone therapy, both or none) MFS was significantly better in patients with high *COX-2* expression ($p=0.007$, HR 1.560 [1.130-2.153]) (Figure 2A) and in patients with *PIK3CA* mutations ($p=0.02$, HR 1.455 [1.058-2.002])

(Figure 2B). *COX-2* expression level had no impact on MFS in the *PIK3CA* mutated patients' subgroup (Figure 2C). However in the *PIK3CA* wild-type patients' subgroup MFS was significantly better in patients with high *COX-2* expression as compared to patients with low *COX-2* expression ($p=0.01$, HR 1.617 [1.113-2.350]) (Figure 2C). Interestingly, the same result was observed in HR⁺ tumors where *PIK3CA* mutations are clearly associated with good prognosis ($p=0.0004$, HR 2.377 [1.473-3.835]) [35–37]. Patients with high *COX-2* expression and *PIK3CA* wild-type had a similar MFS as *PIK3CA* mutated patients ($p=0.07$, HR 1.717 [0.9458-3.116]) (Figure 2D). Low *COX-2* expression and *PIK3CA* wild-type status allowed to identify patients with the worse MFS in the total cohort ($p=0.0004$, HR: 1.761 [1.289 to 2.405]) and among HR⁺ tumors ($p=0.0002$, HR: 2.018 [1.397 to 2.914]) (Figure 3A and 3B). Given the limited number of triple-negative and HR-ERBB2⁺ cases, it was not appropriate to evaluate the prognostic impact of *COX-2* expression according to the *PIK3CA* status in these two subtypes.

COX-2 expression and *PIK3CA* mutational status did not impact overall survival (OS) in this cohort with very long follow-up (Supplementary Figure S2).

In the TCGA validation set, high *COX-2* expression was associated with a better DFS ($p=0.0014$, HR 2.206 [1.356-3.587]) and *PIK3CA* mutations did not have

Table 3: Relationship between COX-2 transcript expression level and classical clinical and biological parameters in a series of 446 breast cancers

	Total population (%)	Number of patients (%)		p value ^a
		COX-2 mRNA expression <0.3 relative to normals	COX-2 mRNA expression >0.3 relative to normals	
Total	446 (100)	332 (74.4)	114 (25.6)	
Age				
≤50	94 (21.1)	67 (71.3)	27 (28.7)	0.59
>50	352 (78.9)	261 (74.1)	91 (25.9)	
SBR histological grade^{b,c}				
I	57 (13)	43 (75.4)	14 (24.6)	0.36
II	223 (51)	172 (77.1)	51 (22.9)	
III	157 (35.9)	111 (71)	46 (29)	
Lymph node status^d				
0	118 (26.5)	89 (75.4)	29 (24.6)	0.69
1-3	232 (52.1)	169 (72.8)	63 (27.2)	
>3	92	71 (77.2)	21 (22.8)	
Macroscopic tumor size^e				
≤25	220 (50.2)	166 (75.4)	54 (24.6)	0.62
>25	218 (49.8)	160 (73.4)	58 (26.6)	
ERα				
Negative	115 (25.8)	64 (55.6)	51 (44.4)	<10 ⁻⁴
Positive	331 (74.2)	264 (79.7)	67 (20.3)	
PR				
Negative	190 (42.6)	125 (65.8)	65 (34.2)	0.0016
Positive	256 (57.4)	203 (79.3)	53 (20.7)	
ERBB2				
Negative	353 (79.1)	263 (74.5)	90 (25.5)	0.95
Positive	93 (20.9)	69 (74.2)	24 (25.8)	
Subgroups				
HR-ERBB2-	68 (15.2)	32 (47)	36 (53)	<10 ⁻⁴
HR-ERBB2+	42 (9.4)	28 (66.7)	14 (33.3)	
HR+ERBB2-	285 (63.9)	231 (81.4)	54 (18.6)	
HR+ERBB2+	51 (11.4)	41 (80.4)	10 (19.6)	
PIK3CA status				
Wild type	298 (66.8)	208 (69.8)	90 (30.2)	<10 ⁻⁴
Mutated	148 (33.2)	120 (81)	28 (19)	
Ki67 mRNA expression				
Median	12.4 (0.80-117)	12 (0.80-117)	13.2 (1.05-94.5)	0.42 ^f
EGFR mRNA expression				
Median	0.22 (0-106)	0.17 (0.02-7.56)	0.47 (0-106)	<10 ^{-4f}
Metastasis				
No	282 (63.2)	198 (70)	84 (30)	<10 ⁻⁴
Yes	164 (36.8)	134 (82)	30 (18)	

^aChi² test

^bScarff bloom Richardson classification.

^cInformation available for 437 patients.

^dInformation available for 442 patients.

^eInformation available for 438 patients.

^fKruskal Wallis's H test.

Table 4: Characteristics of the 446 primary breast tumors and relation to metastasis-free survival

	Number of patients	5 years MFS	p value ^a
Total	446	72.6%	
Age			
≤50	94	70.6%	0.33
>50	352	74.6%	
SBR histological grade^{b,c}			
I	57	92.4%	1.5.10^{-4f}
II	223	76%	
III	157	64.2%	
Lymph node status^d			
0	118	79.4%	1.9.10^{-3f}
1-3	232	76.5%	
>3	92	59.2%	
Macroscopic tumor size^e			
≤25	220	82.9%	1.4.10⁻⁵
>25	218	64.1%	
ERα			
Negative	115	60.2%	8.4.10⁻⁶
Positive	331	78.3%	
PR			
Negative	190	63.5%	8.6.10⁻⁶
Positive	256	81.3%	
ERBB2			
Negative	353	75.1%	0.11
Positive	93	68.5%	
Subgroups			
HR-ERBB2-	68	61.4%	1.5.10^{-5f}
HR-ERBB2+	42	53.8%	
HR+ERBB2-	285	78.3%	
HR+ERBB2+	51	81%	
PIK3CA status			
wild type	298	70.4%	0.02
mutated	148	80.2%	
COX-2 expression			
≤0.22	294	70.6%	0.05
>0.22	152	80.1%	

^aLog-rank test.

^bScarff bloom Richardson classification.

^cInformation available for 437 patients.

^dInformation available for 442 patients.

^eInformation available for 438 patients.

^fGlobal comparison of all subgroups of a category.

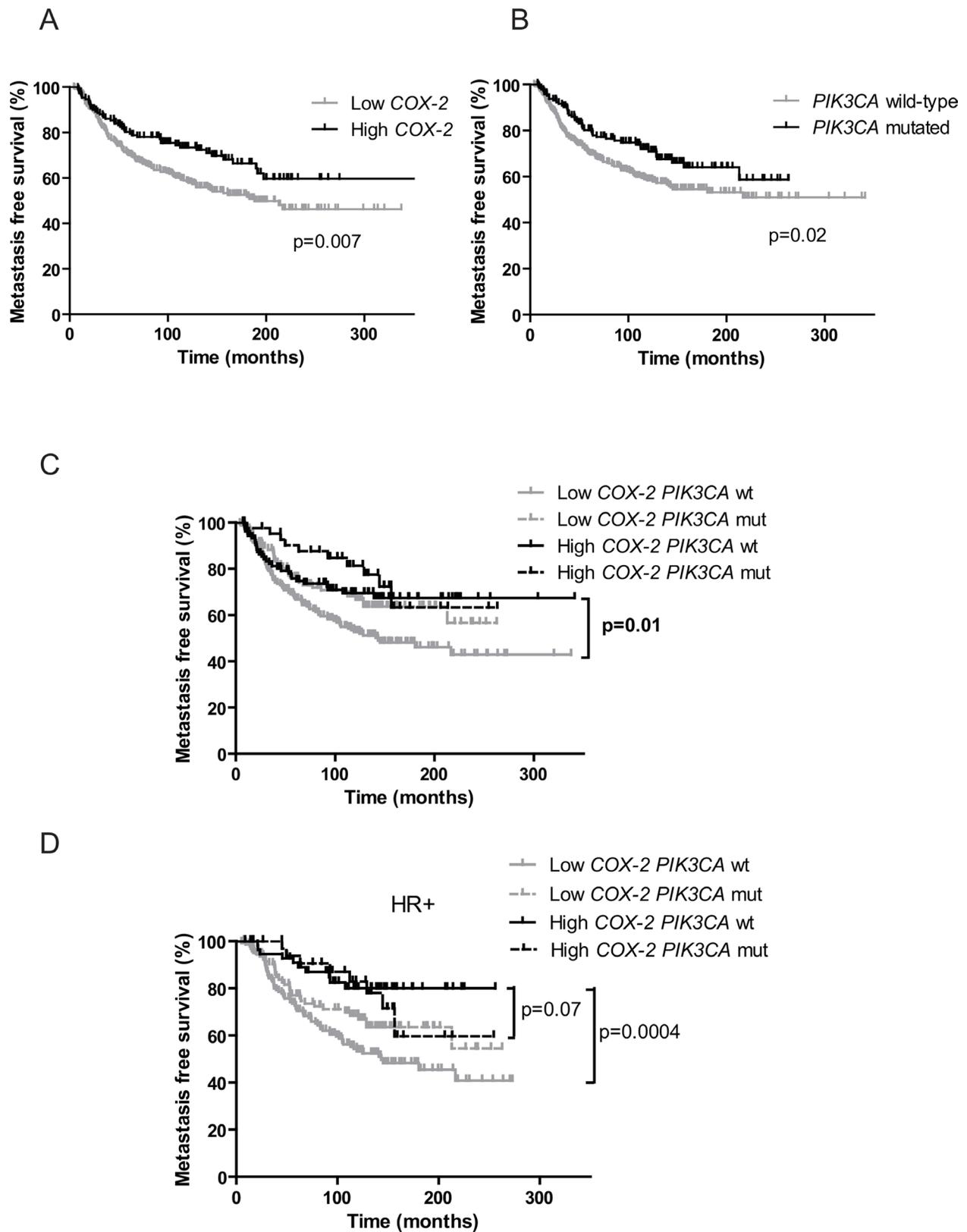


Figure 2: Prognostic value of *COX-2* mRNA expression and *PIK3CA* mutations on patients' metastasis-free survival. A. Kaplan-Meier estimates of metastasis-free survival according to *COX-2* mRNA expression. **B.** Kaplan-Meier estimates of metastasis-free survival according to *PIK3CA* mutations. **C.** Kaplan-Meier estimates of metastasis-free survival according to *COX-2* mRNA expression and *PIK3CA* mutations in the global cohort. wt for wild-type, mut for mutated. **D.** Kaplan-Meier estimates of metastasis-free survival according to *COX-2* mRNA expression and *PIK3CA* mutations in HR+ patients.

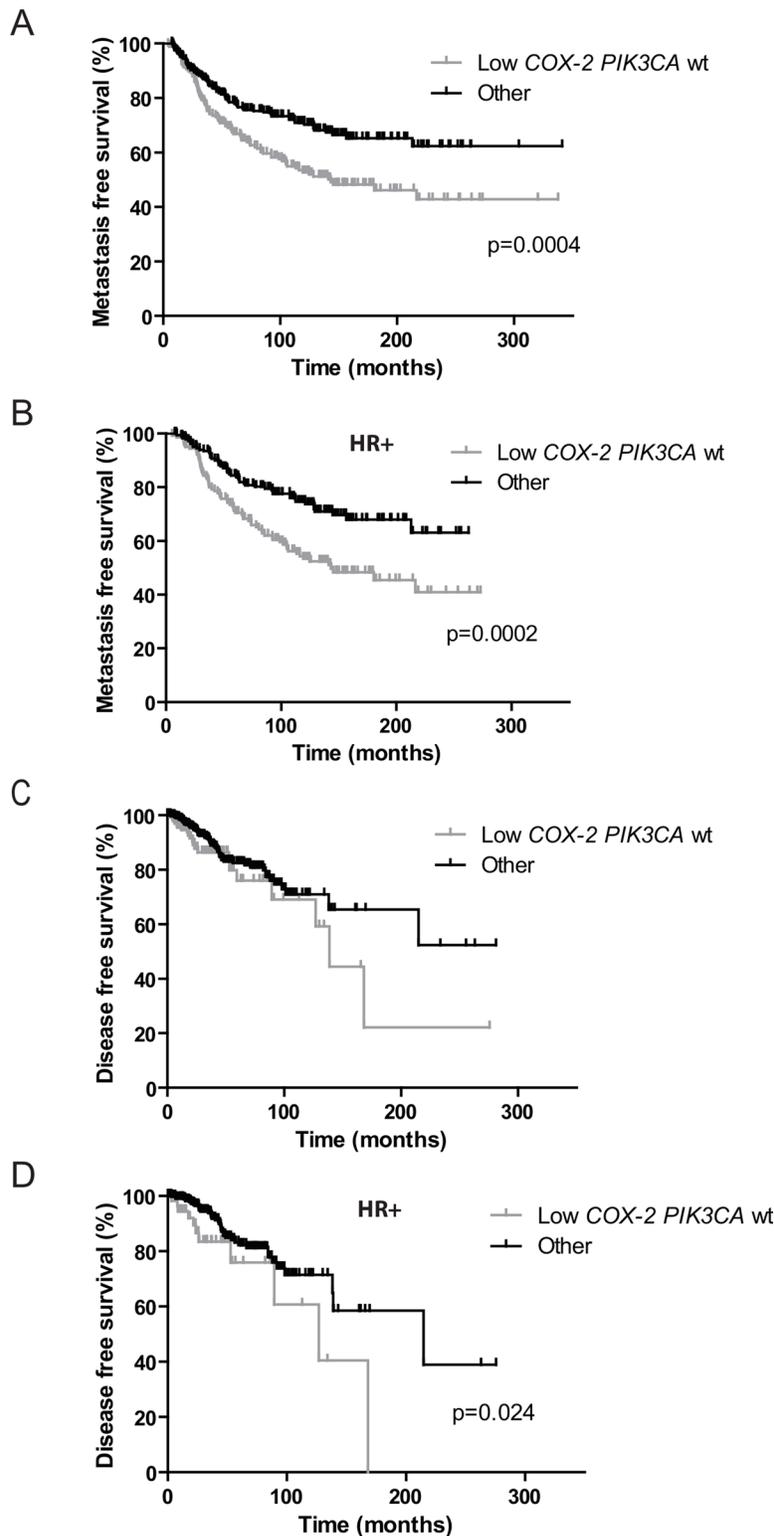


Figure 3: Prognostic value of low *COX-2* mRNA expression and wild-type *PIK3CA* status versus other subgroups on patients' metastasis and disease-free survival. **A.** Kaplan-Meier estimates of metastasis-free survival according to *COX-2* mRNA expression and *PIK3CA* mutations in the global cohort. **B.** Kaplan-Meier estimates of metastasis-free survival according to *COX-2* mRNA expression and *PIK3CA* mutations in HR+ patients. **C.** Kaplan-Meier estimates of disease-free survival according to *COX-2* mRNA expression and *PIK3CA* mutations in the global TCGA cohort. **D.** Kaplan-Meier estimates of disease-free survival according to *COX-2* mRNA expression and *PIK3CA* mutations in HR+ patients. "Other" refers to low *COX-2* *PIK3CA* mutated tumors and high *COX-2* *PIK3CA* wild-type and mutated tumors.

prognostic impact on DFS (Supplementary Figures S3A and S3B). After combination of these two parameters, *COX-2* expression level did not have prognostic impact in *PIK3CA* wild-type patients but high *COX-2* level expression was associated with a better DFS among mutated patients ($p=0.0007$, HR 4.667 [1.917-11.36]) (Supplementary Figure 3C). In the luminal subtype, high *COX-2* expression was associated with a better DFS ($p=0.012$, HR 2.682 [1.243-5.785]) and *PIK3CA* mutations did not have prognostic impact on DFS (data not shown). Among *PIK3CA* wild-type patients, high *COX-2* patients had a better DFS than low *COX-2* patients ($p=0.012$, HR 3.206 [1.287-7.984]) (Supplementary Figure S3D).

Similarly to Institut Curie BC cohort, low *COX-2* expression and *PIK3CA* wild-type status allowed to identify TCGA patients with the worse DFS in the entire cohort (trend, $p=0.105$, HR: 1.584 (0.9090 to 2.758) and among HR+ tumors ($p=0.024$, HR: 2.698 [1.142 to 6.376]) (Figure 3C and 3D).

In the TCGA BC cohort overall survival data showed a better prognosis for high *COX-2* patients ($p=0.0011$, HR 2.452 [1.428-4.208] but not for *PIK3CA* mutated patients (Supplementary Figure S4A and S4B). Among wild-type *PIK3CA* patients, high *COX-2* patients had a better overall survival than low *COX-2* patients ($p=0.018$, HR 2.183 [1.146-4.157]). The same significant difference was observed in *PIK3CA* mutated patients ($p=0.015$, HR 3.494 [1.273-9.592]) (Supplementary Figure S4C). In the luminal subtype, high *COX-2* expression was associated with a better OS ($p=0.0051$, HR 1.831 [0.999-3.357]) and *PIK3CA* mutations did not have a prognostic impact on OS (data not shown). Among mutated *PIK3CA* patients, high *COX-2* patients had a better overall survival than low *COX-2* patients ($p=0.023$, HR 3.206 [1.177-8.734]) (Supplementary Figure S4D).

Celecoxib antitumoral effect is only observed in breast tumor harboring a *PIK3CA* mutation

Since *COX-2* overexpression was associated with TNBC subtype, we chose triple-negative PDX to investigate celecoxib antitumoral effect. Moreover there is a need for targeted therapies in TNBC.

In the *PIK3CA* mutated TNBC PDX model (HBCx-4B) a significant reduction in tumor volume (RTV) was observed in mice receiving celecoxib as compared to control mice from day 22 ($p=0.03$) and until the end of the experiment (day 61, TGI=57%, $p=0.01$) (Figure 4A). These results clearly showed that celecoxib induced a significant antitumor effect in tumors expressing *COX-2* and harboring a *PIK3CA* mutation [31]. The TGI obtained with celecoxib in this model was near to the 60% proposed by Wang *et al.* as a cut-off for mice xenografts likely to lead to a positive clinical outcome [38]. Of note celecoxib is not a chemotherapy and was given as monotherapy in this experiment.

In contrast, in the *PIK3CA* wild-type model (HBCx-52) no significant difference in RTV was observed between the treated and control groups (day 25, $p=0.94$) (Figure 4B). HBCx-4B and HBCx-52 PDX models both express high *COX-2* mRNA level and *COX-2* protein (Table 2). However antitumoral effect was observed in HBCx-4B model only. We can therefore ascertain that celecoxib had no effect on the tumor growth in tumors expressing *COX-2* and *PIK3CA* wild-type.

Antitumoral effect of celecoxib in *PIK3CA* mutated tumors involves phosphorylation of PI3K/AKT pathway main actors

Western-blot analysis in both PDX showed a significant decrease of *COX-2* expression in the celecoxib treated group compared to controls ($p=0.018$ for HBCx-4B and $p=0.02$ for HBCx-52) confirming the pharmacological effect of this molecule. Celecoxib treatment did not affect angiogenesis as shown by the MVD assays in treated and control tumors of both PDX models (Table 5).

In the HBCx-4B responder model, exploration of the PI3K pathway showed a significant decrease of S6 ribosomal protein phosphorylation in the treated group compared to controls ($p=0.0003$) (Figure 5A). We observed a significant increase in the expression of this phospho-protein in two tumors, which progressed under celecoxib as compared to responders ($p=0.02$) (Figure 5A). These two non-responder tumors showed also a significant increase of AKT phosphorylation by comparison to responders ($p=0.02$) (Figure 5A).

In HBCx-52 similar phosphorylation levels of S6 ribosomal protein and AKT were observed in controls and treated tumors (Figure 5B).

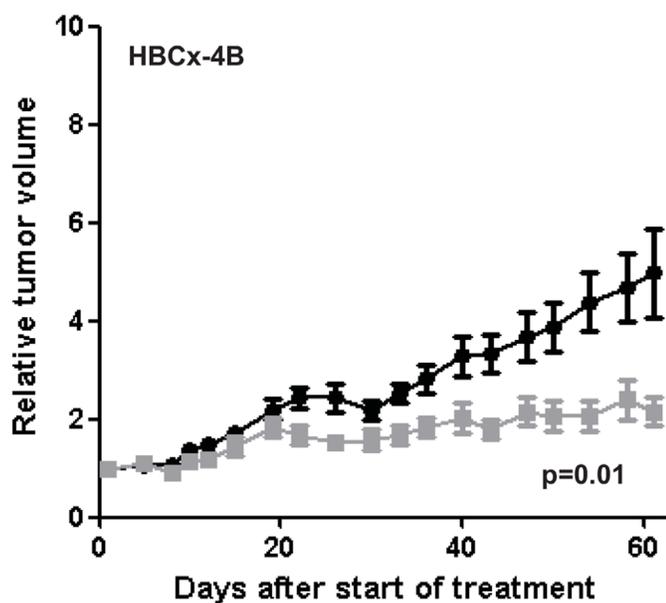
PTEN, another major component of the PI3K/AKT pathway, was not differentially expressed in these two PDX models. INPP4B expression was lost in the HBCx-4B responder model but also in several HBCx-52 tumors (Figure 5A and 5B).

DISCUSSION

In BC, *COX-2* expression level and its prognostic value have been controversial for several decades. In the present study, we observed an overexpression of *COX-2* only in a small percentage of BCs, predominantly belonging to the triple-negative subtype. More importantly the under-expression of *COX-2* is an independent pejorative prognostic factor. Low *COX-2* and *PIK3CA* wild-type status was identified as the worse prognostic factor for MFS in our cohort and confirmed for DFS in an independent validation set.

Several published studies reported variable expression levels of *COX-2* in BCs but most of them assessed *COX-2* expression at the protein level. In the present study, we applied qRT-PCR on RNA extracted

A



B

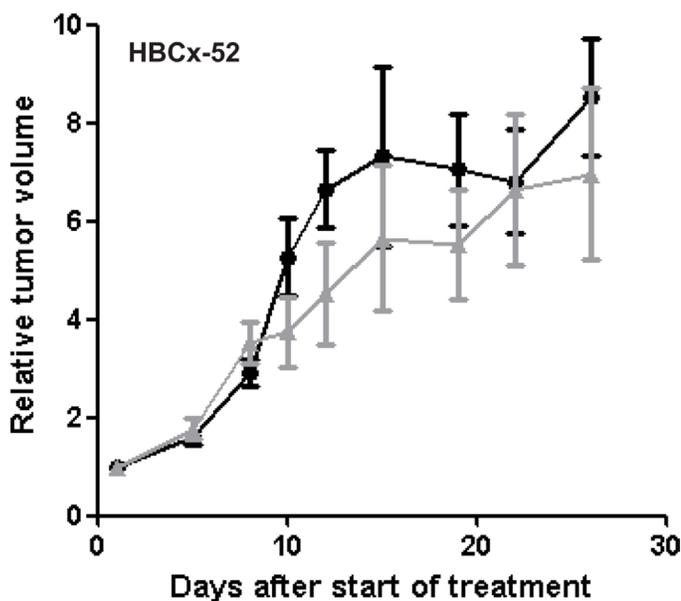


Figure 4: Effect of celecoxib on tumor growth of HBCx-4B and HBCx-52 PDX. Tumor growth was evaluated by plotting the mean of the RTV±SD over time. **A.** HBCx-4B bearing mice were treated with celecoxib (n=18), (40 mg/kg *per os* daily five times a week). Controls (n=20) received MCT (methylcellulose 5% and 0,2% tween *per os* daily five times a week). **B.** HBCx-52 bearing mice were treated with celecoxib (n=6), (40 mg/kg *per os* daily five times a week). Controls (n=7) received MCT (*per os* daily five times a week).

Table 5: Assessment of celecoxib treatment on angiogenesis (microvessel density) performed by ERG immunostaining

	control group	celecoxib-treated group	p value ^c
HBCx-52	40.4 ± 5.8	33.8 ± 9.0	p=0.35
HBCx-4B	23.5 ± 3.4	16.7 ± 3.0 ^a	p=0.058
		18.1 ± 2.2 ^b	p=0.14

Results are expressed as the mean (+/- SD) of the number of blood vessels by mm²

^aResponders tumors

^bNon-responders tumors

^cT-test

from a large cohort of samples all infiltrated with more than 70% of tumor cells to assess COX-2 expression. With the exception of few cases in triple-negative subtype, we showed that the majority of BCs under-express COX-2 mRNA or express COX-2 mRNA at a similar level to normal breast tissue. Few studies assessed COX-2 mRNA expression level. Contrary to our results, several authors found an overexpression of COX-2 mRNA in BC tissues when compared to benign breast lesions [39] or breast normal tissues [2, 40]. All these results were obtained using qualitative RT-PCR in very small cohorts of BCs (13, 10 and 9 cases), which might explain the discrepancy. One report by McCarthy *et al.* using qRT-PCR, showed that the median COX-2 mRNA expression in 45 primary invasive BC samples was not significantly different as compared to the median COX-2 mRNA expression in 22 normal breast tissues [41]. These authors also showed that COX-2 mRNA levels were significantly higher in estrogen ($p < 0.02$) and progesterone ($p < 0.0001$) receptor negative tumors but they did not assess the ERBB2 status of the tumors and consequently no conclusions can be drawn as for the level of mRNA expression of COX-2 in the triple-negative subtype [41]. According to our results Kirkpatrick *et al.* using qRT-PCR found in 40 infiltrating carcinomas and 40 matched adjacent non-cancerous tissue (ANCT) that COX-2 mRNA copy number per µg of RNA was two-fold higher in ANCT compared to the cancerous tissue ($p=0.01$) [42]. Using the robust and reproductive tool of qRT-PCR, our results are therefore similar to the above reports and confirm that COX-2 mRNA overexpression is not a hallmark in all BCs.

We have demonstrated a good correlation between COX-2 mRNA and protein level suggesting that we would expect to find the same minority overexpressed cases with an IHC-based study. This result was obtained by examining few cases using IHC and is contradictory with the majority of published studies describing COX-2 overexpression in invasive BCs [8, 11, 40, 43] but not all [44]. Like the majority of these studies we focused on COX-2 expression in epithelial cells and did not examine stroma. Recently Urban *et al.* analyzed the prognostic value of COX-2 expression not only in breast epithelial

cell but also in stromal cell using different antibodies and scoring algorithms. Although they showed that COX-2 expression in stromal cells and not in epithelial cells is an independent adverse prognostic factor and is relatively insensitive to variations of antibodies used, they finally explained the variability of published results by the use of different antibodies and scoring algorithms [45]. So we prefer to remain cautious about the IHC results and it should be reminded that our conclusions concerning the prognostic value of COX-2 expression is based on COX-2 mRNA expression level.

Published results concerning the changes of COX-2 protein expression during the disease progression and its prognostic significance are contradictory, even in the groups working with the same type of primary antibodies [45]. Miglietta *et al.* published that COX-2 immune positivity and percentage of positive cells correlated significantly with the size, grading, extent of primary tumor and vascular invasion of carcinoma but not with biological parameters (HR and ERBB2 status). Nevertheless, for Park *et al.* there was no significant association between COX-2 over-expression and tumor size, histologic grade, and estrogen receptor expression [46, 47]. In the work of Kim *et al.* COX-2 positivity was significantly correlated with high grade, negative ER, high Ki67, luminal B and triple-negative tumors [48].

To the best of our knowledge we conducted the first COX-2 mRNA expression prognostic impact study in a large cohort of BC patients. Under-expression of COX-2 transcript was associated with poor prognosis and HR status but was not with other classical criteria like high grade, tumor size or lymph node status. Multivariate analysis showed that COX-2 under-expression, high grade, higher tumor size and lymph node involvement were predictive of poor prognosis. COX-2 under-expression should be thus considered as an independent poor prognostic factor. Moreover this under expression combined with a PIK3CA wild-type status allowed to identify a poor prognosis subgroup of patients who might benefit from more intensive treatment regimens. This result was confirmed with the DFS analysis in a TCGA validation cohort. High COX-2 expression is significantly

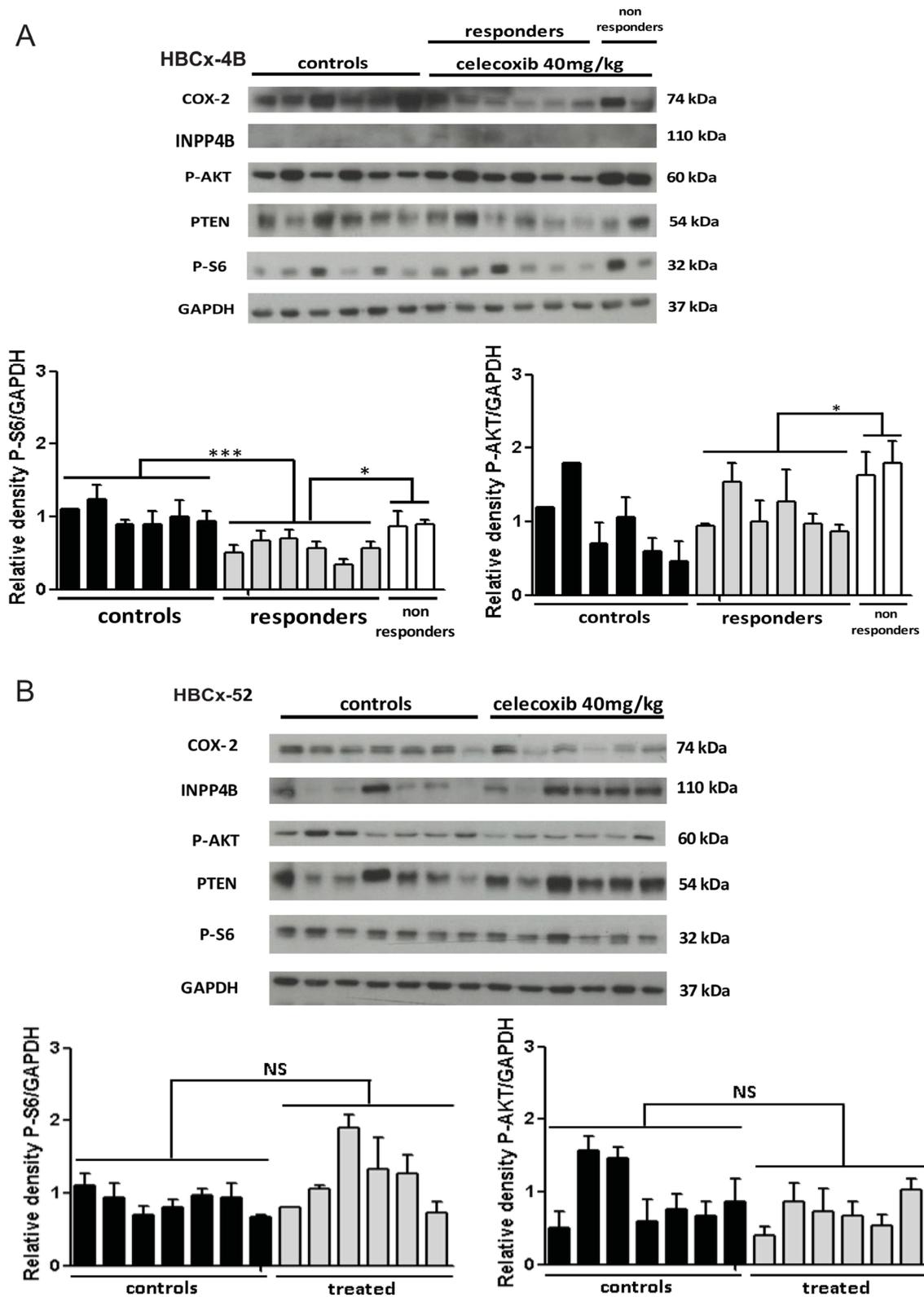


Figure 5: Analysis of tumors. **A.** Western-blot analysis of COX-2, phospho-AKT, phospho-S6 ribosomal protein, PTEN and INPP4B in HBCx-4B control mice (n=6) and celecoxib treated mice: responders (n=6) and non-responders (n=2). **B.** Western-blot analysis of COX-2, phospho-AKT, phospho-S6 ribosomal protein, PTEN and INPP4B in HBCx-52 control mice (n=7) and celecoxib treated mice (n=6). GAPDH served as loading control. One representative blot is presented for each model. The densitometric analysis is the mean \pm SEM. (n=3 experiments). For statistical analysis, treated group was compared with controls, and responders compared with non-responders. *P < 0.05 and ***P < 0.001.

correlated with better OS of *PIK3CA* wild-type and mutated patients in the TCGA cohort but this result was not observed in our BC cohort with a longer median follow-up delay (8.6 years for the Curie cohort *versus* 28.9 months for the TCGA cohort). Interplays between COX-2 and the PI3K/AKT pathway have been already well described in colorectal cancer [49] but still need to be deciphered in BC to explain effects of their combination.

In accordance with the observations made by Liao *et al.* in colorectal cancer, our *in vivo* PDX experiments showed that celecoxib antitumoral effect was restricted to *PIK3CA* mutated breast tumors. *PIK3CA* status has never been explored in BC clinical trials assessing concomitant administration of exemestane or chemotherapy with celecoxib. Our findings led to the hypothesis that the negative results of these clinical trials might come from the fact that patients were not selected according to tumor *PIK3CA* status. Consequently a retrospective analysis of results of these trials regarding *PIK3CA* status could be very interesting. Eventually, new prospective trials combining celecoxib with hormone therapy or chemotherapy may screen patients for tumor *PIK3CA* mutations to confirm its predictive value. It is also important to underline that our *in vivo* experiments were done with TNBC PDX whereas clinical trials were designed for luminal BCs. We cannot exclude that in this subtype of BCs some other unknown factors could interact negatively with antitumoral properties of celecoxib.

There were two non-responders tumors in our *PIK3CA* mutated PDX model. The protein expression analysis on collected tumor xenografts revealed an increase of AKT phosphorylation in these two tumors. In ovarian tumors and MCF-7 breast tumor cell line COX-2 overexpression is associated with an increase of AKT phosphorylation [50, 51], which might explain resistance to celecoxib in both tumors. Antitumoral effect of celecoxib is associated with the inactivation of PI3K/AKT pathway as observed with the decrease of S6 kinase phosphorylation whereas secondary resistance is explained by AKT reactivation.

In conclusion, treatment with celecoxib may be an additional therapeutic option for patients with BCs expressing COX-2 protein and mutated for *PIK3CA* whatever the level of COX-2 mRNA expression. Thus the detection of COX-2 protein should be the only prerequisite criteria for mutated tumors treatment with celecoxib. Noteworthy, *PIK3CA* mutation screening and COX-2 IHC staining are very easy to implement in diagnostic laboratory and could be used routinely for patient selection. These results were obtained with two PDX models only and need to be validated in a clinical trial. In this way we note that no published clinical trials with celecoxib in BC patients reported cardiac toxicity restricting the use of this FDA approved NSAID so it should not be a limiting factor for future trials.

MATERIALS AND METHODS

Patients

Samples of 446 primary breast tumor, excised from women treated at Institut Curie - Hôpital René Huguenin (Saint-Cloud, France) from 1978 to 2008, have been analyzed. All patients treated at Institut Curie before 2007 were informed that their tumor samples might be used for scientific purposes and had the opportunity to decline. Since 2007, patients treated at Institut Curie have given their approval by signing an informed consent. This study was approved by the local ethics committee (Breast Group of René Huguenin Hospital). The samples were immediately stored in liquid nitrogen until RNA extraction. A tumor sample was considered suitable for this study if the proportion of tumor cells exceeded 70%.

All patients (mean age 61.8 years, range 31 – 91 years) met the following criteria: primary unilateral non metastatic breast carcinoma for which complete clinico-pathological data and follow-up were available; no radiotherapy or chemotherapy before surgery; and full follow-up at Institut Curie - Hôpital René Huguenin. Adjuvant therapy was administered to 361 patients, consisting of chemotherapy alone in 87, hormone therapy alone in 175, and both treatments in 99 patients.

Estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (ERBB2) statuses were determined at the protein level by using biochemical methods (Dextran-coated charcoal method, enzyme immunoassay or immunohistochemistry) and confirmed by real-time quantitative RT-PCR assays [22, 23]. The population was divided into 4 groups according to HR (ER and PR) and ERBB2 status as follows: two luminal subtypes [HR⁺ (ER^α⁺ or PR⁺)/ERBB2⁺ (n=51)], and [HR⁺ (ER^α⁺ or PR⁺)/ERBB2⁻ (n=285)]; an ERBB2⁺ subtype [HR⁻ (ER^α⁻ and PR⁻)/ERBB2⁺ (n=42)]; and a triple-negative subtype [HR⁻ (ER^α⁻ and PR⁻)/ERBB2⁻ (n=68)]. Standard prognostic factors are shown in Table 3. Within a median follow-up of 8.6 years (range 6 months to 29 years), 164 patients developed distant metastasis. Ten specimens of adjacent normal breast tissue from BC patients (n=2) and normal breast tissue from women undergoing cosmetic breast surgery (n=8) were used as sources of normal RNA [24].

Public data of 817 breast invasive carcinomas from TCGA were used as a validation set [25]. This cohort was obtained by using www.cbioportal.org. [26, 27]. The population was divided into 5 molecular subtypes: basal (n=136), ERBB2⁺ (n=65), luminal A (n=415), luminal B (n=176) and normal (n=25). Median follow-up was 25 months (range 0 – 281.1 months) and 28.9 months (range 0 - 282.7 months) for disease-free survival and overall survival respectively. COX-2 mRNA expression is expressed in z-Scores (RNA Seq V2 RSEM (RNA-Seq by Expectation-Maximization)).

RNA extraction

Total RNA was extracted from breast tumor samples and PDX tumors by using acid-phenol guanidium as previously described [28]. RNA quality was determined by electrophoresis through agarose gels, staining with ethidium bromide, and visualization of the 18S and 28S RNA bands under ultraviolet light.

Real-time RT-PCR

Quantitative values were obtained from the cycle number (Ct value) at which the increase in the fluorescence signal associated with exponential growth of PCR products started to be detected by the laser detector of the ABI Prism 7900 Sequence Detection System (Perkin-Elmer Applied Biosystems, Foster City, CA), using PE Biosystems analysis software according to the manufacturer's manuals.

The *TBP* gene (Genbank accession NM_003194) encoding the TATA box-binding protein (a component of the DNA-binding protein complex TFIID) was quantified as an endogenous RNA control, and each sample was normalized on the basis of its *TBP* content [22].

Results, expressed as N-fold differences in target gene expression relative to the *TBP* gene and termed "Ntarget", were determined as $N_{\text{target}} = 2^{\Delta C_{\text{t sample}}}$, where the ΔC_{t} value of the sample was determined by subtracting the average Ct value of the target gene from the average Ct value of the *TBP* gene.

The Ntarget values of the samples were subsequently normalized such that the median of the Ntarget values for the ten normal breast tissues was 1. In tumor samples values of 3 or more were therefore considered to represent overexpression, and values of 0.3 or less were considered to represent underexpression of the 10 quantifiable mRNAs, as in previous studies [22, 29]

Primers' sequences are available on request. Agarose gel electrophoresis was used to verify the specificity of PCR amplicons. The conditions of cDNA synthesis and PCR were previously described [22].

Western blot analysis

Proteins were extracted from frozen tumors using RIPA buffer (50 mM Tris-HCl (pH 8), 150 mM NaCl, 0.5% deoxycholic acid, 0.5% triton) supplemented with protease and phosphatase inhibitors. Proteins were separated by SDS-PAGE and then electrophoretically transferred into nitrocellulose membrane and probed using the following primary antibodies: anti-GAPDH (V18 clone, 1/20000) purchased from Santa Cruz Biotechnology, anti-COX-2 (12282, 1/1000), anti-phospho Serin 473-AKT (4060, 1/2000), anti-PTEN (9552, 1/2000), anti-INPP4B (14543, 1/2000) and anti-phospho-S6 ribosomal protein (2211, 1/8000) purchased from Cell Signaling Technology (Ozyme). Proteins were

detected according to the ECL Western Blotting Analysis System procedure (GE Healthcare, Buckinghamshire, UK). The intensity of the protein bands was quantified using ImageJ software.

Immunohistochemical staining

Patient and xenografted tumors were fixed in 10% neutral buffered formalin, paraffin embedded, and hematoxylin-eosin-saffron (HES) stained. The anti-COX-2 antibody (Dako reference M3617) and its isotopic control (Sigma reference F5636) were used on 26 primary breast tumors and 14 xenografted tumors. Staining (intensity and fraction of positive cells) was taken into consideration in the cytoplasm of epithelial cells only.

Assays for microvessel density (MVD) on tumor tissues

Microvessels in tumor tissues were immunostained using anti-ERG antibody (reference AC-0105, clone EP111, Abcam) on 14 xenografts (HBCx-4B, control tumors n=3, celecoxib treated tumors, n=5 (3 responders and 2 non-responders), HBCx-52, control tumors n=3, celecoxib treated tumors n=3). MVD was assessed according to a method adapted of Weidner *et al.*, 1991 [30]. The entire tumor section was first observed at low-power magnification (40x) to select the most vascularized areas (hotspots). Individual microvessels, immunoreactive for ERG, were counted at high-power magnification (400x) within 10 consecutive fields. In each tumor tissue, the microvessel count was expressed by mm².

In vivo experiments

In vivo studies were performed on female Swiss nude mice purchased from Charles River. Mice care and housing were conformed to the institutional guidelines as put forth by the French Ethical Committee. Human TNBC xenografted models were established as previously detailed [31, 32]. The effect of celecoxib (purchased from Pfizer) was evaluated in two PDX: HBCx-4B which presents a *PIK3CA* mutation and HBCx-52, wild-type for this gene, both expressing COX-2. A toxicity study was first performed on mice-bearing human BC xenografts which received 20 or 40 mg/kg of celecoxib by gavage five times a week. As no toxicity was observed, the dose of 40 mg/kg was retained for the next experiments. For HBCx-52, a control group (n=7) received gavage with MCT (methylcellulose 5% + 0.2% tween) five times a week and the treated group received five times a week 40 mg/kg of celecoxib (n=6). For HBCx-4B, the same groups were established: a MCT control group (n=20) and a celecoxib treated group (n=20). Tumor growth was evaluated with a calliper twice a week. Tumor growth inhibition (TGI) of treated tumors *versus* controls was calculated as the ratio of the mean relative tumor volume (RTV) in the treated

group to the mean RTV in the control group at the same time. Statistical significance of TGI was calculated using the paired Student t test comparing the individual RTVs in the treated and control groups.

Statistical analysis

Statistical analyses were performed using GraphPad Prism 5 software. The data are expressed as the mean \pm SEM. The results were considered statistically significant at a p-value <0.05 (*), <0.01 (**), or <0.001 (***)

Relationships between mRNA levels and clinical parameters were identified by using non parametric tests, namely the Chi-square test, Fischer's test and the Mann-Whitney U test.

Metastasis-free survival (MFS) was determined as the interval between initial diagnosis and detection of the first metastasis. Overall survival (OS) was determined as the interval between initial diagnosis and death of any cause. Survival distributions were estimated by the Kaplan-Meier method, and the significance of differences between survival rates was ascertained with the log-rank test. The optimal cut-off value for *COX-2* mRNA expression prognostic value was determined with the AUC-ROC analysis defining "high" *COX-2* mRNA expression >0.22 and "low" *COX-2* mRNA expression <0.22 . The Cox proportional hazards regression model was used to assess prognostic significance in the multivariate analysis and the results are presented as hazard ratios and 95% confidence intervals (CIs).

ACKNOWLEDGMENTS

We thank Martial Caly, Sophie Chateau-Joubert and Jean-Luc Servely for their technical assistance in IHC staining. We thank Rania El Botty for her technical assistance for western-blot.

CONFLICTS OF INTEREST

No potential conflicts of interest are reported by authors.

GRANT SUPPORT

This work was supported by La Ligue contre le Cancer du département des Hauts-de-Seine (grants number WB2014-51) and SIRIC of Curie Institute (grants INCa-DGOS-4654).

REFERENCES

1. Smith WL, Dewitt DL. Prostaglandin endoperoxide H synthases-1 and -2. *Adv Immunol.* 1996; 62:167-215.
2. Parrett M, Harris R, Joarder F, Ross M, Clausen K, Robertson F. Cyclooxygenase-2 gene expression in human breast cancer. *Int J Oncol.* 1997; 10:503-507.
3. Brueggemeier RW, Quinn AL, Parrett ML, Joarder FS, Harris RE, Robertson FM. Correlation of aromatase and cyclooxygenase gene expression in human breast cancer specimens. *Cancer Lett.* 1999; 140:27-35.
4. Glover JA, Hughes CM, Cantwell MM, Murray LJ. A systematic review to establish the frequency of cyclooxygenase-2 expression in normal breast epithelium, ductal carcinoma in situ, microinvasive carcinoma of the breast and invasive breast cancer. *Br J Cancer.* 2011; 105:13-17.
5. Fornetti J, Jindal S, Middleton KA, Borges VF, Schedin P. Physiological COX-2 expression in breast epithelium associates with COX-2 levels in ductal carcinoma in situ and invasive breast cancer in young women. *Am J Pathol.* 2014; 184:1219-1229.
6. van Nes JG, de Kruijf EM, Faratian D, van de Velde CJ, Putter H, Falconer C, Smit VT, Kay C, van de Vijver MJ, Kuppen PJ, Bartlett JM. COX2 expression in prognosis and in prediction to endocrine therapy in early breast cancer patients. *Breast Cancer Res Treat.* 2011; 125:671-685.
7. Denkert C, Winzer KJ, Muller BM, Weichert W, Pest S, Kobel M, Kristiansen G, Reles A, Siegert A, Guski H, Hauptmann S. Elevated expression of cyclooxygenase-2 is a negative prognostic factor for disease free survival and overall survival in patients with breast carcinoma. *Cancer.* 2003; 97:2978-2987.
8. Ristimaki A, Sivula A, Lundin J, Lundin M, Salminen T, Haglund C, Joensuu H, Isola J. Prognostic significance of elevated cyclooxygenase-2 expression in breast cancer. *Cancer Res.* 2002; 62:632-635.
9. Singh-Ranger G, Salhab M, Mokbel K. The role of cyclooxygenase-2 in breast cancer: review. *Breast Cancer Res Treat.* 2008; 109:189-198.
10. Zerkowski MP, Camp RL, Burtness BA, Rimm DL, Chung GG. Quantitative analysis of breast cancer tissue microarrays shows high cox-2 expression is associated with poor outcome. *Cancer Invest.* 2007; 25:19-26.
11. Koki AT, Masferrer JL. Celecoxib: a specific COX-2 inhibitor with anticancer properties. *Cancer Control.* 2002; 9:28-35.
12. Thun MJ, Jacobs EJ, Patrono C. The role of aspirin in cancer prevention. *Nat Rev Clin Oncol.* 2012; 9:259-267.
13. Alshafie GA, Abou-Issa HM, Seibert K, Harris RE. Chemotherapeutic evaluation of Celecoxib, a cyclooxygenase-2 inhibitor, in a rat mammary tumor model. *Oncol Rep.* 2000; 7:1377-1381.
14. Dai ZJ, Ma XB, Kang HF, Gao J, Min WL, Guan HT, Diao Y, Lu WF, Wang XJ. Antitumor activity of the selective cyclooxygenase-2 inhibitor, celecoxib, on breast cancer in Vitro and in Vivo. *Cancer Cell Int.* 2012; 12:53.
15. Bundred NJ, Cramer A, Morris J, Renshaw L, Cheung KL, Flint P, Johnson R, Young O, Landberg G, Grassby S, Turner L, Baidam A, Barr L, Dixon JM. Cyclooxygenase-2 inhibition does not improve the reduction in ductal carcinoma in situ proliferation with aromatase inhibitor therapy: results of the ERISAC

- randomized placebo-controlled trial. *Clin Cancer Res.* 2010; 16:1605-1612.
16. Generali D, Buffa FM, Deb S, Cummings M, Reid LE, Taylor M, Andreis D, Allevi G, Ferrero G, Byrne D, Martinotti M, Bottini A, Harris AL, Lakhani SR, Fox SB. COX-2 expression is predictive for early relapse and aromatase inhibitor resistance in patients with ductal carcinoma in situ of the breast, and is a target for treatment. *Br J Cancer.* 2014; 111:46-54.
 17. Dirix LY, Ignacio J, Nag S, Bapsy P, Gomez H, Raghunadharao D, Paridaens R, Jones S, Falcon S, Carpentieri M, Abbattista A, Lobelle JP. Treatment of advanced hormone-sensitive breast cancer in postmenopausal women with exemestane alone or in combination with celecoxib. *J Clin Oncol.* 2008; 26:1253-1259.
 18. Pierga JY, Delaloge S, Espie M, Brain E, Sigal-Zafrani B, Mathieu MC, Bertheau P, Guinebretiere JM, Spielmann M, Savignoni A, Marty M. A multicenter randomized phase II study of sequential epirubicin/cyclophosphamide followed by docetaxel with or without celecoxib or trastuzumab according to HER2 status, as primary chemotherapy for localized invasive breast cancer patients. *Breast Cancer Res Treat.* 2010; 122:429-437.
 19. Liao X, Lochhead P, Nishihara R, Morikawa T, Kuchiba A, Yamauchi M, Imamura Y, Qian ZR, Baba Y, Shima K, Sun R, Nosho K, Meyerhardt JA, et al. Aspirin use, tumor PIK3CA mutation, and colorectal-cancer survival. *N Engl J Med.* 2012; 367:1596-1606.
 20. Paleari L, Puntoni M, Clavarezza M, DeCensi M, Cuzick J, DeCensi A. PIK3CA Mutation, Aspirin Use after Diagnosis and Survival of Colorectal Cancer. A Systematic Review and Meta-analysis of Epidemiological Studies. *Clin Oncol (R Coll Radiol).* 2015.
 21. Dumont AG, Dumont SN, Trent JC. The favorable impact of PIK3CA mutations on survival: an analysis of 2587 patients with breast cancer. *Chin J Cancer.* 2012; 31:327-334.
 22. Bieche I, Onody P, Laurendeau I, Olivi M, Vidaud D, Lidereau R, Vidaud M. Real-time reverse transcription-PCR assay for future management of ERBB2-based clinical applications. *Clin Chem.* 1999; 45:1148-1156.
 23. Bieche I, Parfait B, Laurendeau I, Girault I, Vidaud M, Lidereau R. Quantification of estrogen receptor alpha and beta expression in sporadic breast cancer. *Oncogene.* 2001; 20:8109-8115.
 24. Finak G, Sadekova S, Pepin F, Hallett M, Meterissian S, Halwani F, Khetani K, Souleimanova M, Zabolotny B, Omeroglu A, Park M. Gene expression signatures of morphologically normal breast tissue identify basal-like tumors. *Breast Cancer Res.* 2006; 8:R58.
 25. Ciriello G, Gatza ML, Beck AH, Wilkerson MD, Rhie SK, Pastore A, Zhang H, McLellan M, Yau C, Kandoth C, Bowlby R, Shen H, Hayat S, et al. Comprehensive Molecular Portraits of Invasive Lobular Breast Cancer. *Cell.* 2015; 163:506-519.
 26. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, Sun Y, Jacobsen A, Sinha R, Larsson E, Cerami E, Sander C, Schultz N. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal.* 2013; 6:pl1.
 27. Cerami E, Gao J, Dogrusoz U, Gross BE, Sumer SO, Aksoy BA, Jacobsen A, Byrne CJ, Heuer ML, Larsson E, Antipin Y, Reva B, Goldberg AP, Sander C, Schultz N. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. *Cancer Discov.* 2012; 2:401-404.
 28. Bieche I, Parfait B, Le Doussal V, Olivi M, Rio MC, Lidereau R, Vidaud M. Identification of CGA as a novel estrogen receptor-responsive gene in breast cancer: an outstanding candidate marker to predict the response to endocrine therapy. *Cancer Res.* 2001; 61:1652-1658.
 29. Bieche I, Onody P, Tozlu S, Driouch K, Vidaud M, Lidereau R. Prognostic value of ERBB family mRNA expression in breast carcinomas. *Int J Cancer.* 2003; 106:758-765.
 30. Weidner N, Semple JP, Welch WR, Folkman J. Tumor angiogenesis and metastasis—correlation in invasive breast carcinoma. *N Engl J Med.* 1991; 324:1-8.
 31. Marangoni E, Vincent-Salomon A, Auger N, Degeorges A, Assayag F, de Cremoux P, de Plater L, Guyader C, De Pinieux G, Judde JG, Rebutti M, Tran-Perennou C, Sastre-Garau X, et al. A new model of patient tumor-derived breast cancer xenografts for preclinical assays. *Clin Cancer Res.* 2007; 13:3989-3998.
 32. Reyat F, Guyader C, Decraene C, Lucchesi C, Auger N, Assayag F, De Plater L, Gentien D, Poupon MF, Cottu P, De Cremoux P, Gestraud P, Vincent-Salomon A, et al. Molecular profiling of patient-derived breast cancer xenografts. *Breast Cancer Res.* 2012; 14:R11.
 33. Cizkova M, Susini A, Vacher S, Cizeron-Clairac G, Andrieu C, Driouch K, Fourme E, Lidereau R, Bieche I. PIK3CA mutation impact on survival in breast cancer patients and in ERalpha, PR and ERBB2-based subgroups. *Breast Cancer Res.* 2012; 14:R28.
 34. Meseure D, Vacher S, Drak Alsibai K, Trassard M, Susini A, Le Ray C, Lerebours F, Le Scodan R, Spyrtos F, Marc Guinebretiere J, Lidereau R and Bieche I. Profiling of EGFR mRNA and protein expression in 471 breast cancers compared with 10 normal tissues: a candidate biomarker to predict EGFR inhibitor effectiveness. *Int J Cancer.* 2012; 131:1009-1010.
 35. Kalinsky K, Jacks LM, Heguy A, Patil S, Drobnjak M, Bhanot UK, Hedvat CV, Traina TA, Solit D, Gerald W, Moynahan ME. PIK3CA mutation associates with improved outcome in breast cancer. *Clin Cancer Res.* 2009; 15:5049-5059.
 36. Loi S, Haibe-Kains B, Majjaj S, Lallemand F, Durbecq V, Larsimont D, Gonzalez-Angulo AM, Pusztai L, Symmans WF, Bardelli A, Ellis P, Tutt AN, Gillett CE, et al. PIK3CA mutations associated with gene signature of

- low mTORC1 signaling and better outcomes in estrogen receptor-positive breast cancer. *Proc Natl Acad Sci U S A*. 2010; 107:10208-10213.
37. Ellis MJ, Lin L, Crowder R, Tao Y, Hoog J, Snider J, Davies S, DeSchryver K, Evans DB, Steinseifer J, Bandaru R, Liu W, Gardner H, et al. Phosphatidylinositol-3-kinase alpha catalytic subunit mutation and response to neoadjuvant endocrine therapy for estrogen receptor positive breast cancer. *Breast Cancer Res Treat*. 2010; 119:379-390.
 38. Wong H, Choo EF, Alicke B, Ding X, La H, McNamara E, Theil FP, Tibbitts J, Friedman LS, Hop CE, Gould SE. Antitumor activity of targeted and cytotoxic agents in murine subcutaneous tumor models correlates with clinical response. *Clin Cancer Res*. 2012; 18:3846-3855.
 39. Yoshimura N, Sano H, Okamoto M, Akioka K, Ushigome H, Kadotani Y, Yoshimura R, Nobori S, Higuchi A, Ohmori Y, Nakamura K. Expression of cyclooxygenase-1 and -2 in human breast cancer. *Surg Today*. 2003; 33:805-811.
 40. Half E, Tang XM, Gwyn K, Sahin A, Wathen K, Sinicrope FA. Cyclooxygenase-2 expression in human breast cancers and adjacent ductal carcinoma in situ. *Cancer Res*. 2002; 62:1676-1681.
 41. McCarthy K, Bustin SA, Ogunkolade B, Khalaf S, Laban CA, McVittie CJ, Carpenter R, Jenkins PJ. Cyclooxygenase-2 (COX-2) mRNA expression and hormone receptor status in breast cancer. *Eur J Surg Oncol*. 2006; 32:707-709.
 42. Kirkpatrick K, Ogunkolade W, Elkak A, Bustin S, Jenkins P, Ghilchik M, Mokbel K. The mRNA expression of cyclooxygenase-2 (COX-2) and vascular endothelial growth factor (VEGF) in human breast cancer. *Curr Med Res Opin*. 2002; 18:237-241.
 43. Watanabe O, Shimizu T, Imamura H, Kinoshita J, Utada Y, Okabe T, Kimura K, Hirano A, Yoshimatsu K, Aiba M, Ogawa K. Expression of cyclooxygenase-2 in malignant and benign breast tumors. *Anticancer Res*. 2003; 23:3215-3221.
 44. Hwang D, Scollard D, Byrne J, Levine E. Expression of cyclooxygenase-1 and cyclooxygenase-2 in human breast cancer. *J Natl Cancer Inst*. 1998; 90:455-460.
 45. Urban J, Kuzbicki L, Szatkowski G, Stanek-Widera A, Lange D, Chwirot BW. Stromal, rather than epithelial cyclooxygenase-2 (COX-2) expression is associated with overall survival of breast cancer patients. *BMC Cancer*. 2014; 14:732.
 46. Miglietta A, Toselli M, Ravarino N, Vencia W, Chiecchio A, Bozzo F, Motta M, Torchio B, Bocca C. COX-2 expression in human breast carcinomas: correlation with clinicopathological features and prognostic molecular markers. *Expert Opin Ther Targets*. 2010; 14:655-664.
 47. Park K, Han S, Shin E, Kim HJ, Kim JY. Cox-2 expression on tissue microarray of breast cancer. *Eur J Surg Oncol*. 2006; 32:1093-1096.
 48. Kim HS, Moon HG, Han W, Yom CK, Kim WH, Kim JH, Noh DY. COX2 overexpression is a prognostic marker for Stage III breast cancer. *Breast Cancer Res Treat*. 2012; 132:51-59.
 49. Cathomas G. PIK3CA in Colorectal Cancer. *Front Oncol*. 2014; 4:35.
 50. Uddin S, Ahmed M, Hussain A, Assad L, Al-Dayel F, Bavi P, Al-Kuraya KS, Munkarah A. Cyclooxygenase-2 inhibition inhibits PI3K/AKT kinase activity in epithelial ovarian cancer. *Int J Cancer*. 2010; 126:382-394.
 51. Bocca C, Ievolella M, Autelli R, Motta M, Mosso L, Torchio B, Bozzo F, Cannito S, Paternostro C, Colombatto S, Parola M, Miglietta A. Expression of Cox-2 in human breast cancer cells as a critical determinant of epithelial-to-mesenchymal transition and invasiveness. *Expert Opin Ther Targets*. 2014; 18:121-135.