

Targeted new peptide based nanoparticles toward high EGFR expressing cancer cells for MRI

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Abstract

In this study, the various core sizes of manganese ferrite nanoparticles (MnFe_2O_4) conjugated with D4 peptide ($\text{MnFe}_2\text{O}_4\text{-PEG-D4}$) were synthesized. The high relaxivity MnFe_2O_4 nanoparticles were obtained by thermal decomposition of Iron acetylacetonate and manganese acetylacetonate in hydrophobic solution at high boiling process. The surface of MnFe_2O_4 nanoparticles were coated with polyethylene glycol (PEG) and EGFR peptide ligand (D4: Leu-Ala-Arg-Leu-Leu-Thr) to improve their dispersion and ability to target EGFR. The negative signal enhancement of EGFR expressing cancer cells (SKBR-3 and PC-3) were significantly higher than that of low EGFR expressing cells (HEK-293).

Introduction

Molecular Imaging has recently been developed very rapidly and extensively in biotechnology [1]. Tumor-targeted drug delivery can enhance the effectiveness of therapy while decreasing the systemic toxicity of these drugs. We synthesized MnFe_2O_4 nanoparticles for MR imaging are coated with polyethylene glycol, which has been frequently used as a drug carrier because of its biocompatibility, water solubility and their ability to escape capture by macrophages. $\text{MnFe}_2\text{O}_4\text{-PEG}$ conjugate with D4 peptide substrate (Fig 1.) were synthesized to examine the binding affinity of EGFR using in vitro MR imaging.

Methods

Iron acetylacetonate ($\text{Fe}(\text{acac})_3$) and manganese acetylacetonate ($\text{Mn}(\text{acac})_2$) precursors were mixed in a molar ratio of 2:1, and reacted in benzyl ether solvent containing oleic acid, oleylamine and, 1,2-hexadecanediol at 300°C for 1 hour to produce manganese ferrite nanoparticles (MnFe_2O_4). The MnFe_2O_4 nanoparticles have various core sizes were synthesized through the same procedure while the amount of solvent was controlled. We tested two cancer cell lines with different levels of EGFR expression: SKBR-3 and PC-3. In addition, we had chosen HEK-293 cells as negative cell for control which lacks of EGFR receptors. All cells were incubated with $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$ (0.5 mM Fe), washed by PBS buffer and scanned by 3.0 T MRI.

Results and Discussion

The manganese ferrite nanoparticles (MnFe_2O_4) was synthesized and characterized by TEM (Fig 2.), SQUID, and FT-IR. Moreover, the TEM results showed that the well-dispersed nanoparticles which coated with PEG. The detection of saturation magnetization by SQUID magnetometry demonstrated that $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$ has high saturation magnetization (84 emu/g) and small coercivity (about 5 G). The internalization of $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$ nanoparticles into positive cells was confirmed by in vitro MR imaging study (Fig 3.). With the $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$ conjugates, the detection of the SKBR-3 cell line occurred with a noticeable negative enhancements (T_2 -weighted MR images). As the relative EGFR expression level increased, the MR contrast increased consistently. The signal intensity of positive cells in the present of $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$ is significantly lower than that of negative cells. No signal intensity change was observed for the negative cells in the presence and absence of $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$.

Conclusion

We have successfully synthesized and characterized $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$ nanoparticles, which was shown well-dispersed, high relaxivity and high saturation magnetization. Moreover, we have demonstrated the D4 peptide substrate that was conjugated to MnFe_2O_4 nanoparticles and shown to enhance binding to and entry into EGFR expressing cells by MR imaging studies.

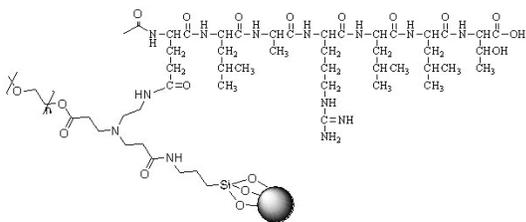


Figure 2. Structure formula of $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$

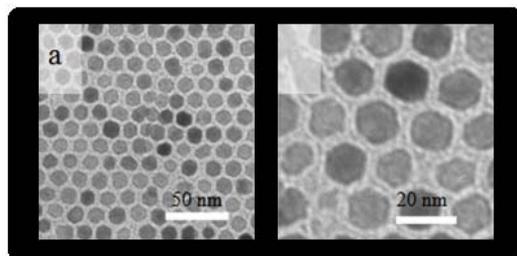


Figure 1. The TEM images of MnFe_2O_4 nanoparticles (A), enlarged images of MnFe_2O_4 nanoparticles 12.4 ± 0.9 nm (B).

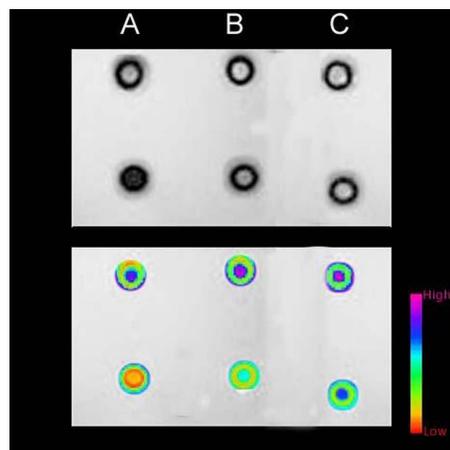


Figure 3. T_2 -weighted images of positive and negative cells after the treatment with or without 0.5 mM $\text{MnFe}_2\text{O}_4\text{-PEG-D4}$. (a) SKBR-3 cells (b) PC-3 cells (c) HEK-293 cells. Upper: cells treatment without contrast agent. Lower: cells treatment with contrast agent.

References

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