



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Genome sequence survey identifies unique sequences and key virulence genes with unusual rates of amino Acid substitution in bovine *Staphylococcus aureus*

Citation for published version:

Herron, LL, Chakravarty, R, Dwan, C, Fitzgerald, JR, Musser, JM, Retzel, E & Kapur, V 2002, 'Genome sequence survey identifies unique sequences and key virulence genes with unusual rates of amino Acid substitution in bovine *Staphylococcus aureus*' *Infection and Immunity*, vol. 70, no. 7, pp. 3978-81.

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

Infection and Immunity

Publisher Rights Statement:

Copyright © 2002, American Society for Microbiology

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



Genome Sequence Survey Identifies Unique Sequences and Key Virulence Genes with Unusual Rates of Amino Acid Substitution in Bovine *Staphylococcus aureus*

Lisa L. Herron,¹ Rajit Chakravarty,^{1†} Christopher Dwan,² J. Ross Fitzgerald,³ James M. Musser,³ Ernest Retzel,² and Vivek Kapur^{1*}

Department of Veterinary Pathobiology and Biomedical Genomics Center, University of Minnesota, St. Paul, Minnesota 55108¹; Computational Biology Center, University of Minnesota, Minneapolis, Minnesota 55455²; and Laboratory of Human Bacterial Pathogenesis, Rocky Mountain Laboratories, National Institute of Allergy and Infectious Disease, National Institutes of Health, Hamilton, Montana 59840³

Received 28 March 2002/Accepted 3 April 2002

***Staphylococcus aureus* is a major cause of mastitis in bovine and other ruminant species. We here present the results of a comparative genomic analysis between a bovine mastitis-associated clone, RF122, and the recently sequenced human-associated clones, Mu50 and N315, of *Staphylococcus aureus*. A shotgun sequence survey of ~10% of the RF122 genome identified numerous unique sequences and those with elevated rates of nonsynonymous substitution. Taken together, these analyses show that there are notable differences in the genomes of bovine mastitis-associated and human clones of *S. aureus* and provide a framework for the identification of specific factors associated with host specificity in this major human and animal pathogen.**

Staphylococcus aureus is a widely distributed animal and human pathogen. Phenotypic analyses of *S. aureus* strains isolated from human and bovine hosts have revealed important differences between the two populations of isolates, but the underlying genetic basis for the variation among *S. aureus* strains remains unknown (10, 14, 15). Multilocus enzyme electrophoretic analysis of 2,077 *S. aureus* isolates from bovine, ovine, and human hosts showed that only 6 of the 33 clusters included isolates from more than one host species (11). Population genetic analysis of 357 *S. aureus* isolates recovered from cases of bovine mastitis from global sources suggested that only eight clones represented 90% of the isolates, and these clones rarely overlapped with those responsible for human disease (5). To begin identifying the nature and scope of the genetic differences between the bovine and human staphylococcal genomes, we present here a comparative analysis of the genome content of the common bovine *S. aureus* clone RF122 (4) and the recently sequenced human pathogenic clones Mu50 and N315 (6).

A random shotgun genomic library was prepared as described previously (7), with the exception that RF122 genomic DNA was mechanically sheared with a Hydroshear device (GeneMachines, San Carlos, Calif.). The first 850 random shotgun clones of RF122, encoding 285,077 unique bases and representing approximately 10% of the total genome, were sequenced by using dideoxy terminator chemistry on ABI 3700 DNA sequencers (ABI, Foster City, Calif.) and assembled into 600 contigs consisting of one to six sequences and ranging in size from 82 to 1,828 bases using phredPhrap (P. Green, Uni-

versity of Washington, Seattle) (details of individual genes, gene names, and categories are available at <http://www.cbc.umn.edu/ResearchProjects/AGAC/Staph/staphhome.html>). The average GC content of the initial sequences was 33.95%, closely approximating the GC content of the completed *S. aureus* genomes (6). The range of GC contents of individual contigs was 21 to 73%. Fourteen contigs had GC contents greater than 45%. Of these, two had no significant hits ($P < e^{-15}$) to any GenBank submission based on tBLASTn searches, five had significant homology to *S. aureus* genomic DNA, and one was homologous to an *S. aureus* plasmid (1). The remaining five contigs had significant ($P < e^{-15}$) homologies to sequences of organisms other than staphylococci and may reflect recent horizontal transfer events.

Identification of unique sequences in RF122. Twelve sequences without homology to any existing sequence in the comprehensive microbial databases were identified in RF122. An additional seven contigs with no homology to *S. aureus* but with close matches to genes from other organisms were identified, including homologs to a *Shigella* transposon, a pathogenicity island element from *Escherichia coli*, a *Rhodospseudomonas palustris* ferredoxin transporter, and a hypothetical protein from *Mesorhizobium loti*. Other orthologs matched the gram-positive organisms *Streptococcus agalactiae* (*sodA*), *Streptococcus pyogenes* (putative phage terminase), *Enterococcus faecalis* (putative invertase), and *Bacillus pumilus* (Tn 10 transposon). Since the present analysis represents sequences from only ~10% of the RF122 genome, these data suggest that there are likely to be 100 or more sequences that differentiate the bovine *S. aureus* clone from its human counterparts.

Recent analysis of RF122 led to the identification of a novel putative pathogenicity island, SaPIbov (4). The 15,981-bp island encodes several virulence factors and toxins (4). As expected, our RF122 contigs included several SaPIbov components. Two of the homologous contigs matched no sequences

* Corresponding author. Mailing address: Department of Veterinary Pathobiology, University of Minnesota, 205 Veterinary Science, 1971 Commonwealth Ave., St. Paul, MN 55108. Phone: (612) 625-7712. Fax: (612) 625-5203. E-mail: vkapur@umn.edu.

† Present address: Biomedical Engineering Center, University of Wisconsin, Madison, WI 53706.

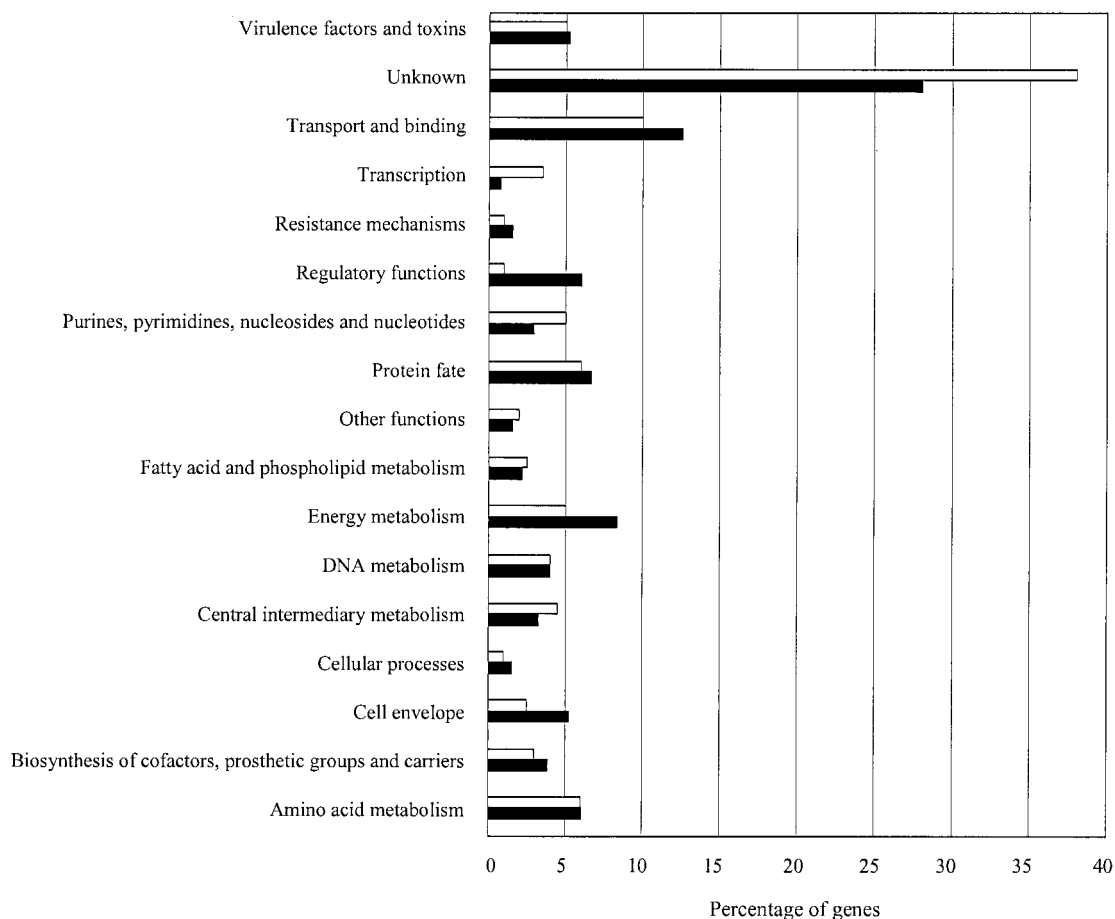


FIG. 1. Gene functions of RF122 (filled bars) were identified based on homology to existing GenBank submissions in tBLASTx analyses. Functional categories were putatively assigned based on the assignment of gene function in *B. subtilis* and the completed *S. aureus* genomes Mu50 and N315. Bars representing Mu50 (open bars) show the approximate percentage of genes in each corresponding category. A breakdown of individual genes, gene names, and categories is available at <http://www.cbc.umn.edu/ResearchProjects/AGAC/Staph/staphhome.html>.

in either of the finished human *S. aureus* genomes, indicating possible bovine-specific components of SaPIbov. The genes in these contigs include homologs to *sel*, whose hypothetical protein product is similar to staphylococcal enterotoxin I, and to a phage terminase homolog.

We were able to assign putative functions to many of the gene products based on motifs identified in the homologs from the finished Mu50 and N315 *S. aureus* genomes and the genome of *Bacillus subtilis* (Fig. 1). In total, 28% of the identified homologs encoded proteins of unknown function. Other genes of interest include *bap*, a biofilm-related gene thus far identified only in bovine isolates, *drp35*, an antibiotic resistance determinant, and *rot*, a transcriptional repressor regulating toxin expression previously characterized only in human strains (2, 8, 9).

Patterns of nucleotide substitution. To understand the level and nature of nucleotide variation between RF122 and the recently sequenced human clones of *S. aureus*, amino acid substitution rates for the aligned sequences were calculated according to the algorithm of Nei and Gojobori (12). For genes with integral roles in bacterial survival and fitness, the rate of nonsynonymous substitution (dN) is often lower

than the rate of synonymous substitution (dS) because these genes are under functional constraint and the majority of phenotypic changes will be disadvantageous. The majority of dN:dS ratios of the RF122 *S. aureus* sequences characterized thus far in comparison with Mu50 and N315 fall in the range of 1:10 to 1:4, corresponding to the most highly conserved sequences (Fig. 2A). The low average ratio of dN:dS for RF122 compared to Mu50 and N315 indicates that the majority of *S. aureus* genes are under functional constraint (Fig. 2B). Several of the highly conserved genes encoded virulence factors, including clumping factor B, as well as metabolic genes from energy, lipid, nucleic acid, and amino acid pathways (Table 1). The analysis also identified 19 orthologs with dN:dS ratios greater than 1, indicating that mutations that result in a change in the primary structure of the proteins have been conserved in these sequences. While the majority of the protein products from these genes were hypothetical, a urease subunit, an ammonium transporter, the cell division protein FtsH, the surface protein and antibiotic resistance determinant Drp35, and a leukotoxin subunit were also included in this category. The leukotoxin subunit is of particular interest in the context of host spec-

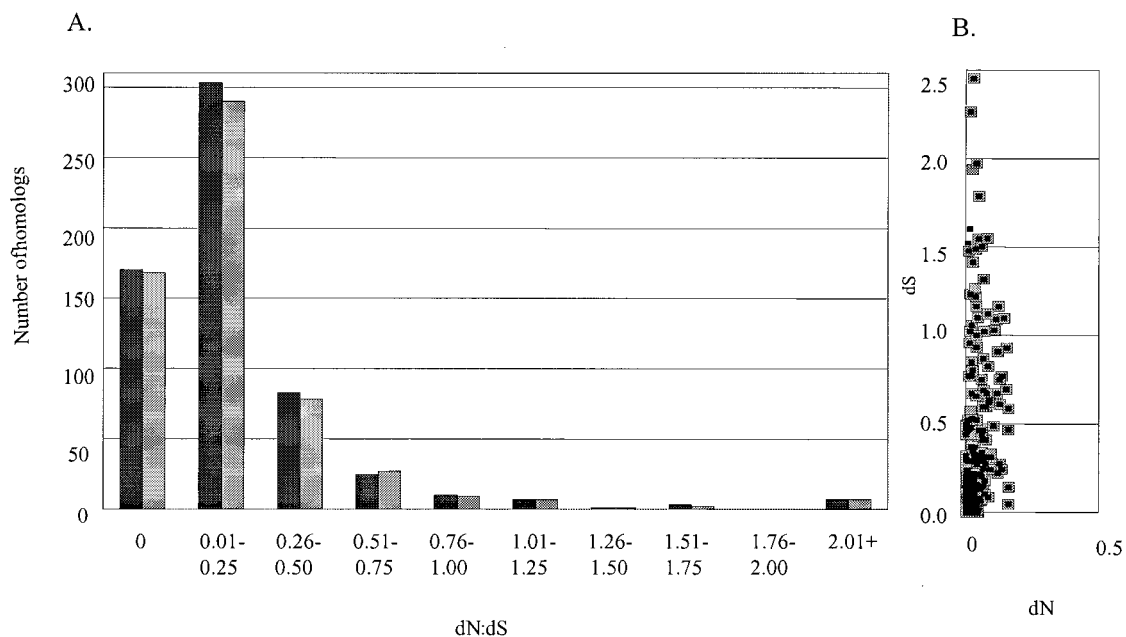


FIG. 2. (A) Distribution of dN:dS ratios among genes, calculated according to the algorithm of Nei and Gojobori (12), using the finished *S. aureus* genomes Mu50 (black bars) and N315 (gray bars) in comparison with the RF122 sequences. Ratios greater than 1.00 represent genes with a higher rate of nonsynonymous than synonymous substitution and constitute possible evidence of positive selection. Ratios of 0.00 result from genes without nonsynonymous substitutions, but the rate of synonymous substitution for these genes varies. (B) Synonymous substitution versus nonsynonymous substitution rates of RF122 compared with Mu50 (black squares) and N315 (gray squares).

ificity, as leukotoxins interact directly with host erythrocytes and phagocytes to cause potentially catastrophic inflammation. It has been suggested that leukotoxins may play a role in causing clinical mastitis, and leukotoxin genes are present in all bovine strains tested, but a causal relationship between leuko-

toxin production and mastitis remains to be established (3, 13). Should leukotoxins be a significant factor in causing mastitis, positive selection of phenotype-altering mutations in the F subunit of *S. aureus* leukotoxin may be related to differences in the bovine and human immunological environments.

TABLE 1. Substitution rate ratios among RF122 genes and corresponding functional categories

dN:dS ratio and gene	Ratio	Product	Category
<0.035 (highly conserved)			
<i>fnb</i>	0.008	Fibrinogen-binding protein	Virulence factors
<i>clpL</i>	0.014	Clp proteinase chain	Protein fate
<i>sdrC</i>	0.016	Conserved hypothetical protein	Putative virulence factors
<i>guaA</i>	0.017	GMP synthase	Purines, pyrimidines, nucleosides, nucleotides
<i>sdrD</i>	0.019	Conserved hypothetical protein	Putative virulence factors
<i>clfB</i>	0.019	Clumping factor B	Virulence factors
<i>gntK</i>	0.024	Glucokinase	Energy metabolism
<i>hlgC</i>	0.025	Gamma hemolysin component C	Virulence factors
<i>opuCD</i>	0.025	Hypothetical protein	Putative transport and binding protein
<i>sdrE</i>	0.026	Conserved hypothetical protein	Putative virulence factors
<i>lpl2</i>	0.030	Hypothetical protein	Putative virulence factors
<i>apt</i>	0.033	Adenine phosphoribosyl transferase	Purines, pyrimidines, nucleosides, nucleotides
<i>aroC</i>	0.033	Chorismate synthase	Amino acid biosynthesis
<i>oppD</i>	0.034	Hypothetical protein	Putative transport and binding proteins
>1.00 (positive selection)			
<i>lukD</i>	1.115	Leukotoxin F subunit	Toxins
<i>ebhB</i>	1.467	Hypothetical cell wall enzyme	Cell surface components
<i>tag</i>	1.560	DNA-3-methyladenine glycosidase	DNA metabolism
<i>ftsH</i>	1.587	Cell division protein	Cellular processes
<i>ureC</i>	2.198	Urease alpha subunit	Central intermediary metabolism
SAB0296	2.450	Similar to exotoxin 4	Putative virulence factors
<i>drp35</i>	3.113	Surface protein	Antibiotic resistance
SAB0711	4.417	Unknown	Unknown
SAB025	5.324	Unknown	Unknown

Concluding comments. In sum, this preliminary analysis of the genome of the common clone of bovine *S. aureus* reveals the presence of numerous genes and sequences that differentiate this isolate from previously characterized strains of the species. Our investigation also provides strong evidence that the whole genome sequencing approach will build a solid foundation for future investigation of the mechanisms of virulence and host specificity of this important human and animal pathogen.

We thank Barbara May, Michael Paustian, and Ling-Ling Li for their assistance with the preparation and analysis of the library and sequences. We also appreciate the support of Megan Lillehei and Todd Markovitz at the Advanced Genetic Analysis Center at the University of Minnesota for assistance with automated DNA sequencing.

This research is funded by a competitive award from the USDA-NRI Sustaining Animal Health and Well-Being Program (to V.K.). L.L.H. is supported by NIH NIGMS Biological Process Technology Institute fellowship GM08347.

REFERENCES

1. Altschul, S. F., T. L. Madden, A. A. Schaffer, J. Zhang, Z. Zhang, W. Miller, and D. J. Lipman. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* **25**:3389–3402.
2. Cucarella, C., C. Solano, J. Valle, B. Amorena, I. Lasa, and J. R. Penades. 2001. Bap, a *Staphylococcus aureus* surface protein involved in biofilm formation. *J. Bacteriol.* **183**:2888–2896.
3. Fitzgerald, J. R., W. J. Meaney, P. J. Hartigan, and C. J. Smyth. 2000. Molecular population and virulence factor analysis of *Staphylococcus aureus* from bovine intramammary infection. *J. Appl. Microbiol.* **88**:1028–1037.
4. Fitzgerald, J. R., S. R. Monday, T. J. Foster, G. A. Bohach, P. J. Hartigan, W. J. Meaney, and C. J. Smyth. 2001. Characterization of a putative pathogenicity island from bovine *Staphylococcus aureus* encoding multiple superantigens. *J. Bacteriol.* **183**:63–70.
5. Kapur, V., W. M. Sischo, R. S. Greer, T. S. Whittam, and J. M. Musser. 1995. Molecular population genetic analysis of *Staphylococcus aureus* recovered from cows. *J. Clin. Microbiol.* **33**:376–380.
6. Kuroda, M., T. Ohta, I. Uchiyama, T. Baba, H. Yuzawa, I. Kobayashi, L. Cui, A. Oguchi, K. Aoki, Y. Nagai, J. Lian, T. Ito, M. Kanamori, H. Matsumaru, A. Maruyama, H. Murakami, A. Hosoyama, Y. Mizutani-Ui, N. K. Takahashi, T. Sawano, R. Inoue, C. Kaito, K. Sekimizu, H. Hirakawa, S. Kuhara, S. Goto, J. Yabuzaki, M. Kanehisa, A. Yamashita, K. Oshima, K. Furuya, C. Yoshino, T. Shiba, M. Hattori, N. Ogasawara, H. Hayashi, and K. Hiramatsu. 2001. Whole genome sequencing of methicillin-resistant *Staphylococcus aureus*. *Lancet* **357**:1225–1240.
7. May, B. J., Q. Zhang, L.-L. Li, M. L. Paustian, T. S. Whittam, and V. Kapur. 2000. Complete genomic sequence of *Pasteurella multocida*, Pm70. *Proc. Natl. Acad. Sci. USA* **98**:3460–3465.
8. McNamara, P. J., K. C. Milligan-Monroe, S. Khalili, and R. A. Proctor. 2000. Identification, cloning, and initial characterization of *rot*, a locus encoding a regulator of virulence factor expression in *Staphylococcus aureus*. *J. Bacteriol.* **182**:3197–3203.
9. Murakami, H., H. Matsumaru, M. Kanamori, H. Hayashi, and T. Ohta. 1999. Cell wall affecting antibiotics induce expression of a novel gene, *drp35*, in *Staphylococcus aureus*. *Biochem. Biophys. Res. Commun.* **264**:348–351.
10. Musser, J. M., P. M. Schlievert, A. W. Chow, P. Ewan, B. N. Kreiswirth, V. T. Rosdahl, A. S. Naidu, W. Witte, and R. K. Selander. 1990. A single clone of *Staphylococcus aureus* causes the majority of cases of toxic shock syndrome. *Proc. Natl. Acad. Sci. USA* **87**:225–229.
11. Musser, J. M., and R. K. Selander. 1990. Genetic analysis of natural populations of *Staphylococcus aureus*, p 59–67. In R. Novick and R. A. Skurray (ed.), *Molecular biology of the staphylococci*. VCH Publishers, New York, N.Y.
12. Nei, M., and T. Gojobori. 1986. Simple methods for estimating the numbers of synonymous and nonsynonymous nucleotide substitutions. *Mol. Biol. Evol.* **3**:418–426.
13. Schuberth, H., C. Krueger, H. Zerbe, E. Bleckmann, and W. Leibold. 2001. Characterization of leukocytotoxic and superantigen-like factors produced by *Staphylococcus aureus* isolates from milk of cows with mastitis. *Vet. Microbiol.* **82**:187–199.
14. Sompolinsky, D., Z. Samra, W. W. Karakawa, W. F. Vann, R. Schneerson, and Z. Malik. 1985. Encapsulation and capsular types in isolates of *Staphylococcus aureus* from different sources and relationship to phage types. *J. Clin. Microbiol.* **22**:828–834.
15. Sordelli, D. O., F. R. Buzzola, M. I. Gomez, L. Steele-Moore, D. Berg, E. Gentilini, M. Catalano, A. J. Reitz, T. Tollersrud, G. Denamiel, P. Jeric, and J. C. Lee. 2000. Capsule expression by bovine isolates of *Staphylococcus aureus* from Argentina: genetic and epidemiologic analyses. *J. Clin. Microbiol.* **38**:846–850.

Editor: V. J. DiRita