

Original Research Article

Evaluation of Apoptotic and Growth Inhibitory Activity of Phloretin in BGC823 Gastric Cancer Cell

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Abstract

Purpose: To evaluate the *in vitro* anti-proliferative activity and probable mechanism of phloretin in human gastric cancer BGC823 cell lines.

Methods: Phloretin was isolated from apple tree leaves and identified by ¹H-Nuclear Magnetic Resonance (NMR), ¹³C-NMR and electrospray ionization tandem mass spectrometry (ESI-MS). The inhibitory activity of the compound in BGC823 gastric cancer cells was determined by Methylthiazolyldiphenyl-tetrazolium bromide (MTT) assay and soft agar colony formation assay while its apoptotic activity was detected by fluorescence staining and Western blotting analysis.

Results: The results show that the inhibitory activity of phloretin in BGC823 gastric cancer cells was mediated by induction of apoptosis and down-regulation of Bcl2 (B-cell lymphoma 2) expression. The anti-proliferative effects of phloretin was dose-dependent and inhibited the growth of BGC823 gastric cancer cells by 73 % at 30 μ M; this effectively induced cleavage of anti-poly (ADP-ribose) polymerase (PARP) as well as downregulation of Bcl2 protein expression in BGC823 cells after 24 h treatment.

Conclusion: Phloretin is a promising preventive and therapeutic agent for gastric cancer.

Keywords: Phloretin, Gastric cancer, Apoptosis, β -Cell lymphoma 2, Anti-poly (ADP-ribose) polymerase

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INTRODUCTION

Gastric cancer is the second leading cause of cancer deaths and remains the fourth most common cancer worldwide [1]. The 5-year survival rate among gastric cancer patients is still < 40 %, which is mainly due to relapse and metastasis. Numerous factors are closely related with a high risk of gastric cancer, such as *Helicobacter pylori* infection, smoking, and iodine deficiency [2]. Furthermore, smoked foods, salt-rich foods and pickled vegetables are associated with a high risk of gastric cancer [3,4]. On the

other hand, fresh fruits and vegetables are associated with a lower risk of gastric cancer [4].

Phloretin is a chalcone, which is a kind of flavonoid recognized to have health-promoting benefits, including antioxidant [5], anti-inflammation [6], disease-preventing and chemopreventive activities [7]. Phloretin is mainly found in apples and strawberries [8-10]. It has numerous biological properties, including reduction of human platelets activity, competitive inhibition of sodium-glucose cotransporters (SGLTs), inhibition of cardiovascular disease and

anticarcinogenic activity [11,12]. Compared with other flavonoids, however, few studies have investigated the anti-tumor potential of phloretin. In this study, we investigated the inhibitory effects of phloretin in BGC823.

EXPERIMENTAL

Plant extracts

Apple leaf extracts (one pack, 500 g) were purchased from Snow biological technology co., LTD (Xi'an, China).

Equipment and chemicals

The ^1H - and ^{13}C -NMR spectra were recorded on a Bruker Avance-400 FT-NMR spectrometer, with TMS internal standard. ESI-MS were recorded on 3200 Q-trap ESI-MS spectrometer (ABI, American). Column chromatography was carried on silica gel (200-300 mesh) and Sephadex LH-20 (Pharmacia Co.). All the organic solvents used were of analytical grade and purchased from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China). Bcl2 and Bax were purchased from Santa Cruz Biotechnology, Inc. Anti- β -actin and Anti-poly (ADP-ribose) polymerase (PARP) were obtained from Sigma Chemical Company (Sigma-Aldrich). Immobilon Western Chemiluminescent HRP Substrate Kit was obtained from Millipore. The cell culture medium (RPMI-1640, Roswell Park Memorial Institute 1640) and fetal bovine serum (FBS) were purchased from Hyclone Company (Logan, Utah, USA). Acrylamide-bis solution (30 %; 29:1) was bought from Bio-Rad laboratories, Inc (Hercules, CA, USA). Protein content was tested using a BCA kit (Thermo Scientific).

Cell line and cultures

Human gastric cancer cell lines, BGC823, were purchased from Institute of Cell Biology (Shanghai, China). Cells were maintained in RPMI-1640 (Roswell Park Memorial Institute 1640).

Extraction and isolation of phloretin

Extract powder (200 g) was suspended in 2 L Millipore-filtered water and successively extracted with chloroform, ethyl acetate and n-butanol (3 x 2 L for each solvent), respectively. The ethyl acetate fraction (EAF) yielded 42 g dry extract. The 40 g EAF were purified on silica gel chromatography column eluted with CHCl_3 -MeOH gradient (from 20:1 to 4:1, v/v) and further purified by C-18 and Sephadex LH-20 to get the

main apple polyphenol compound (2.3 g). The compound was characterized by NMR using CD_3OD as solvent for measurement and ESI-MS (negative ionization TIC modes, the m/z values of the monitored ions were from 100 to 800) spectral analysis and identified as phloretin.

MTT analysis

MTT assay (Cell titer 96 \circ R Aqueous One Solution Cell Proliferation Assay, Promega) was used to assess the inhibition effect of phloretin. Briefly, BGC823 (2×10^3 /well) were seeded in 96-well plates. BGC823 cells were treated with different concentration of phloretin (10, 20 and 30 μM , respectively) for 24 h, then phloretin was removed, fresh culture media added for additional 5 days. The MTT assay was performed using iMarkmicroplate Absorbance Reader (Bio-RAD, Richmond, CA) according to the manufacturer's instructions.

Soft agar colony formation assay

Triplicate samples of cells (1×10^3) were re-suspended in 1 ml of RPMI-1640 medium containing 0.3 % low-melt agarose, supplemented with 10 % fetal bovine serum, 50 units/ml penicillin and 50 $\mu\text{g}/\text{ml}$ streptomycin. BGC823 cells were treated with 10, 20, 30 μM phloretin and plated on top of solidified layer with the same RPMI-1640 medium containing 0.6 % low-met agarose. Plates were incubated for 3 weeks at 37 $^\circ\text{C}$ in 5 % CO_2 in humidified incubator. Then colony formation was stained with 0.01 % crystal violet and photographed and counted.

Cell extraction and western blotting

Western Blots were performed according to the protocols described elsewhere [13]. Cell lysates were prepared with the lysis buffer followed by centrifugation. Approximately 30 μg of denatured protein was resolved on 12 % SDS-PAGE and electroblotted onto nitrocellulose membranes (Amersham, Salt Lake City, UT, USA). After blocking, membranes were incubated with antibodies against PARP at a dilution of 1:1000 or antibodies against Bcl2 at a dilution of 1:1000 or antibodies against Bax at a dilution of 1:1000 or antibodies against β -actin at a dilution of 1:5000 at 4 $^\circ\text{C}$ overnight followed by further incubation with a secondary antibody (1:2000). After washing with Tris-buffer saline containing 0.05 % Tween 20, the blots were detected by chemiluminescence followed by exposure to Kodak-X-Omat film (Shanghai, China).

Statistical analysis

Results are expressed as the means \pm SEM. Statistical significance was determined by Student's t test or a one-way or two-way analysis of variance (ANOVA) followed by Turkey's test, as appropriate using Graphpad Prism statistics software (Graphpad Software). A p -value of < 0.05 was considered statistically significant ($*p < 0.05$, $**p < 0.01$, $***p < 0.001$).

RESULTS

NMR and ESI-MS spectral analysis the main apple polyphenol

The compound was obtained as a white powder, and its UV spectrum showed λ_{max} at 285 nm; ESI-MS yielded a quasi-molecular ion peak $[M-H]^-$ at m/z 273.2 $[M - H]^-$. 1H -NMR (400 MHz, CD_3OD) spectrum showed an AA'BB' system at δ 6.59 ppm (2H, d, $J = 7.0$ Hz, H-3', 5') and 7.01 (2H, d, $J = 7.0$ Hz, H-2', 6'). Two meta coupled protons at δ 5.91 (1H, brs, H-8) and 6.04 (1H, brs, H-6). Two methylene signals at δ 3.20 (2H, t, $J = 7.6$ Hz, H-3) and 2.58 (2H, t, $J = 7.6$ Hz, H-2). ^{13}C -NMR (100 MHz, CD_3OD) δ : 29.4 (C-2), 45.6 (C-3), 114.6 (C-3', 5'), 129.0 (C-2', 6'), 132.4 (C-1'), 154.9 (C-4'), 94.0 (C-6), 96.9 (C-8), 105.3 (C-10), 155.9 (C-5), 164.4 (C-7), 165.8 (C-9), 205.1 (C-4). On the basis of these results, the structure of the apple's main polyphenol compound was identified as phloretin (Fig 1) [10].

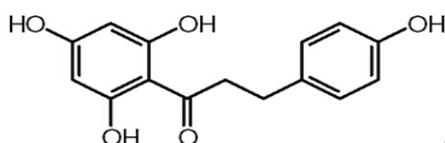


Figure 1: Chemical structure of phloretin

Phloretin inhibits gastric cancer cell growth

The result of MTT assay showed that BGC823 proliferation was inhibited by treatment with phloretin (Fig 2). The phloretin caused a sustained growth arrest of BGC823 in dose-dependent manner.

Soft agar colony formation assay

Formation assay revealed the phloretin's anti-proliferation effects in dose-dependent manner in BGC823 (Fig. 3). Phloretin effectively inhibited soft agar formation in BGC823 gastric cancer cell at the concentration of 10, 20 and 30 μM .

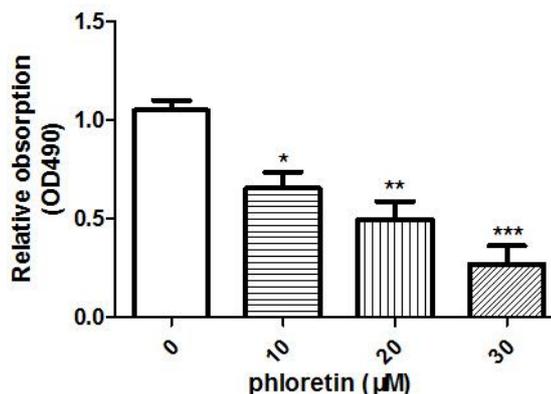


Figure 2: Anti-proliferation activity of phloretin in BGC823 cells. Data represent mean normalized optical densities \pm SEM ($n = 3$); $*p < 0.05$, $**p < 0.01$, $***p < 0.001$ vs control

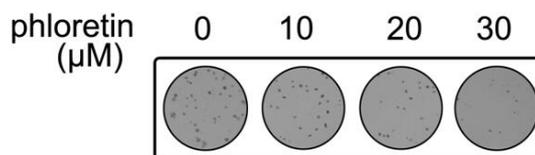


Figure 3: Representative photograph of colony formation. Phloretin inhibited the colony formation capacity in BGC823 in a dose-dependent manner

Phloretin induces apoptosis of BGC823 cells and modulates apoptosis-related protein expression

To investigate whether phloretin induce the apoptosis of gastric cancer cell, BGC823 cells were treated with 20 and 30 μM phloretin for 24 h and then stained by DAPI (4,6-diamidino-2-phenylindole) to investigate nuclear morphological changes under the treatment of phloretin. As observed in Fig 4, the untreated BGC823 cells displayed normal, healthy nuclear shapes, however the cells treated with 20 and 30 μM phloretin for 24 h showed morphological signs of apoptosis, which included chromatin condensation on the nuclear membrane (as indicated by arrow).

To confirm the induction of apoptosis and the anti-proliferation mechanism of phloretin in BGC823 gastric cancer cells, we further analyzed the protein expression of PARP, Bcl2 and Bax. Our results showed that 20 and 30 μM phloretin effectively induced the cleavage of PARP in BGC823 cells after 24 h treatment (Fig 5). Furthermore, Bcl2 and Bax protein expression was detected by western blot. BGC823 treated with 20 and 30 μM phloretin led to downregulation of Bcl2 and upregulation of Bax.

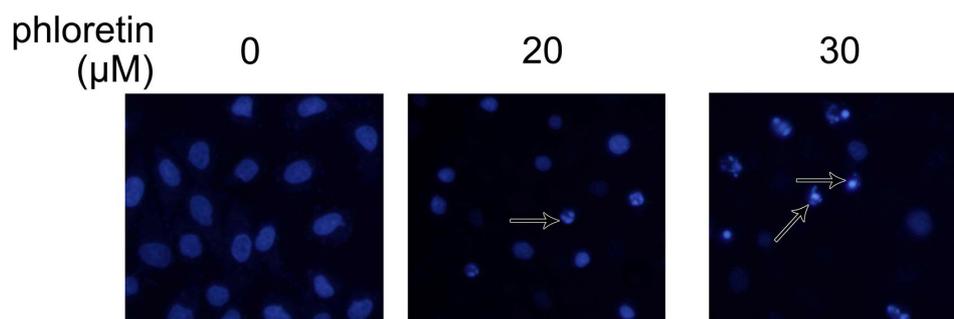


Figure 4: Apoptosis induced by phloretin in BGC823 cells. BGC823 cells treated with 20 and 30 μM phloretin for 24 h and immunostained with DAPI. **Note:** An arrow points to apoptotic cells

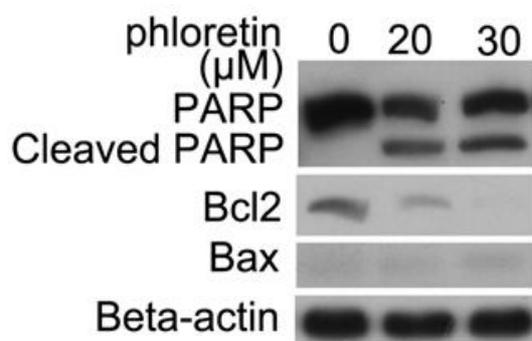


Figure 5: Apoptosis induced by phloretin in BGC823 cells, as detected by Western blot analysis of PARP, Bcl2, Bax in BGC823 cells treated with 20 and 30 μM phloretin for 24 h

lation of anti-apoptotic protein Bcl2, at the same time proapoptotic protein Bax expression was constant (Fig 5).

DISCUSSION

Gastric cancer is still the second leading cause of cancer deaths, in spite of the application of numerous diagnostic tools [14]. Antioxidant intake, especially fresh fruits and vegetables have a preventive effect on stomach tumorigenesis. Phloretin is a chalcone, a type of natural flavonoid found in fruits and vegetables. Flavonoids are well known as antioxidant agents. Phloretin has been reported to inhibit liver cancer [15], breast cancer [16], colon cancer [17], while, its antineoplastic effect on gastric cancer cell is still unclear. In our study we found that 10, 20, 30 μM phloretin significantly inhibit BGC823 gastric cancer cell proliferation by MTT assay. Phloretin (30 μM) almost completely inhibited soft agar colony formation in BGC823. Apoptosis induced by phloretin is an important way to inhibit the cancer cell growth.

Microscopy has been employed to investigate cytological appearance changes and to detect

the cleavage of PARP by Western blotting. Cleavage of PARP, which is a DNA repair enzyme and facilitates cellular disassembly, is the markers of the cells that undergoing apoptosis. The results indicated that incubation of BGC823 cells with 20 and 30 μM phloretin for 24 h resulted in numerous cells that had smaller nuclei with chromatin condensation and perinuclear apoptotic bodies. PARP levels in BGC823 cells were detected after exposure to phloretin by western blotting. The results indicate that 20 and 30 μM phloretin can induce PARP cleavage in a dose-dependent manner. The apoptotic-preventing protein Bcl2 and the pro-apoptotic protein Bax were also detected; the Bcl2/Bax ratios determine sensitivity to different apoptotic stimuli. In present study, 20 and 30 μM phloretin resulted in downregulation of Bcl2 protein expression; however, Bax protein expression showed no change.

Taken together, the results show that phloretin inhibits BGC823 proliferation through downregulation of Bcl2 expression and induction of apoptosis.

CONCLUSION

Phloretin is the main polyphenol compound of apple plant and also is a promising cancer preventive and therapeutic agent for gastric cancer. Further studies are, however, required ascertain the clinical efficacy of the compound in gastric cancer therapy.

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