

Review

The Protein Kinase C Agonist PEP005 (Ingenol 3-Angelate) in the Treatment of Human Cancer: A Balance between Efficacy and Toxicity

Elisabeth Ersvaer ¹, Astrid Olsnes Kittang ^{1,2}, Peter Hampson ³, Kristoffer Sand ¹,
Bjørn Tore Gjertsen ^{1,2}, Janet M. Lord ³ and Øystein Bruserud ^{1,2,*}

¹ Section for Hematology, The University of Bergen, Bergen, Norway;

E-Mails: elisabeth.ersvar@med.uib.no (E.E.); astrid.Olsnes@med.uib.no (A.O.K.);

kesand@gmail.com (K.S.); bjorn.gjertsen@med.uib.no (B.T.G.)

² Division of Hematology, Department of Medicine, Haukeland University Hospital, Norway

³ School of Immunity and Infection, Birmingham University Medical School, Birmingham, UK;

E-Mails: pxh048@bham.ac.uk (P.H.); J.M.Lord@bham.ac.uk (J.M.L.)

* Author to whom correspondence should be addressed; E-Mail: oystein.bruserud@helse-bergen.no;
Tel.: +47-559-72-997; Fax: +47-559-75-890.

Received: 30 November 2009; in revised form: 7 January 2010 / Accepted: 18 January 2010 /

Published: 22 January 2010

Abstract: The diterpene ester ingenol-3-angelate (referred to as PEP005) is derived from the plant *Euphorbia peplus*. Crude euphorbia extract causes local toxicity and transient inflammation when applied topically and has been used in the treatment of warts, skin keratoses and skin cancer. PEP005 is a broad range activator of the classical (α , β , γ) and novel (δ , ϵ , η , θ) protein kinase C isoenzymes. Direct pro-apoptotic effects of this drug have been demonstrated in several malignant cells, including melanoma cell lines and primary human acute myelogenous leukemia cells. At micromolar concentrations required to kill melanoma cells this agent causes PKC-independent secondary necrosis. In contrast, the killing of leukemic cells occurs in the nanomolar range, requires activation of protein kinase C δ (PKC δ) and is specifically associated with translocation of PKC δ from the cytoplasm to the nuclear membrane. However, in addition to this pro-apoptotic effect the agent seems to have immunostimulatory effects, including: (i) increased chemokine release by malignant cells; (ii) a general increase in proliferation and cytokine release by activated T cells, including T cells derived from patients with chemotherapy-induced lymphopenia; (iii) local infiltration of neutrophils after topical application with increased antibody-dependent cytotoxicity; and (iv) development of specific anti-cancer immune responses by

CD8⁺ T cells in animal models. Published studies mainly describe effects from *in vitro* investigations or after topical application of the agent, and careful evaluation of the toxicity after systemic administration is required before the possible use of this agent in the treatment of malignancies other than skin cancers.

Keywords: cancer-protein kinase C-PEP005

1. Introduction

A wide range of carcinogenesis-associated molecules are now investigated as possible therapeutic targets in human malignancies. These possible targets are usually mediators that show an altered expression in cancer cells or they affect essential cancer cell functions, e.g., regulation of proliferation or viability [1,2]. The pharmacological agents investigated can be either molecules known to, or designed to interact with the possible targets. An alternative strategy is to search for new anticancer agents in preparations used in traditional medicine, identify the active compound(s) and characterize their molecular effects [3].

PEP005 (ingenol 3-angelate) is derived from the plant *Euphorbia peplus* and crude euphorbia extracts have been used for centuries in traditional Thai and Australian medicine for treating various skin conditions, including warts, keratoses and cancers [3,4]. Fractionation of the sap yielded several macrocyclic diterpenes with cytotoxic activity or the ability to influence cellular differentiation, and ingenol-3-angelate thus emerged as a possible anti-cancer agent. This hydrophobic diterpene ester is now referred to as PEP005; it is strongly cytotoxic at high concentrations (100 µg/mL) [3] and at lower concentrations of 10–100 ng/mL it is a selective activator of Protein kinase C (PKC) [4].

2. The Protein Kinase C Family

2.1. Classification and Characterization of Protein Kinase C Isoenzymes

The PKC family was first distinguished by their status as cyclic nucleotide-independent kinases [5,6] and is now a complex family of at least 11 phospholipid-dependent serine/threonine kinases with distinct functions and tissue distribution [7–10]. PKC isoenzymes consist of a single polypeptide chain with a C-terminal kinase domain and a regulatory N-terminal domain that interacts with phosphatidylserine, Ca²⁺, diacylglycerol, phorbol ester and/or other lipids [9]. The PKCs can be activated by a wide range of signals, including release of second messengers during lipid-mediated signaling, other signaling pathways like the PI3K-pathway, direct molecular binding to for example ceramide, crosstalk between PKC isoenzymes, reactive oxygen species or proteolytic cleavage by caspases (for references see [8]). PKCs have been regarded as possible participants in carcinogenesis, even though PKC mutations are very uncommon in human cancers [8]. Members of the PKC family are classified as conventional, novel or atypical PKCs, depending upon their co-factor requirements (Table 1) [7]. It can also be seen from the table that the various PKCs have different effects on apoptosis, though most isoenzymes have anti-apoptotic effects [11].

Table 1. Classification of PKC isoforms (8–11).

	Classical isoforms cPKC	Novel isoforms nPKC	Atypical isoforms aPKC
Members	α , β_I , β_{II} , γ	δ , ϵ , η , μ , θ	ζ , ι/λ
Phorbol ester activation	Yes	Yes	No
Regulatory cofactors	Diacylglycerol Phosphatidyl-serine Ca	Diacylglycerol Ca-independent	Independent of Ca and diacylglycerol
Effect on apoptosis	Antiapoptotic: α , β_I , β_{II}	Antiapoptotic: ϵ Proapoptotic: δ	Antiapoptotic: ζ

Although PKC mutations are very uncommon in human cancers, the expression of various PKCs, including PKC δ , is often altered in human cancers, as illustrated by the data summarized in Table 2 [12–36]. Under physiological conditions triggering of phospholipase C activation leads to increased Ca²⁺ and diacylglycerol levels in the cell. These mediators can activate PKC, leading to a wide range of cellular events, depending on the isoenzyme activated [37,38]. In cancer, PKC α and β have been linked to increased invasion, proliferation, drug resistance and genetic instability [37], and like PKC ϵ , they are thought to be oncogenes [39]. PEP005 is a PKC agonist primarily achieving its pro-apoptotic effects through PKC δ , but its effects on intracellular signaling networks will also be influenced by the levels and activation of the other PKCs.

Table 2. Altered PKC expression in human cancers.

PKC isoform	Tumor Type	Expression	References
<i>Classical</i>			
PKC-α	Bladder	Increased	[12]
	Brain	Decreased	[13]
	Brain	Increased	[14]
	Breast	Decreased	[15,16]
	Ovarian	Decreased	[17]
	Renal	Decreased	[18]
	Colon	Decreased	[19]
	T-cell leukemia	Decreased	[20]
PKC-β	Bladder	Decreased	[12]
	Colon	Decreased	[21–23]
	Prostate	Decreased	[24]
	T-cell leukemia	Decreased	[20]
	Melanoma	Decreased	[25]
PKC-β_I	Bladder	Decreased	[26]
PKC-β_{II}	Bladder	Decreased	[27]
	Colon	Decreased	[28]
	DLBCL	Increased	[29]

Table 2. Cont.

<i>Novel</i>			
PKC-δ	Bladder	Decreased	[12,26,27]
	Brain	Decreased	[14]
	Colon	Increased	[23]
	Squamous cell carcinoma	Decreased	[30]
PKC-ϵ	Bladder	Increased	[12]
	Brain	Increased	[31]
	Breast	Increased	[32]
	Colon	Decreased	[23]
	Prostate	Increased	[24]
	Thyroid	Decreased	[33]
PKC-η	Breast	Decreased	[34,35]
	Colon	Decreased	[21]
	Renal	Increased	[18]
PKC-θ	Gastrointestinal stromal tumor	Increased	[36]

2.2. PKC δ and the Effects of PEP005

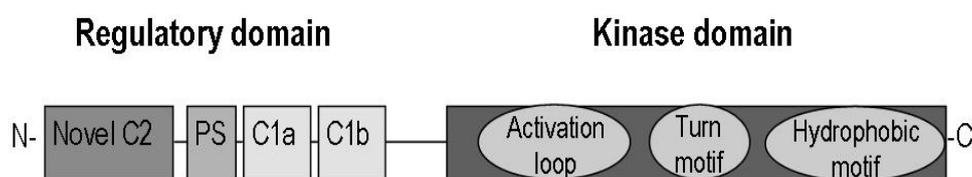
PEP005 is an activator of novel and classical PKC isoenzymes but its pro-apoptotic effects in leukemic cells rely upon the activation of PKC δ and its translocation from the cytoplasm to the plasma membrane, nuclear membrane and mitochondrial membrane in CHO-K1 cells and AML cell lines [4]. PKC δ activation can slow cell proliferation, induce cell cycle arrest and enhance differentiation in various undifferentiated cell lines. It also promotes apoptosis of malignant cells through: (i) activation of caspases and (ii) increased stability of p53 due to activation of I κ B-kinase and thereby increased p53 protein expression despite reduced p53 transcription [37,39–41]. In addition to these functions, PKC δ has been reported to phosphorylate up to 10 different signaling molecules, in addition to mitochondrial and nuclear proteins [39–51]. For example, it has been shown that in response to apoptotic stimuli such as cytarabine, PKC δ translocates to the nucleus where it co-localizes with and phosphorylates lamin B leading to dissolution of the nuclear lamina, and that this could be reduced following PKC δ inhibition. These signals can result in a broad variety of cellular effects, together supporting the hypothesis that PKC δ activity plays a role in regulating the balance between cell proliferation and apoptosis [38–51].

PKC $\delta^{-/-}$ mice develop normally and are fertile, suggesting that PKC δ plays minor roles during development, or that its actions can be taken over by another PKC isoenzyme [42,48]. In contrast, PKC δ seems to play important roles in normal hematopoiesis and oncogenesis. PKC isoforms α , β I, δ , ϵ , ζ and η are all expressed in myeloid cells [38]. Recently, *in vitro* studies have suggested that PKC δ together with PKC α can be essential for monocyte differentiation [42,43]. The human PKC δ gene is located on the short arm of chromosome 3 in a region where there is loss of heterozygosity in many epithelial cancers, suggesting that down regulation of PKC δ contributes to tumor progression [41,43,44,51]. On the other hand, elevated PKC δ expression has been described in multiple myeloma

[40], and overexpression of phosphorylated-PKC is found in nearly half of acute myelogenous leukemia (AML) patients [42].

The molecular structure of the PKC δ isoenzyme is shown in Figure 1 [45]. The intracellular compartmentalization of PKC δ depends upon its post-transcriptional modulation, and PKC δ -mediated signaling has pro-apoptotic effects through several pathways (Figure 2) [11,46,47]. Briefly, translocation of the enzyme from the cytoplasm to the nucleus seems crucial to its pro-apoptotic actions. Initial phosphorylation of the enzyme on tyrosine residues occurs in response to apoptotic stimuli and activated PKC δ accumulates in the nucleus together with activated caspase 3. PKC δ is cleaved by this caspase and a catalytic fragment is thereby formed. This fragment has constitutive activity, remains in the nucleus and induces apoptosis possibly through phosphorylation of apoptosis-regulating proteins. An alternative pro-apoptotic pathway is mediated through the endoplasmic reticulum and mitochondria (Figure 2).

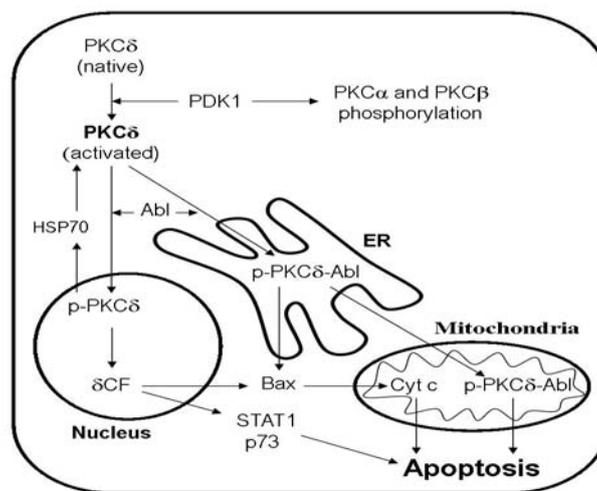
Figure 1. The molecular structure of PKC δ (adapted from [45]). The molecule has a regulatory and a kinase domain. The Novel C2 domain of the regulatory part is Ca²⁺ insensitive in contrast to the conventional PKC's C2 domains. The C1a and C1b parts can bind diacylglycerol (DAG) as well as phorbol esters. The pseudosubstrate (PS) domain has structural similarities to the substrate of the kinase domain and binds to the active site of the kinase domain. Binding of C2 and C1 to membrane structures will release the PS domain from the active site and make substrate binding possible. The Hinge domain is the cleavage site for Caspase 3, this cleavage occurs in the nucleus and results in the release of the δ -catalytic fragment (δ CF) that corresponds to the kinase domain. Phosphorylation of several tyrosine and serine residues both in regulatory and kinase domain has been described. The overall phosphorylation pattern determines the intracellular compartmentalization of the enzyme. Among the kinases involved in phosphorylation of PKC δ are the non-receptor tyrosine kinases Abl and Src like kinase-Lyn.



Several PKC isoenzymes show altered expression in human cancers as summarized in Table 2 [12–36]. Moreover, as some of these isoenzymes can have anti-apoptotic effects, whereas PKC δ is regarded as an important pro-apoptotic mediator, the ultimate outcome of altered expression will depend upon the balance between the activity of pro- and anti-apoptotic PKCs. [11,46,47]. In these models PKC α and PKC β had anti-apoptotic effects, and suppression of these enzymes caused induction of apoptosis with upregulation of pro-apoptotic PKC δ [46]. However, overexpression of PKC δ alone was not sufficient for induction of apoptosis. These observations clearly illustrate that the crosstalk between pro- and anti-apoptotic PKC isoforms is important, and the final effect of the PKC-agonist PEP005 may therefore depend upon the balance between the various isoenzymes present

within a tumor and the drug may be less effective in those tumors with increased levels of the anti-apoptotic isoforms (Table 2).

Figure 2. Intracellular compartmentalization of PKC δ . Phosphoinositide dependent kinase 1 (PDK1) is responsible for the initial activating phosphorylation of PKC δ ; this enzyme can also phosphorylate PKC α and PKC β as an initial activating event for these enzymes. If a pro-apoptotic signal is involved the activated PKC δ is thereafter translocated either to the nucleus or to the endoplasmic reticulum (ER). After nuclear translocation caspase 3 cleavage results in the formation of the δ CF fragment that has a pro-apoptotic effect either (i) *via* upregulation of Bax and subsequent mitochondrial release of cytochrome c, or (ii) *via* the cytoplasmic mediators STAT1 and p73. Alternatively, the activated PKC δ can be exported from the nucleus by a mechanism involving dephosphorylation and subsequent molecular stabilization by Heat shock protein 70 (HSP70). The translocation to ER is initiated through cytoplasmic association of activated PKC δ with the Abl kinase; this results in PKC δ phosphorylation and translocation of the p-PKC δ -Abl complex to ER where pro-apoptotic signaling is initiated either through Bax or through further translocation of the complex to the mitochondria [11,45–47].



2.3. The Phenotype of PKC δ Null Mice

As stated above PKC $\delta^{-/-}$ mice develop normally and are fertile [42–44]. However, studies using PKC δ null mice (PKC $\delta^{-/-}$) have given important insight into the role of PKC δ *in vivo*. Studies by Leitges and colleagues showed that vein segments from PKC $\delta^{-/-}$ mice, subsequently grafted onto the carotid arteries of recipient mice (either PKC $\delta^{-/-}$ or PKC $\delta^{+/+}$), lead to more severe atherosclerosis than was seen with PKC $\delta^{+/+}$ vein grafts [44]. The authors went on to show that atherosclerotic lesions in PKC $\delta^{-/-}$ mice contained significantly more smooth muscle cells (SMCs) than were found in the wild-type animals, and that this increased number of cells correlated with decreased SMC death in the lesions of PKC $\delta^{-/-}$ mice [44]. Finally, the authors demonstrated that SMCs from PKC $\delta^{-/-}$ mice were resistant to cell death after treatment with a number of apoptosis-inducing stimuli, including UV light, H₂O₂, and TNF- α [44]. A more recent study by Humphries and colleagues showed that γ -irradiation

induced apoptosis of parotid glands was reduced by 60% in PKC $\delta^{-/-}$ mice when compared to wild-type mice [48]. It was shown that primary parotid cells from PKC $\delta^{-/-}$ mice were defective in mitochondrial dependent apoptosis, as shown by a suppression of etoposide-induced cytochrome-c release. Moreover, apoptotic responsiveness was restored by re-introduction of PKC δ [49]. Both of these studies demonstrate a pro-apoptotic role of PKC δ *in vivo*. Other work with PKC $\delta^{-/-}$ mice has demonstrated a role for PKC δ in the negative regulation of B-cell proliferation [50]. In this study, mice that lacked PKC δ exhibited an expansion of B-lymphocytes leading to the formation of germinal centres in the absence of stimulation, and the rate of proliferation of B-lymphocytes in response to stimulation was greater in the PKC $\delta^{-/-}$ mice [49–50]. Similar studies showed that PKC δ deficiency prevented B-cell tolerance, allowing maturation and terminal differentiation of self reactive B-cells in the presence of tolerizing antigens [50]. Whether this was due to diminished apoptosis was not investigated.

3. The Importance of Neutrophil Recruitment and Humoral Immunity after Topical Application of Pep005 for Skin Cancer

Human neutrophils express the conventional PKCs α , β I, β II, the novel PKC δ and the atypical PKC ζ , and PKCs are important in neutrophil function [9]. PKC is involved in the activation of integrins as well as other adhesion molecules; it associates with several cytoskeletal components and thereby forms a functional bridge between the plasma membrane and the cytoskeleton.

3.1. PEP005 EFFECTS on Endothelial Cells

The recruitment of neutrophils to sites of inflammation usually occurs across the endothelial cells in postcapillary venules [52–54]. PEP005 induces the expression of IL1 β , TNF- α and the neutrophil chemotactic chemokine CXCL8 in mouse normal skin and skin tumors as well as in human keratinocytes, fibroblasts and melanoma cells [55]. These cytokines may then: (i) activate neighboring endothelial cells and thereby favor adhesion and transendothelial migration of circulating leukocytes; and (ii) create a chemotactic CXCL8 gradient that favors local recruitment of neutrophils [56].

An additional mechanism for PEP005-induced recruitment of neutrophils could be direct effects on the endothelial cells with increased expression of adhesion molecules and/or the induction of neutrophil-chemotactic cytokines. A recent study described increased transcriptional upregulation of the expression of E-selectin, ICAM-1 and CXCL8 in umbilical vein endothelial cells after exposure to PEP005 [56]. When using a flow-based adhesion assay PEP005 then caused increased adhesion of neutrophils to a level that was comparable to endothelial cells activated with TNF- α . The adhesion was dependent on E-selectin, was accompanied by a translocation of PKC δ from the cytosol to the perinuclear membrane, and siRNA knockdown of PKC δ abolished neutrophil recruitment [56].

Taken together these results suggest that PEP005 causes local recruitment of neutrophils through: (i) direct effects on endothelial cells with increased adhesion; (ii) indirect effects on the endothelial cells through local release of activating cytokines from neighboring cells; and (iii) the release of neutrophil chemotactic CXCL8 by endothelial cells and perivascular cells. Although one cannot exclude that umbilical cord and microvascular endothelial cells show functional differences, it seems

likely that all three mechanisms contribute to the inflammatory response to topical application of PEP005.

3.2. *The Anticancer Effect of Neutrophils after Topical Application of PEP005*

Topical treatment of skin tumors with PEP005 induces cancer cell necrosis followed by local inflammation characterized by neutrophil infiltration and release of reactive oxygen species [55]. The treatment also increases the levels of antitumor antibodies and thereby enhances tumor cell killing by antibody-dependent neutrophil cytotoxicity. The following observations have been made after implantation of tumor cells in mice followed by topical PEP005 treatment [3,55]:

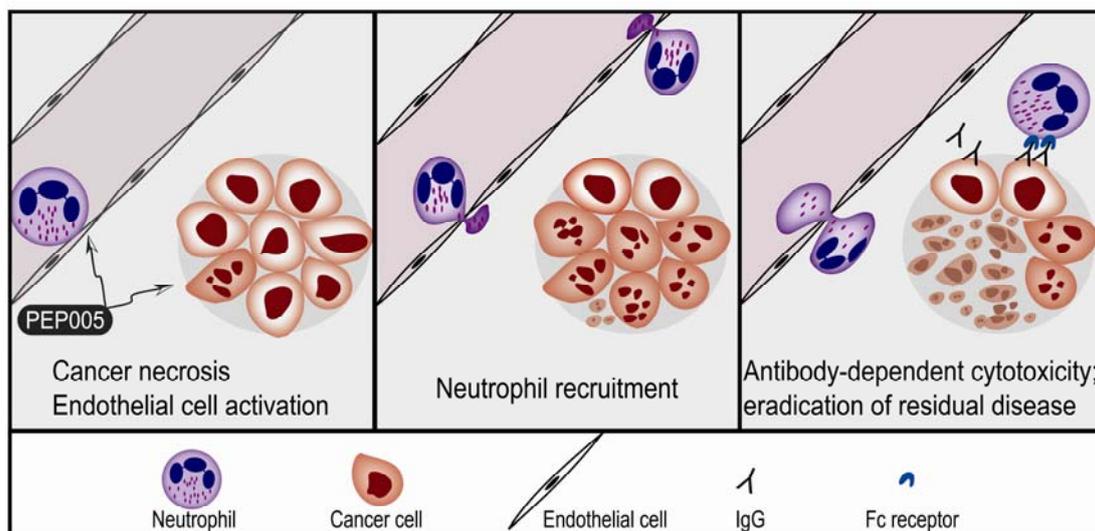
- Topical application of PEP005 can cure implanted skin cancers without later relapse in the T cell deficient Foxn1^{nu} mice. This effect is associated with local macroscopic inflammation due to leukocyte infiltration dominated by neutrophils. After antibody-depletion of neutrophils topical PEP005 treatment caused a similar initial ablation, but tumors later re-emerged.
- The neutrophil extravasation into the inflamed sites is severely impaired in CD18-deficient mice; topical treatment of implanted tumors in these animals was associated with initial cure followed by a weak local inflammation and later tumor relapse.
- NK cells and macrophages are present in Foxn1^{nu} mice, and macrophages are seen in PEP005 induced infiltrates. The local inflammation and relapse rate were not altered by depletion of NK cells. Neither inflammation nor relapse risk was altered for tumors implanted in Csfmop/Csfmop mice that lack functional M-CSF and therefore are severely monocytopenic.
- The effect of topical PEP005 was investigated for LK2 tumors implanted in SCID mice that lack a humoral immune system [55]. Tumors grew at similar rates and the initial tumor-ablative effect and local inflammatory reactions were similar to Foxn1^{nu} mice, but a high relapse rate was observed for the B cell-depleted mice.

Taken together these results suggest that topical application of PEP005 to skin tumors mediates anticancer effects through three distinct phases (Figure 3). Firstly, the initial tumor ablation is caused by a direct effect of the drug and local production of inflammatory cytokines [55]. The second phase is characterized by local inflammation due to neutrophil infiltration. During the third and last phase tumor-reactive antibodies are induced and relapses are avoided through antibody-dependent neutrophil cytotoxicity that eliminates remaining cancer cells [55].

3.3. *Clinical Studies of PEP005 in the Treatment of Skin Cancer*

Two randomized studies have investigated the short-course use of topical PEP005 in the treatment of actinic keratosis, a premalignant lesion that can progress to invasive squamous cell carcinoma [57,58]. Both studies concluded that topical application was effective and caused by local induction of necrosis and inflammation. The safety profile seems favorable, and treatment-related scarring was not a major problem. Thus, these studies support the conclusions from animal studies that topical PEP005 in the treatment of skin cancer is safe and effective and without systemic toxicity [3].

Figure 3. Effects of PEP005 in experimental skin cancer. (LEFT) Topical application of PEP005 causes high local drug concentrations with two direct effects; endothelial cell activation with neutrophil adhesion; and (ii) direct induction of necrosis in the malignant cells. (MIDDLE) There is transmigration and local neutrophil recruitment with a local inflammation. (RIGHT) Finally there is an antitumor humoral immune response leading to antibody-dependent cytotoxicity and eradication of residual cancer cells.



4. Antileukemic Effects of Pep005

4.1. Effects of PEP005 on Acute Myelogenous Leukemia Cells

Acute myelogenous leukemia (AML) is the human malignancy where the effects of low-dose PEP005 have been most extensively studied [59,60]. In most of these experiments PEP005 was used at a final concentration of 20 nM. The following effects were described for primary AML cells [59,60]:

- *Chemokine release.* Primary human AML cells show constitutive release of a wide range of chemokines [61,62]. PEP005 causes increased release of both CCL and CXCL chemokines, including CXCL8 that also was released at increased levels by skin cells after topical application (see above). The chemokines released at increased levels are pro-angiogenic and chemotactic not only for neutrophils but also for T cells and monocytes.
- *Chemokine receptor expression.* PEP005 has only minor effects on the expression of most CCR and CXCR receptors (CCR1-3, CCR5, CXCR2, 3), the only exception being CXCR4 that shows decreased expression. CXCR4 is one of the two receptors for the CXCL12 chemokine that is usually not released or only released at low levels by primary human AML cells [61]. However, it is released by bone marrow stromal cells [62]. The CXCL12/CXCR4 system is important for AML cell migration and CXCR4 expression seems to have an adverse prognostic impact in AML [63]. For this reason the PEP005 induced reduction in CXCR4 expression should possibly be regarded as an anti-leukemic effect.

- *Cytokine release.* Other cytokines are also released at higher levels, including Hepatocyte growth factor (HGF) and Granulocyte-macrophage colony-stimulating factor (GM-CSF) [59,60].
- *Differentiation.* PEP005 decreases the expression of stem cell markers (including CXCR4) and increases the expression of lineage-associated markers, an observation consistent with differentiation induction.
- *Apoptosis regulation.* PEP005 increases the expression of Bax and the activation of caspase 3. These pro-apoptotic effects are seen over a wide concentration range, whereas no induction of apoptosis was evident for normal CD34⁺ hematopoietic cells when testing concentrations up to 200 nM.
- *Intracellular signaling.* The effect in AML cells is mediated through a PKC δ agonistic effect. The ERK1/2 pathway then seems to be important for the increased chemokine release together with increased expression of the NF κ B subunits p50, p52 and p65.

Thus, the effect of PEP005 at these relatively low concentrations is mediated through induction of apoptosis and differentiation.

4.2. The Role of PKC in Other Leukemias

The effect of PEP005 has been investigated only in AML, but various PKC isoenzymes also seem to be important in other leukemias as reviewed by Redig and Plataniak [8]. In chronic lymphocytic leukemia (CLL) several PKCs are expressed in cells from most patients, including PKC β , PKC γ , PKC δ and PKC ζ and for some patients also PKC α , PKC ι and PKC ϵ . Global PKC inhibition induces apoptosis in CLL cells. So far there is no evidence for an important role of PKC δ in CLL and PKC α seems to be more important in regulation of proliferation and apoptosis in these cells [64]. Thus, the balance between pro- and anti-apoptotic isoforms may be important not only in solid tumors (Table 2) but also in hematological malignancies. The possible importance of PKC for disease development or chemosensitivity in other leukemias remains to be clarified.

5. Effects of PEP005 in Solid Tumors

5.1. Pharmacological *in Vitro* Studies

The studies described above demonstrate that PEP005 has an anticancer effect in different malignancies, but it should be emphasized that human cancer cells can also be generally resistant or the pro-apoptotic effect may be context-dependent [65]. In a recent study the effect of PEP005 on TRAIL (Tumor necrosis factor related apoptosis inducing ligand)-induced apoptosis was examined in human melanoma cell lines [66]. Enhancing or inhibitory effects on TRAIL-induced apoptosis were then observed depending on the cell line investigated, and the authors suggested that the effect of PEP005 in these models is not dependent on PKC δ alone but also on low expression of PKC ϵ .

Another study described induction of senescence in melanoma cells after *in vitro* exposure to PEP005; this additional pharmacological effect was observed for 20% of the cell lines [67]. This growth arrest involved signaling through ERK, the same pathway that seems responsible for the pro-

apoptotic and chemokine-increasing effects in AML cells (see above). The growth arrest seen with PEP005 treatment consisted of accumulation of cells in G₁ phase for up to 24 hours after *in vitro* exposure. Optimal combination of PEP005 with conventional cytotoxic drugs therefore seems to require a lag-time between exposure to the different drugs [67].

Resistance mechanisms to PEP005 have also been investigated in colon cancer cell lines that were cross-resistant to several chemotherapeutics [68]. PEP005 resistance seemed dependent on high expression of the small vasoactive peptide E1 that stimulates proliferation of colorectal cancer cells *via* the ETRA receptor. Other studies in colon cancer cells suggest that PEP005 can affect signaling through several pathways with increased phosphorylation of PKC δ , Raf1, ERK1/2, c-jun, p38, mitogen-activated protein kinase (MAPK) and PTEN [69]. These authors also described that PEP005 reduced the expression of PKC α and reduced the levels of the active phosphorylated form of Protein kinase B.

Taken together these observations suggest that PEP005 can affect several intracellular signaling pathways and that resistance may occur dependent upon the differential activity of pro- and anti-apoptotic pathways in individual patients and between different malignancies.

5.2. Studies in Animal Models

The effect of topical PEP005 has also been tested for other malignancies after skin implantation in Foxn1^{nu} mice [70]. These experiments demonstrated that PEP005 was effective not only against squamous cell carcinoma cells but also cells derived from human and murine melanoma, murine lung carcinoma, human prostate cancer and human cervical carcinoma [3]. Additional *in vitro* studies demonstrated that the drug could kill human breast cancer cells and T-leukemia cells, and for all these cell types the LD90s seemed comparable. The mechanisms behind the effects seem to be destabilization of endocytosed vesicles followed by endosome disruption with release of calcium into the cytoplasm and thereby mitochondrial swelling, disturbed energy metabolism, loss of mitochondrial membrane potential, rapid plasma membrane perturbation and cell death due to necrosis [3].

5.3. The Possibility of Topical Application for Other Cancers

Another possibility for topical treatment is bladder cancer [3]. The experience so far is limited, but *in vitro* studies suggest that normal urothelial cells may be less sensitive than bladder cancer cells. However, initial animal studies will be required because frequent inspection of the local inflammation is not possible in bladder cancer, and if severe hematuria occurred this complication may require specific therapeutic intervention.

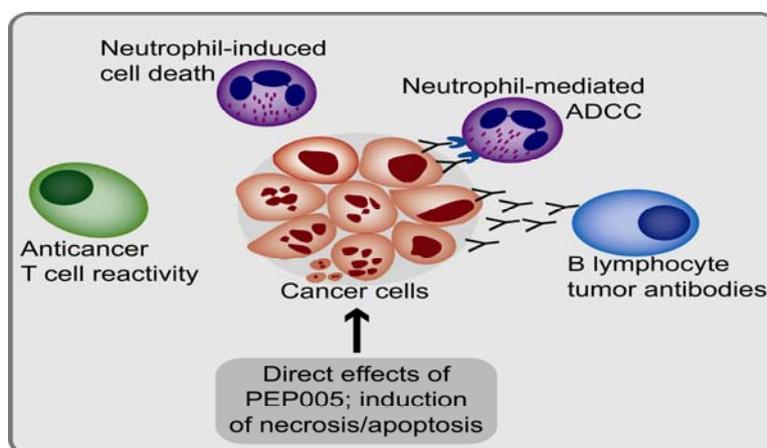
6. Immunomodulatory Effects of PEP005

The increased release of chemotactic chemokines that enhance recruitment of various leukocytes by PEP005 treated AML cells must be regarded as an immunostimulatory effect (Figure 3). The effects on neutrophils are described above. In addition PEP005 has direct T cell effects in AML derived cells resulting in: (i) increased proliferative T cell responses in cells from patients with untreated disease and patients with severe chemotherapy-induced panleukopenia, including severe T lymphopenia

[3,71]; and (ii) increased release of several cytokines by activated T cells, including IFN γ , GM-CSF, IL-2, IL-10, IL-13 and TNF- α , in cells from AML patients with chemotherapy-induced cytopenia (72). Thus, PEP005 *in vitro* seems to have both anti-leukemic and immunostimulatory effects in cells from AML patients and if this is extended to the *in vivo* situation, the immunostimulation could also include indirect effects through increased T cell recruitment [59] and direct T cell stimulatory effects [71,72].

Even though the early animal studies concluded that elimination of implanted tumors in mice were dependent on neutrophils and B cells (see above), a recent report reported that specific T cell responses can also be induced following local treatment [73]. Induction of a tumor-specific CD8⁺ response by PEP005 was observed, and this response contributed to regression of distant metastases. PEP005 was also found to have adjuvant properties and upregulated the expression of T cell costimulatory molecules CD80 and CD86 on dendritic cells. These observations further demonstrate that PEP005 has a broad immunostimulatory effect (Figure 4).

Figure 4. A summary of direct and indirect anticancer effects of PEP005. PEP005 has direct effects on malignant cells leading to either necrosis or apoptosis depending on the drug concentration. High concentrations are relevant for topical application, whereas lower concentrations are more relevant for leukemic disease. The indirect effects that may contribute to the anticancer effects are: (i) increased T cell reactivity, including increased cytokine release; (ii) local recruitment of neutrophils, endothelial cell activation contributes to this; (iii) induction of anticancer humoral immune responses with enhanced antibody-dependent cellular cytotoxicity (ADCC).



7. Concluding Remarks: Efficiency *versus* Toxicity in the future Use of PEP005 in Cancer Treatment

The overall literature described above suggests that PEP005 can mediate anticancer effects in different malignancies (Figure 4), but it should be emphasized that except for local application in skin tumors most of the present evidence comes from experimental *in vitro* studies. In contrast, the immunostimulatory effect is documented both in experimental models, *in vitro* studies of human T cells and after topical application in humans. In systemic therapy the immunostimulatory effects represent a beneficial effect with regard to anticancer activity but also a potential risk of toxicity if pro-inflammatory effects predominate. Testing of PEP005 in animal models of leukemia is now required to

determine if the compound applied systemically can achieve its anti-leukemic effects without significant toxicity.

7.1. Combination of PEP005 with Conventional Chemotherapy

PEP005 has been used as a single agent therapy in the topical treatment of skin diseases. Preclinical studies suggest that a combination of PKC δ agonists with conventional chemotherapy should be considered in human cancer therapy. PKC δ activation is induced after exposure of leukemia cells to etoposide, this is also observed after exposure of myeloid leukemia cells to Interferon- α , and in addition PKC δ seems important for anthracyclin-induced pro-apoptotic signaling [74,75]. Both etoposide and anthracyclines are widely used in the treatment of several other malignancies, including lymphomas and solid tumors, and combination therapy may therefore result in additive or supra-additive effects.

7.2. The proinflammatory Effects of PEP005, A Possible Risk during Systemic Therapy

The proinflammatory effect of PEP005 is clearly seen after topical therapy and involves neutrophils, B cells and T cells. Generally, great care should be taken if a drug with known proinflammatory effects is tried in systemic therapy. A dramatic example was the monoclonal antibody TGN1412, an anti-CD28 specific IgG₄ antibody [76–79]. Its preclinical screening showed no evidence for severe proinflammatory reactivity, but the phase I study in healthy volunteers resulted in severe multiorgan failure within hours after administration. The pathogenesis was massive cytokine release. Similar reactions have also been observed in other patients and with other agents, e.g., the use of the CD20 specific antibody rituximab in the induction treatment of patients with lymphocytic leukemia [80]. Thus, such reactions are not specific for the TGN1412 antibody but can also be seen with other agents. PEP005 is a drug with known proinflammatory effects, and for these reasons systemic administration of PEP005 has to be done with great care.

The clinical studies of topical PEP005 therapy showed no evidence for systemic effects. However, when using topical application to other body surfaces the risk of increased absorption must be considered. As an example, PEP005 is now considered for the treatment of bladder cancer [3]. The absorption from a relatively large urothelial surface may differ from the skin, and the risk of systemic effects has to be considered in the design of future clinical studies.

The increased chemokine release by AML cells after PEP005 exposure will also affect NK cells that express receptors for several CCL and CXCL chemokines [81]. It is not known whether this potentially proinflammatory effect will increase the risk of systemic toxicity when tried in AML.

7.3. Cancer-Directed Delivery of PEP005 in Systemic Therapy

One possible approach to avoid severe adverse events during systemic administration of proinflammatory drugs could be to direct the drug release towards the disease compartment. Several strategies may then be possible. In hematological malignancies the disease is usually detected throughout a large part of the body. These disorders often infiltrate diffusely throughout the bone marrow or affect several lymph node regions. Coupling of drugs to a disease-reactive monoclonal

antibody has been used to direct anticancer therapy, e.g., coupling to antibodies against the myeloid marker CD33 or the CD20 lymphoid marker [82–84]. Another possibility is to administer the drug in a form where drug release is only seen after therapeutic intervention; examples are drug release in a visualized tumor through local ultrasound treatment or photochemical therapy [85,86].

7.4. Sequential Treatment with Intensive Chemotherapy and PEP005; Decreased Risk for Proinflammatory-Induced Adverse Events?

Another possibility to avoid adverse events due to proinflammatory effects would be to administer the drug to severely immunocompromised cancer patients. Patients receiving treatment for acute leukemia develop a period of 2-3 weeks with severe leukopenia, and the risk of developing severe side effects may be less in such patients [87]. Furthermore, conventional cytotoxic drugs are often most effective against proliferating cells, and due to its antiproliferative effects PEP005 should possibly be administered sequentially with conventional chemotherapy to achieve a maximal anticancer effect. However, it should be emphasized that even patients with severe chemotherapy-induced cytopenia have an operative immune system [88], and a risk of proinflammatory side effects, though much reduced, is probably present even in such patients.

7.5. PEP005 effects on the Chemokine System—Advantage or Disadvantage?

Several chemokines that show altered release after PEP005 exposure have angioregulatory effects [59], but whether the drug will have pro- or anti-angiogenic effects will probably depend on several factors including: (i) additional local chemokine release by stromal cells, (ii) the overall cancer cell chemokine release profile, (iii) genetic polymorphisms within chemokine or chemokine receptor genes; and (iv) the concomitant expression of chemokine decoy receptors [59,61,89–93]. Matrix metalloproteinase (MMP) 2, 9 and 10 can also be constitutively released by primary human AML cells and may also contribute to leukemia-associated bone marrow angiogenesis, but PEP005 has only minor effects on this release [94].

The increased chemokine release may have proinflammatory effects that may represent a risk of toxicity. However, in certain clinical contexts the combination of anticancer and proinflammatory effects may be an advantage. Antileukemic immune reactivity is important for the reduced relapse risk after allogeneic stem cell transplantation, and increased T cell reactivity may then strengthen this antileukemic effect [90,93,95]. Whether modulation of the chemokine system will alter humoral immune reactivity is not known [90,96]. Finally, the possible leukemia-enhancing effect by increased CXCL12/CXCR4 expression may be counteracted by specific inhibitors [97], and this may become true also for other chemokines/chemokine receptors.

7.6. Final Comment

PEP005 has both anticancer and proinflammatory effects (Figure 4). These dual effects are an advantage in topical skin application, but it is not known whether the proinflammatory effects will represent an advantage or a disadvantage with risk of severe systemic toxicity after systemic therapy. Only extensive preclinical evaluation in relevant experimental models and careful design of clinical

studies can clarify whether systemic use of this drug will be acceptable with regard to the risk of toxicity.

Acknowledgements

This work was supported by the Norwegian Cancer Society (EE, AOK, BTG and ØB), Solveig and Ove Lunds Foundation (ØB) and a European commission integrated project LSHB-CT-2004-503467 (PH and JML).

References and Notes

1. Reikvam, H.; Ersvaer, E.; Bruserud, O. Heat shock protein 90 - a potential target in the treatment of human acute myelogenous leukemia. *Curr. Cancer Drug Targets* **2009**, *9*, 761–776.
2. Reikvam, H.; Olsnes, A.M.; Gjertsen, B.T.; Ersvær, E.; Bruserud, Ø. Nuclear Factor- κ B signaling - a contributor in leukemogenesis and a target for pharmacological intervention in human acute myelogenous leukemia. *Curr. Rev. Oncogen.* **2009**, in press.
3. Ogbourne, S.M.; Hampson, P.; Lord, J.M.; Parsons, P.; De Witte, P.A.; Suhrbier, A. Proceedings of the First International Conference on PEP005. *Anticancer Drugs* **2007**, *18*, 357–362.
4. Kedei, N.; Lundberg, D.J.; Toth, A.; Welburn, P.; Garfield, S.H.; Blumberg, P.M. Characterization of the interaction of ingenol 3-angelate with protein kinase C. *Cancer Res.* **2004**, *64*, 3243–3255.
5. Hug, H.; Sarre, T.F. Protein kinase C isoenzymes: Divergence in signal transduction? *Biochem. J.* **1993**, *291*, 329–343.
6. Ohno, S.; Akita, Y.; Konno, Y.; Imajoh, S.; Suzuki, K. A novel phorbol ester receptor/protein kinase, nPKC, distantly related to the protein kinase C family. *Cell* **1988**, *53*, 731–741.
7. Mellor, H.; Parker, P.J. The extended protein kinase C superfamily. *Biochem. J.* **1998**, *332*, 281–292.
8. Redig, A.J.; Plataniias, L.C. Protein kinase C signaling in leukemia. *Leuk. Lymphoma* **2008**, *49*, 1255–1262.
9. Selvatici, R.; Falzarano, S.; Mollica, A.; Spisani, S. Signal transduction pathways triggered by selective formylpeptide analogues in human neutrophils. *Eur. J. Pharmacol.* **2006**, *534*, 1–11.
10. Le Good, J.A.; Ziegler, W.H.; Parekh, D.B.; Alessi, D.R.; Cohen, P.; Parker, P.J. Protein kinase C isoforms controlled by phosphoinositide 3-kinase through the protein kinase PDK1. *Science* **1998**, *281*, 2042–2045.
11. Reyland, M.E. Protein kinase C δ and apoptosis. *Biochem. Soc. Transact.* **2007**, *35*, 1001–1004.
12. Varga, A.; Czifra, G.; Tállai, B.; Németh, T.; Kovács, I.; Kovács, L.; Bíró, T. Tumor grade-dependent alterations in the protein kinase C isoform pattern in urinary bladder carcinomas. *Eur. Urol.* **2004**, *46*, 462–465.
13. Benzil, D.L.; Finkelstein, S.D.; Epstein, M.H.; Finch P.W. Expression pattern of alpha-protein kinase C in human astrocytomas indicates a role in malignant progression. *Cancer Res.* **1992**, *52*, 2951–2956.

14. Mandil, R.; Ashkenazi, E.; Blass, M.; Kronfeld, I.; Kazimirsky, G.; Rosenthal, G.; Umansky, F.; Lorenzo, P.S.; Blumberg, P.M.; Brodie, C. Protein kinase Calpha and protein kinase Cdelta play opposite roles in the proliferation and apoptosis of glioma cells. *Cancer Res.* **2001**; *61*, 4612–4619.
15. Ainsworth, P.D.; Winstanley, J.H.; Pearson, J.M.; Bishop, H.M.; Garrod, D.R. Protein kinase C alpha expression in normal breast, ductal carcinoma *in situ* and invasive ductal carcinoma. *Eur. J. Cancer* **2004**, *40*, 2269–2273.
16. Kerfoot, C.; Huang, W.; Rotenberg, S.A. Immunohistochemical analysis of advanced human breast carcinomas reveals downregulation of protein kinase C alpha. *J. Histochem. Cytochem.* **2004**, *52*, 419–422.
17. Weichert, W.; Gekeler, V.; Denkert, C.; Dietel, M.; Hauptmann, S. Protein kinase C isoform expression in ovarian carcinoma correlates with indicators of poor prognosis. *Int. J. Oncol.* **2003**, *23*, 633–639.
18. Brenner, W.; Färber, G.; Herget, T.; Wiesner, C.; Hengstler, J.G.; Thüroff, J.W. Protein kinase C eta is associated with progression of renal cell carcinoma (RCC). *Anticancer Res.* **2003**, *23*, 4001–4006.
19. Verstovsek, G.; Byrd, A.; Frey, M.R.; Petrelli, N.J.; Black, J.D. Colonocyte differentiation is associated with increased expression and altered distribution of protein kinase C isozymes. *Gastroenterology* **1998**, *115*, 75–85.
20. Hidaka, M.; Nakakuma, H.; Kawaguchi, T.; Nagakura, S.; Horikawa, K.; Okuno, Y.; Kagimoto, T.; Takatsuki, K. Altered expression of protein kinase C in adult T-cell leukemia cells. *Int. J. Hematol.* **1992**, *56*, 135–141.
21. Doi, S.; Goldstein, D.; Hug, H.; Weinstein, I.B. Expression of multiple isoforms of protein kinase C in normal human colon mucosa and colon tumors and decreased levels of protein kinase C beta and eta mRNAs in the tumors. *Mol. Carcinog.* **1994**, *11*, 197–203.
22. Levy, M.F.; Pocsidio, J.; Guillem, J.G.; Forde, K.; LoGerfo, P.; Weinstein, I.B. Decreased levels of protein kinase C enzyme activity and protein kinase C mRNA in primary colon tumors. *Dis. Colon. Rectum.* **1993**, *36*, 913–921.
23. Pongracz, J.; Clark, P.; Neoptolemos, J.P.; Lord, J.M. Expression of protein kinase C isoenzymes in colorectal cancer tissue and their differential activation by different bile acids. *Int. J. Cancer* **1995**, *61*, 35–39.
24. Cornford, P.; Evans, J.; Dodson, A.; Parsons, K.; Woolfenden, A.; Neoptolemos, J.; Foster, C.S. Protein kinase C isoenzyme patterns characteristically modulated in early prostate cancer. *Am. J. Pathol.* **1999**, *154*, 137–144.
25. Gilhooly, E.M.; Morse-Gaudio, M.; Bianchi, L.; Reinhart, L.; Rose, D.P.; Connolly, J.M.; Reed, J.A.; Albino, A.P. Loss of expression of protein kinase C beta is a common phenomenon in human malignant melanoma: A result of transformation or differentiation? *Melanoma Res.* **2001**, *11*, 355–369.
26. Koren, R.; Langzam, L.; Paz, A.; Livne, P.M.; Gal, R.; Sampson, S.R. Protein kinase C (PKC) isoenzymes immunohistochemistry in lymph node revealing solution-fixed, paraffin-embedded bladder tumors. *Appl. Immunohistochem. Mol. Morphol.* **2000**, *8*, 166–171.

27. Langzam, L.; Koren, R.; Gal, R.; Kugel, V.; Paz, A.; Farkas, A.; Sampson, S.R. Patterns of protein kinase C isoenzyme expression in transitional cell carcinoma of bladder. Relation to degree of malignancy. *Am. J. Clin. Pathol.* **2001**, *116*, 377–385.
28. Kuranami, M.; Powell, C.T.; Hug, H.; Zeng, Z.; Cohen, A.M.; Guillem, J.G. Differential expression of protein kinase C isoforms in human colorectal cancers. *J. Surg. Res.* **1995**, *58*, 233–239.
29. Hans, C.P.; Weisenburger, D.D.; Greiner, T.C.; Chan, W.C.; Aoun, P.; Cochran, G.T.; Pan, Z.; Smith, L.M.; Lynch, J.C.; Bociek, R.G.; Bierman, P.J.; Vose, J.M.; Armitage, J.O. Expression of PKC-beta or cyclin D2 predicts for inferior survival in diffuse large B-cell lymphoma. *Mod. Pathol.* **2005**, *18*, 1377–1384.
30. D'Costa, A.M.; Robinson, J.K.; Maududi, T.; Chaturvedi, V.; Nickoloff, B.J.; Denning, M.F. The proapoptotic tumor suppressor protein kinase C-delta is lost in human squamous cell carcinomas. *Oncogene* **2006**, *25*, 378–386.
31. Sharif, T.R.; Sharif, M. Overexpression of protein kinase C epsilon in astroglial brain tumor derived cell lines and primary tumor samples. *Int. J. Oncol.* **1999**, *15*, 237–243.
32. Pan, Q.; Bao, L.W.; Kleer, C.G.; Sabel, M.S.; Griffith, K.A.; Teknos, T.N.; Merajver, S.D. Protein kinase C epsilon is a predictive biomarker of aggressive breast cancer and a validated target for RNA interference anticancer therapy. *Cancer Res.* **2005**, *65*, 8366–8371.
33. Knauf, J.A.; Ward, L.S.; Nikiforov, Y.E.; Nikiforova, M.; Puxeddu, E.; Medvedovic, M.; Liron, T.; Mochly-Rosen, D.; Fagin, J.A. Isozyme-specific abnormalities of PKC in thyroid cancer: Evidence for post-transcriptional changes in PKC epsilon. *J. Clin. Endocrinol. Metab.* **2002**, *87*, 2150–2159.
34. Beck, J.; Bohnet, B.; Brügger, D.; Bader, P.; Dietl, J.; Scheper, R.J.; Kandolf, R.; Liu, C.; Niethammer, D.; Gekeler, V. Multiple gene expression analysis reveals distinct differences between G2 and G3 stage breast cancers, and correlations of PKC eta with MDR1, MRP and LRP gene expression. *Br. J. Cancer* **1998**, *77*, 87–91.
35. Masso-Welch, P.A.; Winston, J.S.; Edge, S.; Darcy, K.M.; Asch, H.; Vaughan, M.M.; Ip, M.M. Altered expression and localization of PKC eta in human breast tumors. *Breast Cancer Res. Treat* **2001**, *68*, 211–223.
36. Blay, P.; Astudillo, A.; Buesa, J.M.; Campo, E.; Abad, M.; García-García, J.; Miquel, R.; Marco, V.; Sierra, M.; Losa, R.; Lacave, A.; Braña, A.; Balbín, M.; Freije, J.M. Protein kinase C theta is highly expressed in gastrointestinal stromal tumors but not in other mesenchymal neoplasias. *Clin. Cancer Res.* **2004**, *10*, 4089–4095.
37. Koivunen, J.; Aaltonen, V.; Peltonen, J. Protein kinase C (PKC) family in cancer progression. *Cancer letters* **2006**, *235*, 1–10.
38. Bassini, A.; Zauli, G.; Migliaccio, G.; Migliaccio, A.R.; Pascuccio, M.; Pierpaoli, S.; Guidotti, L.; Capitani, S.; Vitale, M. Lineage-restricted expression of protein kinase C isoforms in hematopoiesis. *Blood* **1999**, *93*, 1178–1188.
39. Steinberg S.F. Distinctive activation mechanisms and functions for protein kinase Cdelta. *Biochem. J.* **2004**, *384*, 449–459.
40. Yamaguchi, T.; Miki, Y.; Yoshida K. Protein kinase C delta activates IkappaB-kinase alpha to induce the p53 tumor suppressor in response to oxidative stress. *Cell Signal* **2007**, *19*, 2088–2097.

41. Abbas, T.; White, D.; Hui, L.; Yoshida, K.; Foster, D.A.; Bargonetti, J. Inhibition of human p53 basal transcription by down-regulation of protein kinase Cdelta. *J. Biol. Chem.* **2004**, *279*, 9970–9977.
42. Pearn, L.; Fisher, J.; Burnett, A.K.; Darley, R.L. The role of PKC and PDK1 in monocyte lineage specification by Ras. *Blood* **2007**, *109*, 4461–4469.
43. Jackson, D.N.; Foster, D.A. The enigmatic protein kinase Cdelta: Complex roles in cell proliferation and survival. *FASEB J.* **2004**, *18*, 627–636.
44. Leitges, M.; Mayr, M.; Braun, U.; Mayr, U.; Li, C.; Pfister, G.; Ghaffari-Tabrizi, N.; Baier, G.; Hu, Y.; Xu, Q. Exacerbated vein graft arteriosclerosis in protein kinase Cdelta-null mice. *J. Clin. Invest* **2001**, *108*, 1505–1512.
45. Newton, A.C. Regulation of the ABC kinases by phosphorylation: Protein kinase C as a paradigm. *Biochem. J.* **2003**, *370*, 361–371.
46. Zhu, T.; Tsuji, T.; Chen, C. Roles of PKC isoforms in the induction of apoptosis elicited by aberrant Ras. *Oncogene* **2009**, doi:10.1038/onc.2009.344.
47. Qi, X.; Mochly-Rosen, D. The PKC δ -Abl complex communicates ER stress to the mitochondria – an essential step in subsequent apoptosis. *J. Cell Sci.* **2007**, *121*, 804–813.
48. Humphries, M.J.; Limesand, K.H.; Schneider, J.C.; Nakayama, K.I.; Anderson, S.M.; Reyland, M.E. Suppression of apoptosis in the protein kinase Cdelta null mouse *in vivo*. *J. Biol. Chem.* **2006**, *281*, 9728–9737.
49. Miyamoto, A.; Nakayama, K.; Imaki, H.; Hirose, S.; Jiang, Y.; Abe, M.; Tsukiyama, T.; Nagahama, H.; Ohno, S.; Hatakeyama, S.; Nakayama, K.I. Increased proliferation of B cells and auto-immunity in mice lacking protein kinase Cdelta. *Nature* **2002**, *416*, 865–869.
50. Mecklenbräuer, I.; Saijo, K.; Zheng, N.Y.; Leitges, M.; Tarakhovsky, A. Protein kinase Cdelta controls self-antigen-induced B-cell tolerance. *Nature* **2002**, *416*, 860–865.
51. Zabarovsky, E.R.; Lerman, M.I.; Minna, J.D. Tumor suppressor genes on chromosome 3p involved in the pathogenesis of lung and other cancers. *Oncogene* **2002**, *21*, 6915–6935.
52. Zimmerman, G.A.; Prescott, S.M.; McIntyre, T.M. Endothelial cell interactions with granulocytes: Tethering and signaling molecules. *Immunol. Today* **1992**, *13*, 93–100.
53. Imhof, B.A.; Dunon, D. Leukocyte migration and adhesion. *Adv. Immunol.* **1995**, *58*, 345–416.
54. Springer, T.A. Traffic signals on endothelium for lymphocyte recirculation and leukocyte emigration. *Annu. Rev. Physiol.* **1995**, *57*, 827–872.
55. Challacombe, J.M.; Suhrbier, A.; Parsons, P.G.; Jones, B.; Hampson, P.; Kavanagh, D.; Rainger, G.E.; Morris, M.; Lord, J.M.; Le, T.T.; Hoang-Le, D.; Ogbourne, S.M. Neutrophils are a key component of the antitumor efficacy of topical chemotherapy with ingenol-3-angelate. *J. Immunol.* **2006**, *177*, 8123–8132.
56. Hampson, P.; Kavanagh, D.; Smith, E.; Wang, K.; Lord, J.M.; Ed Rainger, G. The anti-tumor agent, ingenol-3-angelate (PEP005), promotes the recruitment of cytotoxic neutrophils by activation of vascular endothelial cells in a PKC-delta dependent manner. *Cancer Immunol. Immunother.* **2008**, *57*, 1241–1251.
57. Siller, G.; Gebauer, K.; Welburn, P.; Katsamas, J.; Ogbourne, S.M. PEP005 (ingenol mebutate) gel, a novel agent for the treatment of actinic keratosis: Results of a randomized, double-blind, vehicle-controlled, multicentre, phase IIa study. *Austral. J. Dermatol.* **2009**, *50*, 16–22.

58. Anderson, L.; Schmieder, G.J.; Werschler, W.P.; Tschen, E.H.; Ling, M.R.; Stough, D.B.; Katsamas, J. Randomized, double-blind, double-dummy, vehicle-controlled study of ingenol mebutate gel 0.025% and 0.05% for actinic keratosis. *J. Am. Acad. Dermatol.* **2009**, *60*, 934–943.
59. Olsnes, A.M.; Ersvaer, E.; Rynningen, A.; Paulsen, K.; Hampson, P.; Lord, J.M.; Gjertsen, B.T.; Kristoffersen, E.K.; Bruserud, Ø. The protein kinase C agonist PEP005 increases NF-kappaB expression, induces differentiation and increases constitutive chemokine release by primary acute myeloid leukemia cells. *Br. J. Haematol.* **2009**, *145*, 761–774.
60. Hampson, P.; Chahal, H.; Khanim, F.; Hayden, R.; Mulder, A.; Assi, L.K.; Bunce, C.M.; Lord, J.M. PEP005, a selective small-molecule activator of protein kinase C, has potent antileukemic activity mediated *via* the delta isoform of PKC. *Blood* **2005**, *106*, 1362–1368.
61. Bruserud, Ø.; Rynningen, A.; Olsnes, A.M.; Stordrange, L.; Øyan, A.M.; Kalland, K.H.; Gjertsen, B.T. Subclassification of patients with acute myelogenous leukemia based on chemokine responsiveness and constitutive chemokine release by their leukemic cells. *Haematologica* **2007**, *92*, 332–341.
62. Olsnes, A.M.; Hatfield, K.J.; Bruserud, Ø. The chemokine system and its contribution to leukemogenesis and treatment responsiveness in patients with acute myelogenous leukemia. *J. BUON.* **2009**, *14*, 131–140.
63. Kittang, A.M.O.; Hatfield, K.J.; Sand, K.E.; Reikvam, H.; Bruserud, Ø. The chemokine network in acute myelogenous leukemia: Molecular mechanisms involved in leukemogenesis and their therapeutic implications. *Cur. Microbiol. Immunol. Rev.* **2010**, in press.
64. Nakagawa, R.; Soh, J.W.; Michie, A.M. Subversion of protein kinase C alpha signaling in hematopoietic progenitor cells results in the generation of a B –cell chronic lymphocytic leukemia-like population *in vivo*. *Cancer Res.* **2006**, *66*, 527–534.
65. Ghoul, A.; Serova, M.; Astorgues-Xerri, L.; Bieche, I.; Bousquet, G.; Varna, M.; Vidaud, M.; Phillips, E.; Weill, S.; Benhadji, K.A.; Lokiec, F.; Cvitkovic, E.; Faivre, S.; Raymond, E. Epithelial-to-mesenchymal transition and resistance to ingenol 3-angelate, a novel protein kinase C modulator, in colon cancer cells. *Cancer Res.* **2009**, *69*, 4260–4269.
66. Gillespie, S.K.; Zhang, X.D.; Hersey, P. Ingenol 3-angelate induces dual modes of cell death and differentially regulates tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis in melanoma cells. *Mol. Cancer Ther.* **2004**, *3*, 1651–1658.
67. Cozzi, S.J.; Parsons, P.G.; Ogbourne, S.M.; Pedley, J.; Boyle, G.M. Induction of senescence in diterpene ester-treated melanoma cells *via* protein kinase C-dependent hyperactivation of the mitogen-activated protein kinase pathway. *Cancer Res.* **2006**, *66*, 10083–10091.
68. Benhadji, K.A.; Serova, M.; Ghoul, A.; Cvitkovic, E.; Le Tourneau, C.; Ogbourne, S.M.; Lokiec, F.; Calvo, F.; Hammel, P.; Faivre, S.; Raymond, E. Antiproliferative activity of PEP005, a novel ingenol angelate that modulates PKC functions, alone and in combination with cytotoxic agents in human colon cancer cells. *Br. J. Cancer* **2008**, *99*, 1808–1815.
69. Serova, M.; Ghoul, A.; Benhadji, K.A.; Faivre, S.; Le Tourneau, C.; Cvitkovic, E.; Lokiec, F.; Lord, J.; Ogbourne, S.M.; Calvo, F.; Raymond, E. Effects of protein kinase C modulation by PEP005, a novel ingenol angelate, on mitogen-activated protein kinase and phosphatidylinositol 3-kinase signaling in cancer cells. *Mol. Cancer Ther.* **2008**, *7*, 915–922.

70. Ogbourne, S.M.; Suhrbier, A.; Jones, B.; Cozzi, S.J.; Boyle, G.M.; Morris, M.; McAlpine, D.; Johns, J.; Scott, T.M.; Sutherland, K.P.; Gardner, J.M.; Le, T.T.; Lenarczyk, A.; Aylward, J.H.; Parsons, P.G. Antitumor activity of 3-ingenyl angelate: Plasma membrane and mitochondrial disruption and necrotic cell death. *Cancer Res.* **2004**, *64*, 2833–2839.
71. Ersvaer, E.; Hampson, P.; Wendelbo, Ø.; Lord, J.M.; Gjertsen, B.T.; Bruserud, Ø. Circulating T cells in patients with untreated acute myelogenous leukemia are heterogeneous and can be activated through the CD3/TCR complex. *Hematology* **2007**, *12*, 199–207.
72. Ersvaer, E.; Hampson, P.; Hatfield, K.; Ulvestad, E.; Wendelbo, Ø.; Lord, J.M.; Gjertsen, B.T.; Bruserud, Ø. T cells remaining after intensive chemotherapy for acute myelogenous leukemia show a broad cytokine release profile including high levels of interferon-gamma that can be further increased by a novel protein kinase C agonist PEP005. *Cancer Immunol. Immunother.* **2007**, *56*, 913–925.
73. Le, T.T.; Gardner, J.; Hoang-Le, D.; Schmidt, C.W.; MacDonald, K.P.; Lambley, E.; Schroder, W.A.; Ogbourne, S.M.; Suhrbier, A. Immunostimulatory cancer chemotherapy using local ingenol-3-angelate and synergy with immunotherapies. *Vaccine* **2009**, *27*, 3053–3062.
74. Shin, S.Y.; Kim, C.G.; Ko, J.; Min, D.S.; Chang, J.S.; Ohba, M.; Kuroki, T.; Choi, Y.B.; Kim, Y.H.; Na, D.S.; Kim, J.W.; Lee, Y.H. Transcriptional and post-transcriptional regulation of the PKC delta gene by etoposide in L1210 murine leukemia cells: Implication of PKC delta autoregulation. *J. Mol. Biol.* **2004**, *340*, 681–693.
75. Kaur, S.; Parmar, S.; Smith, J.; Katsoulidis, E.; Li, Y.; Sassano, A.; Majchrzak, B.; Uddin, S.; Tallman, M.S.; Fish, E.N.; Plataniias, L.C. Role of protein kinase C-delta (PKC-delta) in the generation of the effects of IFN-alpha in chronic myelogenous leukemia cells. *Exp. Hematol.* **2005**, *33*, 550–557.
76. Wood, A.J.; Darbyshire, J. Injury to research volunteers--the clinical-research nightmare. *N. Engl. J. Med.* **2006**, *354*, 1869–1871.
77. Farzaneh, L.; Kasahara, N.; Farzaneh, F. The strange case of TGN1412. *Cancer Immunol. Immunother.* **2007**, *56*, 129–134.
78. Dayan, C.M.; Wraith, D.C. Preparing for first-in-man studies: The challenges for translational immunology post-TGN1412. *Clin. Exp. Immunol.* **2008**, *151*, 231–234.
79. Kenter, M.J.; Cohen, A.F. Establishing risk of human experimentation with drugs: Lessons from TGN1412. *Lancet* **2006**, *368*, 1387–1391.
80. Winkler, U.; Jensen, M.; Manzke, O.; Schulz, H.; Diehl, V.; Engert, A. Cytokine-release syndrome in patients with B-cell chronic lymphocytic leukemia and high lymphocyte counts after treatment with an anti-CD20 monoclonal antibody (rituximab, IDEC-C2B8). *Blood* **1999**, *94*, 2217–2224.
81. Maghazachi, A. Role of chemokines for natural killer cells. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
82. Carter, P.J.; Senter P.D. Antibody-drug conjugates for cancer therapy. *Cancer J.* **2008**, *14*, 154–169.
83. Stasi, R. Gemtuzumab ozogamicin: An anti-CD33 immunoconjugate for the treatment of acute myeloid leukemia. *Expert Opin. Biol. Ther.* **2008**, *8*, 527–540.

84. Wong, J.Y. Systemic targeted radionuclide therapy: potential new areas. *Int. J. Radiat. Oncol. Biol. Phys.* **2006**, *66*, 74–82.
85. Postema, M.; Gilja, O.H. Ultrasound-directed drug delivery. *Curr. Pharm. Biotechnol.* **2007**, *8*, 355–361.
86. Berg, K.; Folini, M.; Prasmickaite, L.; Selbo, P.K.; Bonsted, A.; Engesaeter, B.Ø.; Zaffaroni, N.; Weyergang, A.; Dietze, A.; Maeldandsmo, G.M.; Wagner, E.; Norum, O.J.; Høgset, A. Photochemical internalization: A new tool for drug delivery. *Curr. Pharm. Biotechnol.* **2007**, *8*, 362–372.
87. Bruserud, Ø.; Foss, B.; Petersen H. Hematopoietic growth factors in patients receiving intensive chemotherapy for malignant disorders: Studies of granulocyte-colony stimulating factor (G-CSF), granulocyte-macrophage colony stimulating factor (GM-CSF), interleukin-3 (IL-3) and Flt-3 ligand (Flt3L). *Eur. Cytokine Netw.* **2001**, *12*, 231–238.
88. Ersvaer, E.; Olsnes, A.M.; Bruserud, Ø. The immunological dilemma: Cellular innate and adaptive immune response *versus* human acute myeloid leukemia. *Open Hematol Reviews* **2007**, *1*, 1–14.
89. Dimberg, A. Chemokines in angiogenesis. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
90. Bruserud, Ø. The chemokine system in experimental and clinical hematology. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
91. Bonecchi, R.; Savino, B.; Borroni, E.M.; Mantovani, A.; Locati, M. Chemokine decoy receptors: Structure-function and biological properties. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
92. Løffler, J.; Mezger, M.; Ok, M.; Oliver Morton, C.; Einsele, H. Genetic polymorphisms in the cytokine and chemokine system – their possible importance in allogeneic stem cell transplantation. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
93. Kittan, N.A.; Hildebrandt, G.C. The chemokine system, a possible therapeutic target in acute graft *versus* host disease. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
94. Reikvam, H.; Hatfield, K.J.; Øyan, A.; Kalland, K.H.; Kittang, A.O.; Bruserud Ø. Primary human acute myelogenous leukemia cells release matrix metalloproteases and their inhibitors: Release profile and pharmacological modulation. *Eur. J. Haematol.* **2010**, in press.
95. Engelhardt, B.G.; Crowe, J.E. Homing in acute graft-versus-host disease: Tissue-specific T regulatory and Th17 cells. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
96. Sandset, P.M. The immunobiology of heparin-induced thrombocytopenia. *Curr. Topics Microbiol. Immunol.* **2010**, in press.
97. Calandra, G. CXCR4 in clinical hematology. *Curr. Topics Microbiol. Immunol.* **2010**, in press.