

Assessment of urinary tract infection and their resistance to antibiotics in diabetic and non-diabetic patients

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Article Info

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Received: 2 September 2016
Accepted: 4 September 2016
Available Online: 6 September 2016

ISSN: 2224-7750 (Online)
2074-2908 (Print)

DOI: 10.3329/bsmmuj.v9i3.29511

Cite this article:

Zahra N, Rehman K, Aqeel R, Parveen A, Akash MSH. Assessment of urinary tract infection and their resistance to antibiotics in diabetic and non-diabetic patients. *Bangabandhu Sheikh Mujib Med Univ J* 1016; 9: 151-155.

Available at:

www.banglajol.info

A Journal of Bangabandhu Sheikh Mujib Medical University, Dhaka, Bangladesh



Abstract

The study was undertaken to determine the influence of diabetes mellitus on the uropathogens and antibiotic sensitivity pattern among patients with urinary tract infections (UTIs). A cross-sectional study was conducted on 150 diabetic and 250 non-diabetic patients. Out of 160 uropathogenic isolates, *Escherichia coli* was found as a leading pathogen i.e. 46.2% followed by *Candida spp.* 30.6%, *Streptococcus faecalis* 15.6%, *Pseudomonas aeruginosa* 3.1%, *Pneumococcus* 1.2%, methicillin sensitive *S. aureus* 1.2%, methicillin resistant *S. aureus* 0.6%, *Proteus spp.* 0.6% and vancomycin resistant *Enterococcus spp.* 0.6%. The results indicated that prevalence of UTIs was significantly higher in diabetic patients than in non-diabetic subjects. *E. coli* was found to be the most common isolate. It was observed that UTIs in diabetic patients was more in female diabetic patients than in male patients. Investigation of bacteriuria in diabetic patients for UTIs is an important tool for the treatment and prevention of renal complications.

Introduction

Diabetes mellitus (DM) has been characterized by a metabolic disorder of multiple etiology including hyperglycemia and dyslipidemia along with disturbance in carbohydrate, fat and protein metabolism.^{1,2} DM and its associated complications are one of the most prevalent diseases worldwide. Development of multidrug resistant uropathogenic strains in associated with DM is being escalated that helps to determine the prevalence of urinary tract infections (UTIs) among diabetic patients and sensitivity of bacterial isolates against various types of antimicrobial agents. UTIs is one of the most common diseases that encounters in clinical practice today.³ It is particularly common infection in diabetic patients that occurs in the all ages of both males and females⁴ and if it is left untreated, causes considerable morbidity.^{5,7}

UTIs are mainly originated by the bacterial species. *Escherichia coli* belongs to the specific serogroups of uropathogenic and is considered as frequently identified organism. Serogroups have many virulence factors that are specific for invasion of urinary epithelium.⁸ *E. coli* is the most common cause of the uncomplicated UTI and account for about 95% of all infections.⁹ *E. coli* is the main causative factor for the induction of UTIs in women and also increases the likelihood of persistent UTIs.

DM has a number of effects on genitourinary

system and has long been considered to be a predisposing factor for UTIs. A characteristic feature observed in UTIs in diabetic patients in the presence of asymptomatic bacteriuria, is more in female than in male patients. The exact reason is not clear, but may be attributed to a number of factors. These include impairment of granulocyte function, increased adherence of uropathogens to uroepithelial cells, dysfunctional bladder and increased in sugar content of urine.¹⁰ Meiland et al. found that longer duration of DM was associated with the risk of asymptomatic bacteriuria.¹¹ The prevalence of asymptomatic bacteriuria is 15 to 30% higher in diabetic than in non-diabetic women.

To the best of our knowledge, limited data regarding the correlation of UTIs and their susceptibility to antibacterial agents in diabetic and non-diabetic patients is available online. The aim of present study was the assessment of UTIs and their susceptibility to various antibacterial agents among diabetic and non-diabetic agents in one of the most populated cities i.e. Lahore of Punjab province, in Pakistan.

Materials and Methods

Study design

A cross-sectional study was carried out at one of the largest government hospital located in

Lahore, Punjab province, in Pakistan and experimental analysis was conducted at department of microbiology, Institute of Molecular Biology and Biotechnology, The University of Lahore. Consecutive diabetic and non-diabetic patients of any sex who visited the hospital, were approached to participate in this study. A total of 400 urine samples from the outdoor and indoor patients were collected for specimen culturing. According to the clean-catch procedure, midstream urine samples were collected using sterile container on the same day of enrolment. After collection, urine samples were immediately brought to the microbiology laboratory for further analysis. In this study, the exclusion criteria included the chronic renal disease, known underlying renal pathology, use of antimicrobial therapy during the last month and pregnancy. Informed written consent was also obtained from individual participant and data regarding the clinical characteristics were collected on pre-tested questionnaire.

Bacterial isolation

Urinalysis was performed for all urine samples. Centrifuged urine was taken in a dropper and put a drop on clean slide for microscopy. By using standard quantitative loop, 0.001 mL was used to inoculate urine samples on cystein lactose electrolyte deficient Agar and MacConkey's agar plates and incubated at 37°C for 24 hours. When at least 10⁵ colony forming unit (CFU)/mL of urine was present, UTI was considered to be occurred. After incubation, we examined each plate for etiological agent and colony count. We also isolated different colonies present on the culture plate and performed further tests for bacterial identification.

Bacterial identification

We identified the bacteria isolated with the help of Gram's stain, rapid tests (catalase, oxidase, coagulase) using API-20E test kit.

Determination of antimicrobial susceptibility

We determined the antimicrobial susceptibility of bacteriuria using disc diffusion method. Nutrient agar (Merk, Germany) at the rate of 14.5 g/liter was prepared. Antibiotic susceptibility test was performed by Kirby Bauer modified disc diffusion method. Following antibiotic discs were used against bacterial pathogens: ampicillin, amoxicillin, clavulanic acid, cefepime, cefoperazone, imipenem, meropenem, vancomycin, amikacin, gentamycin, doxycycline, ciprofloxacin, levofloxacin, sulfamethoxazole, nitrofurantion, pipemedic acid and nalidixic acid.

We standardized the turbidity of test inoculums using McFarland nephelometer tube (# 0.5). As the inoculum was prepared in Mueller Hinton broth, in order to estimate bacterial cell density, we prepared 1% (v/v) sulfuric acid in Mueller Hinton broth and

1.175% aqueous solution of barium chloride. Standard solution of turbidity was prepared by adding 0.05 mL 1.175% barium chloride solution in 9.9 mL 1% sulfuric acid. A loop full from colony was taken and transferred to 5 mL of Mueller Hinton broth and broth was incubated at 37°C for 24 hours. To get appropriate cell density (150 × 10⁶ CFU/mL), we compared the turbidity with 0.5 McFarland standardized nephelometer tube and standardized inoculum suspension was inoculated within 15-20 min. Antibiotic discs released impregnated antibiotic into the surrounding medium when placed on the plates containing uniformly inoculated and actively growing microorganisms. The plates were inverted and placed in an incubator at 37°C.

Interpretation of inhibition zones

Results were reported either as sensitive (S), resistant (R) and intermediate (I) according to the interpretation table supplied by the company (Oxoid limited, England).

Results

Correlation of UTIs among diabetic and non-diabetic patients

A total 160 patients of UTIs have been studied, out of which 80 were diabetic and 80 were non-diabetic. Out of 160 patients, 69 were males and 91 were females, which shows that more number of females suffered from UTIs than males. Out of 80 diabetic patients, 29 were males and 51 were females. Whereas, out of 80 non-diabetic patients, 40 were males and 40 were females.

Bacterial isolation

The common organism isolated from the urine of all patients was *E. coli*. In diabetic patients, *E. coli* was found in 48 non-diabetic patients (Table I). Similarly, *Candida spp.* and *Streptococcus faecalis* were found to be 23.7 and 11.2% in diabetic patients and 37.5 and 20% in non-diabetic patients respectively (Table I). *Pseudomonas aeruginosa* and *Pneumococcus* were isolated only in non-diabetic patients, whereas *methicilline resistant S. aureus* (MRSA), *Proteus spp.* and vancomycin resistant *Enterococcus spp.* were found only in diabetic patients (Table I).

Determination of antimicrobial susceptibility

We used disc diffusion method to determine the antibacterial susceptibility of isolated bacteriuria. From the results mentioned in Table II, it has been clearly found that *E. coli* exhibited maximum sensitivity against imipenem and meropenem (96%) followed by amikacin that was 74%. Whereas, *E. coli* exhibited antimicrobial resistance to all other antibacterial agents. We also found that *E. coli* did not show any kind of susceptibility to some anti-

| Table I | | | |
|--|---------------------|--------------|---------|
| Bacterial isolation and characterization from diabetic and non-diabetic UTI patients | | | |
| Bacteria found | Number (percentage) | | p value |
| | Diabetic | Non-diabetic | |
| <i>Escherichia coli</i> | 48 (60) | 26 (32.5) | 0.007 |
| <i>Candida spp.</i> | 19 (23.7) | 30 (37.5) | |
| <i>Streptococcus faecalis</i> | 9 (11.2) | 16 (20) | |
| <i>Pseudomonas aeruginosa</i> | 0 | 5 (6.2) | |
| <i>Pneumococcus</i> | 0 | 2 (2.5) | |
| Methicillin-susceptible <i>Staphylococcus aureus</i> | 1 (1.2) | 1 (1.2) | |
| Methicillin-resistance <i>Staphylococcus aureus</i> | 1 (1.2) | 0 | |
| <i>Proteus spp.</i> | 1 (1.2) | 0 | |
| Vancomycin resistant <i>Enterococcus spp.</i> | 1 (1.2) | 0 | |

iotics namely ampicillin, amoxicillin, vancomycin and doxycycline. *S. faecalis* was susceptible to all antimicrobial agents that are mentioned in Table II. Vancomycin showed 88% sensitivity to *S. faecalis* while all other antibiotics found resistant to *S. faecalis* in greater ratio as shown in Table II. Imipenem, meropenem and amikacin exhibited over 80% sensitivity for *P. aeruginosa* and remaining antibiotics showed greater ratio of resistance to *P. aeruginosa*. *Pneumococcus* showed 100% sensitivity to imipenem, meropenem, amikacin, ciprofloxacin and levofloxacin while it showed resistance to other antibiotics used (Table II).

We found that methicillin sensitive *S. aureus* (MSSA) exhibited its sensitivity (100%) to clavulanic acid, cefepime, cefoperazone, imipenem, meropenem, amikacin, gentamycin, doxycycline, nitrofurantoin and nalidixic acid while other antibiotics used were resistant to MSSA as shown in Table II. MRSA exhibited 100% sensitivity to doxycycline and nitrofurantoin while all other antibiotics used were found resistant. Cefepime, cefoperazone, imipenem, meropenem and nitrofurantoin were found 100% sensitive to *proteus spp.* while remaining antibiotics used were found resistant. Nitrofurantoin was the only antibiotic that was found be sensitive to vancomycin resistant *Enterococcus spp.* and remain-ing antibiotics used were found resistant (Table II).

| Table II | | | | | | | | | |
|---|------------------------|----------------|--------------------|----------------------|---------------------|------------|------------|---------------------|-----------------|
| Antimicrobial sensitivity of UTI isolates | | | | | | | | | |
| Anti-microbial agents | Susceptibility pattern | <i>E. coli</i> | <i>S. faecalis</i> | <i>P. aeruginosa</i> | <i>Pneumococcus</i> | MSSA | MRSA | <i>Proteus spp.</i> | <i>VRE spp.</i> |
| Ampicillin | S | - | 10 (40) | - | - | 0 | 0 | - | 0 |
| | R | - | 15 (60) | - | - | 2 (100) | 1 (100) | - | 1 (100) |
| Amoxicillin | S | - | 10 (40) | - | - | 0 | 0 | - | 0 |
| | R | - | 15 (60) | - | - | 2 (100) | 1 (100) | - | 1 (100) |
| Clavulanic acid | S | 14 (19) | 7 (28) | 1 (20) | 0 | 2 (100) | 0 | - | 0 |
| | R | 60 (81) | 18 (72) | 4 (80) | 2 (100) | 0 | 1 (100) | | 01 (100) |
| Cefepime | S | 7 (10) | 3 (12) | 2 (40) | 0 | 2 (100) | 0 | 1 (100) | - |
| | R | 67 (90) | 22 (88) | 3 (60) | 2 (100) | 0 | 1 (100) | 0 | - |
| Cefoperazone | S | 7 (10) | 3 (12) | 2 (40) | 0 | 2 (100) | 0 | 1 (100) | - |
| | R | 67 (90) | 22 (88) | 3 (60) | 2 (100) | 0 | 1 (100) | 0 | - |

Data are expressed as number (% within the parenthesis)

Cont.

Table II

Antimicrobial sensitivity of UTI isolates (Cont.)

| Anti-microbial agents | Susceptibility pattern | <i>E. coli</i> | <i>S. faecalis</i> | <i>P. aeruginosa</i> | <i>Pneumococcus</i> | MSSA | MRSA | <i>Proteus spp.</i> | <i>VRE spp.</i> |
|-----------------------|------------------------|----------------|--------------------|----------------------|---------------------|------------|------------|---------------------|-----------------|
| Imipenem | S | 71 (96) | 10 (40) | 4 (80) | 2 (100) | 2 (100) | 0 | 1 (100) | - |
| | R | 3 (04) | 15 (60) | 1 (20) | 0 | 0 | 1 (100) | 0 | - |
| Meropenem | S | 71 (96) | 10 (40) | 4 (80) | 2 (100) | 2 (100) | 0 | 1 (100) | - |
| | R | 3 (04) | 15 (60) | 1 (20) | 0 | 0 | 1 (100) | 0 | - |
| Vancomycin | S | - | 22 (88) | - | - | 2 (100) | - | - | 0 |
| | R | - | 3 (12) | - | - | 0 | - | - | 1 (100) |
| Amikacin | S | 55 (74) | - | 4 (80) | 2 (100) | 2 (100) | - | 0 | - |
| | R | 19 (26) | - | 1 (20) | 0 | 0 | - | 1 (100) | - |
| Gentamycin | S | 17 (23) | 3 (12) | 1 (20) | 0 | 2 (100) | 0 | 0 | 0 |
| | R | 57 (77) | 22 (88) | 4 (80) | 2 (100) | 0 | 1 (100) | 1 (100) | 1 (100) |
| Doxycycline | S | - | 4 (16) | 1 (20) | - | 2 (100) | 1 (100) | - | 0 |
| | R | - | 21 (84) | 4 (80) | - | 0 | 0 | - | 1 (100) |
| Ciprofloxacin | S | 14 (19) | 5 (20) | 1 (20) | 2 (100) | 1 (50) | 0 | 0 | 0 |
| | R | 60 (81) | 20 (80) | 4 (80) | 0 | 1 (50) | 1 (100) | 1 (100) | 1 (100) |
| Levofloxacin | S | 14 (19) | 5 (20) | 1 (20) | 2 (100) | 1 (50) | 0 | 0 | 0 |
| | R | 60 (81) | 20 (80) | 4 (80) | 0 | 1 (50) | 1 (100) | 1 (100) | 1 (100) |
| Sulfamethoxazole | S | 11 (15) | 10 (40) | 2 (40) | - | 1 (50) | 0 | - | - |
| | R | 63 (85) | 15 (60) | 3 (60) | - | 1 (50) | 1 (100) | - | - |
| Nitrofurantion | S | 11 (15) | 7 (28) | 1 (20) | 0 | 2 (100) | 1 (100) | 1 (100) | 1 (100) |
| | R | 63 (85) | 18 (72) | 4 (80) | 2 (100) | 0 | 0 | 0 | 0 |
| Pipemedic acid | S | 7 (10) | 4 (16) | 1 (20) | 0 | 1 (50) | 0 | 0 | 0 |
| | R | 67 (90) | 21 (84) | 4 (80) | 2 (100) | 1 (50) | 1 (100) | 1 (100) | 1 (100) |
| Nalidixic acid | S | 71 (96) | 3 (12) | 2 (40) | 0 | 2 (100) | 0 | 0 | 0 |
| | R | 3 (04) | 22 (88) | 3 (60) | 2 (100) | 0 | 1 (100) | 1 (100) | 1 (100) |

Data are expressed as number (% within the parenthesis)

Discussion

In the present study, we found that incidences of UTIs in diabetic patients are maximum than non-diabetics. Prevalence of UTIs were found more in female patients (63.7%) than in male patients (36.2%) which indicate that female patients (particularly diabetic) are more susceptible to UTIs as compared to male patients. The prevalence of UTIs among the study participants of present study was in accordance with already published reports.¹²⁻¹⁴ The prevalence of UTI was also described in renal transplant recipients.¹⁵

Moreover, in present study, the most common uropathogen that was detected in the urine of all participants was *E. coli*. In diabetic patients, *E. coli* was detected in 60% patients, whereas in non-diabetic patients, it was detected in up to 32% patients. The increased prevalence of *E. coli* in UTIs indicates that *E. coli* is the leading and most significant isolate that is responsible to cause UTIs more significantly in diabetic patients especially females than non-diabetic patients.

Conclusion

Both diabetic and non-diabetic patients were susceptible to UTIs in which *E. coli* was found to be the most frequent uropathogen that exhibited its maximum susceptibility against tested antibacterial agents. Among the diabetic patient having UTIs, female diabetic patients were more susceptible to UTIs as compared to male diabetics which indicate that prevalence of UTIs in female diabetic patients is high as compared to that of male diabetic patients. Therefore, continued surveillance of prevalence of bacteriuria mandatory to ensure the appropriate diagnosis and recommendations for treatment of UTIs.

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