

Cytosolic Iron-Sulfur Protein Assembly 1 (*CIAO1*) Downstream Activation of Phospholipase A2 and Hormone-Mediated Signaling-Induced Cell Death Network in Human Hepatocellular Carcinoma (HCC) by Systems-Theoretical Analysis

Lianxiu Qi¹, Lin Wang^{1*}, Minghu Jiang², Juxiang Huang¹ and Hong Lin¹

¹Biomedical Center, School of Electronic Engineering, Beijing University of Posts and Telecommunications, Beijing, 100876, China

²Lab of Computational Linguistics, School of Humanities and Social Sciences, Tsinghua University, Beijing, 100084, China

Abstract

We constructed the significant high expression (fold change ≥ 2) cytosolic iron-sulfur protein assembly 1 (*CIAO1*) downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network in human Hepato Cellular Carcinoma (HCC), compared with low expression no-tumor hepatitis/cirrhotic tissues (HBV or HCV infection) in GEO data set, by using integration of gene regulatory activated and inhibited network inference method. Our result showed that *CIAO1* downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death upstream network had no result, and downstream *CIAO1*-activated *PLA2G1B*, *NUP62* in HCC. By integrative analysis of biological processes simultaneous occurrence between the different *CIAO1* activated downstream cell death gene ontology (GO) network of HCC compared with *CIAO1* activated downstream cell death GO network of no-tumor hepatitis/cirrhotic tissues, and the same compared *CIAO1* inhibited downstream cell death GO network of no-tumor hepatitis/cirrhotic tissues, or the different compared *CIAO1* inhibited downstream cell death GO network of HCC, we proposed and verified that *CIAO1* activated upstream network had no result; Downstream network consisted of activation of phospholipase A2, cell death, protein kinase cascade, regulation of signal transduction, hormone-mediated signaling, negative regulation of epidermal growth factor receptor signaling pathway, negative regulation of MAPK (Mitogen Activated Protein Kinase) activity, negative regulation of Ras protein signal transduction in HCC, as a result of downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death in HCC.

Keywords: Cytosolic iron-sulfur protein assembly 1 (*CIAO1*); Human hepatocellular carcinoma (HCC); Downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network; Systems-theoretical analysis

Introduction

Cytosolic iron-sulfur protein assembly 1 (*CIAO1*) is our identified significant high expression (fold change ≥ 2) gene in human hepatocellular carcinoma (HCC) compared with low expression no-tumor hepatitis/cirrhotic tissues (HBV or HCV infection) from GEO (Gene Expression Omnibus) data set GSE10140-10141 (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE10140>, <http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE10141>) [1].

We interpreted *PLA2G1B* and *NUP62* by using gene ontology (GO). *PLA2G1B* cellular component, molecular function and biological process are relevant to extracellular region, extracellular space; receptor binding, calcium ion binding, hydrolase activity, bile acid binding, cell surface binding, calcium-dependent phospholipase A2 activity; activation of MAPK activity, neutrophil mediated immunity, fatty acid biosynthesis, phospholipid metabolism, actin filament organization, signal transduction, protein kinase cascade, glucose transport, leukotriene biosynthesis, neutrophil chemotaxis, organismal lipid catabolism, positive regulation of DNA replication, phosphatidylcholine metabolism, positive regulation of fibroblast proliferation, arachidonic acid secretion, positive regulation of protein secretion, positive regulation of immune response, activation of NF-kappaB transcription factor, positive regulation of calcium ion transport into cytosol, positive regulation of specific transcription from RNA polymerase II promoter, activation of phospholipase A2, interleukin-8 production, cellular response to insulin stimulus (GO database). *NUP62* cellular component, molecular function and biological process are relevant to nucleus, nuclear pore, nucleolus, cytoplasm, centrosome, nucleocytoplasmic shuttling complex, nuclear membrane; chromatin

binding, protein serine/threonine kinase activity, nucleocytoplasmic transporter activity, protein binding, structural constituent of nuclear pore, receptor signaling complex scaffold activity, transcription regulator activity, SH2 domain binding, ubiquitin binding, thyroid hormone receptor binding, PTB (Phosphotyrosine-binding) domain binding; cell surface receptor linked signal transduction, cell aging, cell death, negative regulation of cell proliferation, hormone-mediated signaling, regulation of signal transduction, protein transport, negative regulation of epidermal growth factor receptor signaling pathway, negative regulation of apoptosis, positive regulation of I-kappaB kinase/NF-kappaB cascade, negative regulation of MAPK activity, positive regulation of epidermal growth factor receptor signaling pathway, positive regulation of transcription, negative regulation of Ras protein signal transduction, mRNA transport, intracellular protein transmembrane transport (GO database).

Study of cytosolic iron-sulfur protein assembly 1 (*CIAO1*) is presented in several papers as follows: Mouse knock-out of IOP1 protein reveals its indispensable role in mammalian cytosolic iron-sulfur protein biogenesis [2]; Tah18 transfers electrons to Dre2 in cytosolic iron-sulfur protein biogenesis [3]; A role for IOP1 in mammalian

*Corresponding author: Lin Wang, Biomedical Center, School of Electronics Engineering, Beijing University of Posts and Telecommunications, Beijing, 100876, China, Tel: 0086-13240981826; Fax: 8610-62785736; E-mail: wanglin98@tsinghua.org.cn

Received March 26, 2012; Accepted April 21, 2012; Published April 27, 2012

Citation: Qi L, Wang L, Jiang M, Huang J, Lin H (2012) Cytosolic Iron-Sulfur Protein Assembly 1 (*CIAO1*) Downstream Activation of Phospholipase A2 and Hormone-Mediated Signaling-Induced Cell Death Network in Human Hepatocellular Carcinoma (HCC) by Systems-Theoretical Analysis. Mol Biol 1:105. doi:10.4172/2168-9547.1000105

Copyright: © 2012 Qi L, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

cytosolic iron-sulfur protein biogenesis [4]; The essential cytosolic iron-sulfur protein Nbp35 acts without Cfd1 partner in the green lineage [5]; Role of human mitochondrial Nfs1 in cytosolic iron-sulfur protein biogenesis and iron regulation [6]; Activation of the iron regulon by the yeast Aft1/Aft2 transcription factors relies on mitochondrial but not cytosolic iron-sulfur protein biogenesis [7]; Gene structure and mutation causing X-linked sideroblastic anemia with ataxia with cytosolic iron-sulfur protein maturation disruption [8]. Downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death is presented in some papers. Such as, Cytoplasmic phospholipase A2 levels correlate with apoptosis in human colon tumorigenesis [9]; Potentiation by vitamin D analogs of TNF α and ceramide-induced apoptosis in MCF-7 cells is related to activation of cytosolic phospholipase A2 [10]; 1,25-Dihydroxyvitamin D3 protects human leukemic cells from tumor necrosis factor-induced apoptosis through inactivation of cytosolic phospholipase A2 [11]; Phospholipase A2-activating protein (PLAA) enhances cisplatin-induced apoptosis in HeLa cells [12]; Alpha-Tocopheryl succinate contributes to apoptosis in erbB2-expressing breast cancer cell via NF-kappaB pathway [13]; Gene network signaling in hormone responsiveness modifies autophagy and apoptosis in breast cancer cells [14]; Rho/ROCK/actin signaling regulates membrane androgen receptor induced apoptosis in prostate cancer cells [15]; Gonadotropin-releasing hormone type II contributes to apoptosis of human endometrial cancer cells by activating GADD45 α [16]. Yet the distinct high expression *CIAO1* downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network in HCC remains to be elucidated. Here we constructed the high expression *CIAO1* activated downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network in HCC from GEO data set by gene regulatory network inference method based on linear programming and decomposition procedure.

In this study, we constructed *CIAO1* upstream and downstream activated and inhibited downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network in no-tumor hepatitis/cirrhosis tissues and HCC. The biological process and data analysis of the low- and high- expression *CIAO1* downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network was done in no-tumor hepatitis/cirrhosis tissues (HBV or HCV infection) and HCC by gene ontology (GO) database. By comparison with the same and different upstream and downstream GO and numbers of *CIAO1* activated and inhibited downstream cell death network between no-tumor hepatitis/cirrhosis tissues and HCC, we put forward hypothesis of *CIAO1* activated downstream cell death network of downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death in HCC.

Materials and Methods

Microarrays 6,144 genes were used for analyzing *CIAO1* downstream cell death mechanism and constructing molecular network of HCC from our total network of 225 significant high expression (fold change ≥ 2) molecules in HCC compared with no-tumor hepatitis/cirrhosis tissues (HBV or HCV infection) based on GEO data set GSE10140-10141 (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE10140>, <http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE10141>). The raw microarray data was preprocessed by log base 2.

Significant expressed genes for studying *CIAO1* downstream cell death mechanism and molecular network were identified using significant analysis of microarrays (SAM) (<http://www-stat.stanford.edu/~tibs/SAM/>) [17]. We selected two classes unpaired and minimum fold change ≥ 2 and chose the significant highly expressed value genes of HCC compared with that of no-tumor hepatitis/cirrhosis tissues (HBV or HCV infection) under the false-discovery rate was 0%.

CIAO1 downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network was constructed based on GRNInfer and GVedit tools (<http://www.graphviz.org/About.php>). GRNInfer is a novel mathematic method called GNR (Gene Network Reconstruction tool) based on linear programming and a decomposition procedure for inferring gene network [18]. We established *CIAO1* activated network of HCC based on the fold change ≥ 2 distinguished genes and selected parameters as lambda 0.0 because we used one data set. Lambda was a positive parameter which balanced the matching and sparsity terms in the objective function. Using different thresholds, we could predict various networks with the different edge density. The threshold parameters make the edge whose strength of link is smaller than threshold not shown in the network graph. The smaller this parameter, the more edges in the network graph. We selected threshold as 1.0e-7.

CIAO1 downstream cell death mechanism of HCC was analyzed using Molecule Annotation System, MAS (CapitalBio Corporation, Beijing, China; <http://bioinfo.capitalbio.com/mas3/>). MAS is a Web-based software toolkit for a whole data mining and function annotation solution to extract and analyze biological molecules relationships from public databases. The primary databases of MAS integrated various well-known biological resources, such as Gene Ontology (<http://www.geneontology.org>), KEGG (<http://www.genome.jp/kegg/>), BioCarta (<http://www.biocarta.com/>), GenMapp (<http://www.genmapp.org/>), HPRD (<http://www.hprd.org/>), MINT (<http://mint.bio.uniroma2.it/mint/Welcome.do>), BIND (<http://www.blueprint.org/>), Intact (<http://www.ebi.ac.uk/intact/>), UniGene (www.ncbi.nlm.nih.gov/UniGen), OMIM (<http://www.ncbi.nlm.nih.gov/sites/entrez?db=omim>) and disease (<http://bioinfo.capitalbio.com/mas3/>). MAS offers various query entries and graphics. The algorithm is P, Q value in GO and pathway of module was presented in reference [19].

Results

We analyzed *CIAO1* downstream cell death mechanism and constructed molecular network of HCC from our total network of 225 significant high expression molecules (fold change ≥ 2) from 6,144 genes of HCC compared with no-tumor hepatitis/cirrhosis tissues (HBV or HCV infection) by GRNInfer, respectively.

Results

We extracted the biological process of GO terms and did numbers data analysis of the different biological processes of *CIAO1* activated downstream cell death network in HCC compared with activated network of no-tumor hepatitis/cirrhosis tissues, the same biological processes of *CIAO1* activated downstream cell death network in HCC compared with inhibited network of no-tumor hepatitis/cirrhosis tissues, as shown in Table 1. GO terms and numbers data was analyzed the different biological processes of *CIAO1* activated compared with inhibited downstream cell death network in HCC, as shown in Table 2.

CIAO1 activated downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network was constructed in HCC. Our result showed that upstream had no result, and the downstream *CIAO1*-activated *PLA2G1B*, *NUP62* in HCC, as shown in Figure 1 and Figure 2.

Discussion

Our aim is to study novel high expression *CIAO1* downstream

The Different Biological Processes of <i>CIAO1</i> Activated Downstream Cell Death Network of HCC Compared with Activated Network of No-tumor Hepatitis/cirrhotic Tissues	
Terms	Numbers
protein kinase cascade	1
activation of phospholipase A2	1
regulation of signal transduction	1
cell death	1
hormone-mediated signaling	1
negative regulation of epidermal growth factor receptor signaling pathway	1
negative regulation of MAPK activity	1
negative regulation of Ras protein signal transduction	1
fatty acid biosynthesis	1
organismal lipid catabolism	1
phosphatidylcholine metabolism	1
phospholipid metabolism	1
The Same Biological Processes of <i>CIAO1</i> Activated Downstream Cell Death Network of HCC Compared with Inhibited Network of No-tumor Hepatitis/cirrhotic Tissues	
Terms	Numbers
None	0

Table 1: GO Terms and numbers data analysis of the different biological processes of *CIAO1* activated downstream cell death network of HCC compared with activated network of no-tumor hepatitis/cirrhotic tissues, the same biological processes of *CIAO1* activated downstream cell death network of HCC compared with inhibited network of no-tumor hepatitis/cirrhotic tissues.

The Different Biological Processes of <i>CIAO1</i> Activated Downstream Cell Death Network of HCC Compared with Inhibited Network of HCC	
Terms	Numbers
negative regulation of epidermal growth factor receptor signaling pathway	1
negative regulation of MAPK activity	1
negative regulation of Ras protein signal transduction	1
protein kinase cascade	1
regulation of signal transduction	1
activation of phospholipase A2	1
cell death	1
hormone-mediated signaling	1
fatty acid biosynthesis	1
organismal lipid catabolism	1
phosphatidylcholine metabolism	1
phospholipid metabolism	1

Table 2: GO Terms and numbers data analysis of the different biological processes of *CIAO1* activated downstream cell death network of HCC compared with inhibited network of HCC.

cell death mechanism and molecular network in HCC. We have already constructed and analyzed some novel molecular network from different databases presented in our articles [19-32]. In this study, we constructed *CIAO1* upstream and downstream activated and inhibited downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network in no-tumor hepatitis/cirrhotic tissues and HCC. The biological process and data analysis of the low and high expression *CIAO1* downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network was done in no-tumor hepatitis/cirrhotic tissues (HBV or HCV infection) and HCC by GO database. By comparison with the same and different upstream and downstream GO and numbers of *CIAO1* activated and inhibited downstream cell death network between no-tumor hepatitis/cirrhotic tissues and HCC, we put forwards hypothesis of *CIAO1* activated downstream cell death network of downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death in HCC.

We extracted the biological process of GO terms and did numbers data analysis of the different biological processes of *CIAO1* activated downstream cell death network in HCC compared with activated network of no-tumor hepatitis/cirrhotic tissues, the same biological processes of *CIAO1* activated downstream cell death network in HCC

compared with inhibited network of no-tumor hepatitis/cirrhotic tissues (Table 1 and Table 2). We constructed the high expression (fold change ≥ 2) *CIAO1* activated downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death network in human hepatocellular carcinoma (HCC) compared with low expression no-tumor hepatitis/cirrhotic tissues (HBV or HCV infection) in GEO data set using integration of gene regulatory network inference method. Our result showed that *CIAO1* downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death upstream network had no result, and downstream *CIAO1*-activated *PLA2G1B*, *NUP62* (Figure 1 and Figure 2) in HCC.

By further comparison with the same biological processes of and different gene ontology (GO) of *CIAO1* activated and inhibited downstream cell death network between no-tumor hepatitis/cirrhotic tissues and HCC, we found that the different biological processes of *CIAO1* activated upstream network had no result; Downstream network consisted of protein kinase cascade, activation of phospholipase A2, regulation of signal transduction, cell death, hormone-mediated signaling, negative regulation of epidermal growth factor receptor signaling pathway, negative regulation of MAPK activity, negative regulation of Ras protein signal transduction in HCC compared with activated network of no-tumor hepatitis/cirrhotic tissues.



Figure 1: *CIAO1* upstream activated phospholipase A2 and hormone-mediated signaling-induced cell death network in HCC by GRNInfer and our programming.

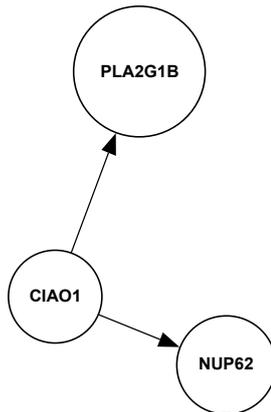


Figure 2: *CIAO1* downstream activated phospholipase A2 and hormone-mediated signaling-induced cell death network in HCC by GRNInfer and our programming.

The same biological processes of *CIAO1* activated upstream network had no result; downstream network had no result in HCC compared with inhibited network of no-tumor hepatitis/cirrhosis tissues.

The different biological processes of *CIAO1* activated network had no result; Downstream network included negative regulation of epidermal growth factor receptor signaling pathway, negative regulation of MAPK activity, negative regulation of Ras protein signal transduction, protein kinase cascade, regulation of signal transduction, activation of phospholipase A2, cell death, hormone-mediated signaling compared with inhibited network of HCC.

By integrative analysis of biological processes simultaneous occurrence between the different *CIAO1* activated downstream cell death gene ontology (GO) network of HCC compared with *CIAO1* activated downstream cell death GO network of no-tumor hepatitis/cirrhosis tissues, and the same compared *CIAO1* inhibited downstream cell death GO network of HCC, we proposed and verified that *CIAO1* activated upstream network had no result; Downstream network included activation of phospholipase A2, cell death, protein kinase cascade, regulation of signal transduction, hormone-mediated signaling, negative regulation of epidermal growth factor receptor signaling pathway, negative regulation of MAPK activity, negative regulation of Ras protein signal transduction in HCC, as a result of downstream activation of phospholipase A2 and hormone-mediated signaling-induced cell death in HCC.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (No.61171114), the Returned Overseas Chinese Scholars for Scientific research Foundation of State Education Ministry, Significant Science and Technology

Project for New Transgenic Biological Species (2009ZX08012-001B), Automatical Scientific Planning of Tsinghua University (20111081023 and 20111081010), State Key Lab of Pattern Recognition Open Foundation.

References

1. Hoshida Y, Villanueva A, Kobayashi M, Peix J, Chiang DY, et al. (2008) Gene expression in fixed tissues and outcome in hepatocellular carcinoma. *N Engl J Med* 359: 1995-2004.
2. Song D, Lee FS (2012) Mouse knock-out of IOP1 protein reveals its essential role in mammalian cytosolic iron-sulfur protein biogenesis. *J Biol Chem* 286: 15797-15805.
3. Netz DJ, Stumpf M, Dore C, Muhlenhoff U, Pierik AJ, et al. (2012) Tah18 transfers electrons to Dre2 in cytosolic iron-sulfur protein biogenesis. *Nat Chem Biol* 6: 758-765.
4. Song D, Lee FS (2008) A role for IOP1 in mammalian cytosolic iron-sulfur protein biogenesis. *J Biol Chem* 283: 9231-9238.
5. Bych K, Netz DJ, Vigani G, Bill E, Lill R, et al. (2008) The essential cytosolic iron-sulfur protein Nbp35 acts without Cfd1 partner in the green lineage. *J Biol Chem* 283: 35797-35804.
6. Biederick A, Stehling O, Rosser R, Niggemeyer B, Nakai Y, et al. (2006) Role of human mitochondrial Nfs1 in cytosolic iron-sulfur protein biogenesis and iron regulation. *Mol Cell Biol* 26: 5675-5687.
7. Rutherford JC, Oieda L, Balk J, Muhlenhoff U, Lill R, et al. (2005) Activation of the iron regulon by the yeast Aft1/Aft2 transcription factors depends on mitochondrial but not cytosolic iron-sulfur protein biogenesis. *J Biol Chem* 280: 10135-10140.
8. Bekri S, Kispal G, Lange H, Fitzsimons E, Tolmie J, et al. (2000) Human ABC7 transporter: gene structure and mutation causing X-linked sideroblastic anemia with ataxia with disruption of cytosolic iron-sulfur protein maturation. *Blood* 96: 3256-3264.
9. Dong M, Johnson M, Rezaie A, Ilsley JN, Nakanishi M, et al. (2005) Cytoplasmic phospholipase A2 levels correlate with apoptosis in human colon tumorigenesis. *Clin Cancer Res* 11: 2265-2271.
10. Pirianov G, Danielsson C, Carlberg C, James SY, Colston KW (1999) Potentiation by vitamin D analogs of TNF α and ceramide-induced apoptosis in MCF-7 cells is associated with activation of cytosolic phospholipase A2. *Cell Death Differ* 6: 890-901.
11. Wu YL, Jiang XR, Lillington DM, Allen PD, Newland AC, et al. (1998) 1,25-Dihydroxyvitamin D3 protects human leukemic cells from tumor necrosis factor-induced apoptosis via inactivation of cytosolic phospholipase A2. *Cancer Res* 58: 633-640.
12. Zhang F, Suarez G, Sha J, Sierra JC, Peterson JW, et al. (2009) Phospholipase A2-activating protein (PLAA) enhances cisplatin-induced apoptosis in HeLa cells. *Cell Signal* 21: 1085-1099.
13. Wang XF, Xie Y, Wang HG, Zhang Y, Duan XC, et al. (2010) α -Tocopheryl succinate induces apoptosis in erbB2-expressing breast cancer cell via NF- κ B pathway. *Acta Pharmacol Sin* 31: 1604-1610.
14. Clarke R, Shahjahan AN, Riggins RB, Cho Y, Crawford A, et al. (2009) Gene network signaling in hormone responsiveness modifies apoptosis and autophagy in breast cancer cells. *J Steroid Biochem Mol Biol* 114: 8-20.
15. Papadopoulou N, Charalampopoulos I, Alevizopoulos K, Gravanis A, Stournaras C, et al. (2008) Rho/ROCK/actin signaling regulates membrane androgen receptor induced apoptosis in prostate cancer cells. *Exp Cell Res* 314: 3162-3174.
16. Wu HM, Cheng JC, Wang HS, Huang HY, MacCalman CD, et al. (2009) Gonadotropin-releasing hormone type II induces apoptosis of human endometrial cancer cells by activating GADD45 α . *Cancer Res* 69: 4202-4208.
17. Storey JD (2002) A direct approach to false discovery rates. *J R Stat Soc Series B Stat Methodol* 64: 479-498.
18. Wang Y, Joshi T, Zhang XS, Xu D, Chen L (2006) Inferring gene regulatory networks from multiple microarray datasets. *Bioinformatics* 22: 2413-2420.
19. Wang L, Sun L, Huang J, Jiang M (2011) Cyclin-dependent kinase inhibitor 3 (CDKN3) novel cell cycle computational network between human non-malignancy associated hepatitis/cirrhosis and hepatocellular carcinoma (HCC) transformation. *Cell Prolif* 44: 291-299.

20. Wang L, Sun Y, Jiang M, Zheng X (2009) Integrative decomposition procedure and Kappa statistics for the distinguished single molecular network construction and analysis. *J Biomed Biotechnol* 2009: 726728.
21. Wang L, Sun Y, Jiang M, Zhang S, Wolf S (2009) FOS proliferating network construction in early colorectal cancer (CRC) based on integrative significant function cluster and inferring analysis. *Cancer Invest* 27: 816-824.
22. Wang L, Huang J, Jiang M, Zheng X (2010) AFP computational secreted network construction and analysis between human hepatocellular carcinoma (HCC) and no-tumor hepatitis/cirrhotic liver tissues. *Tumour Biol* 31: 417-425.
23. Wang L, Huang J, Jiang M, Sun L (2011) MYBPC1 computational phosphoprotein network construction and analysis between frontal cortex of HIV encephalitis (HIVE) and HIVE-control patients. *Cell Mol Neurobiol* 31: 233-241.
24. Wang L, Huang J, Jiang M, Sun L (2011) Survivin (BIRC5) cell cycle computational network in human no-tumor hepatitis/cirrhosis and hepatocellular carcinoma transformation. *J Cell Biochem* 112: 1286-1294.
25. Wang L, Huang J, Jiang M, Lin H (2012) Signal Transducer and Activator of Transcription 2 (STAT2) Metabolism Coupling Postmitotic Outgrowth to Visual and Sound Perception Network in Human Left Cerebrum by Biocomputation. *J Mol Neurosci*.
26. Wang L, Huang J, Jiang M (2010) RRM2 computational phosphoprotein network construction and analysis between no-tumor hepatitis/cirrhotic liver tissues and human hepatocellular carcinoma (HCC). *Cell Physiol Biochem* 26: 303-310.
27. Wang L, Huang J, Jiang M (2011) CREB5 computational regulation network construction and analysis between frontal cortex of HIV encephalitis (HIVE) and HIVE-control patients. *Cell Biochem Biophys* 60: 199-207.
28. Sun Y, Wang L, Liu L (2008) Integrative decomposition procedure and Kappa statistics set up ATF2 ion binding module in Malignant Pleural Mesothelioma (MPM). *Frontiers of Electrical and Electronic Engineering in China* 3: 381-387.
29. Sun Y, Wang L, Jiang M, Huang J, Liu Z, et al. (2010) Secreted Phosphoprotein 1 Upstream Invasive Network Construction and Analysis of Lung Adenocarcinoma Compared with Human Normal Adjacent Tissues by Integrative Biocomputation. *Cell Biochem Biophys* 56: 59-71.
30. Sun L, Wang L, Jiang M, Huang J, Lin H (2011) Glycogen debranching enzyme 6 (AGL), enolase 1 (ENOSF1), ectonucleotide pyrophosphatase 2 (ENPP2_1), glutathione S-transferase 3 (GSTM3_3) and mannosidase (MAN2B2) metabolism computational network analysis between chimpanzee and human left cerebrum. *Cell Biochem Biophys* 61: 493-505.
31. Huang JX, Wang L, Jiang MH (2010) TNFRSF11B computational development network construction and analysis between frontal cortex of HIV encephalitis (HIVE) and HIVE-control patients. *J Inflamm (Lond)* 7: 50.
32. Huang J, Wang L, Jiang M, Zheng X (2010) Interferon α -Inducible Protein 27 Computational Network Construction and Comparison between the Frontal Cortex of HIV Encephalitis (HIVE) and HIVE-Control Patients. *The Open Genomics Journal* 3: 1-8.

Submit your next manuscript and get advantages of OMICS Group submissions

Unique features:

- User friendly/feasible website-translation of your paper to 50 world's leading languages
- Audio Version of published paper
- Digital articles to share and explore

Special features:

- 200 Open Access Journals
- 15,000 editorial team
- 21 days rapid review process
- Quality and quick editorial, review and publication processing
- Indexing at PubMed (partial), Scopus, DOAJ, EBSCO, Index Copernicus and Google Scholar etc
- Sharing Option: Social Networking Enabled
- Authors, Reviewers and Editors rewarded with online Scientific Credits
- Better discount for your subsequent articles

Submit your manuscript at: <http://www.omicsonline.org/submit/>