



Deposition and metabolism of inhaled ciclesonide in the human lung

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ABSTRACT: Ciclesonide is an inhaled corticosteroid, administered as a prodrug *via* a metered-dose inhaler. Following deposition in the lung, ciclesonide is hydrolysed by esterases to form the pharmacologically active metabolite desisobutyryl-ciclesonide (des-CIC). Formation of des-CIC, as well as reversible esterification of des-CIC with fatty acids, has been demonstrated *in vitro*. The aim of this study was to investigate the *in vivo* metabolism of ciclesonide in the human lung.

This single-dose, open-label, nonrandomised study was performed in 20 patients undergoing planned lung surgery for treatment of malignant pulmonary lesions. Patients inhaled a single dose of 1,280 µg ciclesonide at various time-points between 2 and 24 h prior to lung tissue resection. The concentration of ciclesonide, des-CIC and fatty acid conjugates of des-CIC in tissue samples was determined. Serum samples for pharmacokinetic analysis were taken at several time-points after inhalation.

The pharmacokinetics in serum indicated that the inhalation by the patients was adequate. Metabolites (des-CIC, des-CIC oleate and des-CIC palmitate) were detected in the resected central and peripheral lung tissues. A substantial portion of ciclesonide was already activated to des-CIC at the first time-point of tissue analysis.

Activation of ciclesonide and formation of des-CIC fatty acid conjugates was confirmed *in vivo* in the human lung.

KEYWORDS: Ciclesonide, human, *in vivo*, lung metabolism

Asthma is one of the most common chronic inflammatory disorders of the airways and is characterised by airway obstruction, inflammation and hyperresponsiveness resulting from complex interactions among inflammatory cells, mediators, and the cells and tissues of the airways [1, 2]. International and national treatment guidelines recommend the use of inhaled corticosteroids (ICS) as first-line therapy for patients with asthma [3, 4].

Ciclesonide is a novel ICS administered as a prodrug *via* a metered-dose inhaler using hydrofluoroalkane (HFA) as the propellant. It is available as Alvesco® (Nycomed GmbH, Konstanz, Germany) in many markets. Ciclesonide is converted to the active metabolite desisobutyryl-ciclesonide (des-CIC), by esterases that cleave isobutyrate at the C21 position [5–7]. Des-CIC has a much higher affinity for the glucocorticoid receptor than ciclesonide (~100 times higher) [5, 8]. Alvesco® has a high pulmonary deposition of 52% [9, 10]. Both oropharyngeal deposition of ciclesonide and activation of ciclesonide in the oropharynx to des-CIC are low [10–12].

In vitro studies in human tissue and *in vivo* studies in rats have reported that des-CIC forms reversible

conjugates with fatty acids, which are pharmacologically inactive and do not bind to the glucocorticoid receptor [7, 13–15]. The findings regarding the metabolism of CIC derived from *in vitro* and animal studies have yet to be confirmed by *in vivo* studies in human subjects. It was the aim of the present study to investigate for the first time the *in vivo* deposition and metabolism of ciclesonide by applying a comprehensive analysis of CIC, des-CIC, and its reversible conjugates in central and peripheral human lung tissue.

METHODS

Subjects

Inclusion criteria

Male and female patients undergoing elective lung surgery for malignant pulmonary lesions, aged 18–70 yrs were eligible for inclusion in the study. Patients had to demonstrate an acceptable inhalation technique at screening and have a forced expiratory volume in 1 s (FEV₁) ≥70% predicted. In addition, the therapeutic surgical strategy for the underlying disease had to require the removal of lung tissue.

Lung tissue prepared for pharmacological analysis in this study had to be ventilated prior to

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resection (*i.e.* without relevant proximal bronchial obstruction), vital at the time of resection, and noncancerous. The tissue used for analysis was removed for therapeutic reasons only and was not required for any other therapeutic or diagnostic procedures.

Exclusion criteria

Patients with diseases contraindicating the use of ICS (*e.g.* active pulmonary tuberculosis or relevant fungal, bacterial or viral infections of the lower respiratory tract), those with a history of allergic reactions to ICS or HFA-solution aerosols, and those participating in a clinical study ≤ 30 days before the start of the current study were excluded. Pregnant females, nursing mothers or females of childbearing potential who were not using a medically acceptable and reliable method of contraception for the entire study duration were not eligible to participate.

Ethics

The study was performed in compliance with the International Conference on Harmonization (ICH) Guidelines for Good Clinical Practice (ICH Consolidated Guideline E6) and the Declaration of Helsinki in its revised form (Somerset West, South Africa, October 1996). The protocol was approved by the Independent Ethics Committee at the Faculty of Medicine of Heidelberg University. All patients gave written informed consent.

Study design

This was an open-label, single-dose, nonrandomised pilot study. Patients underwent a screening examination 2–7 days prior to the treatment day. On the treatment day, patients inhaled a single dose of ciclesonide (Alvesco®) (1,280 μg ex-actuator; equivalent to 1,600 μg ex-valve; to be inhaled by eight puffs of 200 μg within 4 min) 2–24 h prior to scheduled surgical removal of the lung tissue. The time between inhalation of ciclesonide and scheduled surgical removal (*i.e.* clipping the resected tissue from the remaining part) of the lung tissue was planned to be divided into four groups of five patients each, to cover the complete period between 2 and 24 h. The intended grouping was $>2\text{--}4$ h, $>4\text{--}8$ h, $>8\text{--}16$ h and $>16\text{--}24$ h.

In addition, three patients did not take any study medication but donated lung tissue as controls for bioanalytic testing of matrix effects. Patients who received ciclesonide underwent a post-study examination 3–5 days after receiving treatment.

All measures that were required for the pre-operative management of the patient were performed irrespective of the current study, according to the medical needs of the patient. Concomitant medication for the adequate clinical management of the patient was allowed.

Serum samples

To assess adequate inhalation, blood samples for pharmacokinetic analysis were taken pre-dose and at 0.75, 1.5, 3, 6, 12 and 24 h post-dose. Blood (4 mL) was sampled into nonheparinised serum monovettes and immediately stored at $2\text{--}8^\circ\text{C}$ for 60–90 min to allow clotting. Samples were then centrifuged ($2,000 \times g$, 4°C) for 10 min to obtain serum, which was

transferred into polypropylene tubes. All samples were stored at -20°C until further analysis.

Lung tissue preparation

The lung tissue sample for analyses was excised from the surgical lung specimen immediately after resection. The sample had to measure at least $2.5 \times 2.5 \times 2.5$ cm (equivalent to a volume of ~ 16 mL). If a single piece of tissue of this size was not available, several smaller pieces from one anatomical area were dissected until a total volume of ~ 16 mL was reached. A central airway had to be identified within the sample and the bronchial tree was dissected removing the adjacent tissue, as long as the small bronchi could be clearly identified macroscopically. For the purpose of this study, a central airway was defined as having a diameter of $\sim 1\text{--}3.5$ mm in order to start the dissection with cartilaginous airways, which helps to clearly identify bronchi within the sample. Peripheral airways with cross-sections < 1 mm, together with adjacent lung tissue, represented the peripheral lung tissue.

All lung tissue samples (two or three aliquots per sample) were stored at -80°C after dissection until further analysis (pharm-analyt Labor GmbH, Baden, Austria). Lung samples were weighed, cut into smaller pieces, thawed, homogenised in ethanol and extracted in cold conditions ($8\text{--}15^\circ\text{C}$). The samples were centrifuged ($3,000 \times g$) and extracts were collected.

Histological specimens of lung tissue from patients treated with the study drug and those in the control group were also prepared and stained with haemalaun and eosin.

Analysis of ciclesonide and metabolites

For the lung samples, the concentrations of ciclesonide, des-CIC and the two fatty acid conjugates of des-CIC (des-CIC oleate and des-CIC palmitate) were determined using high-performance liquid chromatography with tandem mass spectrometry (LC-MS/MS). The analytical method for the tissue samples was established at pharm-analyt Labor GmbH using lung tissue samples from pigs, as well as from patients in the control group who received no ciclesonide. For des-CIC in serum, analyses were performed by Nycomed GmbH. The analytical methods are based on validated LC-MS/MS methods used previously in other pharmacokinetic studies [16–19].

Due to the different molecular weights of the metabolites, a comparison based on molar concentration was considered most appropriate. The lower limits of quantitation (LLOQs), based on tissue samples of 0.5 g, were $0.563 \text{ pmol}\cdot\text{g}^{-1}$ for ciclesonide, $0.644 \text{ pmol}\cdot\text{g}^{-1}$ for des-CIC, $0.136 \text{ pmol}\cdot\text{g}^{-1}$ for des-CIC oleate and $0.0424 \text{ pmol}\cdot\text{g}^{-1}$ for des-CIC palmitate. Dilution factors were taken into account when a sample of < 0.5 g was provided.

Safety

Safety and tolerability assessments were based on adverse events reported during the entire study, clinical laboratory values, lung function tests (spirometry), physical examinations, ECGs, blood pressure, pulse rate and body temperature. Vital signs were measured as close as possible to the time-points scheduled for the pharmacokinetic measurements.

Pharmacokinetic evaluation

The sample size of 20 patients receiving ciclesonide and three control patients was chosen on grounds of feasibility. The primary variables in this study were the lung-tissue concentrations of ciclesonide, des-CIC and the fatty acid conjugates of des-CIC. The total concentration of ciclesonide-related compounds (sum of ciclesonide, des-CIC, des-CIC oleate and des-CIC palmitate) was also calculated, and compared between central and peripheral lung tissue. The following pharmacokinetic parameter characteristics were also assessed for the active metabolite, des-CIC, based on the concentrations in serum at 0.75, 1.5, 3, 6, 12 and 24 h post-dose: maximum serum concentration (C_{max}), time to reach C_{max} (t_{max}), terminal elimination half-life ($t_{1/2}$) and area under the serum concentration–time profile extrapolated to infinity (AUC_{0-inf}). AUC_{0-inf} was calculated using the trapezoidal formula up to the last sampling time with a concentration above the LLOQ ($10 \text{ pg}\cdot\text{mL}^{-1}$) and extrapolated to infinity using standard techniques. C_{max} and t_{max} were obtained directly from the serum concentration–time profiles. The pharmacokinetic evaluation was performed using WinNonlin professional, Version 4.1 (Pharsight Corporation, Mountain View, CA, USA). All variables were analysed in a descriptive manner using summary statistics including mean, SD or SEM. Secondary safety variables were analysed in a descriptive manner.

RESULTS

Subjects

A total of 20 patients (15 males and five females) with a median age of 58 yrs were included in the study (table 1). All of the patients were undergoing surgery for pulmonary malignant lesions (lung cancer, $n=17$; pulmonary metastases of breast cancer, $n=2$; pulmonary metastases of sarcoma, $n=1$) and underwent anatomical segmentectomy or lobectomy. All patients were Caucasian and had a median height of 174 cm and a median weight of 74 kg. The majority of patients ($n=19$) were ex- ($n=11$) or current ($n=8$) smokers. 10 patients were assigned to have mild to moderate chronic obstructive pulmonary disease according to their medical records. All patients had an $FEV_1 \geq 70\%$ predicted.

TABLE 1 Patient characteristics and lung function parameters

Characteristic	Patients undergoing elective surgery [#]
Sex	
Male	15
Female	5
Age yrs	58 (28–69)
Height cm	174 (158–185)
Weight kg	74 (53–103)
FEV₁ L	2.9 (1.8–4.2)
FEV₁ % pred	89 (70–119)
FVC L	4.1 (2.7–6.4)

Data are presented as n or median (range). FEV₁: forced expiratory volume in 1 s; % pred: % predicted; FVC: forced vital capacity. [#]: $n=20$.

Assay performance

Serum concentrations of des-CIC were determined using a previously validated bioanalytical method. The LLOQ was $10 \text{ pg}\cdot\text{mL}^{-1}$. The interbatch precision and accuracy of the quality control (QC) samples for des-CIC in serum were 1.88–6.52% and 97.3–97.9%, respectively.

The bioanalytical evaluations of ciclesonide, des-CIC and the oleate and palmitate of des-CIC in lung tissue were performed using a validated LC-MS/MS method (LLOQ provided previously). The interbatch precision and accuracy results of the QC samples for all analytes in tissue were 2.79–5.44% and 93.9–105.6%, respectively.

Pharmacokinetics

The pharmacokinetic evaluation focused on the pharmacologically active metabolite des-CIC, which is the major metabolite in serum. The C_{max} of des-CIC was attained between 0.75–1.5 h after ciclesonide inhalation, and the mean C_{max} of des-CIC was $3.29 \text{ pmol}\cdot\text{mL}^{-1}$ ($1.55 \text{ }\mu\text{g}\cdot\text{L}^{-1}$). Serum concentrations of des-CIC decreased with a mean $t_{1/2}$ of 5.7 h (table 2).

Lung tissue

There were no interfering matrix effects in the bioanalysis as ciclesonide, des-CIC, des-CIC oleate and des-CIC palmitate were not detectable in the control samples from the three patients who did not inhale ciclesonide. Histological evaluation of all peripheral lung tissue samples revealed normal lung parenchyma.

For patients who had inhaled ciclesonide, a total of 39 central and peripheral lung tissue samples were available for the bioanalysis (no central tissue sample was obtained in one patient dosed 2.08 h prior to surgery), and three aliquots per sample were obtained for 19 patients. Because the concentrations of analytes in the aliquots were below the LLOQ in some cases, median concentrations were provided for the corresponding sample.

Ciclesonide, des-CIC, des-CIC oleate and des-CIC palmitate were detectable in the tissue samples and the individual results are given in figure 1 and figure 2.

A substantial proportion of ciclesonide was hydrolysed to des-CIC even at the earliest available time-point of 2.08 h after

TABLE 2 Pharmacokinetic parameter estimates of desisobutryl-ciclesonide in serum of 20 patients after a single dose of 1,280 μg ciclesonide

	Mean	SD	SEM
AUC_{0-inf} pmol·h·mL⁻¹	15.49	7.33	1.64
C_{max} pmol·mL⁻¹	3.29	1.68	0.38
t_{max} h	0.86	0.27	0.06
$t_{1/2}$ h	5.70	1.39	0.31

Data are presented as molar concentrations. AUC_{0-inf} : area under the serum concentration–time profile extrapolated to infinity; C_{max} : maximum serum concentration; t_{max} : time to reach maximum serum concentration; $t_{1/2}$: terminal elimination half-life.

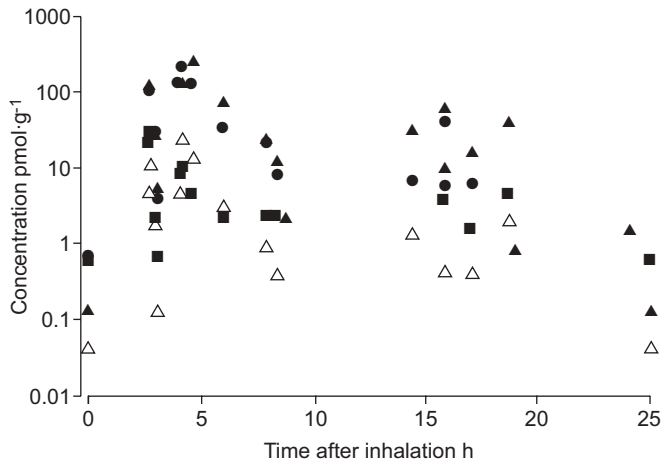


FIGURE 1. Concentrations of ciclesonide (CIC; ■), and its active metabolite desisobutryl-CIC (des-CIC; ●) and fatty acid ester conjugates (des-CIC oleate (▲) and des-CIC palmitate (Δ)) in central lung tissue. The lower limit of quantitation of the compounds is provided for illustration at 0 and 25 h.

inhalation of ciclesonide. In peripheral lung tissue, the highest concentration of des-CIC ($30.2 \text{ pmol}\cdot\text{g}^{-1}$) was measured at this time-point. In almost all samples with detectable des-CIC concentrations, the level of the metabolite was higher than that of the parent compound. The concentrations of ciclesonide were below the LLOQ in the majority of central lung tissue samples collected ≥ 14 h after inhalation. The same was true for all samples of peripheral lung tissue obtained ≥ 4.5 h after inhalation. After 8 h, des-CIC oleate was the main metabolite in both the central and peripheral lung tissue samples. At least one of the metabolites of ciclesonide was present in the lung tissue samples at all study time-points up to 24 h.

The distribution of time-points of resection following inhalation was well balanced within the investigational period, as five patients were included per group (>2 –4 h, >4 –8 h, >8 –16 h and >16 –24 h). The earliest time-point was 2.08 h and the latest time-point after inhalation was 24.08 h. Table 3 presents the median concentrations of ciclesonide and its metabolites in lung tissue.

After inhalation of $1,280 \mu\text{g}$ ciclesonide, the concentrations of ciclesonide and its metabolites (des-CIC, des-CIC oleate and des-CIC palmitate) were higher in the central airway tissue than in the peripheral lung tissue. Des-CIC and des-CIC oleate were the main metabolites in both lung areas. The levels of des-CIC palmitate were considerably lower than those of des-CIC oleate overall and in both lung areas (table 3).

Comparison of total ciclesonide-related compounds

Overall, higher concentrations of total ciclesonide-related compounds were found in the central airways compared with peripheral lung tissue. No time-dependent changes were noted in the ratio of total concentrations in central to peripheral lung tissue during the 24 h after inhalation of ciclesonide (fig. 3). Regression analysis of the two sets of data resulted in apparent $t_{1/2}$ of the total ciclesonide-related concentrations of ~ 5 h in the lung tissue.

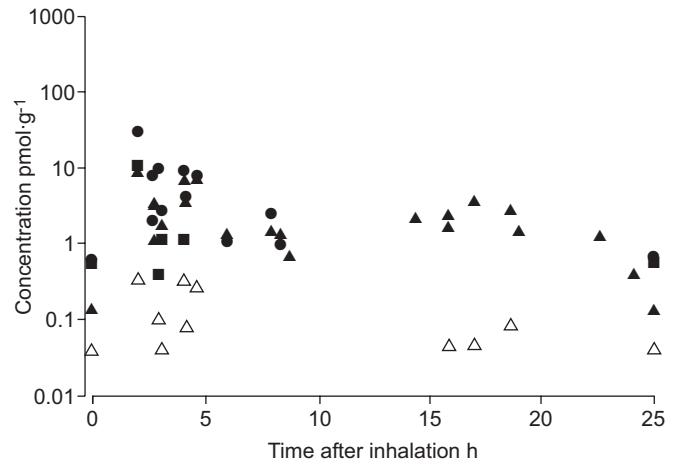


FIGURE 2. Concentrations of ciclesonide (CIC; ■), and its active metabolite desisobutryl-CIC (des-CIC; ●) and fatty acid ester conjugates (des-CIC oleate (▲) and des-CIC palmitate (Δ)) in peripheral lung tissue. The lower limit of quantitation of the compounds is provided for illustration at 0 and 25 h. The lung resections of two patients were performed at the same time (4.17 h following inhalation); therefore, a slight offset for different time-points in these patients was used for better illustration.

The concentration of des-CIC in serum decreased similarly over time, resulting in a $t_{1/2}$ of 5.7 h (table 2); however, the concentrations were lower. The mean concentration of the major metabolite des-CIC in serum at 1.5 and 24 h was 2.5 and $0.1 \text{ pmol}\cdot\text{mL}^{-1}$, respectively.

Safety analysis

A total of seven treatment-emergent adverse events (TEAEs) (atrial fibrillation, acute respiratory distress syndrome, pneumonia, sepsis, enteritis, QTc interval prolongation and pyothorax) were reported by four patients. Three TEAEs were mild or moderate in intensity and four (in two patients) were severe. In all cases, TEAEs were determined to be unrelated to the study medication. None of the TEAEs led to study discontinuation. Laboratory abnormalities and findings at physical examination were surgery-related and assessed by the investigators not to be clinically relevant.

DISCUSSION

After inhalation of a single dose of $1,280 \mu\text{g}$ ciclesonide by patients undergoing elective lung surgery, ciclesonide was rapidly hydrolysed to des-CIC in both central airways and peripheral lung tissue. Des-CIC and des-CIC oleate were the main metabolites in both compartments, and at least one of the metabolites of ciclesonide (des-CIC, des-CIC oleate or des-CIC palmitate) was present in the lung tissue samples at all study time-points up to 24 h. The results are of relevance as the study was performed in humans who inhaled ciclesonide and tissue samples of the target organ of the drug were taken from living patients. These *in vivo* findings confirm the two major metabolic pathways of ciclesonide in the human lung: the rapid, on-site conversion of ciclesonide to des-CIC and the conjugation of des-CIC with fatty acids, in particular oleic acid. Such findings have previously been shown *in vitro* in human tissue and *in vivo* in animals [13–15].

TABLE 3 Median concentrations ($\text{pmol}\cdot\text{g}^{-1}$) of ciclesonide and its metabolites in lung tissue after inhalation of 1,280 μg ciclesonide

Group	n	Median time h	Peripheral lung tissue				Central lung tissue			
			CIC	Des-CIC	Des-CIC oleate	Des-CIC palmitate	CIC	Des-CIC	Des-CIC oleate	Des-CIC palmitate
>2–4 h	5	2.8	<0.56	8.33	2.85	0.04	11.7	62.5	78.0	3.36
>4–8 h	5	4.7	<0.56	4.43	3.64	0.08	4.21	120	134	4.51
>8–16 h	5	14.4	<0.56	<0.64	1.69	<0.04	<0.56	6.60	12.5	0.43
>16–24 h	5	19.0	<0.56	<0.64	1.50	<0.04	<0.56	<0.64	1.49	<0.04

In the >2–4 h group, no central tissue sample was obtained from one patient (2.08 h), resulting in apparently lower median concentrations. CIC: ciclesonide; des-CIC: desisobutyryl-CIC.

A 1,280 μg dose of ciclesonide was selected in the current study as this was expected to be sufficient for detection in the pharmacokinetic analysis. Previous studies have indicated the safety of 1,280 $\mu\text{g}\cdot\text{day}^{-1}$ ciclesonide [20, 21]. As ciclesonide was always inhaled ≥ 2 h prior to anaesthesia, the absorption of the drug is assumed not to be influenced by the anaesthetics. The pharmacokinetic profiles of serum des-CIC concentrations measured over time indicated that drug inhalation by patients (of whom, the majority were ex- or current smokers) was adequate; the AUC of des-CIC in this study was 6.68 $\mu\text{g}\cdot\text{h}\cdot\text{L}^{-1}$ following a single dose of 1,280 μg ciclesonide. The dose-adjusted AUC values in healthy subjects were in the range of 4.11 $\mu\text{g}\cdot\text{h}\cdot\text{L}^{-1}$ and 7.32 $\mu\text{g}\cdot\text{h}\cdot\text{L}^{-1}$ [16, 17]. The pharmacokinetic and pharmacodynamic properties of inhaled ciclesonide have been described in recent publications [22–24].

Total lung tissue concentrations of ciclesonide, des-CIC and the fatty acid conjugates of des-CIC were higher in the central airways than in the peripheral-lung tissue. However, the amount of the drug that reached the peripheral lung was substantial, which is consistent with previous findings of high peripheral lung deposition of ciclesonide [10]. Overall, 55.8%

of the deposited dose was found in the peripheral lung that represented $\sim 75\%$ of the lung volume. Effects of ciclesonide on small airways were recently demonstrated in a clinical study [25]. The elimination of the total concentrations was similar in peripheral and central lung tissue. Therefore, the ratio between peripheral and central lung appeared to be stable. At all time-points, the total ciclesonide-related concentrations in the central and peripheral lung tissue were clearly higher than those of des-CIC in serum. Clearance appeared to occur in parallel for the lung tissue and the systemic circulation.

Previous studies investigating the distribution of other ICS (fluticasone propionate, beclomethasone dipropionate and budesonide) have used samples from resected pulmonary tissue and confirmed that the experimental approach is appropriate to characterise the pulmonary disposition of ICS [26–29]. One study showed high fluticasone propionate concentrations in central lung tissue for a longer time period compared to the tissue concentration in peripheral lung tissue, as well as higher concentrations in the lung tissue than in serum [26]. Similarly, concentrations of budesonide in blood plasma were lower than those in lung tissue and concentration levels fall almost in parallel over time [28]. Furthermore, following inhalation of beclomethasone dipropionate *via* a HFA metered-dose inhaler, serum concentrations of beclomethasone-17-monopropionate were lower compared to the lung tissue concentration of this metabolite [27]. In a recent study, the deposition of budesonide and fluticasone propionate were directly compared [29]. Patients were given single 1,000- μg doses of both budesonide and fluticasone propionate *via* dry-powder inhalers before lung surgery. In addition to tissue samples from the central and peripheral lung, *ex vivo* bronchial brush samples were taken during surgery. Interestingly, the highest concentration of fluticasone propionate was detected in bronchial brush samples and detectable levels of this drug for ≤ 18 h, suggesting the presence of undissolved drug-powder particles in the airway lumen. The mean concentrations of budesonide and fluticasone propionate in the peripheral lung tissue at the interval 1–6 h were 4.3 $\text{pmol}\cdot\text{g}^{-1}$ and 18.5 $\text{pmol}\cdot\text{g}^{-1}$, respectively [29]. Overall, this is a similar order of magnitude compared to des-CIC with a median value of 8.33 $\text{pmol}\cdot\text{g}^{-1}$ in the interval of 2–4 h. However, interstudy comparisons should be interpreted with caution, due to variables such as the bioanalytical assays used,

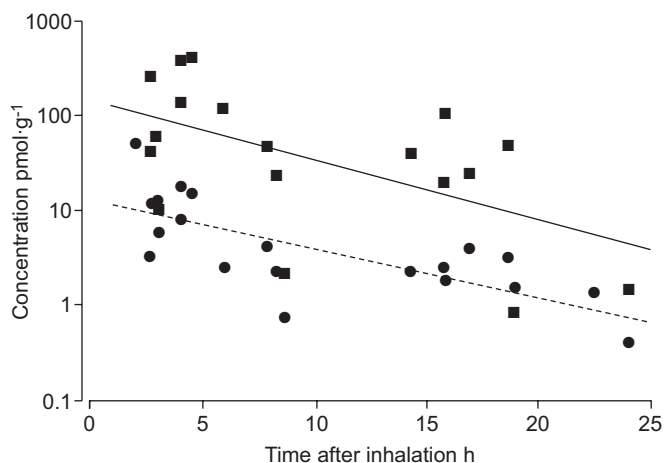


FIGURE 3. Total concentrations (sum of the concentrations for ciclesonide, desisobutyryl-ciclesonide, oleate and palmitate) in central (■) and peripheral (●) lung tissue. The corresponding linear regression lines are shown (—: central; ---: peripheral).

the special populations studied, the low number of patients included and the devices used. In the previously mentioned study, the intracellular esterification of budesonide *in vivo* was confirmed but was not observed for fluticasone propionate [29]. This is in line with *in vitro* investigation, using human precision-cut lung slices, demonstrating that des-CIC and budesonide, in contrast to fluticasone propionate, form fatty acid esters [15]. Fatty acid esters of budesonide were formed rapidly after inhalation of the drug and budesonide oleate is the major metabolite in the lung. A similar metabolic pathway was confirmed in humans for intracellularly formed des-CIC *in vitro*, as well as in this *in vivo* investigation.

The current study has confirmed rapid activation of ciclesonide in the lung *in vivo*. For example, at the earliest time-point in peripheral lung tissue, the parent compound ciclesonide represented 21.5% of the total drug concentration ($10.9 \text{ pmol}\cdot\text{g}^{-1}$), compared with 78.5% ($39.75 \text{ pmol}\cdot\text{g}^{-1}$) for the metabolites. *In vitro* investigation using precision-cut human lung slices has demonstrated that ciclesonide is initially converted to des-CIC and subsequently conjugated with fatty acids, with des-CIC oleate as the main metabolite [14, 15]. Using ^{14}C -ciclesonide *in vitro* and detecting radioactive ciclesonide and metabolites, a complete picture of metabolism can be obtained [14]. For all metabolites, reference compounds and sensitive bioanalytical assays are available that were used as the basis for the current study to investigate *in vivo* metabolism without using radiolabelled drug. It should also be noted that the LLOQ for des-CIC oleate was surprisingly low ($0.136 \text{ pmol}\cdot\text{g}^{-1}$ based on tissue samples of 0.5 g) and, therefore, was detectable even in very small quantities.

Due to the special patient population in the current study (*i.e.* patients undergoing elective surgery), a similar study for steady state conditions is not feasible. However, studies in rats following repeated inhalation of ciclesonide for 4 weeks have clearly demonstrated that the active metabolite, as well as the fatty acid ester conjugate, were formed and still detected 27 h following the last inhalation [13].

In conclusion, ciclesonide is rapidly converted to the active metabolite, des-CIC, in central and peripheral human lung tissue *in vivo*, and des-CIC forms conjugates with fatty acids, in particular oleic acid. The formation and the depot-like storage of reversibly formed des-CIC fatty acid conjugates ≤ 24 h after inhalation may support the once-daily dosing regimen of ciclesonide.

STATEMENT OF INTEREST

Statements of interest for R. Nave, H. Boss and H. Magnussen can be found at www.erj.ersjournals.com/site/misc/statements.xhtml

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REFERENCES

1 Holgate ST, Polosa R. The mechanisms, diagnosis, and management of severe asthma in adults. *Lancet* 2006; 368: 780–793.

- 2 Canonica GW. Treating asthma as an inflammatory disease. *Chest* 2006; 130: Suppl. 1, 21S–28S.
- 3 National Asthma Education and Prevention Program, Expert Panel Report: Guidelines for the Diagnosis and Management of Asthma Update on Selected Topics – 2002. *J Allergy Clin Immunol* 2002; 110: Suppl. 5, S141–S219.
- 4 Global Initiative for Asthma. Global Strategy for Asthma Management and Prevention. www.ginasthma.org/Guidelineitem.asp?i1=2&i2=1&intId=1561 Date last accessed: 2008. Date last updated: January 12, 2010.
- 5 Dietzel K, Engelstätter R, Keller A. Ciclesonide: an on-site activated steroid. *Prog Respir Res* 2001; 31: 91–93.
- 6 Mutch E, Nave R, McCracken N, *et al.* The role of esterases in the metabolism of ciclesonide to desisobutyryl-ciclesonide in human tissue. *Biochem Pharmacol* 2007; 73: 1657–1664.
- 7 Nave R, McCracken N. Metabolism of ciclesonide in the upper and lower airways. Review of available data. *J Asthma Allergy* 2008; 1: 11–18.
- 8 Belvisi MG, Bundschuh DS, Stoeck M, *et al.* Preclinical profile of ciclesonide, a novel corticosteroid for the treatment of asthma. *J Pharmacol Exp Ther* 2005; 314: 568–574.
- 9 Leach CL, Bethke TD, Boudreau RJ, *et al.* Two-dimensional and three-dimensional imaging show ciclesonide has high lung deposition and peripheral distribution: a nonrandomized study in healthy volunteers. *J Aerosol Med* 2006; 19: 117–126.
- 10 Newman S, Salmon A, Nave R, *et al.* High lung deposition of (^{99m}Tc)-labeled ciclesonide administered *via* HFA-MDI to patients with asthma. *Respir Med* 2006; 100: 375–384.
- 11 Nave R, Zech K, Bethke TD. Lower oropharyngeal deposition of inhaled ciclesonide *via* hydrofluoroalkane metered-dose inhaler compared with budesonide *via* chlorofluorocarbon metered-dose inhaler in healthy subjects. *Eur J Clin Pharmacol* 2005; 61: 203–208.
- 12 Richter K, Kannies F, Biberger C, *et al.* Comparison of the oropharyngeal deposition of inhaled ciclesonide and fluticasone propionate in patients with asthma. *J Clin Pharmacol* 2005; 45: 146–152.
- 13 Nave R, Meyer W, Fuhst R, *et al.* Formation of fatty acid conjugates of ciclesonide active metabolite in the rat lung after 4-week inhalation of ciclesonide. *Pulm Pharmacol Ther* 2005; 18: 390–396.
- 14 Nave R, Fisher R, Zech K. *In vitro* metabolism of ciclesonide in human lung and liver precision-cut tissue slices. *Biopharm Drug Dispos* 2006; 27: 197–207.
- 15 Nave R, Fisher R, McCracken N. *In vitro* metabolism of beclomethasone dipropionate, budesonide, ciclesonide, and fluticasone propionate in human lung precision-cut tissue slices. *Respiratory Research* 2007; 8: 65.
- 16 Nave R, Gunawardena KA, Zech K, *et al.* Pharmacokinetic disposition of inhaled ciclesonide and its metabolite desisobutyryl-ciclesonide in healthy subjects and patients with asthma are similar. *Int J Clin Pharmacol Ther* 2006; 44: 1–7.
- 17 Nave R, Drollmann A, Steinijans VW, *et al.* Lack of pharmacokinetic drug–drug interaction between ciclesonide and erythromycin. *Int J Clin Pharmacol Ther* 2005; 43: 264–270.
- 18 Drollmann A, Nave R, Steinijans VW, *et al.* Equivalent pharmacokinetics of the active metabolite of ciclesonide with and without use of the AeroChamber Plus spacer for inhalation. *Clin Pharmacokinet* 2006; 45: 729–736.
- 19 Mascher HJ, Zech K, Mascher DG. Sensitive simultaneous determination of ciclesonide, ciclesonide-M1-metabolite and fluticasone propionate in human serum by HPLC-MS/MS with APPI. *J Chromatogr B Analyt Technol Biomed Life Sci* 2008; 869: 84–92.
- 20 Derom E, van De Velde V, Marissens S, *et al.* Effects of inhaled ciclesonide and fluticasone propionate on cortisol secretion and airway responsiveness to adenosine 5′monophosphate in asthmatic patients. *Pulm Pharmacol Ther* 2005; 18: 328–336.
- 21 Szeffler S, Rohatagi S, Williams J, *et al.* Ciclesonide, a novel inhaled steroid, does not affect hypothalamic–pituitary–adrenal axis function in patients with moderate-to-severe persistent asthma. *Chest* 2005; 128: 1104–1114.

- 22** Nave R. Clinical pharmacokinetic and pharmacodynamic profile of inhaled ciclesonide. *Clin Pharmacokinet* 2009; 48: 243–252.
- 23** Derom E, Louis R, Tiesler C, *et al.* Effects of ciclesonide and fluticasone on cortisol secretion in patients with persistent asthma. *Eur Respir J* 2009; 33: 1277–1286.
- 24** Xu J, Nave R, Lahu G, *et al.* Population pharmacokinetics and pharmacodynamics of inhaled ciclesonide and fluticasone propionate in patients with persistent asthma. *J Clin Pharmacol* 2010; [Epub ahead of print DOI: 10.1177/0091270009354994].
- 25** Cohen J, Douma WR, ten Hacken NHT, *et al.* Ciclesonide improves measures of small airway involvement in asthma. *Eur Respir J* 2008; 31: 1213–1220.
- 26** Esmailpour N, Hogger P, Rabe KF, *et al.* Distribution of inhaled fluticasone propionate between human lung tissue and serum *in vivo*. *Eur Respir J* 1997; 10: 1496–1499.
- 27** Holz O, Zuhlke I, Einhaus M, *et al.* Direct measurement of BDP and 17-BMP in bronchial and peripheral lung tissue after inhalation of HFA- vs CFC-driven aerosols. *Pulm Pharmacol Ther* 2004; 17: 233–238.
- 28** Van den Bosch JM, Westermann CJ, Aumann J, *et al.* Relationship between lung tissue and blood plasma concentrations of inhaled budesonide. *Biopharm Drug Dispos* 1993; 14: 455–459.
- 29** Van den Brimk KIM, Boorsma M, Staal-van den Brekel AJ, *et al.* Evidence of the *in vivo* esterification of budesonide in human airways. *Br J Pharmacol* 2008; 66: 27–35.