

Heterotrimerization of HSF1 and HSF2 provides a transcriptional switch in response to distinct stimuli

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Short title: HSF1-2 heterotrimers and transcription

Abbreviations:

HSF - heat shock factor

Hsp - heat shock protein

nSB – nuclear stress body

Sat III – satellite III

Abstract

Organisms respond to circumstances threatening the cellular protein homeostasis by activation of heat shock transcription factors (HSFs), which play important roles in stress resistance, development and longevity. Of the four HSFs in vertebrates (HSF1-4), HSF1 is activated by stress whereas HSF2 lacks intrinsic stress responsiveness. The mechanism by which HSF2 is recruited to stress-inducible promoters and how HSF2 is activated is not known. However, changes in the HSF2 expression occur, coinciding with the functions of HSF2 in development. Here, we demonstrate that HSF1 and HSF2 form heterotrimers when bound to satellite III DNA in nuclear stress bodies, subnuclear structures where HSF1 induces transcription. By depleting HSF2, we show that HSF1-HSF2 heterotrimerization is a mechanism regulating transcription. Upon stress, HSF2 DNA-binding is HSF1 dependent. Intriguingly, when the elevated expression of HSF2 during development is mimicked, HSF2 binds to DNA and becomes transcriptionally competent. HSF2 activation leads to activation of also HSF1, revealing a functional interdependency that is mediated through the conserved trimerization domains of these factors. We propose that heterotrimerization of HSF1 and HSF2 integrates transcriptional activation in response to distinct stress and developmental stimuli.

Introduction

Cells react to stressful conditions by activation of heat shock factors (HSFs), of which there are three mammalian isoforms; HSF1, HSF2 and HSF4 (Pirkkala *et al.*, 2001). Activated HSFs bind to heat shock elements (HSEs) within the promoters of their target genes and induce synthesis of protective molecular chaperones called heat shock proteins (Hsps). Hsps prevent protein misfolding and are required for stress resistance and healthy cell growth, development and aging (Bukau *et al.*, 2006, Morimoto, 2008). They also protect against metabolic, neurodegenerative and cardiovascular disorders (Balch *et al.*, 2008). In addition to Hsps, HSFs are associated with expression of noncoding satellite III (sat III) RNA in nuclear stress bodies (nSBs) (Jolly *et al.*, 2004, Rizzi *et al.*, 2004), to which HSF1 and HSF2 translocate upon stress (Jolly *et al.*, 1997, Alastalo *et al.*, 2003). The nSBs form on the locus 9q12 consisting of pericentromeric heterochromatin, and the sat III transcripts provide scaffolds for docking of other nSB components, as shown for the splicing factor ASF/SF2 (Chiodi *et al.*, 2004, Metz *et al.*, 2004). Besides ASF/SF2, the RNA processing factors hnRNP A1-associated protein (HAP), hnRNPM, Sam68, and SRp30c localize to the nSBs (Weighardt *et al.*, 1999, Denegri *et al.*, 2001).

HSF1 is activated by classical stresses such as heat shock and heavy metals and responds to elevated temperatures *in vitro* (Ahn *et al.*, 2001, Anckar and Sistonen, 2007). HSF1 is also involved in development and has critical roles in longevity and cancer (Xiao *et al.*, 1999, Hsu *et al.*, 2003, Morley *et al.*, 2004, Dai *et al.*, 2007). Unlike HSF1, HSF2 lacks intrinsic stress responsiveness (Ahn *et al.*, 2001). Another major difference between these factors is that while HSF1 is evenly expressed, the levels of HSF2 fluctuate. These changes in expression coincide temporally with HSF2 DNA-binding activity during

developmental processes (Rallu *et al.*, 1997, Min *et al.*, 2000). The function of HSF2 in development was revealed by *hsf2*^{-/-} mice, which display neurological and reproductive abnormalities in both genders (Kallio *et al.*, 2002, Wang *et al.*, 2003, Chang *et al.*, 2006, Åkerfelt *et al.*, 2008). In addition, the stress-induced expression of *hsps* in *hsf2*^{-/-} cells is altered (Östling *et al.*, 2007). However, the mechanism by which HSF2 is recruited to stress-inducible promoters is not known. How HSF2 is activated, and the functional relationship between HSF1 and HSF2 also remain to be elucidated.

In this study, we use the nSBs as a model system to show that HSF1 and HSF2 interact as DNA-bound heterotrimers. When HSF1-HSF2 heterotrimerization is inhibited by depletion of HSF2, the transcription of sat III DNA is enhanced, indicating that HSF1-HSF2 heterotrimerization regulates transcription. We mimic the elevated HSF2 concentration during development and demonstrate that increased HSF2 expression induces transcription of sat III DNA and localization of both HSF1 and HSF2 to the nSBs. In testis, where HSF2 is abundantly expressed and plays a role in spermatogenesis (Sarge *et al.*, 1994, Fiorenza *et al.*, 1995, Kallio *et al.* 2002, Wang *et al.*, 2003, Åkerfelt *et al.*, 2008), we show interaction between HSF1 and HSF2. Increased HSF2 expression also induces transcription of the classical HSF target *hsp70*, suggesting that HSF2 is activated by its elevated expression. Importantly, while the stress-induced DNA binding of HSF2 is dependent on HSF1 activity, induced HSF2 expression converts HSF1 to a transcriptionally competent state. We propose that heterotrimerization is a transcriptional switch at the interface of activation by either HSF1 or HSF2.

Materials and Methods

Cell culture, treatments, plasmids and transfections

HeLa and HEK293T cells were cultured in Dulbecco's modified Eagle's medium and K562 cells in RPMI-1640 medium in 5% CO₂ at 37°C. The media were supplemented with 10% fetal calf serum, 2 mM L-glutamine, penicillin and streptomycin. Heat shocks were performed at 42°C in a water bath for the indicated time periods. HeLa and HEK293T cells were transfected by electroporation (975 μF, 220 V, Bio-Rad Gene Pulser). Plasmid DNA and 5×10⁶ cells in 0.4 ml of Optimem (Gibco-BRL) were added to a 0.4-cm-gap electroporation cuvette (BTX) and subjected to a single electric pulse. The mouse HSF1-YFP was generated as the mouse HSF1-CFP, described in Jolly *et al.*, 2002. The human HSF2-YFP, containing amino acids 1-214, was constructed by PCR and cloned into the BamHI and XhoI sites of pEYFP-N1 (Clontech). The tandem CFP-YFP construct was a kind gift of Richard I. Morimoto (Northwestern University, Evanston, IL). The mHSF2 α and mHSF2 β plasmids used for overexpression of HSF2 were described in Alastalo *et al.*, 2003. All constructs were sequenced.

RNA interference

Transient downregulation of HSF1 was performed by electroporation of Scramble and HSF1 RNAi plasmids in HEK293T (Östling *et al.*, 2007). The cells were harvested after 72 h incubation. The stable scrambled and HSF1-downregulating cell lines were generated by transfection of the pSuper shRNA Scrambled and HSF1 RNAi plasmids (Östling *et al.*, 2007) to HeLa cells, and single clones were established after selection

with neomycin. For downregulation of HSF2, siRNA against HSF2 or AllStars negative control siRNA was transfected using HiPerFect Transfection reagent (all from Qiagen).

Chromatin immunoprecipitation (ChIP)

ChIP was performed as in Östling *et al.*, 2007. K562 cells were cross-linked with 1% formaldehyde. Chromatin was sonicated and immunoprecipitated with antibodies against HSF1 (SPA-901, Stressgen), HSF2 SFI58 (Östling *et al.*, 2007) and normal rabbit serum (NS, Jackson Immuno Research Laboratory). The following primers were used for ChIP: *sat III* (based on clone17 in Valgardsdottir *et al.*, 2005), For 5'- AAT GAA CCC GAT GCA AT -3', Rev 5'- CCA TTC TTG TTG AAT CCA TT -3', *β-actin* For 5'-AAC TCT CCC TCC TCC TCT TCC TC-3', Rev 5'-GAG CCA TAA AAG GCA ACT TTC GG-3'.

Immunofluorescence

For immunofluorescence analysis, HeLa cells cultured on coverslips were fixed with -20°C methanol for 6 min or with 3% paraformaldehyde in PBS for 15 min. After three washes with PBS-0.5% Tween 20, the cells were incubated in blocking solution (1% BSA PBS-0.5% Tween 20) for 1 h. Rabbit anti-HSF1 (Holmberg *et al.*, 2000), rat anti-HSF1 (Neomarkers), rabbit anti-HSF2 (Sarge *et al.*, 1993) or rat anti-HSF2 (Neomarkers) antibodies were diluted in blocking solution and added for 1 h. Secondary antibodies, anti-rabbit Alexa 488 and anti-rat Alexa 568 (Molecular Probes), were incubated for 1 h. The coverslips were mounted and DNA was visualized using Vectashield mounting medium with DAPI (Vector Laboratories). The cells were analyzed with a Zeiss LSM510-Meta scanning confocal microscope equipped with the SP2 (version 3.2)

software. The images were acquired using a Plan-Apochromat 63x/1.4 Oil DIC objective and further processed using Adobe Photoshop and CorelDraw software.

Western blot analysis

Soluble cell extracts were prepared and subjected to SDS-PAGE followed by transfer to nitrocellulose membrane (Protran nitrocellulose, Schleicher & Schuell). HSF1 was detected by polyclonal anti-HSF1 antibodies (Sarge *et al.*, 1993; Holmberg *et al.*, 2000), HSF2 by polyclonal anti-HSF2 antibodies (Sarge *et al.*, 1993; Östling *et al.*, 2007) and Hsc70 by SPA-815 (StressGen). Secondary antibodies were horseradish peroxidase-conjugated and purchased from Promega or Amersham Biosciences. The immunoblots were developed with an enhanced chemiluminescence method (ECL kit, Amersham Biosciences).

Immunoprecipitation

Male hybrid mice of the B6129SF2/J strain were used in co-immunoprecipitation experiments. HSF2 knockout mice were obtained by matings of heterozygous mice that have been described earlier (Kallio *et al.*, 2002), and were maintained in a C57BL/6N background. The pathogen-free mice were housed under controlled environmental conditions and fed with complete pellet chow and allowed tap water. The mice were killed by CO₂ asphyxiation. All mice were handled in accordance with the institutional animal care policies of the Åbo Akademi University (Turku, Finland). For co-immunoprecipitation experiments, testes were isolated from 60-80 days old mice and lysed in 2 ml of lysis buffer (Alastalo *et al.*, 2003). The precleared cellular lysate was

incubated with anti-HSF1 (Neomarkers), anti-HSF2 (Neomarkers) or anti-Flag M2 (Sigma) antibodies at 4°C for 1 h under rotation, after which 40 µl of a 50% slurry of protein-G/Sepharose was added to the reaction mixture and incubated for 1 h at 4°C under rotation. After centrifugation, the Sepharose beads were washed with supplemented TEG buffer and the immunoprecipitated proteins were run on 8% SDS-PAGE and transferred to nitrocellulose filter for immunoblotting as described above.

Semiquantitative RT-PCR and real-time RT-PCR

RNA was isolated with the RNAeasy kit (Qiagen). Contaminating genomic DNA was removed with two DNase I treatments according to the RNAeasy protocol (Qiagen). Of each sample, 1 µg RNA was subjected to reverse transcription using the High Capacity cDNA Reverse Transcription Kit (Applied Biosystems). For semiquantitative RT-PCR, ABsolute Rox mix (Advanced Biotechnologies) was used and the PCR was run 40 cycles. The same sat III primers as for ChIP were used. The GAPDH primers were *GAPDH* For 5'- ACC CAC TCC TCC ACC TTT GA -3', *GAPDH* Rev 5'- TTG CTG TAG CCA AAT TCG TTG T-3'. Real-time RT-PCR analyses were performed with ABsolute cybrgreen mix (Advanced Biotechnologies Ltd.) and the ABI prism 5700 and 7900 (Applied Biosystems). Relative RNA quantities were normalized to GAPDH. For real-time RT-PCR, the following primers and probes were used: *sat III* For 5'-AAT GGA ATG CAA TGG AAT GG-3', *sat III* Rev 5'-CCT GTA CTC GGG TTG ATT CC-3', *GAPDH* For 5'-ACC CAC TCC TCC ACC TTT GA-3', *GAPDH* Rev 5'-CTG TTG CTG TAG CCA AAT TCG T-3' (Shumaker *et al.*, 2006). *hsp70* Probe 5'-FAM TTACACACCTGCTCCAGCTCCTTCCTCTT TAMRA-3', *hsp70* For 5'-

GCCGAGAAGGACGAGTTTGA-3', *hsp70* Rev 5'-CCTGGTACAGTCCGCTGATGA-3', *GAPDH* Probe 5'-FAM ACCAGGCGCCCAATACGACCAA TAMRA-3', *GAPDH* For 5'-GTTTCGACAGTCAGCCGCATC-3', *GAPDH* Rev 5'-GGAATTTGCCATGGGTGGA-3'.

Structural modeling

The structural model of the human HSF heterotrimer of two HSF1 (aa 16-205) molecules and one HSF2 (aa 8-194) was done in three steps. Firstly, a template of the DNA-binding domain of six *K. lactis* HSF monomers bound to a 32-bp DNA was generated using SYBYL[®] 7.3 (Tripos Inc., St. Louis, Missouri, USA) by aligning three dimers of the crystal structure of *K. lactis* HSF bound to DNA next to each other as suggested by Littlefield and Nelson (1999). Secondly, the HR-A domain was aligned against the *E. coli* Lpp-56 X-ray structure (Shu *et al.*, 2000) and the HR-B domain was aligned against the mH38-P1 GCN4 Leucine Zipper X-ray structure (Shu *et al.*, 1999), resulting in the template structure for the HR-A/B trimerization domain. The alignments were done according to the characteristic heptad repeat sequence (abcdefg)_n seen in coiled coil structures (Suppl. Figure 3). Thirdly, the final template used for modeling the heterotrimer of the DNA-binding and HR-A/B domain was generated by linking the two domains using the X-ray structure of human GABP α protein (Batchelor *et al.*, 1998). In the final model of the heterodimer, HSF2 makes both head-to-head and tail-to-tail contacts with HSF1. For sequence alignments, MALIGN and MALFORM (Johnson *et al.*, 1993) were used within the Bodil visualization and modeling package (Lehtonen *et al.*, 2004). Ten models were generated with Modeller (Sali *et al.*, 1993) and the model

with the lowest objective function was chosen. Sequence alignment in Supplementary Figure 3 was done with ALSRIPT (Barton, 1993), and Figure 2A and B were created with the PYMOL Molecular Graphics System (DeLano Scientific).

Confocal microscopy and two-photon fluorescence lifetime imaging

The two-photon and confocal microscopy on HeLa cells was performed with an inverted two-photon laser scanning microscope Axiovert 200M (LSM510 NLO META, Carl Zeiss). During the experiment, cells were maintained at 37°C in a humidified atmosphere containing 5% CO₂ using an on-stage incubator (PeCon, Germany). All measurements were performed using a 63×/1.4 oil immersion plan-apochromat objective. In the fluorescence lifetime imaging (FLIM) experiments, the fluorescence decays were measured by the time-correlated single photon counting technique. Fluorescence decays were fitted using a bi-exponential model and the corresponding mean decay time in each pixel was color coded to obtain FLIM images (SPCImage software, Becker&Hickl). FRET was identified by the shorter lifetime of donor (CFP) in the presence of acceptor (τ_{DA}) as compared to that (τ_D) in the control donor-only cells. The FLIM/FRET efficiency

was calculated as: $E_{FLIM/FRET} = \left(1 - \frac{\tau_{DA}}{\tau_D}\right)$.

Additional acceptor photobleaching experiments were carried out on the same cell and completed with FLIM measurements to confirm FRET. At least five cells were measured for each experimental condition.

Results

Stress-induced translocation of HSF2 in the nSBs is HSF1 dependent

The colocalization of HSF1 and HSF2 into the nSBs (Alastalo *et al.*, 2003) prompted us to further investigate their functional relationship. HSF1 binds to DNA in the nSBs (Jolly *et al.*, 2002), and we performed chromatin immunoprecipitation (ChIP) in K562 cells to examine the binding of HSF2 to sat III DNA. Heat shock-induced binding of HSF1 and HSF2 was detected (Figure 1A), indicating that both HSFs occupy the same sat III DNA sequences (see also Suppl. Figure 1). To investigate how HSF1 affects the localization of HSF2, we generated a HeLa cell line stably downregulating HSF1 using vector-based RNAi (Figure 1C). In HSF1-depleted cells, the localization of both HSF1 and HSF2 into nSBs was abrogated (Figure 1B, for additional time points, see Suppl. Figure 2A), inferring that stress-induced HSF2 DNA-binding activity is dependent on HSF1. This result is supported by the finding that transient downregulation of HSF1 by shRNAs in HEK293T cells (Figure 1E) abolishes sat III transcription (Figure 1D, Suppl. Figure 2B).

HSF1 and HSF2 interact as DNA-bound heterotrimers

The dependency of HSF2 on HSF1 activity for localization to the nSBs (Figure 1B) raised the question of the underlying mechanism. Upon activation, both HSF1 and HSF2 form homotrimers through their trimerization domain consisting of the heptad repeats A and B (HR-A/B) (Sarge *et al.*, 1993). As HSF2 interacts with HSF1 via the HR-A/B domain (Alastalo *et al.*, 2003), we studied if HSF1 and HSF2 can form heterotrimers. We aligned the HR-A/B sequences of HSF1 and HSF2 and found that the amino acids involved in trimerization are conserved, especially well within the mid section of the HR-

A/B domains of HSF1 and HSF2 (Suppl. Figure 3). Using two HSF1 HR-A/B helices and one HSF2 HR-A/B, we made a structural model of the left-handed coiled coil that constitutes the trimer interface (Figure 2A). All buried polar residues, which have been suggested to play a role in partner verification, are conserved in the HSF1-HSF2 coiled coil structure (Figure 2A). Based on the crystal structure of DNA-bound *K. lactis* (KLUELA) HSF (Littlefield *et al.*, 1999), we generated a model of a human HSF1-HSF2 heterotrimer. The heterotrimer is bound to DNA and composed of the DNA-binding and HR-A/B domains of HSF1 and HSF2 (Figure 2B). Adjacent to the HSF1-HSF2 heterotrimer, an HSF1 homotrimer is shown. The trimers are bound to a 32-bp DNA double-stranded helix, with two nucleotide spacers as in a canonical HSE. We measured a distance of ~ 40 Å between the HR-A/B coiled coils of two HSF trimers (Figure 2B). Noncovalent contacts require proximities of <4 Å (Laberge, 1998), and at a distance of 40 Å, electrostatic interactions are unlikely to occur (Creighton, 1993), excluding interactions between separate trimers. This suggests that interactions between HSF1 and HSF2 on DNA is mediated through heterotrimerization.

Fluorescence lifetime imaging microscopy-based FRET (FLIM-FRET) was employed to investigate whether HSF1 and HSF2 interact on DNA within the nSBs. We could not detect FRET when fusing CFP and YFP to the C-termini of full-length HSF1 and HSF2, presumably because the C-termini of HSF1 and HSF2 are highly mobile, preventing FRET. Therefore, to facilitate the proper positioning of the CFP and YFP moieties, we used HSF1 and HSF2 C-terminal deletion constructs (Figure 2C), that translocate into nSBs spontaneously (Suppl. Figure 4A, Jolly *et al.*, 2004). These C-terminal deletion constructs contain the DNA-binding and HR-A/B domains of HSF1 and

HSF2, and should be apt for the study of interaction between the proteins as HSF1 and HSF2 interact through the HR-A/B domains (Alastalo *et al.*, 2003). Upon transfection of HeLa cells with the HSF1 and HSF2 constructs, the fluorescence lifetime of the donor (HSF1-CFP) was shorter in the presence of acceptor (HSF2-YFP) compared to that in cells with only HSF1-CFP (Figure 2D), indicating that FRET occurs. The FRET signal was predominantly localized to the nSBs with a mean FRET efficiency of 10%, a significant number considering that the FRET efficiency of an HSF1-CFP/HSF1-YFP pair representing HSF1 homotrimers was 14% (Figure 2E). Additional FACS-FRET (Suppl. Figure 4B) and acceptor photobleaching experiments (data not shown) confirmed the FRET results, showing that HSF1 and HSF2 interact as DNA-bound heterotrimers.

Stress-induced HSF activity is regulated through HSF1-HSF2 heterotrimerization

To investigate the impact of HSF1-HSF2 heterotrimerization on stress-induced transcription, we abrogated heterotrimer formation by depleting HSF2. HSF2-specific siRNAs were transfected to HEK293T cells (Figure 3B), and the transcription of sat III DNA was measured by real-time RT-PCR. A robust increase in the accumulation of sat III transcripts was evident when HSF2 was depleted (Figure 3A), demonstrating that HSF1-HSF2 heterotrimerization regulates HSF-mediated transcription. As expected, HSF2 knockdown did not alter the stress-induced relocalization of HSF1 to the nSBs (Figure 3C).

Elevated HSF2 expression induces HSF1-HSF2 heterotrimerization and activates transcription

HSF2 is involved in development, and several reports show a correlation between increased HSF2 expression and DNA-binding activity (Murphy *et al.*, 1994, Rallu *et al.*, 1997, Min *et al.*, 2000, Chang *et al.*, 2006, Åkerfelt *et al.*, 2008). Abrogation of HSF2 DNA binding in the nSBs was associated with reduced HSF2 protein levels (Figure 1C), indicating that HSF2 is regulated in a concentration-dependent manner. We investigated the impact of elevated HSF2 expression on sat III transcription in HeLa and HEK293T cells by real-time RT-PCR, and found that increased HSF2 concentration (Figure 4C, Suppl. Figure 5C) led to a prominent induction (~500 fold in HeLa cells, ~150 fold in HEK293T cells) of sat III transcription in the absence of stress (Figure 4A, Suppl. Figure 5A). To extend the study to other HSF targets, we measured the impact of HSF2 overexpression on the transcription of *hsp70*. In HeLa cells, transcription of also *hsp70* was induced ~2 fold (Figure 4B), suggesting that HSF2 is activated when abundantly expressed. No similar induction of *hsp70* was detected in HEK293T cells (Suppl. Figure 5B), which is probably due to the constitutive HSF activity in these cells, caused by the adenoviral transactivator E1A (Phillips *et al.*, 1991).

The effect of elevated HSF2 expression on the localization of HSF1 was studied with immunofluorescence in HeLa and HEK293T cells. When abundantly expressed, HSF2 translocated to the nSBs (Figure 4D, Suppl. Figure 5D), whereas it remained dispersed in the nucleoplasm of moderately overexpressing HeLa cells (Figure 4E), further suggesting that HSF2 is regulated by its concentration. Intriguingly, HSF2 recruited also HSF1 to the nSBs (Figure 4D), implying that the increased expression of HSF2 seen during development leads to activation of HSF1.

To provide more evidence for HSF1-HSF2 heterotrimerization as a regulatory mechanism of transcription during developmental processes, we chose to investigate interaction between HSF1 and HSF2 in mouse testis, a tissue undergoing active differentiation. Moreover, HSF2 is abundantly expressed in testis and has been shown to be active during spermatogenesis (Sarge *et al.*, 1994, Fiorenza *et al.*, 1995, Alastalo *et al.*, 1998, Kallio *et al.*, 2002, Wang *et al.*, 2003, Åkerfelt *et al.*, 2008). We performed co-immunoprecipitation of HSF1 and HSF2 in both wild-type and HSF2 knockout testis, and found that HSF2 could be co-immunoprecipitated with HSF1 antibodies and *vice versa* (Figure 4F). These results indicate that HSF1 and HSF2 form heterotrimers during spermatogenesis.

Discussion

Integration of HSF activity in response to stress and developmental stimuli

Trimerization is a crucial step in the activation process of HSFs that greatly increases the affinity for DNA (Xiao *et al.*, 1991). In this study, we show that HSF1 and HSF2 form heterotrimers and propose that HSF1-HSF2 heterotrimerization provides a switch that integrates transcriptional activation in response to stress and developmental stimuli (Figure 5). HSF1 is known to be activated by stress and responds to elevated temperatures *in vivo* and *in vitro*. We suggest that when activated, HSF1 does not only form homotrimers but also trimerizes with HSF2, which itself is inert to stress. This model explains why the translocation of HSF2 into the nSBs is abrogated when HSF1 is depleted, and also the previously suggested dependency of HSF2 on HSF1 for stress-induced DNA-binding activity (Östling *et al.*, 2007). Moreover, we show that elevated expression of HSF2 leads to its activation, suggesting that concentration regulates HSF2 during development (Figure 5). When activated, HSF2 incorporates HSF1 into a transcriptionally competent heterotrimer, illustrating the capacity of HSFs to initiate heterotrimerization. This shows, for the first time, an interdependent cooperativity between HSF1 and HSF2 that is mediated through the conserved HR-A/B domains. We propose that HSF1 needs cooperation from HSF2 in responding to developmental stimuli, and that some of the functions during development earlier ascribed to HSF1 are in fact a consequence of HSF1-HSF2 heterotrimer activity.

Implications of HSF1-HSF2 heterotrimerization

Our results show that HSF1-HSF2 heterotrimerization regulates stress-induced transcription, as demonstrated by increased expression of *sat III* transcripts when HSF2 is depleted. Also, the positive impact on *hsp70* and *hsp25*, and the negative impact on *hsp40* and *hsp110* transcription seen in *hsf2^{-/-}* cells (Östling *et al.*, 2007), can now be explained by HSF1-HSF2 heterotrimerization. HSF1 and HSF2 prefer architecturally different HSEs (Kroeger *et al.*, 1993). This suggests that HSF1-HSF2 heterotrimers bind DNA differently than homotrimers, and that the distinct regulation of HSF target genes could arise from binding of HSF1-HSF2 heterotrimers to specific sites. Another possibility is that HSF1-HSF2 heterotrimers compete with homotrimers for common binding sites. For example, the clusterin promoter contains an HSE which binds only one trimer, and on which HSF1-HSF2 heterotrimerization has been proposed to occur (Loison *et al.*, 2006). However, no formal proof for HSF1-HSF2 heterotrimerization on the clusterin promoter has yet been provided.

Another example of a protein family employing heterotrimerization is the matrilins. The matrilin heterotrimers have a variable stoichiometry that has been suggested to be determined by the concentration of the individual monomers (Frank *et al.*, 2002). Our model does not exclude variations in the stoichiometry of the HSF1-HSF2 heterotrimers, which may change similarly to that of the matrilin analogs. As the expression of HSF2 varies between different cell types and tissues (Rallu *et al.*, 1997, Fiorenza *et al.*, 1995), the HSF1-HSF2 heterotrimer stoichiometry could be modified accordingly, allowing tissue-specific regulation of HSF-mediated transcription.

Interestingly, HSF2 levels are reduced when HSF1 is downregulated (Figure 1C), a phenomenon that has been noted also by others (Rossi *et al.*, 2006) and is not due to

unspecific downregulation by RNAi (Suppl. Figure 6). Besides HSF1 downregulation, heat shock reduces the levels of HSF2 (Figure 3B). It is plausible that the HSF-mediated stress-induced transcription upon prolonged stress is determined by the receding amounts of HSF2 available for heterotrimerization with HSF1, and that HSF1-HSF2 heterotrimerization regulates transcription in a temporal manner.

The nSBs – versatile centers for regulation of gene expression?

The localization of HSF1 and HSF2 to the locus 9q12 is followed by expression of sat III transcripts and formation of nSBs. Transcription of sat III DNA is a general response to stress (Valgardsdottir *et al.*, 2008), and the sat III transcripts are noncoding and heterogenous in size (Jolly *et al.*, 2004, Rizzi *et al.*, 2004). The transcripts bind a subset of mRNA-processing factors that localize to the nSBs (Weighardt *et al.*, 1999, Denegri *et al.*, 2001). As the ratio between splicing factors determines the choice of splicing site, the nSBs are thought to induce alternative splicing upon stress (Jolly and Lakhotia, 2006). However, the nSBs may have multiple functions. The sat III transcripts could be involved in genomic silencing by incorporation into the RNAi system (Biamonti, 2004). Possibly, the sat III transcripts may play a role in the regulation of gene expression. In *Drosophila*, noncoding RNAs (ncRNAs) can activate transcription (Sanchez-Elsner *et al.*, 2006). Interestingly, this regulation is mediated via binding of Ash1 to the ncRNA molecules, which bind to the same sequences from which they are transcribed, a feature they share with the sat III transcripts in the nSBs. Moreover, it has been proposed that the expression of human and chick coding mRNAs containing α -like sequences in their UTRs is controlled by small and developmentally expressed ncRNAs derived from α -

satellite DNA (Li *et al.*, 2003). In human genes, segments of sat III DNA have been detected in the flanking regions and introns (Borstnik *et al.*, 1994). It is tempting to speculate that an analogous control system of gene expression, involving the sat III transcripts and regulated by HSF1 and HSF2 in response to distinct stimuli, is employed in humans.

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References

Ahn, S. G., Liu, P. C., Klyachko, K., Morimoto, R. I. and Thiele, D. J. (2001) The loop domain of heat shock transcription factor 1 dictates DNA-binding specificity and responses to heat stress. *Genes Dev.* *15*, 2134-2145.

Åkerfelt, M., Henriksson, E., Laiho, A., Vihervaara, A., Rautoma, K., Kotaja, N. and Sistonen, L. (2008). Promoter ChIP-chip analysis in mouse testis reveals Y chromosome occupancy by HSF2. *Proc. Natl. Acad. Sci. USA.* *105*, 11224-11229.

Alastalo, T-P., Hellesuo, M., Sandqvist, A., Hietakangas, V., Kallio, M. and Sistonen, L. (2003). Formation of nuclear stress granules involves HSF2 and coincides with the nucleolar localization of Hsp70. *J. Cell Sci.* *116*, 3557-3570.

Alastalo, T-P., Lönnström, M., Leppä, S., Kaarniranta, K., Peltö-Huikko, M., Sistonen, L. and Parvinen, M. (1998). Stage-specific expression and cellular localization of the heat shock factor 2 isoforms in the rat seminiferous epithelium. *Exp. Cell Res.* *240*, 16-27.

Anckar, J. and Sistonen, L. (2007). Heat shock factor 1 as a coordinator of stress and developmental pathways. *Adv. Exp. Med. Biol.* *594*, 78-88.

Balch, W. E., Morimoto, R. I., Dillin, A. and Kelly, J. W. (2008). Adapting proteostasis for disease intervention. *Science.* *319*, 916-919.

Barton, G. J. (1993). ALSCRIPT: a tool to format multiple sequence alignments. *Protein Eng.* *6*, 37-40.

Batchelor, A. H., Piper, D. E., de la Brousse, F. C., McKnight, S. L. and Wolberger, C. (1998). The structure of GABPalpha/beta: an ETS domain- ankyrin repeat heterodimer bound to DNA. *Science*. 279, 1037-1041.

Biamonti, G. (2004). Nuclear stress bodies: a heterochromatin affair? *Nat. Rev. Mol. Cell Biol.* 5, 493-498.

Borstnik, B., Pumpernik, D., Lukman, D., Ugarkovic, D. and Plohl, M. (1994). Tandemly repeated pentanucleotides in DNA sequences of eucaryotes. *Nucleic Acids Res.* 22, 3412-3417.

Bukau, B., Weissman, J. and Horwich, A. (2006). Molecular chaperones and protein quality control. *Cell*. 125, 443-451.

Chang, Y., *et al* (2006). Role of heat-shock factor 2 in cerebral cortex formation and as a regulator of p35 expression. *Genes Dev.* 20, 836-847.

Chiodi, I., Corioni, M., Giordano, M., Valgardsdottir, R., Ghigna, C., Cobianchi, F., Xu, R. M., Riva, S. and Biamonti, G. (2004). RNA recognition motif 2 directs the recruitment of SF2/ASF to nuclear stress bodies. *Nucleic Acids Res.* 32, 4127-4136.

Creighton, E. T. (1993) *Proteins: Structures and Molecular Properties*, W.H Freeman and Company.

Dai, C., Whitesell, L., Rogers, A. B. and Lindquist, S. (2007). Heat shock factor 1 is a powerful multifaceted modifier of carcinogenesis. *Cell*. 130, 1005-1018.

Denegri, M., Chiodi, I., Corioni, M., Cobianchi, F., Riva, S. and Biamonti, G. (2001). Stress-induced nuclear bodies are sites of accumulation of pre-mRNA processing factors. *Mol. Biol. Cell.* *12*, 3502-3514.

Fiorenza, M. T., Farkas, T., Dissing, M., Kolding, D. and Zimarino, V. (1995). Complex expression of murine heat shock transcription factors. *Nucleic Acids Res.* *23*, 467-474.

Frank, S., Schulthess, T., Landwehr, R., Lustig, A., Mini, T., Jenö, P., Engel, J. and Kammerer, R. A. (2002). Characterization of the matrilin coiled-coil domains reveals seven novel isoforms. *J. Biol. Chem.* *277*, 19071-19079.

Holmberg, C. I., Illman, S. A., Kallio, M., Mikhailov, A. and Sistonen, L. (2000). Formation of nuclear HSF1 granules varies depending on stress stimuli. *Cell Stress Chap.* *5*, 219-228.

Hsu, A. L., Murphy, C. T. and Kenyon, C. (2003). Regulation of aging and age-related disease by DAF-16 and heat-shock factor. *Science.* *300*, 1142-1145.

Johnson, M. S. and Overington, J. P. (1993). A structural basis for sequence comparisons. An evaluation of scoring methodologies. *J. Mol. Biol.* *233*, 716-738.

Jolly, C., Konecny, L., Grady, D. L., Kutsikova, Y. A., Cotto, J. J., Morimoto, R. I. and Vourc'h, C. (2002). In vivo binding of active heat shock transcription factor 1 to human chromosome 9 heterochromatin during stress. *J. Cell Biol.* *156*, 775-781.

Jolly, C. and Lakhota, S. C. (2006). Human sat III and Drosophila hsr omega transcripts: a common paradigm for regulation of nuclear RNA processing in stressed cells. *Nucleic Acids Res.* *34*, 5508-5514.

Jolly, C., Metz, A., Govin, J., Vigneron, M., Turner, B. M., Khochbin, S. and Vourc'h, C. (2004). Stress-induced transcription of satellite III repeats. *J. Cell Biol.* *164*, 25-33.

Jolly, C., Morimoto, R., Robert-Nicoud, M. and Vourc'h, C. (1997). HSF1 transcription factor concentrates in nuclear foci during heat shock: relationship with transcription sites. *J. Cell Sci.* *110*, 2935.

Kallio, M., *et al* (2002). Brain abnormalities, defective meiotic chromosome synapsis and female subfertility in HSF2 null mice. *EMBO J.* *21*, 2591-2601.

Kroeger, P. E., Sarge, K. D. and Morimoto, R. I. (1993). Mouse heat shock transcription factors 1 and 2 prefer a trimeric binding site but interact differently with the HSP70 heat shock element. *Mol. Cell. Biol.* *13*, 3370-3383.

Laberge, M. (1998). Intrinsic protein electric fields: basic non-covalent interactions and relationship to protein-induced Stark effects. *Biochim. Biophys. Acta.* *1386*, 305-330.

Lehtonen, J. V., *et al* (2004). BODIL: a molecular modeling environment for structure-function analysis and drug design. *J. Comput. Aided Mol. Des.* *18*, 401-419.

Li, Y. X. and Kirby, M. L. (2003). Coordinated and conserved expression of alphoid repeat and alphoid repeat-tagged coding sequences. *Dev. Dyn.* *228*, 72-81.

Littlefield, O. and Nelson, H. C. M. (1999). A new use for the 'wing' of the 'winged' helix-turn-helix motif in the HSF-DNA cocrystal. *Nat. Struct. Biol.* *6*, 464-470.

Loison, F., Debure, L., Nizard, P., Le Goff, P., Michel, D. and Le Drean, Y. (2006). Up-regulation of the clusterin gene after proteotoxic stress. Implication of HSF1/HSF2 heterocomplexes. *Biochem. J.* *395*, 223-231.

Metz, A., Soret, J., Vourc'h, C., Tazi, J. and Jolly, C. (2004). A key role for stress-induced satellite III transcripts in the relocalization of splicing factors into nuclear stress granules. *J. Cell Sci.* *117*, 4551-4558.

Min, J. N., Han, M. Y., Lee, S. S., Kim, K. J. and Park, Y. M. (2000). Regulation of rat heat shock factor 2 expression during the early organogenic phase of embryogenesis. *Biochim. Biophys. Acta.* *1494*, 256-262.

Morimoto, R. I. (2008). Proteotoxic stress and inducible chaperone networks in neurodegenerative disease and aging. *Genes Dev.* *22*, 1427-1438.

Morley, J. F. and Morimoto, R. I. (2004). Regulation of longevity in *Caenorhabditis elegans* by heat shock factor and molecular chaperones. *Mol. Biol. Cell.* *15*, 657-664.

Murphy, S. P., Gorzowski, J. J., Sarge, K. D. and Phillips, B. (1994). Characterization of constitutive HSF2 DNA-binding activity in mouse embryonal carcinoma cells. *Mol. Cell. Biol.* *14*, 5309-5317.

Östling, P., Björk, J. K., Roos-Mattjus, P., Mezger, V. and Sistonen, L. (2007). Heat shock factor 2 (HSF2) contributes to inducible expression of hsp genes through interplay with HSF1. *J. Biol. Chem.* *282*, 7077-7086.

Phillips, B., Abravaya, K. and Morimoto, R. I. (1991). Analysis of the specificity and mechanism of transcriptional activation of the human hsp70 gene during infection by DNA viruses. *J. Virol.* *65*, 5680-5692.

Pirkkala, L., Nykänen, P. and Sistonen, L. (2001). Roles of the heat shock transcription factors in regulation of the heat shock response and beyond. *FASEB J.* *15*, 1118-1131.

Rallu, M., Loones, M., Lallemand, Y., Morimoto, R., Morange, M. and Mezger, V. (1997). Function and regulation of heat shock factor 2 during mouse embryogenesis. *Proc. Natl. Acad. Sci. U.S.A.* *94*, 2392-2397.

Rizzi, N., Denegri, M., Chiodi, I., Corioni, M., Valgardsdottir, R., Cobianchi, F., Riva, S. and Biamonti, G. (2004). Transcriptional activation of a constitutive heterochromatic domain of the human genome in response to heat shock. *Mol. Biol. Cell.* *15*, 543-551.

Rossi, A., Ciafre, S., Balsamo, M., Pierimarchi, P. and Santoro, M. G. (2006). Targeting the heat shock factor 1 by RNA interference: a potent tool to enhance hyperthermochemotherapy efficacy in cervical cancer. *Cancer Res.* *66*, 7678-7685.

Sali, A. and Blundell, T. L. (1993). Comparative protein modelling by satisfaction of spatial restraints. *J. Mol. Biol.* *234*, 779-815.

Sanchez-Elsner, T., Gou, D., Kremmer, E. and Sauer, F. (2006). Noncoding RNAs of trithorax response elements recruit Drosophila Ash1 to Ultrabithorax. *Science*. *311*, 1118-1123.

Sarge, K. D., Murphy, S. P. and Morimoto, R. I. (1993). Activation of heat shock gene transcription by heat shock factor 1 involves oligomerization, acquisition of DNA-binding activity, and nuclear localization and can occur in the absence of stress. *Mol. Cell. Biol.* *13*, 1392-1407.

Sarge, K. D., Park-Sarge, O-K., Kirby, J. D., Mayo K. E. and Morimoto, R. I. (1994). Expression of heat shock factor 2 in mouse testis: potential role as a regulator of heat-shock protein gene expression during spermatogenesis. *Biol. Reprod.* *50*, 1334-13343.

Shu, W., Ji, H. and Lu, M. (1999). Trimerization specificity in HIV-1 gp41: analysis with a GCN4 leucine zipper model. *Biochemistry*. *38*, 5378-5385.

Shu, W., Liu, J., Ji, H. and Lu, M. (2000). Core structure of the outer membrane lipoprotein from Escherichia coli at 1.9 Å resolution. *J. Mol. Biol.* *299*, 1101-1112.

Shumaker, D. K., *et al* (2006). Mutant nuclear lamin A leads to progressive alterations of epigenetic control in premature aging. *Proc. Natl. Acad. Sci. U.S.A.* *103*, 8703-8708.

Valgardsdottir, R., Chiodi, I., Giordano, M., Cobianchi, F., Riva, S. and Biamonti, G. (2005). Structural and functional characterization of noncoding repetitive RNAs transcribed in stressed human cells. *Mol. Biol. Cell.* *16*, 2597-2604.

Valgardsdottir, R., Chiodi, I., Giordano, M., Rossi, A., Bazzini, S., Ghigna, C., Riva, S. and Biamonti, G. (2008). Transcription of Satellite III non-coding RNAs is a general stress response in human cells. *Nucleic Acids Res.* *36*, 423-434.

Wang, G., Zhang, J., Moskophidis, D. and Mivechi, N. F. (2003). Targeted disruption of the heat shock transcription factor (hsf)-2 gene results in increased embryonic lethality, neuronal defects, and reduced spermatogenesis. *Genesis.* *36*, 48-61.

Weighardt, F., Cobianchi, F., Cartegni, L., Chiodi, I., Villa, A., Riva, S. and Biamonti, G. (1999). A novel hnRNP protein (HAP/SAF-B) enters a subset of hnRNP complexes and relocates in nuclear granules in response to heat shock. *J. Cell Sci.* *112*, 1465-1476.

Xiao, H., Perisic, O., Lis, J. T. (1991). Cooperative binding of *Drosophila* heat shock factor to arrays of a conserved 5 bp unit. *Cell.* *64*, 585-593.

Xiao, X., Zuo, X., Davis, A. A., McMillan, D. R., Curry, B. B., Richardson, J. A. and Benjamin, I. J. (1999). HSF1 is required for extra-embryonic development, postnatal growth and protection during inflammatory responses in mice. *EMBO J.* *18*, 5943-5952.

Figure legends

Figure 1. Stress-induced localization of HSF2 into the nSBs is HSF1 dependent. (A) HSF1 and HSF2 binding to sat III in untreated (C) and 1 h heat-shocked (HS) K562 cells was analyzed with ChIP. *β-actin* was used as a control promoter. Input represents 1% of the total material and a nonspecific antibody (NS) was used as a negative control. The ChIP assay on sat III was performed on four biological samples. (B) HSF1 was downregulated in HeLa cells (HSF1 RNAi) and the nSB formation followed by staining of HSF1 (green) and HSF2 (red) (upper panel). As a control, a scrambled cell line (Scr.) was used. Note that the settings used for image acquisition of HSF1 and HSF2 in Scr. cells were reused for HSF1 RNAi cells. For staining, polyclonal and monoclonal antibodies were used against HSF1 and HSF2, respectively. The nucleus is shown in blue by staining of DNA with DAPI. Heat shock and control is indicated by HS and C, respectively. For additional time points, see Suppl. Figure 2A. (C) Western blot analysis of HSF1 and HSF2 in the stable Scr. (-) and HSF1-RNAi (+) cell lines. The retarded mobility of HSF1 in heat-shocked (HS) samples is due to hyperphosphorylation (Sarge *et al.*, 1993). Hsc70 serves as a loading control. (D) Real-time RT-PCR analysis of sat III transcription in Scr. (-) and HSF1-RNAi (+) cells. The results are shown as fold induction upon 30 min and 1 h of heat shock (HS). Fold induction was calculated by comparing the induction in HSF1 RNAi samples to the induction in scrambled samples, which were arbitrarily set to 1. The data represent three biological samples, and relative quantities of sat III RNA were normalized to GAPDH. Error bars indicate standard deviation (SD). (E) Western blot analysis of HSF1 downregulation in HEK293T cells. HS indicates a 1 h heat shock.

Figure 2. HSF1 and HSF2 interact as DNA-bound heterotrimers. (A) Structural model of a heterotrimer formed by the HR-A/B domains of two HSF1 molecules (green) and one HSF2 molecule (beige). The conserved amino acids found at positions a and d in the heptad repeat (Suppl. Figure 3) are shown as spheres. (B) Surface representation of the structural model of an HSF1-HSF2-HSF1 heterotrimer and an HSF1 homotrimer bound to DNA. HSF2 is colored beige and the different HSF1 molecules are colored in different shades of green. The HSF trimers are separated on DNA by two nucleotides as in a consensus HSE. The height and width of the complex as well as the distance between the two coiled coils are indicated. (C) Schematic presentation of the HSF1 and HSF2 C-terminal deletion constructs used for FRET. The position of amino acids are shown, and the DNA-binding domain (DBD) and trimerization domains HR-A/B are indicated. The deleted C-terminal regions are illustrated by dashed lines. (D) Interaction between HSF1 and HSF2 on DNA in the nSBs was detected with FLIM-FRET on live cells. HeLa cells were transfected with C-terminal HSF1 and HSF2 deletions fused to CFP and YFP, and the fluorescence lifetime of the donor (HSF1-CFP) was measured before or after photobleaching of the acceptor (HSF2-YFP). The fluorescence lifetime after photobleaching of the acceptor is indicated by a color scale bar. (E) A mean FLIM-FRET efficiency was calculated for the FRET pairs HSF1-CFP - HSF1-YFP and HSF1-CFP - HSF2-YFP. The data for each experimental condition represents measurements from at least five cells, and the SD is indicated.

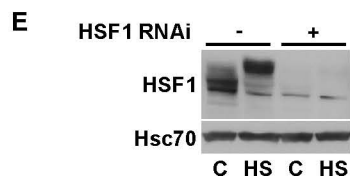
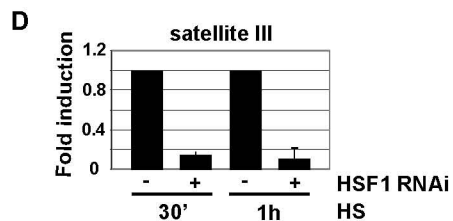
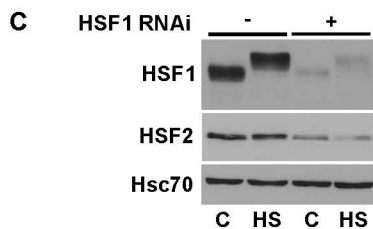
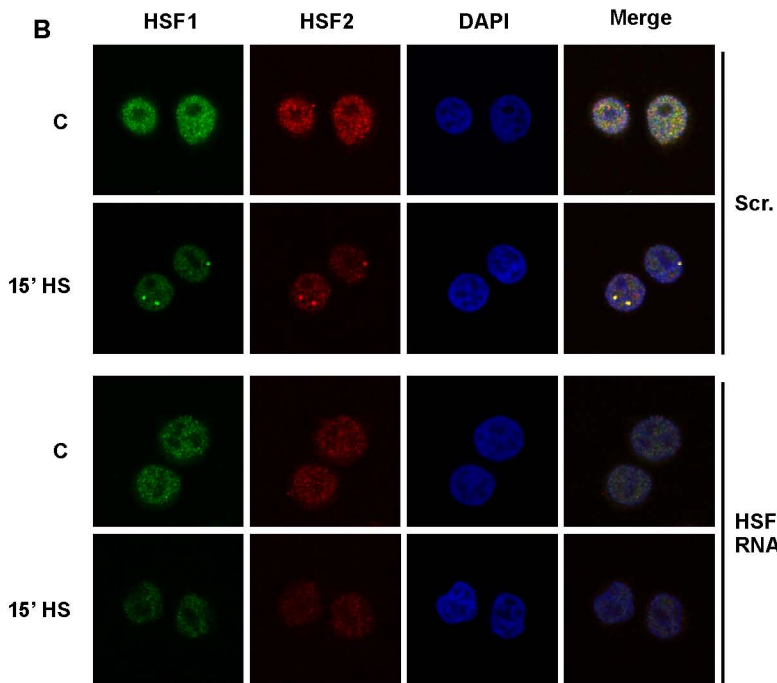
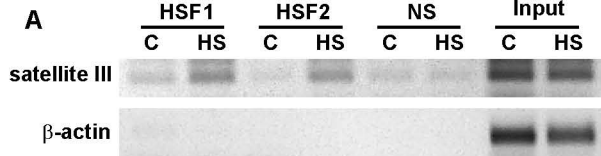
Figure 3. Stress-induced HSF activity is regulated through HSF1-HSF2 heterotrimerization. (A) Real-time RT-PCR was used to assess the impact of HSF2 downregulation on the transcriptional activity during heat shock (HS) in the nSBs. The fold induction was calculated by comparing the induction in HSF2 RNAi (+) samples to the induction in Scr. (-) samples, which were arbitrarily set to 1. The data represent three biological samples, and relative quantities of sat III RNA were normalized to GAPDH. Error bars indicate SD. (B) Western blot analysis of Scr. (-) and HSF2 RNAi (+) HEK293T cells. HS indicates a 1 h heat shock. (C) The localization of HSF1 (green) and HSF2 (red) to nSBs in Scrambled (Scr.) or HSF2-downregulating cells (HSF2 RNAi) was followed by staining. The settings used for acquiring images of HSF1 and HSF2 in Scr. cells were reused for HSF2 RNAi cells. For staining, polyclonal and monoclonal antibodies were used against HSF1 and HSF2, respectively. The nucleus is shown in blue by staining of DNA with DAPI. Heat shock and control is indicated by HS and C, respectively.

Figure 4. Elevated HSF2 expression activates transcription and induces HSF1-HSF2 heterotrimerization. (A) Real-time RT-PCR analysis of sat III transcription upon HSF2 overexpression (+) in untreated HeLa cells. To exclude the possibility of unspecific activation of the heat shock response, GFP was overexpressed as a control (-). (B) *hsp70* transcription upon HSF2 overexpression (+) was measured in untreated HeLa cells by real-time RT-PCR. GFP was overexpressed as a control (-). In panels A and B, fold induction was calculated by comparing transcription in the HSF2-overexpressing cells to the control cells, in which transcription was arbitrarily set to 1. The data represent three

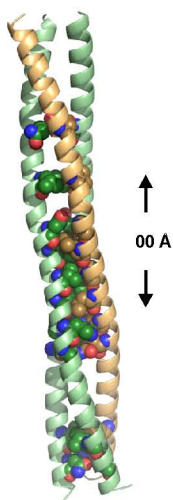
(panel A) and five (panel B) biological samples, and the relative quantities of RNA were normalized to GAPDH. Error bars indicate SD. (C) Western blot analysis of HSF1 (upper panel) and HSF2 (lower panel) in control (-) and HSF2-overexpressing (+) HeLa cells. (D) HSF2 was overexpressed in HeLa cells and the localization of HSF1 (green) and HSF2 (red) was monitored in the absence of stress. The arrow marks a cell overexpressing HSF2. (E) HSF1 and HSF2 localization in untreated HSF2-overexpressing HeLa cells. Moderate and robust HSF2 expression is indicated by a thick and thin arrow, respectively. In panels D and E, monoclonal anti-myc and polyclonal anti-HSF1 antibodies were used for staining. The nucleus is shown in blue by staining of DNA with DAPI. (F) Co-immunoprecipitation of endogenous HSF1 and HSF2 in mouse wild-type testis (HSF2 WT). To show interaction during spermatogenesis, antibodies against HSF1 and HSF2 were used for immunoprecipitation. In HSF2 knockout testis (HSF2 KO), no interaction could be detected. As a negative control, anti-Flag antibodies were used. The asterisk indicates small molecular weight isoforms of HSF2, as described in Alastalo *et al.*, 1998.

Figure 5. Schematic illustration of HSF1-HSF2 heterotrimerization as a mechanism integrating HSF activity. HSF1 and HSF2 are indicated as 1 and 2, respectively. Upon stress, HSF1 is activated, leading to formation of HSF1-HSF2 heterotrimers. Stress-induced HSF activity is regulated through HSF1-HSF2 heterotrimerization, a mechanism that probably provides also temporal regulation as heat shock diminishes HSF2 levels, thereby restricting heterotrimerization through limited availability of HSF2. During development, HSF2 levels are increased at certain stages and in a tissue-specific manner,

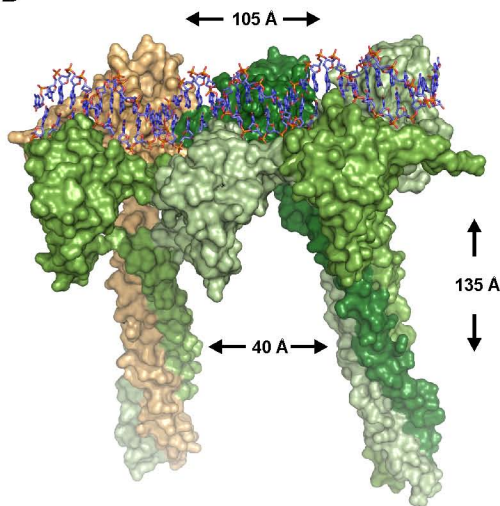
leading to activation of HSF2. Elevated HSF2 expression in turn induces HSF1-HSF2 heterotrimerization, illustrating the integrating role for HSF1-HSF2 heterotrimerization in response to distinct stimuli.



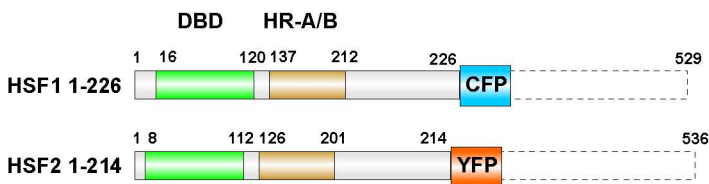
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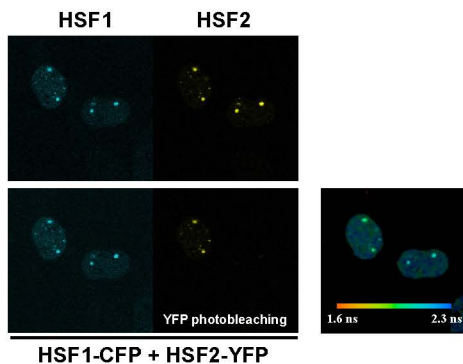
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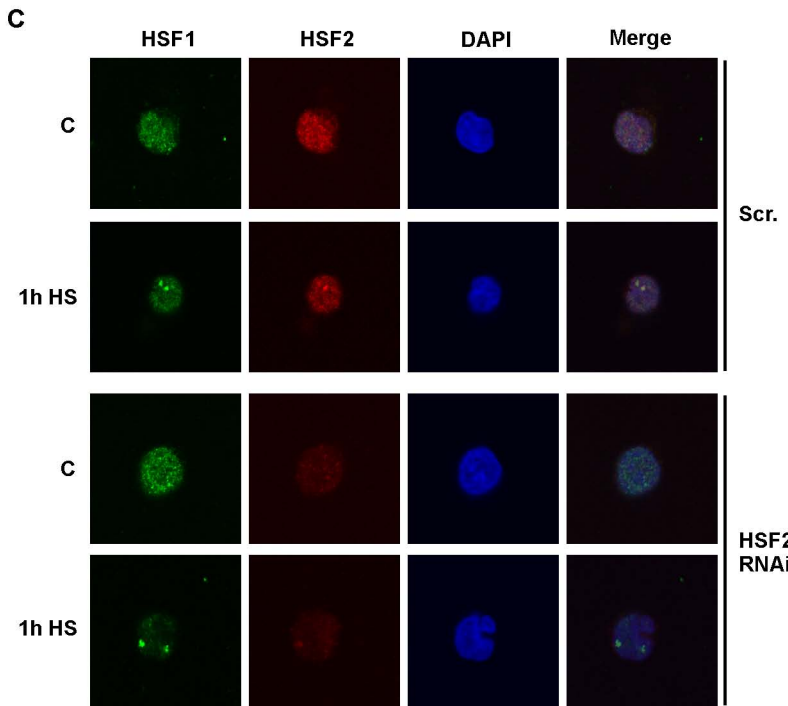
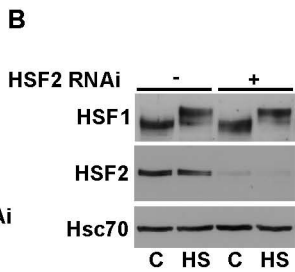
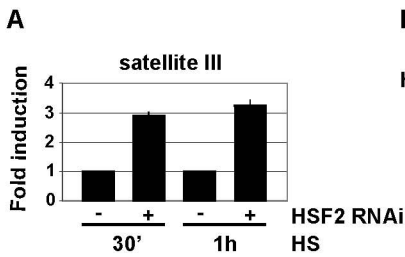


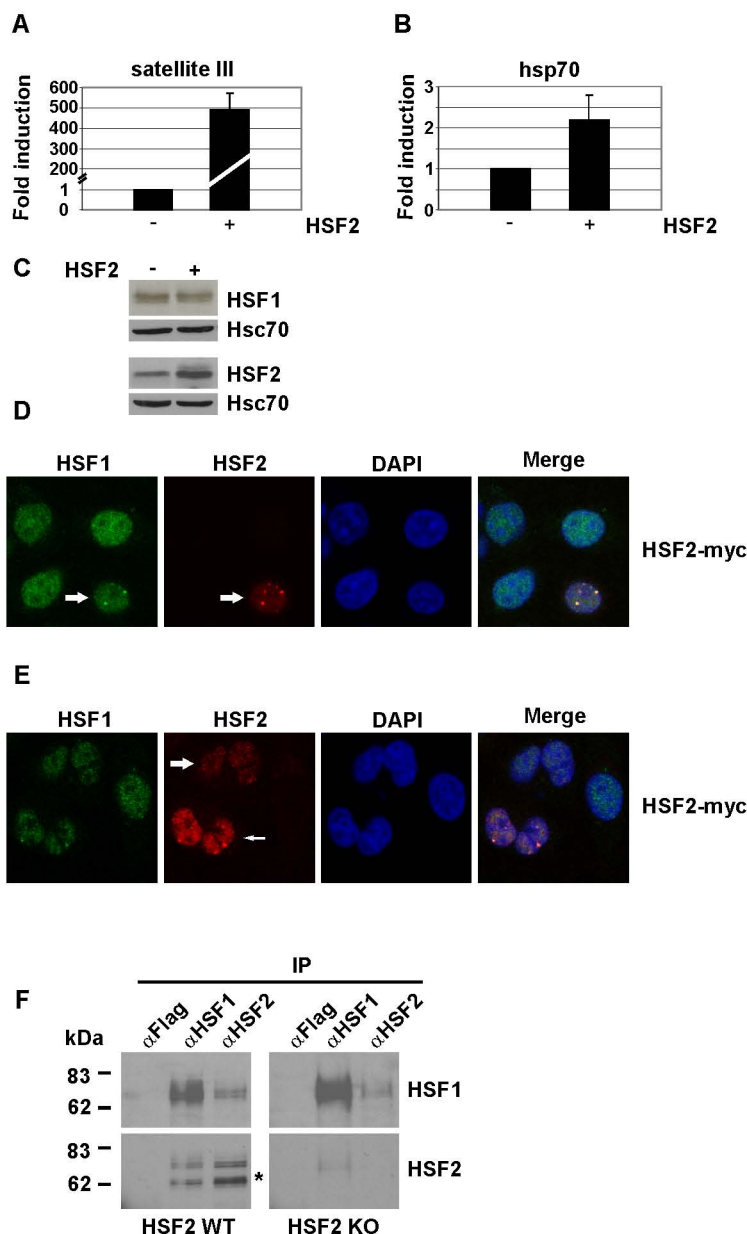
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E

	Lifetime before photobleaching	Lifetime after photobleaching	FRET efficiency
HSF1-HSF1	1.82 ± 0.05 ns	2.12 ± 0.06 ns	14%
HSF1-HSF2	1.94 ± 0.06 ns	2.15 ± 0.05 ns	10%





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Figure 5

