

# Toxicity Assessment of Titanium (IV) Oxide Nanoparticles Using *Daphnia magna* (Water Flea)

Seung Hyuck Bang<sup>1</sup>, Thai-Hoang Le<sup>1</sup>, Sung Kyu Lee<sup>2</sup>, Pil Kim<sup>3</sup>, Jong Soo Kim<sup>1</sup>, Jiho Min<sup>1,3</sup>

<sup>1</sup>Department of Bioprocess Engineering, Chonbuk National University, Jeonju; <sup>2</sup>Korea Institute of Technology, Daejeon;

<sup>3</sup>Division of Chemical Engineering, Chonbuk National University, Jeonju, Korea

**Objectives:** Titanium dioxide (TiO<sub>2</sub>), a common nanoparticle widely used in industrial production, is one of nano-sized materials. The purpose of this study was to determine the acute and chronic toxicity of TiO<sub>2</sub> using different size and various concentrations on *Daphnia magna*.

**Methods:** In the acute toxicity test, four concentrations (0, 0.5, 4, and 8 mM) for TiO<sub>2</sub> with 250 or 500 nm and five concentrations (0, 0.25, 0.5, 0.75, and 1 mM) for TiO<sub>2</sub> with 21 nm were selected to analyze the toxic effect to three groups of ten daphnia neonates over 96 hours. In addition, to better understand their toxicity, chronic toxicity was examined over 21 days using 0, 1, and 10 mM for each type of TiO<sub>2</sub>.

**Results:** Our results showed that all organisms died before the reproduction time at a concentration of 10 mM of TiO<sub>2</sub>. In addition, the exposure of anatase (21 nm) particles were more toxic to *D. magna*, comparing with that of anatase (250 nm) and rutile (500 nm) particles.

**Conclusions:** This study indicated that TiO<sub>2</sub> had adverse impacts on the survival, growth and reproduction of *D. magna* after the 21 days exposure. In addition, the number of test organisms that were able to reproduce neonates gradually were reduced as the size of TiO<sub>2</sub> tested was decreased.

**Key words:** Titanium dioxide, *Daphnia magna*, Size-dependent toxicity, Acute toxicity, Chronic toxicity

## INTRODUCTION

Nanomaterials, such as nanoparticles, nanotubes, fullerenes, nanowires, and quantum dots, have been developed in numerous research laboratories because they have a much greater surface area to mass ratio than micro-scale material, and they have been adopted in many new technologies [1]. The unique properties and applications of nanoparticles derive from numerous attributes; biomolecules, such as proteins, polynucleic acids, and nanoparticles made of metals and semiconductor materials, can serve as magnetic carriers and can provide fluorescence [2]. The use of nanoparticles for clinical applications presents a risk and has the potential for exposure to a toxic hazard [3].

Acute aquatic methods are a convenient analysis of nanoparticle toxicity, and the method uses various organisms including bacteria *Vibrio fischeri*, *Daphnia magna*, and *thamnocephalus platyurus* [4]. Further, to analyze the *in vitro* toxicity of nanoparticles and micro-scale materials, the interactions between particles and bacteria and their surface properties were investigated [5].

Titanium dioxide (TiO<sub>2</sub>) is widely used and is high insoluble and thermally stable. Since it has excellent optical

performance and electrical properties, TiO<sub>2</sub> has been used for the development solar cell energy [6]. In addition, it is used as a cosmetic to protect the skin from sunlight (e.g. a sunblock) and as a pigment to block water from penetrating a house [7]. Based on these characteristics of TiO<sub>2</sub>, we investigated toxicity via cellular response by the injection of various nanoparticles and solutions [8] in organisms (e.g. mice, *Daphnia magna*) [9].

In this study, we conducted an experiment to measure the toxicity of TiO<sub>2</sub> nanoparticles via a size- and concentration-dependent manner through *in vivo* acute and chronic toxicity testing using a *D. magna* according to international standards. The aim of this study was to analyze both the acute and chronic toxicity using *D. magna* and to meet these goals, we have used three different types of TiO<sub>2</sub> particles,

**Correspondence:** Jiho Min, Ph.D.

664-14 Deokjin-dong, 1Ga Deokjin-Gu, Jeonju 561-756, Korea

Tel: +82-63-270-2436, Fax: +82-63-270-2306

E-mail: jihomin@jbnu.ac.kr

Received: Nov 8, 2010, Accepted: Nov 30, 2010, Published Online: Mar 14, 2011

This article is available from: <http://e-eht.org/>

i.e. anatase sized 250 nm and 21 nm, and rutile sized 500 nm.

## MATERIALS AND METHODS

### I. *Daphnia magna* Culture

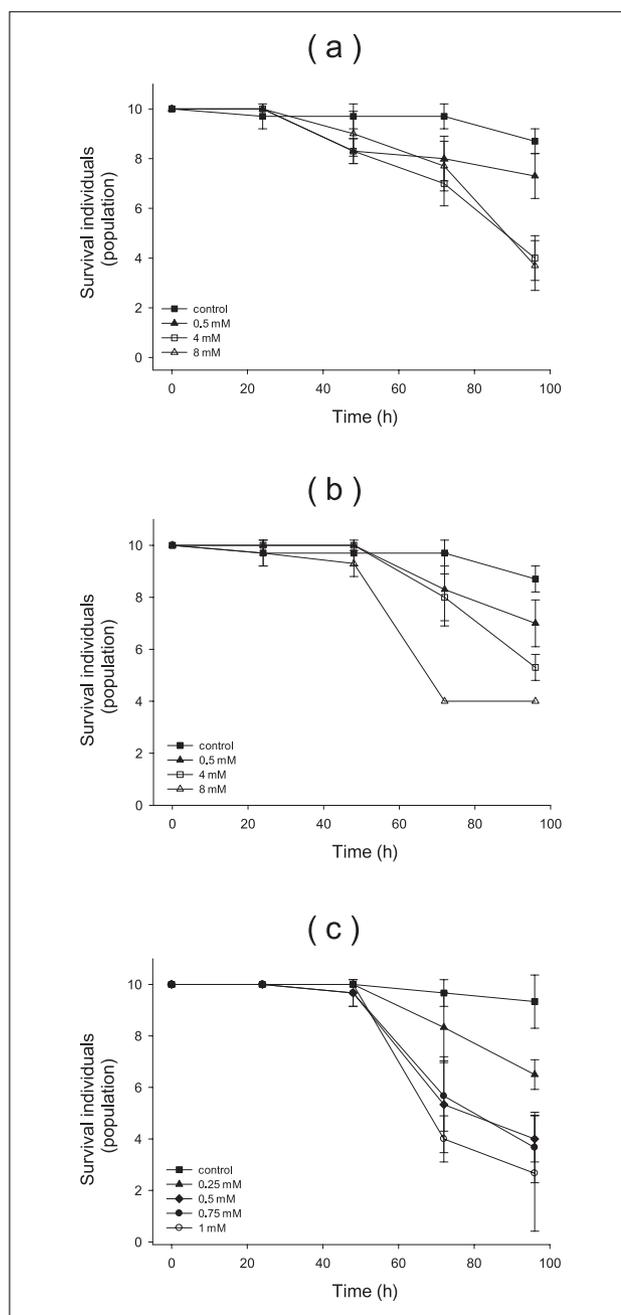
The *D. magna* used in this study was provided by the Korea Institute of Toxicology (Daejeon, Korea). The *D. magna* was cultured and treated as described in the US Environmental Protection Agency (US EPA) manual [10]. *D. magna* cultured and raised in a chamber were retained at  $20 \pm 1^\circ\text{C}$  in 2 L beakers containing 1.5 L of hard, reconstituted water ( $0.008 \text{ g L}^{-1}$  KCl,  $0.12 \text{ g L}^{-1}$   $\text{CaCO}_3$ ,  $0.12 \text{ g L}^{-1}$   $\text{MgSO}_4$ , and  $0.192 \text{ g L}^{-1}$   $\text{NaHCO}_3$  in distilled water filtered through a Minipore Milli-Q apparatus). For performance of the reproduction tests the pH of the medium was adjusted to  $8.2 \pm 0.2$ . Cultures were maintained under an 18 hours light: 6 hours dark photoperiod. The medium for the *D. magna* culture was changed three times per week with fresh HRW (hard reconstituted water) and fed with green algae (*Selenastrum capricornutum*) and YTC (a mixture of yeast, cerophyll, and Trout chow) purchased from Aquatic Biosystem Inc., (Colorado, US). *D. magna* culture was maintained in 2-liter glass beakers containing 1.6 L of cultured medium and 30-50 daphnids.

### II. Acute Toxicity Test

The acute toxicity test was performed according to the standard protocol for *D. magna* acute test [10]. Approximately 30 neonates aged less than 24 hours were separated in to three groups and exposed to titanium (iv) oxide (Sigma, US) with different sizes and several concentrations for 96 hours in a static test. All test concentrations were prepared using a volume of 50 mL in triplicate and maintained at  $20 \pm 1^\circ\text{C}$  during a 96-hours photoperiod of 16 hours light: 8 hours dark without any feeding. Test beakers were covered with vinyl wrap.

### III. Chronic Toxicity Test

The chronic toxicity test was performed according to the standard protocol through *D. magna* chronic toxicity testing [11]; neonates aged less than 24 hours at the start of the test were exposed to approximately three concentrations of each size of the nanoparticles for a period of 21 days. Neonates were fed two to three times per week, and the media and nanoparticle solution were changed, simultaneously. Survival and offspring were observed and monitored every day. The test beakers were maintained in a chamber at  $20 \pm 1^\circ\text{C}$  during a 96 hours photoperiod of 16 hours light: 8 hours dark.



**Figure 1.** Cumulative parent survival for each of the tested nanoparticles in the 96-hours acute toxicity test: (a) anatase (250 nm), (b) rutile (500 nm), and (c) anatase (21 nm) particles.

### IV. Data Analysis

All of the data were obtained from three independent samples carried out simultaneously for error analysis, and the results were shown along with the standard deviation and the correlation between the mortality under several experimental conditions. The data were analyzed using Sigma Plot (SPSS Inc., Chicago, IL, USA). A  $p$  value  $< 0.05$  was considered significant.

## RESULTS

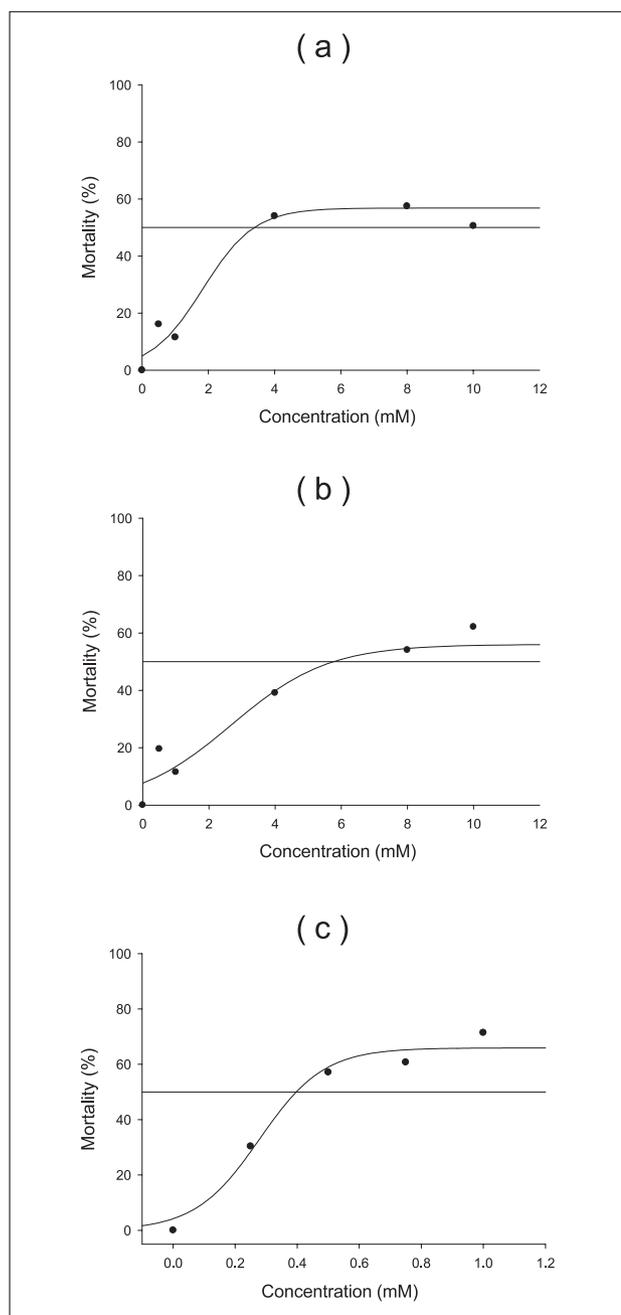
The acute toxicity of the three other sizes of titanium (iv) oxide was investigated using a 96-hours toxicity test with 10 neonates of *D. magna* (less than 24-hours old) in one beaker. The chronic toxicity test of the three other sizes of titanium (iv) oxide was performed for 21 days using 10 neonates with one neonate in each beaker. The experiments were conducted according to EPA guidelines. The pH of these toxicity tests was unchanged during the tests. All tests were performed in triplicate.

### I. Acute Toxicity of TiO<sub>2</sub> Particles

We performed the acute toxicity test for 96 hours with various concentrations and sizes of TiO<sub>2</sub> particles. The acute experiment determined the concentration range to be used in the survival experiments. Sodium phosphate buffer (pH 6, 0.1 M) was used as the control, and its mortality was shown in Figure 1. All sizes and concentrations of TiO<sub>2</sub> were not acutely toxic over 48 hours. After 96 hours, four neonates treated with anatase (250 nm) and rutile (500 nm) particles at 8 mM survived, but of those treated with anatase (21 nm) at 0.75 mM, four survived. From these results, we determined that the higher concentration led to a higher mortality (see all graphs). Furthermore, the smaller size led to a higher mortality (Figure 1c). Given these results, we found that the regression curves of the acute toxicity data, LC50 and LC20 were determined as the lethal concentrations of a chemical when the percentage of *D. magna* mortality was 50% and 20%, respectively. In this study, *D. magna* was exposed to two different LC values (LC20, LC50) of anatase (250 nm) 1.29 and 3.47 mM, respectively, for 96 hours to examine the effects of the anatase form (250 nm) of TiO<sub>2</sub> (Figure 2a). Likewise, characteristics of the rutile (500 nm) and anatase (21 nm) varieties were measured. The LC20 and 50 of the rutile (500 nm) and anatase type (21 nm) particles were 1.98, 5.94, 0.19, and 0.42 mM, respectively (Figure 2b, c). Based on the above results, the *D. magna* toxicity of the three types of TiO<sub>2</sub> particles decreased from anatase (21 nm) to anatase (250 nm) to rutile (500 nm) particles.

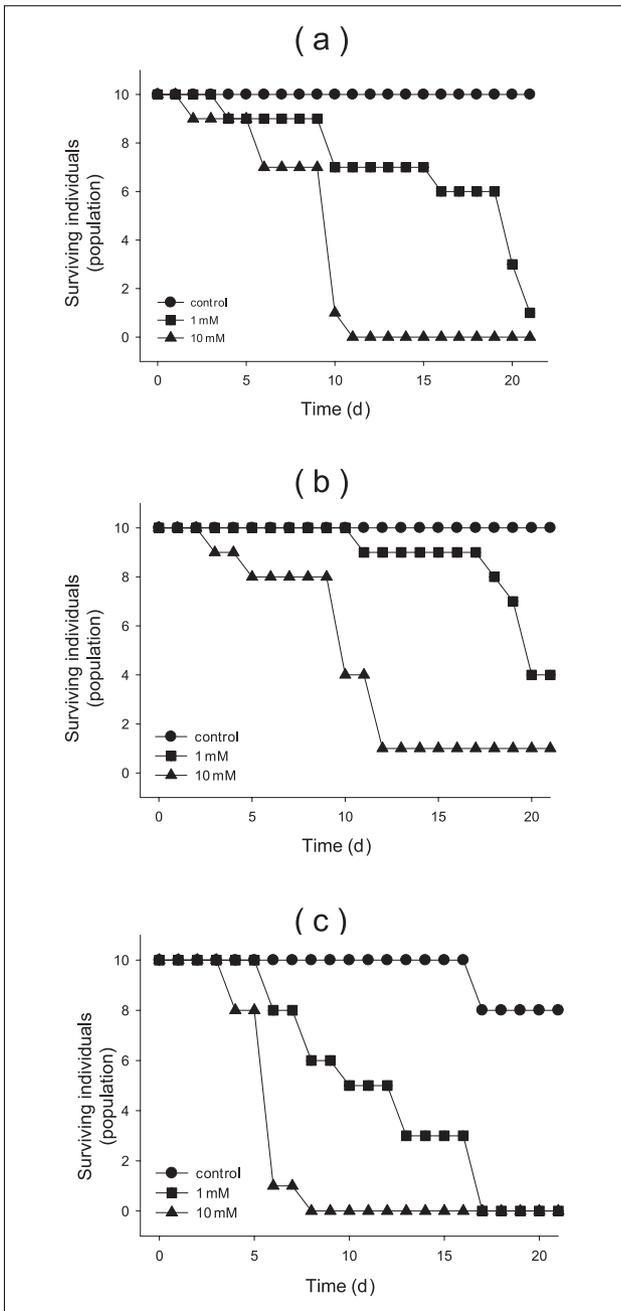
### II. Chronic Toxicity of TiO<sub>2</sub> Particles

In addition, the chronic toxicity over 21 days using various concentrations and sizes of TiO<sub>2</sub> particles determined in this study. Moreover, we used and compared the concentrations (0, 1, 10 mM) in all chronic toxicity tests. We measured the concentration- and size-dependent survival of neonates over 21 days. Survival of anatase (250 nm) and rutile (500 nm) were observed at approximately 10, 40% at 1 mM (Figure 3a, b). In contrast, the survival for 1 mM of anatase (21 nm) particles was not measured because there were killed after the cells were exposed to the solution for 16 days (Figure 3c).



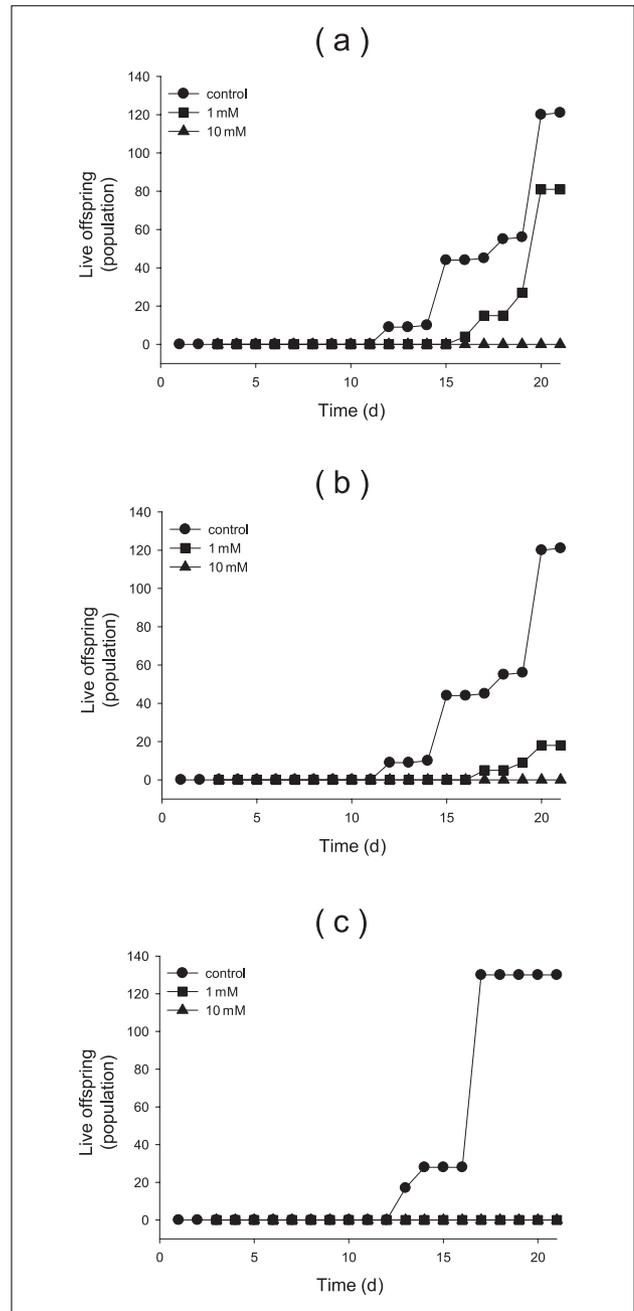
**Figure 2.** Effects of the two types of titanium (iv) oxide nanoparticles on *Daphnia magna* in the 96-hours acute toxicity test: (a) anatase (250 nm), (b) rutile (500 nm), and (c) anatase (21 nm) nanoparticles.

From these results, we determined that the toxic response of anatase (250 nm) and rutile (500 nm) progressed slowly, but the response of anatase (21 nm) particles progressed rapidly. There were more than 120 live offspring of the control after 21 days of testing (Figure 4a). That of the anatase (250 nm) and rutile (500 nm) particles with 1 mM was approximately 80 and 20%, respectively. However we did not find any neonates after exposure to anatase (21 nm) particles at 1 mM (Figure 4). Further, we found that no offspring were found at



**Figure 3.** Cumulative live offspring produced per female for each of the tested nanoparticulates after 21 days of exposure: (a) anatase (250 nm), (b) rutile (500 nm), and (c) anatase (21 nm) nanoparticulates.

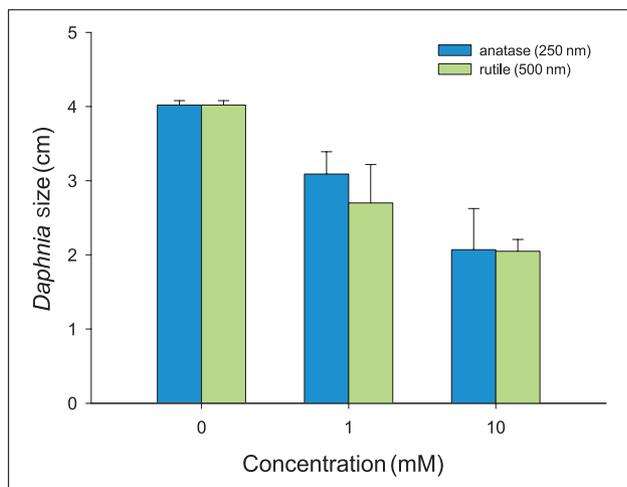
the 10 mM concentration of additional sizes of TiO<sub>2</sub> (Figure 4c). After finishing the chronic toxicity tests, we measured the size of neonates living until last day using microscope. Averagely, the size of neonates with 0, 1, and 10 mM anatase (250 nm) materials is 4, 3, and 2 centimeters, respectively. For rutile (500 nm) nanoparticulates, the sizes are 4, 2.5, and 2 centimeters, respectively (Figure 5). However, the size of anatase (21 nm) particles could not be measured because all neonates died.



**Figure 4.** Cumulative parent survival for each of the tested nanoparticles subjected to a chronic toxicity test for 21 days: (a) anatase (250 nm), (b) rutile (500 nm), and (c) anatase (21 nm) nanoparticulates.

## DISCUSSION

The *D. magna* showed a very sensitive response dependent upon the size and concentration of TiO<sub>2</sub> nanoparticles in both acute and chronic toxicity tests. In particular, the size difference of the nanoparticles demonstrated a greater response than the concentration. As shown in Figure 1, while the toxicity of anatase (250 nm)



**Figure 5.** Size comparison of daphnia exposed to two types of the titanium(IV) oxide nanoparticles during a chronic toxicity test for 21 days (■) anatase (250 nm) and (■) rutile (500 nm) nanoparticles.

and rutile (500 nm) particles demonstrated similarities, the smallest size indicated TiO<sub>2</sub> induced the greatest toxicity despite the low concentrations of the solution. In a study similar to ours, *D. magna* was used to confirm that toxicity is influenced by the size of TiO<sub>2</sub> nanoparticles [12]. Furthermore, we investigated the same concentration of anatase (250 nm) and rutile (50 nm) particles to compare the toxicity of three types of TiO<sub>2</sub>. However, no neonates survived the addition of anatase (21 nm) particles (data not shown). The result is likely a rapid absorption of the anatase (21 nm) particles in the neonate because of the small size [13]. Furthermore, oxidative DNA damage occurs in *D. magna* with TiO<sub>2</sub> exposure [14]. We suggest that the neonates have a higher potential for reaching deep into the lungs or damaging internal organs because of the size of TiO<sub>2</sub> particles.

## CONCLUSIONS

Our innovative approach based on a combination of traditional ecotoxicology methods allowed the clear differentiation of the toxic effects of nonmetal oxide nanoparticles. In addition, this is the first 96-hours evaluation of an acute toxicity test of the size dependence of TiO<sub>2</sub> using *D. magna*. All TiO<sub>2</sub> nanoparticles tested by *D. magna* were sensitive to concentration, and *D. magna* is widely used in aquatic risk assessment. On the basis of these acute toxicity test results, we conducted a chronic, *in vivo* toxicity test to confirm dependence on the TiO<sub>2</sub> particle size. Acute and chronic effects on *D. magna* were observed in this investigation.

These responses are due to adaptations, such as changes of state, which happen after exposure to toxicant nano-sized

materials. *D. magna* showed specific response patterns according to toxic effects caused by the size and concentration of TiO<sub>2</sub> nanoparticles. The behavior manners of *D. magna* exposing by different sized TiO<sub>2</sub> were considerably different. Therefore, the size effect of TiO<sub>2</sub> nanoparticles should be considered in hazard evaluations for the potential to impact aquatic life.

## ACKNOWLEDGEMENTS

This work was supported by Mid-career Researcher Program through NRF grant funded by the MEST (No. R01-2008-000-20773-0). The authors are grateful for their support.

## CONFLICT OF INTEREST

The authors have no conflict of interest to declare on this study.

## REFERENCES

- Aitken RJ, Chaudhry MQ, Boxall AB, Hull M. Manufacture and use of nanomaterials: current status in the UK and global trends. *Occup Med (Lond)* 2006; 56(5): 300-306.
- De M, Ghosh PS, Rotello VM. Applications of nanoparticles in biology. *Adv Mater* 2008; 20: 4225-4241.
- Wiesner MR, Lowry GV, Alvarez P, Dionysiou D, Biswas P. Assessing the risks of manufactured nanomaterials. *Environ Sci Technol* 2006; 40(14): 4336-4345.
- Heinlaan M, Ivask A, Blinova I, Dubourguier HC, Kahru A. Toxicity of nanosized and bulk ZnO, CuO and TiO<sub>2</sub> to bacteria *Vibrio fischeri* and crustaceans *Daphnia magna* and *Thamnocephalus platyurus*. *Chemosphere* 2008; 71(7): 1308-1316.
- Jiang W, Mashayekhi H, Xing B. Bacterial toxicity comparison between nano- and micro-scaled oxide particles. *Environ Pollut* 2009; 157(5): 1619-1625.
- Mor GK, Varghese OK, Paulose M, Shankar K, Grimes CA. A review on highly ordered, vertically oriented TiO<sub>2</sub> nanotube arrays: fabrication, material properties, and solar energy applications. *Sol Energy Mater Sol Cells* 2006; 90(14): 2011-2075.
- Jaroenworarluck A, Sunsaneeyametha W, Kosachan N, Stevens R. Characteristics of silica-coated TiO<sub>2</sub> and its UV absorption for sunscreen cosmetic applications. *Surf Interface Anal* 2006; 38(4): 473-477.
- Le TH, Lim ES, Lee SK, Choi YW, Kim YH, Min J. Effects of glyphosate and methidathion on the expression of the Dhb, Vtg, Arnt, CYP4 and CYP314 in *Daphnia magna*. *Chemosphere* 2010; 79(1): 67-71.
- Chen J, Dong X, Zhao J, Tang G. *In vivo* acute toxicity of titanium dioxide nanoparticles to mice after intraperitoneal injection. *J Appl Toxicol* 2009; 29(4): 330-337.

10. US EPA. Methods for measuring the acute toxicity of effluents and receiving waters to freshwater, marine organisms. [cited 2011 Feb 07]. Available from: <http://www.epa.gov/water-science/methods/wet/disk2/atx.pdf>.
11. US EPA. Ecological effects test guidelines (OPPTS 850.1300 Daphnid Chronic Toxicity test). [cited 2010 Dec 5]. Available from: [http://www.epa.gov/ocspp/pubs/frs/publications/OPPTS\\_Harmonized/850\\_Ecological\\_Effects\\_Test\\_Guidelines/Drafts/850-1300.pdf](http://www.epa.gov/ocspp/pubs/frs/publications/OPPTS_Harmonized/850_Ecological_Effects_Test_Guidelines/Drafts/850-1300.pdf).
12. Wiench K, Wohlleben W, Hisgen V, Radke K, Salinas E, Zok S, et al. Acute and chronic effects of nano-and non-nano-scale TiO<sub>2</sub> and ZnO particles on mobility and reproduction of the freshwater invertebrate *Daphnia magna*. *Chemosphere* 2009; 76(10): 1356-1365.
13. Kim KT, Klaine SJ, Cho J, Kim SH, Kim SD. Oxidative stress responses of *Daphnia magna* exposed to TiO<sub>2</sub> nanoparticles according to size fraction. *Sci Total Environ* 2010; 408(10): 2268-2272.
14. Karlsson HL, Gustafsson J, Cronholm P, Möller L. Size-dependent toxicity of metal oxide particles--a comparison between nano-and micrometer size. *Toxicol Lett* 2009; 188(2): 112-118.