

Role of Heparanase on Hepatic Uptake of Intestinal Derived Lipoprotein and Fatty Streak Formation in Mice

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Abstract

Background: Heparanase modulates the level of heparan sulfate proteoglycans (HSPGs) which have an important role in multiple cellular processes. Recent studies indicate that HSPGs have an important function in hepatic lipoprotein handling and processes involving removal of lipoprotein particles.

Principal Findings: To determine the effects of decreased HSPGs chain length on lipoprotein metabolism and atherosclerosis, transgenic mice over-expressing the human heparanase gene were studied. Hepatic lipid uptake in hpa-Tg mice were evaluated by giving transgenic mice oral fat loads and labeled retinol. Sections of aorta from mice over-expressing heparanase (hpa-Tg) and controls (C57/BL6) fed an atherogenic diet were examined for evidence of atherosclerosis. Heparanase over-expression results in reduced hepatic clearance of postprandial lipoproteins and higher levels of fasting and postprandial serum triglycerides. Heparanase over-expression also induces formation of fatty streaks in the aorta. The mean lesion cross-sectional area in heparanase over-expressing mice was almost 6 times higher when compared to control mice ($23,984 \mu\text{m}^2 \pm 5,922$ vs. $4,189 \mu\text{m}^2 \pm 1,130$, $p < 0.001$).

Conclusions: Over-expression of heparanase demonstrates the importance of HSPGs for the uptake of intestinal derived lipoproteins and its role in the formation of fatty streaks.

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Introduction

Heparan sulfate proteoglycans (HSPGs) are major components of the extracellular matrix and have a major role in signal transduction and modulating cell interactions with the microenvironment [1,2]. Recent studies indicate that HSPGs have an important function in hepatic lipoprotein handling and processes involving removal of lipoprotein particles [3,4,5].

The involvement of HSPGs in multiple physiologic processes has spurred interest in identifying factors that regulate cell surface heparan sulfate (HS) content. A key regulatory enzyme is heparanase, a HS degrading endoglycosidase produced by different cell types [6,7]. The function of the HSPG-heparanase system in modulating hepatic lipoprotein handling especially in the post prandial state has been previously studied. In the liver, infusion of heparinase (the bacterial equivalent of heparanase) into the portal vein lowers hepatic HSPG with an attendant reduction in triglyceride rich particle (TRP) clearance [8]. In diabetics, there is a reduced level of the HSPG assembly enzyme *N*-deacetylase/*N*-sulfotransferase-1 (Ndst1) that is associated with decreased postprandial clearance of triglyceride rich particles (TRPs) and hypertriglyceridemia [9]. Based on experiments with syndecan-1 deficient mice that exhibited delayed clearance of TRLs (Triglyceride Rich Lipoproteins), Stanford et al concluded that

syndecan-1 is the primary hepatic proteoglycan receptor mediating TRL clearance [10].

To study the effect of heparanase on lipid metabolism and atherosclerosis development *in vivo*, an experimental model in which heparanase is over-expressed was utilized. We have found that over-expression of heparanase induces inhibition of postprandial lipoprotein clearance and uptake by the liver and is associated with formation of fatty streaks.

Materials and Methods

Animals

The study was conducted according to protocols approved by the Animal Care Committee at the Hadassah - Hebrew University Medical Center. Homozygous transgenic mice over-expressing the human heparanase gene (hpa-Tg) were produced as previously described [11]. The transgenic mice were backcrossed to C57BL/6 mice for 7 generations and maintained as homozygotes for the transgene. C57BL/6 mice were used as controls. Mice were maintained in a SPF animal facility and fed *ad libitum* either with normal chow or an atherogenic diet (42% of calories from fat, 43% from carbohydrates, and 15% from protein, TD 88137, Harlan Teklad) for six months. Food consumption was measured and there were no changes in eating

behavior and body weight between pure bred hpa-Tg and control C57BL/6 mice.

Genotyping of transgenic mice

To ensure that the animals were homozygous for the human heparanase gene, DNA was isolated from the tail tip [11] and real time PCR was performed using human hpa specific PCR primers: F-5'-TACCTTCATTGCAACACTG-3' and R-5'-GTGACAT-TATGGAGGTT-3'. The PCR was performed using Syber universal PCR Master Mix (ABI, Warrington, UK). L19 cDNA was used as an internal standard.

Assessment of atherosclerosis in the aortic root

hpa-Tg and control C57BL/6 male mice were analyzed for atherosclerosis. Following perfusion of the heart, the upper section of the heart was placed in embedding medium (Tissue-Tek, Satura FineTek, Torrance, CA), frozen in methyl butane and stored at -80°C . Every other section (10 microns thick) throughout the aortic sinus (400 μm) was taken for atherosclerosis analysis. Frozen sections were stained with Oil red O and three representative sections from each mouse were analyzed by image analysis using ImagePro Plus software [12].

Analytic procedures

Following an overnight fast, retro-orbital blood samples were taken from mice for lipid profile evaluation [13]. Cholesterol and triglyceride distribution among lipoproteins fractions was determined by fast pressure liquid chromatography (FPLC) after loading 200 μl of pooled plasma from control and hpa-Tg fasting mice onto a superose 6HR 10/30 FPLC column (Piscataway, NJ) and 0.5 ml fractions were eluted with 0.15 M NaCl, 0.01 M Na_2HPO_4 , 0.1 mM EDTA and 0.02 mM NaN_3 . Cholesterol and triglycerides (TG) were determined using commercial kits (Roche/Hitachi, Mannheim, Germany and Thermo, Waltham, MA, respectively).

Lipoprotein lipase (LPL) activity

Mice were injected i.p. with 0.5 U/gr heparin. Post heparin plasma was withdrawn 10 minutes later, and LPL activity was determined as described [14]. LPL activity is given in mU/ml, where 1 mU represents 1 nmol of free fatty acid released from the triglyceride substrate per minute.

Fat and retinol load

An oral fat load test was performed with hpa-Tg and their controls. Mice were fasted overnight and then administered a bolus of 100 μl corn oil via a stomach tube. Triglyceride and cholesterol levels were determined in the plasma at specific time points as indicated in the text [15]. The same experiment was repeated with 100 μl corn oil containing 10 μCi of ^3H -retinol ([11,12- ^3H (N)], Perkin Elmer, Boston) and 3000 units of unlabeled retinol (Sigma-Aldrich) [16]. Radioactivity in the plasma was determined 1.5, 3 and 6 h following fat administration. Radioactivity distribution among lipoproteins fractions was determined by fast pressure liquid chromatography (FPLC) loading a superpose column (described above) with 200 μl of radiolabeled plasma from control and hpa-Tg mice 3 h after retinol administration.

Body distribution of radioactive retinol

Three hours after intragastric administration of bolus of 100 μl corn oil containing 10 μCi of ^3H -retinol and 3000 units of unlabeled retinol, mice were anesthetized, bled, and perfused with a normal saline solution. Fat extraction from liver and carcass samples was done with a solution of methanol:chloroform 1:1, and

radioactivity was measured in these samples and serum. Absorption of retinol from the intestine and its clearance from the circulation by liver and carcass were calculated.

Statistical analysis

Data is reported as mean \pm SEM. Comparisons between groups were made utilizing the Student's *t* test (two-tailed).

Results

Effect of heparanase over-expression on serum lipids

To determine the metabolic effects of increased heparanase expression, the lipoprotein profile in hpa-Tg mice fed a high fat diet was investigated. No significant difference was detected in total serum cholesterol levels between overnight fasted hpa-Tg mice (223 mg/dl \pm 30) and C57BL/6 mice (177 mg/dl \pm 18) although cholesterol levels in the transgenic mice tended to be higher (Table 1). However, there was a mild, but significant, elevation in TG levels in hpa-Tg mice compared to control mice (102 mg/dl \pm 11 vs. 75 mg/dl \pm 6, respectively; $p=0.04$) as shown in Table 1. FPLC analysis revealed that the AUC of TG in the transgenic mice was increased by 1.4 fold compared to control C57BL/6 mice. A 3.7 fold increased in AUC of VLDL cholesterol was observed in the hpa-Tg mice. As most of the plasma cholesterol in mice is located in the HDL fraction, the AUC of total cholesterol didn't vary significantly between the two groups. (Figure 1A, B). There was no significant difference in post heparin plasma lipoprotein lipase activity between control and hpa-Tg mice, (40 \pm 3 mU/ml in control vs. 54 \pm 9 mU/ml in hpa-Tg mice) indicating that the effect of heparanase over-expression on remnant lipoprotein metabolism is not due to reduced lipoprotein lipase activity.

Effect of heparanase over-expression on lipoprotein metabolism

The effect of elevated heparanase on post-prandial lipoprotein accumulation in the plasma was studied by oral administration of corn oil to fasted mice. As shown in Figure 2A, the differences in serum TG between hpa-Tg and control mice increased during the first 3 h following the fat load. TG levels were significantly higher at all time points in the hpa-Tg mice (Figure 2A; $p<0.005$ for all points), and these differences were maintained up until 6 h after the fat load. For example, TG levels increased from 120 mg/dl \pm 10 at baseline to 320 mg/dl \pm 50 at 1.5 h after fat administration. This is in comparison to control C57BL/6 mice in which there was only a mild elevation in plasma TG levels measured 1.5 h post-prandially (from 90 mg/dl \pm 4 to 160 mg/dl \pm 10). We have also determined plasma radioactivity 1.5, 3 and 6 h following retinol load. Plasma vitamin A (^3H -retinol) radioactivity was higher at all time points in the hpa-Tg compared to C57BL/6 mice (Figure 3A; $p<0.01$ for all points). FPLC analysis of plasma taken 3 h after retinol load revealed higher peaks of labeled VLDL and remnant particles in the hpa-Tg mice (Figure 3B).

The accumulation of TG in plasma of hpa-Tg mice suggests that over-expression of heparanase decreases remnant particle clearance by the liver. To test this hypothesis, ^3H -retinol was administered orally and the distribution of retinol was assayed in blood, liver and carcass of hpa-Tg and C567BL/6 mice. The intestinal absorption of labeled retinol was calculated by adding the radioactivity recovered in the plasma, liver and carcass (excluding other organs and the entire GI tract) and expressed as the percent of the total administered dose of radioactivity. Extraction of the administered label from liver, blood and carcass, 3 h following retinol load, revealed that the absorption was

Table 1. Serum lipid profile of hpa-Tg and control C57BL/6 mice.

	hpa-Tg (n = 15)	C57BL/6 (n = 15)	P value
Total cholesterol (mean±SE) mg/dl	223±30	177±18	ns
Triglycerides (mean±SE) mg/dl	102±11*	75±6	0.04

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approximately 12% of the total administered dose for both the control and hpa-Tg animals (Figure 4A). However, there were differences in blood and liver distribution of labeled retinol

between the two groups indicating that remnant particle clearance by the liver was reduced in hpa-Tg mice (Figure 4B). Fat extracted from liver of hpa-Tg mice contained 32% ±2 of the absorbed retinol, while fat extracted from liver of C57BL/6 mice contained 64% ±2 of the absorbed radioactivity ($p < 0.0001$, Figure 4B). The decrease in liver uptake of retinol was paralleled by an equivalent increase in the level of labeled retinol in blood of the hpa-Tg mice. The percent of the absorbed retinol found in blood was 41% ±1 in hpa-Tg mice compared to 9% ±1 in controls ($p < 0.0001$).

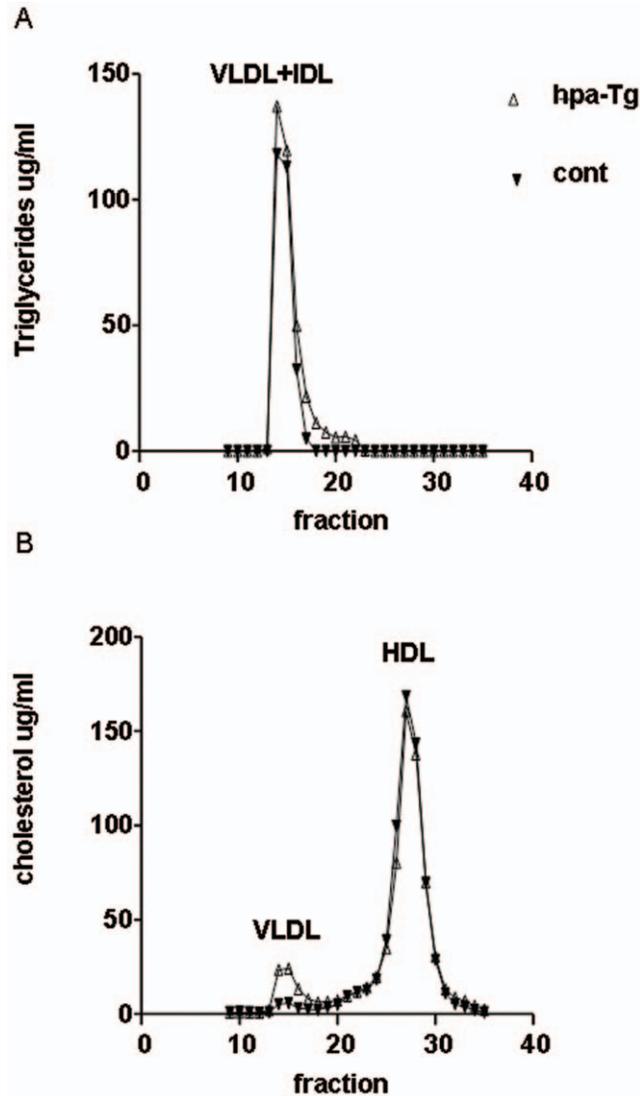


Figure 1. Distribution of triglyceride (A) and cholesterol (B) in the plasma of hpa-Tg (open triangles) and control C67BL/6 mice (closed triangles). Pooled plasma samples of 4 fasting mice from each group were applied to FPLC analysis. As indicated in the figure, triglyceride levels are increased in the non-HDL fractions (A), and cholesterol level is slightly increased in the VLDL+IDL fractions (B) of the hpa-Tg mice compared to controls. AUC ($\mu\text{g/ml} \times \text{fraction}$) of C57BL/6 triglyceride was 281 compared to 393 of hpa-Tg whereas the AUC of C57BL/6 total cholesterol was 662 compared to 705 of hpa-Tg mice. The AUC of VLDL-cholesterol peak was 20 compared to 74 of hpa-Tg mice. doi:10.1371/journal.pone.0018370.g001

Fatty streaks formation in heparanase over-expressing mice

The effect of over-expression of heparanase on the development of atherosclerotic lesions was studied in hpa-Tg and control C57BL/6 male mice fed a high fat diet for 6 months. Frozen sections of aortic root were stained with Oil-Red-O for lipid content and fatty streak formation. Figure 5A shows representative sections of aorta from hpa-Tg and control mice. Quantitative analysis of fatty streaks was performed by measuring the extent of Oil-red-O staining of cryosections from the aortic root. Mean lesion cross-sectional area in heparanase over-expressing mice was almost 6 times higher when compared to control mice ($23984 \mu\text{m}^2 \pm 5922$ vs. $4189 \mu\text{m}^2 \pm 1130$, $p < 0.001$; Figure 5B).

Discussion

By manipulating the expression of heparanase, the only HSPG degrading enzyme, it is possible to determine the effect of decreased intact HSPG content on both liver uptake of TRPs

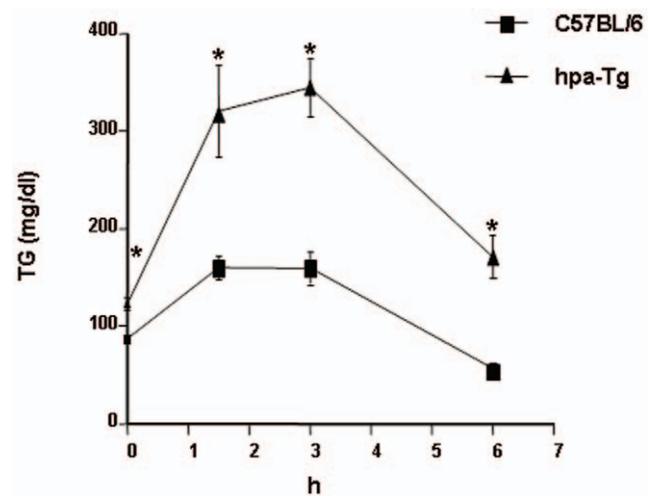


Figure 2. TG accumulation in plasma following oral administration. Mice were fasted overnight and then administered a bolus of 100 μl corn oil via a stomach tube. TG levels in serum of hpa-Tg mice (n = 6) were significantly higher compared to TG levels in C57BL/6 mice (n = 6) during the first 3 hrs ($p < 0.005$ for all measured time points). doi:10.1371/journal.pone.0018370.g002

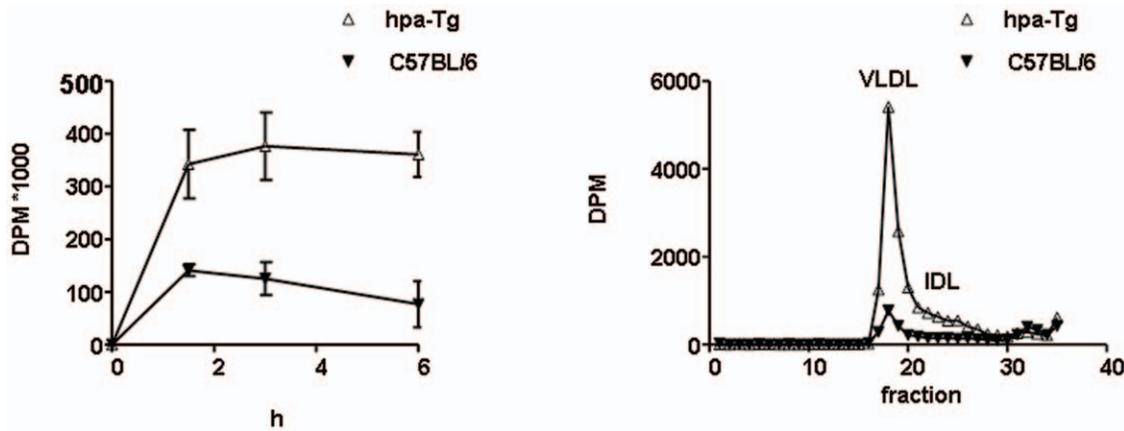


Figure 3. ^3H -retinol accumulation (A) and distribution (B) in plasma following oral administration. (A) Mice were fasted overnight and then administered a bolus of 100 μl corn oil containing ^3H -retinol via a stomach tube. Radioactivity in plasma of hpa-Tg mice ($n = 3$) was significantly higher compared to C57BL/6 ($n = 3$) during the 6 h following retinol administration ($p < 0.01$). (B) 200 μl of plasma from hpa-Tg and C57BL/6 mice, taken 3 h following retinol load were analyzed by FPLC. As indicated in the figure, retinol radioactivity was increased in the VLDL and IDL fractions of hpa-Tg compared to control mice. AUC (dpm \times fraction) of hpa-Tg was 16703 compared to 4640 in C57BL/6 mice. doi:10.1371/journal.pone.0018370.g003

and the development of atherosclerotic lesions. Our studies show that when HS chain length is decreased due to over-expression of heparanase, there is increased fasting TG and reduced hepatic uptake/clearance of TRPs. At the same time, the cross sectional area of fatty streaks was increased.

HSPGs have been shown to regulate the clearance of TRPs via binding, uptake and degradation of these particles in the liver. The importance of HSPG mediated pathways of TRP clearance was demonstrated by several studies including Ebara et al [9] who demonstrated in a diabetic murine model that delayed clearance of

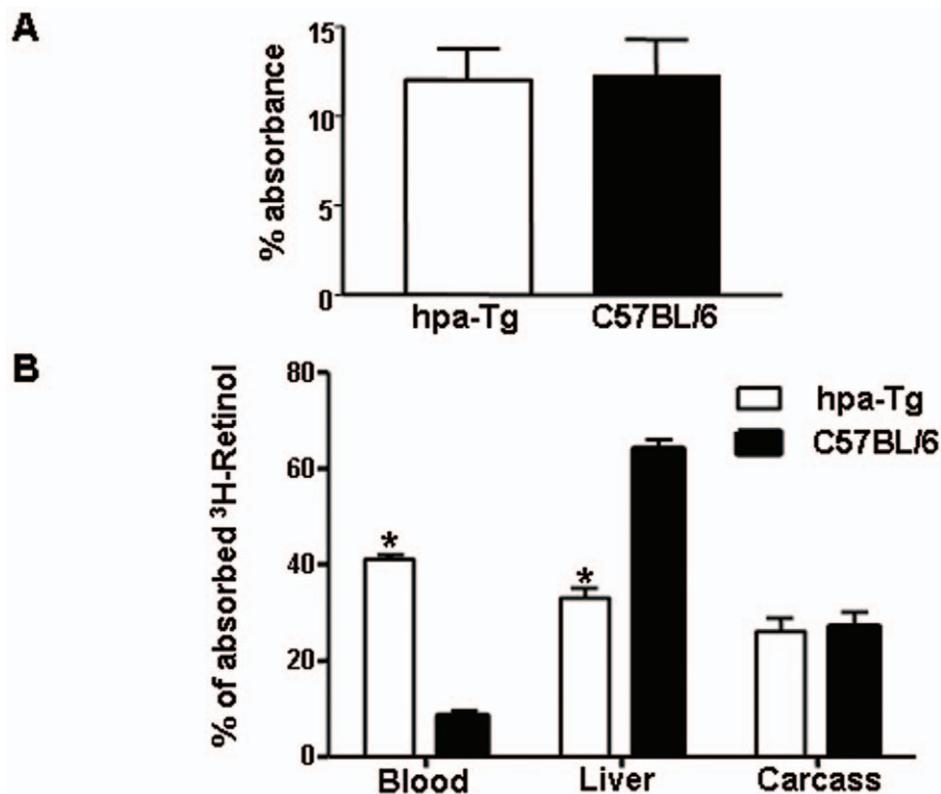


Figure 4. Post-prandial lipid handling in hpa-Tg and C57BL/6 mice. (A) Absorption of radio-labeled vitamin A (^3H -retinol). Intestinal absorption was 12% of the total administered dose of ^3H -retinol in both hpa-Tg ($n = 4$) and C57BL/6 mice ($n = 4$). (B) Distribution of absorbed vitamin A (^3H -retinol) in blood, liver and carcass. 41% of the radioactive retinol were detected in serum of hpa-Tg mice compared to 9% detected in serum of C57BL/6 mice, $p < 0.0001$. No difference was detected in radioactivity extracted from carcasses of the two experimental groups. The decrease in liver uptake of retinol was paralleled by an equivalent increase in the level of labeled retinol in the blood of hpa-Tg mice. doi:10.1371/journal.pone.0018370.g004

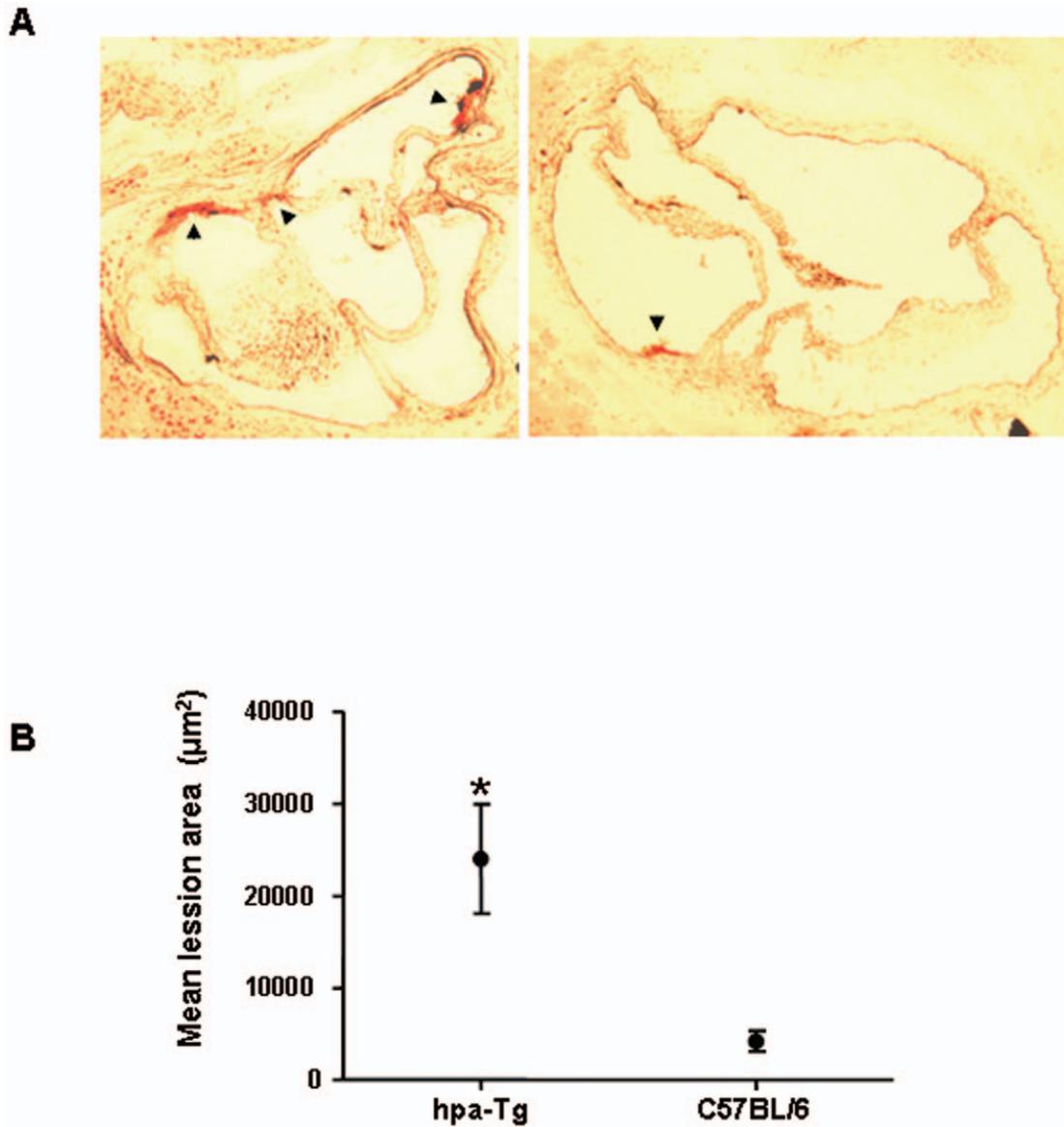


Figure 5. Fatty streaks in hpa-Tg and C57BL/6 mice. (A) Frozen sections of aortic root from hpa-Tg and C57BL control mice (n = 15 for each group) were stained with Oil-Red-O for lipid content. (B) Mean lesion area in heparanase over-expressing mice was significantly higher compared to control mice, $p < 0.001$.

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post-prandial apoB-48-containing lipoproteins is due to decreased hepatic HS. Reduced hepatic HS in diabetes was attributed to decreased expression of hepatic N-deacetylase/N-sulfotransferase 1 (NDST) which is a key enzyme in the biosynthesis of HSPG [9,17]. Our data demonstrates that over-expression of heparanase is associated with delayed clearance of TRPs and reduced uptake by the liver resulting in elevated plasma levels of these atherogenic particles.

The specific role of HSPGs in the hepatic uptake of lipoproteins, especially TRPs, has been elucidated by selectively modifying different receptors and enzymes involved in lipoprotein metabolism. In the most recently proposed models, apoE mediates the binding of remnants to HSPGs leading to sequestration of these particles in the space of Disse. Internalization of remnants occurs via several pathways including those transduced by LDL receptor (LDLR), LDLR-related protein (especially in the absence of

functional LDLR) and HSPG [3,4,18]. HSPGs also facilitate internalization of apoE-enriched remnant lipoproteins by a LDLR independent pathway [2]. At least two distinct HSPGs, syndecan and perlecan, have been shown to be involved in the internalization of remnants in tissue culture system [19,20]. Stanford et al. provided direct genetic evidence *in vivo* that syndecan-1 is the primary hepatic proteoglycan receptor mediating TRL clearance. Their conclusion was based on the observation that TRLs resembling VLDL and VLDL remnants accumulate in fasted syndecan-1 deficient mice and that these mice exhibited delayed clearance of TRLs derived from dietary fat [10].

The significance of the contribution of HSPGs to remnant clearance was shown by Ji et al. [8]. Infusion of heparinase into the portal vein of mice reduced the level of hepatic HSPG (as measured by sulfation) leading to a significant decrease in

clearance and uptake (up to 80%) of remnants. Similarly in our experiments with human heparanase over-production, there was a significant reduction in remnant clearance and internalization. Interestingly, in contrast to Ji et al., we have previously shown that overproduction of heparanase does not lead to overall decrease in hepatic HSPG content but to a reduction in the length of the HS chains attached to the proteoglycan core protein [21]. Shortening of the HS chains may have a significant impact on remnant clearance as there are fewer sites available for the binding of lipoproteins resulting in less sequestration and internalization.

LPL is known to accelerate the clearance of post-prandial lipoprotein by hydrolyzing the TG content of the TRPs. It also binds to lipoprotein *in vivo*, independent of its lipolytic activity and serves as a bridge between lipoprotein and HS chains [22]. The association between hypertriglyceridemia and atherosclerosis development has been demonstrated recently in mice that are mutated in chylomicron-processing due to mutations in LPL binding or activity and accumulation of large TG-rich particles [23,24]. We investigated whether over-expression of heparanase impacts on the availability of LPL on the endothelial surface thereby reducing LPL mediated lipolysis of TRPs and binding to hepatic HSPGs. Our data showed that there was no difference in post heparin plasma LPL activity between hpa-Tg and control mice indicating that the effect of heparanase over-expression on remnant lipoprotein clearance is not due to reduced lipoprotein lipase activity and availability. This is consistent with recent findings that glycosylphosphatidylinositol-HDL binding protein 1 (gpihbp1) is the main protein that binds LPL to endothelial cell surface rather than HSPG as has been previously reported [5,25,26].

Thirty years ago, Zilvermit proposed that post-prandial TRPs promote the formation of atherosclerotic lesions [27] based on experimental data showing that cholesterol fed rabbits which develop atherosclerosis have a defect in chylomicron clearance by the liver. Subsequent human studies indicated that triglyceridemia is an independent risk factor for early atherosclerosis and subjects with coronary artery diseases exhibit delayed metabolism of TRPs [28,29,30]. Part of the increased susceptibility to atherosclerosis of patients with type 2 diabetes mellitus and the metabolic syndrome has been attributed to the decreased clearance of TRPs [5,31,32].

References

- Vlodavsky I, Korner G, Ishai-Michaeli R, Bashkin P, Bar-Shavit R, et al. (1990) Extracellular matrix-resident growth factors and enzymes: possible involvement in tumor metastasis and angiogenesis. *Cancer Metastasis Rev* 9: 203–226.
- Williams KJ, Fuki IV (1997) Cell-surface heparan sulfate proteoglycans: dynamic molecules mediating ligand catabolism. *Curr Opin Lipidol* 8: 253–262.
- MacArthur JM, Bishop JR, Stanford KI, Wang L, Bensadoun A, et al. (2007) Liver heparan sulfate proteoglycans mediate clearance of triglyceride-rich lipoproteins independently of LDL receptor family members. *J Clin Invest* 117: 153–164.
- Mahley RW, Huang Y (2007) Atherogenic remnant lipoproteins: role for proteoglycans in trapping, transferring, and internalizing. *J Clin Invest* 117: 94–98.
- Williams KJ (2008) Molecular processes that handle — and mishandle — dietary lipids. *J Clin Invest* 118: 3247–3259.
- Toyoshima M, Nakajima M (1999) Human heparanase. Purification, characterization, cloning, and expression. *J Biol Chem* 274: 24153–24160.
- Vlodavsky I, Friedmann Y, Elkin M, Aingorn H, Atzmon R, et al. (1999) Mammalian heparanase: gene cloning, expression and function in tumor progression and metastasis. *Nat Med* 5: 793–802.
- Ji ZS, Sanan DA, Mahley RW (1995) Intravenous heparinase inhibits remnant lipoprotein clearance from the plasma and uptake by the liver: *in vivo* role of heparan sulfate proteoglycans. *J Lipid Res* 36: 583–592.
- Ebara T, Conde K, Kako Y, Liu Y, Xu Y, et al. (2000) Delayed catabolism of apoB-48 lipoproteins due to decreased heparan sulfate proteoglycan production in diabetic mice. *J Clin Invest* 105: 1807–1818.
- Stanford KI, Bishop JR, Foley EM, Gonzales JC, Niesman IR, et al. (2009) Syndecan-1 is the primary heparan sulfate proteoglycan mediating hepatic clearance of triglyceride-rich lipoproteins in mice. *J Clin Invest*.
- Zcharia E, Metzger S, Chajek-Shaul T, Aingorn H, Elkin M, et al. (2004) Transgenic expression of mammalian heparanase uncovers physiological functions of heparan sulfate in tissue morphogenesis, vascularization, and feeding behavior. *Faseb J* 18: 252–263.
- George J, Afek A, Shaish A, Levkovitz H, Bloom N, et al. (2001) 12/15-Lipoxygenase gene disruption attenuates atherogenesis in LDL receptor-deficient mice. *Circulation* 104: 1646–1650.
- Metzger S, Hassin T, Barash V, Pappo O, Chajek-Shaul T (2001) Reduced body fat and increased hepatic lipid synthesis in mice bearing interleukin-6-secreting tumor. *Am J Physiol Endocrinol Metab* 281: E957–965.
- Goldberg DM, Chajek-Shaul T (1990) Effect of chronic heparin administration on serum lipolytic activity and some aspects of lipid metabolism. *Biochim Biophys Acta* 1047: 103–111.
- Ishibashi S, Perrey S, Chen Z, Osuga J, Shimada M, et al. (1996) Role of the low density lipoprotein (LDL) receptor pathway in the metabolism of chylomicron remnants. A quantitative study in knockout mice lacking the LDL receptor, apolipoprotein E, or both. *J Biol Chem* 271: 22422–22427.
- Chang S, Zhang SH, Maeda N, Borenstajn J (1994) Hepatic clearance of chylomicron remnants in mice lacking apoprotein E. *Biochim Biophys Acta* 1215: 205–208.
- Williams KJ, Liu ML, Zhu Y, Xu X, Davidson WR, et al. (2005) Loss of heparan N-sulfotransferase in diabetic liver: role of angiotensin II. *Diabetes* 54: 1116–1122.
- Williams KJ, Chen K Recent insights into factors affecting remnant lipoprotein uptake. *Curr Opin Lipidol* 21: 218–228.
- Fuki IV, Iozzo RV, Williams KJ (2000) Perlecan heparan sulfate proteoglycan: a novel receptor that mediates a distinct pathway for ligand catabolism. *J Biol Chem* 275: 25742–25750.

Author Contributions

Conceived and designed the experiments: DP SM EZ DW TCS. Performed the experiments: DP SM EZ TCS. Analyzed the data: DP SM EZ DW IV TCS. Contributed reagents/materials/analysis tools: EZ IV. Wrote the paper: DP SM EZ IV DW TCS.

20. Fuki IV, Kuhn KM, Lomazov IR, Rothman VL, Tuszynski GP, et al. (1997) The syndecan family of proteoglycans. Novel receptors mediating internalization of atherogenic lipoproteins in vitro. *J Clin Invest* 100: 1611–1622.
21. Li JP, Galvis ML, Gong F, Zhang X, Zcharia E, et al. (2005) In vivo fragmentation of heparan sulfate by heparanase overexpression renders mice resistant to amyloid protein A amyloidosis. *Proc Natl Acad Sci U S A* 102: 6473–6477.
22. Zheng C, Murdoch SJ, Brunzell JD, Sacks FM (2006) Lipoprotein lipase bound to apolipoprotein B lipoproteins accelerates clearance of postprandial lipoproteins in humans. *Arterioscler Thromb Vasc Biol* 26: 891–896.
23. Weinstein MM, Yin L, Tu Y, Wang X, Wu X, et al. Chylomicronemia elicits atherosclerosis in mice—brief report. *Arterioscler Thromb Vasc Biol* 30: 20–23.
24. Zhang X, Qi R, Xian X, Yang F, Blackstein M, et al. (2008) Spontaneous atherosclerosis in aged lipoprotein lipase-deficient mice with severe hypertriglyceridemia on a normal chow diet. *Circ Res* 102: 250–256.
25. Beigneux AP, Davies BS, Gin P, Weinstein MM, Farber E, et al. (2007) Glycosylphosphatidylinositol-anchored high-density lipoprotein-binding protein 1 plays a critical role in the lipolytic processing of chylomicrons. *Cell Metab* 5: 279–291.
26. Franssen R, Young SG, Peelman F, Hertecant J, Sierts JA, et al. (2010) Chylomicronemia with low postheparin lipoprotein lipase levels in the setting of GPIIb/IIIa defects. *Circ Cardiovasc Genet* 3: 169–178.
27. Zilversmit DB (1979) Atherogenesis: a postprandial phenomenon. *Circulation* 60: 473–485.
28. Boquist S, Ruotolo G, Tang R, Björkegren J, Bond MG, et al. (1999) Alimentary lipemia, postprandial triglyceride-rich lipoproteins, and common carotid intima-media thickness in healthy, middle-aged men. *Circulation* 100: 723–728.
29. Noto AT, Mathiesen EB, Brox J, Björkegren J, Hansen JB (2008) Delayed metabolism of postprandial triglyceride-rich lipoproteins in subjects with echolucent carotid plaques. *Lipids* 43: 353–360.
30. Patsch JR, Miesenböck G, Hopferwieser T, Muhlberger V, Knapp E, et al. (1992) Relation of triglyceride metabolism and coronary artery disease. Studies in the postprandial state. *Arterioscler Thromb* 12: 1336–1345.
31. Mero N, Malmstrom R, Steiner G, Taskinen MR, Syvanne M (2000) Postprandial metabolism of apolipoprotein B-48- and B-100-containing particles in type 2 diabetes mellitus: relations to angiographically verified severity of coronary artery disease. *Atherosclerosis* 150: 167–177.
32. Tabas I, Williams KJ, Boren J (2007) Subendothelial lipoprotein retention as the initiating process in atherosclerosis: update and therapeutic implications. *Circulation* 116: 1832–1844.
33. Chung BH, Tallis G, Yalamoori V, Anantharamaiah GM, Segrest JP (1994) Liposome-like particles isolated from human atherosclerotic plaques are structurally and compositionally similar to surface remnants of triglyceride-rich lipoproteins. *Arterioscler Thromb* 14: 622–635.
34. Rapp JH, Lespine A, Hamilton RL, Colyvas N, Chaumeton AH, et al. (1994) Triglyceride-rich lipoproteins isolated by selected-affinity anti-apolipoprotein B immunosorption from human atherosclerotic plaque. *Arterioscler Thromb* 14: 1767–1774.
35. Kudchadkar R, Gonzalez R, Lewis KD (2008) PI-88: a novel inhibitor of angiogenesis. *Expert Opin Investig Drugs* 17: 1769–1776.
36. Vlodaysky I, Ilan N, Naggi A, Casu B (2007) Heparanase: structure, biological functions, and inhibition by heparin-derived mimetics of heparan sulfate. *Curr Pharm Des* 13: 2057–2073.