

RESEARCH ARTICLE

Plant flavones enhance antimicrobial activity of respiratory epithelial cell secretions against *Pseudomonas aeruginosa*

Benjamin M. Hariri¹, Derek B. McMahon¹, Bei Chen¹, Nithin D. Adappa¹, James N. Palmer¹, David W. Kennedy¹, Robert J. Lee^{1,2*}

1 Department of Otorhinolaryngology—Head and Neck Surgery, University of Pennsylvania Perelman School of Medicine, Philadelphia, Pennsylvania, United States of America, **2** Department of Physiology, University of Pennsylvania Perelman School of Medicine, Philadelphia, Pennsylvania, United States of America

* rjl@mail.med.upenn.edu



OPEN ACCESS

Citation: Hariri BM, McMahon DB, Chen B, Adappa ND, Palmer JN, Kennedy DW, et al. (2017) Plant flavones enhance antimicrobial activity of respiratory epithelial cell secretions against *Pseudomonas aeruginosa*. PLoS ONE 12(9): e0185203. <https://doi.org/10.1371/journal.pone.0185203>

Editor: Hiroaki Matsunami, Duke University, UNITED STATES

Received: May 23, 2017

Accepted: September 7, 2017

Published: September 20, 2017

Copyright: © 2017 Hariri et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by grants from the National Institutes of Health (R03DC013862) and Cystic Fibrosis Foundation (LEER16G0) to R.J. L. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

Flavones are a class of natural plant secondary metabolites that have anti-inflammatory and anti-bacterial effects. Some flavones also activate the T2R14 bitter taste receptor, which is expressed in motile cilia of the sinonasal epithelium and activates innate immune nitric oxide (NO) production. Flavones may thus be potential therapeutics for respiratory infections. Our objective was to examine the anti-microbial effects of flavones on the common sinonasal pathogens *Candida albicans*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*, evaluating both planktonic and biofilm growth. Flavones had only very low-level anti-bacterial activity alone. They did not reduce biofilm formation, but did reduce production of the important *P. aeruginosa* inflammatory mediator and ciliotoxin pyocyanin. However, flavones exhibited synergy against *P. aeruginosa* in the presence of antibiotics or recombinant human lysozyme. They also enhanced the efficacy of antimicrobials secreted by cultured and primary human airway cells grown at air-liquid interface. This suggests that flavones may have anti-gram-negative potential as topical therapeutics when combined with antibiotics or in the context of innate antimicrobials secreted by the respiratory or other epithelia. This may have an additive effect when combined with T2R14-activated NO production. Additional studies are necessary to understand which flavone compounds or mixtures are the most efficacious.

Introduction

Chronic rhinosinusitis (CRS) is a syndrome of chronic inflammation and/or infection of the upper respiratory tract (nose and sinuses, termed the sinonasal cavity), which leads to substantial decreases in patient quality of life, creates >8 billion dollars in direct healthcare costs in the US alone, and can seed lower respiratory infections and exacerbate lung diseases [1–3]. CRS is also an important public health concern, as it accounts for ~20% of antibiotic prescriptions in adults in the US [1, 4–8], making it a significant driver for the emergence of antibiotic

Competing interests: The authors have declared that no competing interests exist.

resistant organisms [9–15]. Identification of novel compounds with antibacterial and immunomodulatory activity that can be used as topical therapeutics is of paramount importance to combating CRS and other types of respiratory diseases.

Plants produce thousands of polyphenolic flavonoid compounds [16–19], which are of great biomedical interest because they have biological effects on both eukaryotic and prokaryotic cells [17–28]. Flavones are a sub-group of flavonoids that have been demonstrated to have antibacterial, antioxidant, and anti-inflammatory effects in various *in vitro* models [18, 21, 23, 25, 27, 29, 30]. The flavone apigenin is found in bee propolis and chamomile flowers [20]. Apigenin can inhibit the enzyme D-alanine:D-alanine ligase [22], which catalyzes the production of the peptidoglycan precursor D-ala-D-ala, an essential component of the bacterial cell wall. Apigenin may also increase susceptibility of methicillin-resistant *Staphylococcus aureus* (MRSA) to β-lactam antibiotics [23]. Apigenin can inhibit inflammatory protein kinase C (PKC) [31, 32] and nuclear-factor-kappa B (NFκB) [33] signaling in cells *in vitro*. The flavone chrysanthemum is from *Passiflora* and chamomile flowers as well as the *Pleurotus ostreatus* mushroom [18, 21]. Chrysanthemum was previously demonstrated to have both antifungal and antibacterial effects, either alone or in combination with antibiotics [18, 23, 25, 27, 29, 30]. Chrysanthemum may also inhibit inflammatory responses by interfering with cyclooxygenase-2 production of prostaglandins [24, 34]. Wogonin is from *Scutellaria baicalensis*, one of the 50 fundamental herbs of traditional Chinese medicine [35]. Wogonin has antibacterial effects against *Flavobacterium* fish pathogens [36, 37] and also inhibits inflammatory signaling by TNFα [38], PKC [39], and NFκB [40–42] in mammalian cells *in vitro*. Tangeritin is a flavone found in the peels of tangerines and other citrus fruits [18, 21] which may have antibiofilm effects [43].

Moreover, many flavones activate bitter “taste” G-protein-coupled receptor [44–46], known as taste family 2 receptors or T2Rs. T2Rs are now known to be expressed in many tissues outside of the tongue [47, 48], including in motile cilia of the sinonasal cavity, where they modulate mucociliary clearance through activation of nitric oxide (NO) synthase (NOS) [2, 48–53]. We recently demonstrated that several flavones activate the bitter taste receptor isoform T2R14, which is expressed in sinonasal motile cilia [32]. Innate antimicrobial activities of flavones combined with their ability to activate airway T2Rs may increase their potential efficacy as therapeutics to eradicate infections independent of antibiotics. We thus sought to test the effects of flavones on three representative nasal pathogens frequently found in CRS patients [54, 55]: (1) *Candida albicans*, a fungus, (2) *Staphylococcus aureus*, a gram-positive bacteria species, and (3) *Pseudomonas aeruginosa*, a gram-negative bacteria species. Understanding how flavones interact with different types of sinonasal pathogens will begin to elucidate the potential clinical utility of these compounds against bacterial or fungal respiratory infections.

Results

We studied several representative naturally-occurring flavones: apigenin, chrysanthemum, wogonin, and tangeritin (Fig 1). These flavones have been determined in studies from our own lab [32] and others [44–46] to activate T2Rs and also were determined to have antibacterial effects in other studies (described above). We observed only very subtle effects of individual flavones or a mixture of flavones on planktonic growth of the fungal nasal pathogen *C. albicans* (S1 Fig), measured by changes in optical density (OD) at 600 nm (OD_{600}). A mixture of apigenin, chrysanthemum, tangeritin, and wogonin (100 μM each) slowed planktonic growth by approximately 50% ($p < 0.01$ vs. control). Wogonin was the only individual flavone that significantly reduced OD_{600} after 6 hrs ($p < 0.05$ vs. control) (S1 Fig). The flavone mixture did not significantly enhance the activity of a low concentration of the antifungal antibiotic amphotericin B (0.25 μg/mL). No effects were observed on *C. albicans* hyphae growth (S1 Fig), measured using

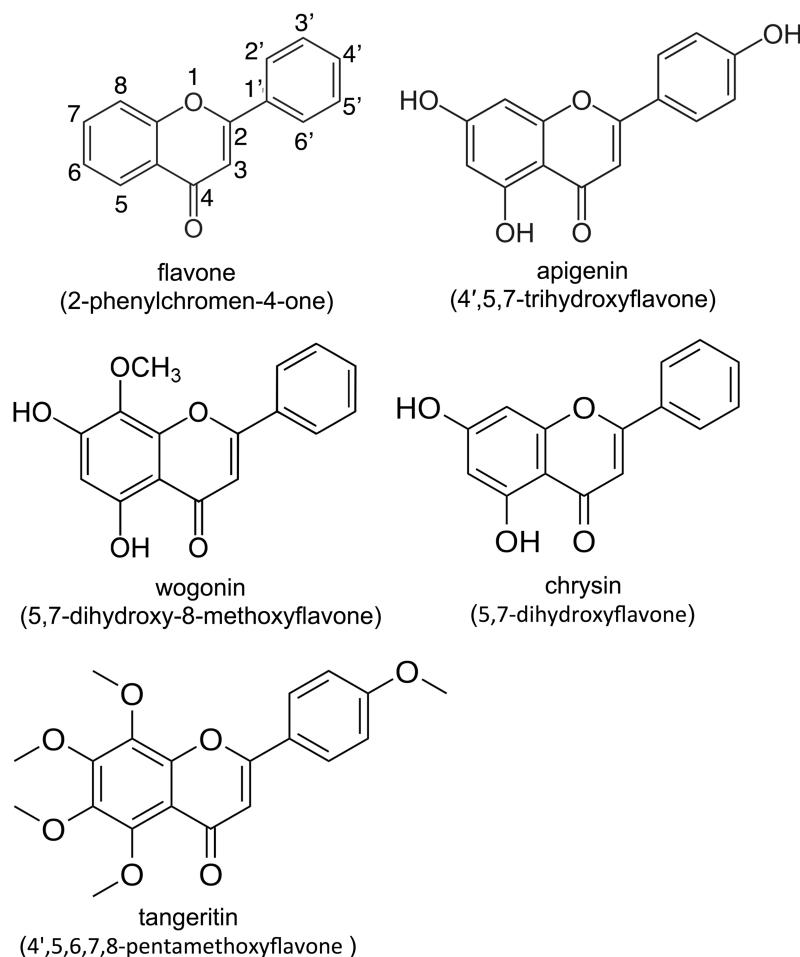


Fig 1. Flavone backbone structure and specific compounds used in this study.

<https://doi.org/10.1371/journal.pone.0185203.g001>

a strain of *C. albicans* (HGFP3) expressing GFP under a hyphae-specific promoter [56]. We also found that flavones likewise had only small effects on the growth of two coagulase-negative *Staphylococcus* (CNS) and two methicillin-resistant *S. aureus* (MRSA) clinical isolate strains (S2 Fig). Flavones did not appear to enhance the activity of a penicillin/streptomycin mix (S2 Fig). However, we did observe a small increase in growth inhibition of MRSA by a low concentration of methicillin (3 µg/mL; S2 Fig) when combined with this flavone mixture. There was no significant effect of the flavone mixture when combined with a higher concentration (30 µg/mL) methicillin (S2 Fig).

In contrast with the more subtle effects observed with *C. albicans* and *Staphylococcus*, we saw more marked effects with *P. aeruginosa*, an important opportunistic gram-negative airway pathogen [55]. A flavone mixture (apigenin, chrysin, and wogonin; 100 µM each) and some individual flavones significantly reduced planktonic growth, but more interestingly also caused a substantial reduction of culture optical density at 600 nm (OD_{600}) when combined with penicillin/streptomycin (Fig 2A and 2B) in wild-type (Wt) *P. aeruginosa* strains PAO1 and ATCC 27853. This suggested that flavones may enhance lysis of *P. aeruginosa* under certain conditions, possibly by destabilizing cell wall components such as D-ala-D-ala, as previously shown with apigenin [22]. We performed planktonic growth/lysis assays in media of varying ionic

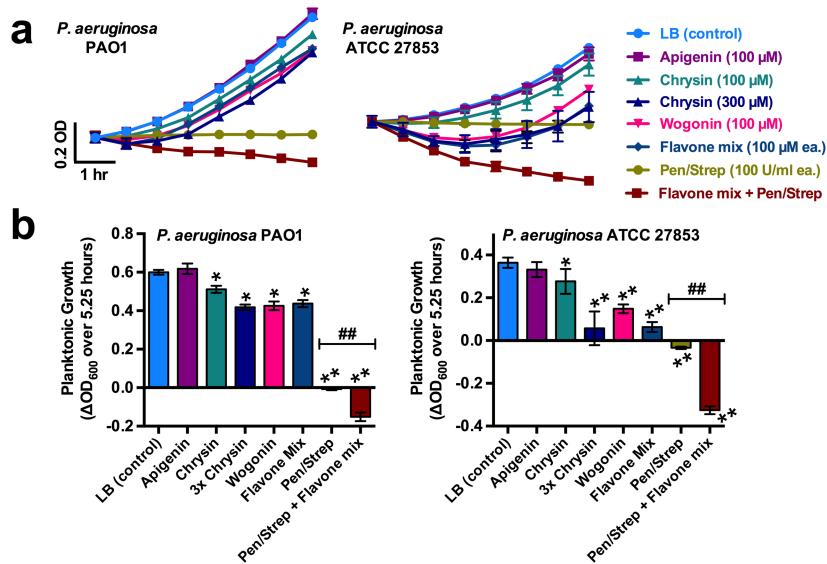


Fig 2. Synergistic anti-bacterial effects of flavones in combination with antibiotics. (A) Planktonic growth traces (OD_{600}) of 2 strains of *P. aeruginosa* (PAO1 and ATCC 27853) under the indicated conditions. Note reduction of OD_{600} in the presence of penicillin/streptomycin plus flavone mixture (apigenin, chrysin, wogonin; 100 μM each). (B) Bar graphs showing ΔOD_{600} over 5.25 hrs from A ($n = 4$ experiments for each condition). Asterisks denote significance vs. control (LB only; one-way ANOVA, Dunnett's post-test; * = $p < 0.05$, ** = $p < 0.01$); ## indicates $p < 0.01$ between bracketed bars (one-way ANOVA, Bonferroni post-test).

<https://doi.org/10.1371/journal.pone.0185203.g002>

strength (100% LB or 25% LB, diluted from 100% with sterile DI-water), and found that lysis of bacteria (evidenced by reduction in OD_{600} from the starting OD of 0.1) in the presence of gentamicin \pm flavones was enhanced by low ionic strength (S3 Fig). In the absence of flavones, a drop in OD_{600} was not observed over 6 hrs (S3 Fig), but lysis was observed when gentamicin was combined with flavones. Reduced ionic strength caused a lower OD_{600} (i.e. a greater decrease from the starting OD_{600} of 0.1) at 30 min (S3 Fig).

We also tested flavones against *P. aeruginosa* biofilm formation using a 96-well plate-based crystal violet assay, but no effects on biofilms were observed (S4 Fig). However, individual flavones, as well as a flavone mixture, significantly decreased production of the ciliotoxin and inflammatory stimulant pyocyanin [57–61] at effective concentrations (ECs) of 1–10 μM for individual flavones and 0.1 μM for the mixture in planktonic PAO1 and ATCC 27853 cultures (S4 Fig). Despite a lack of effect on *P. aeruginosa* biofilms, the data above suggest that flavones have some small but statistically significant effects on bacterial growth alone. More intriguingly, these compounds may significantly enhance the efficacy of antibiotics under certain conditions, potentially by contributing to a disruption of cell wall integrity.

Because many innate defense proteins secreted by the airway epithelium act in part through bacterial lysis or permeabilization (e.g. lysozyme [62–67] and β -defensins [68–71]), we sought to test whether flavones may enhance the efficacy of airway antimicrobials. We tested the effects of flavones combined with the important airway antimicrobial lysozyme, which is secreted by airway submucosal gland serous cells [72–74] and catalyzes the hydrolysis of 1,4-beta-linkages between N-acetyl-D-glucosamine and N-acetylmuramic acid residues of peptidoglycan in the bacterial cell wall [75, 76]. We focused on *P. aeruginosa*, as it appeared to be the most sensitive to flavone effects among the pathogens tested here. While lysozyme is more effective against gram-positive bacteria, it does have effects against gram-negative bacteria. In

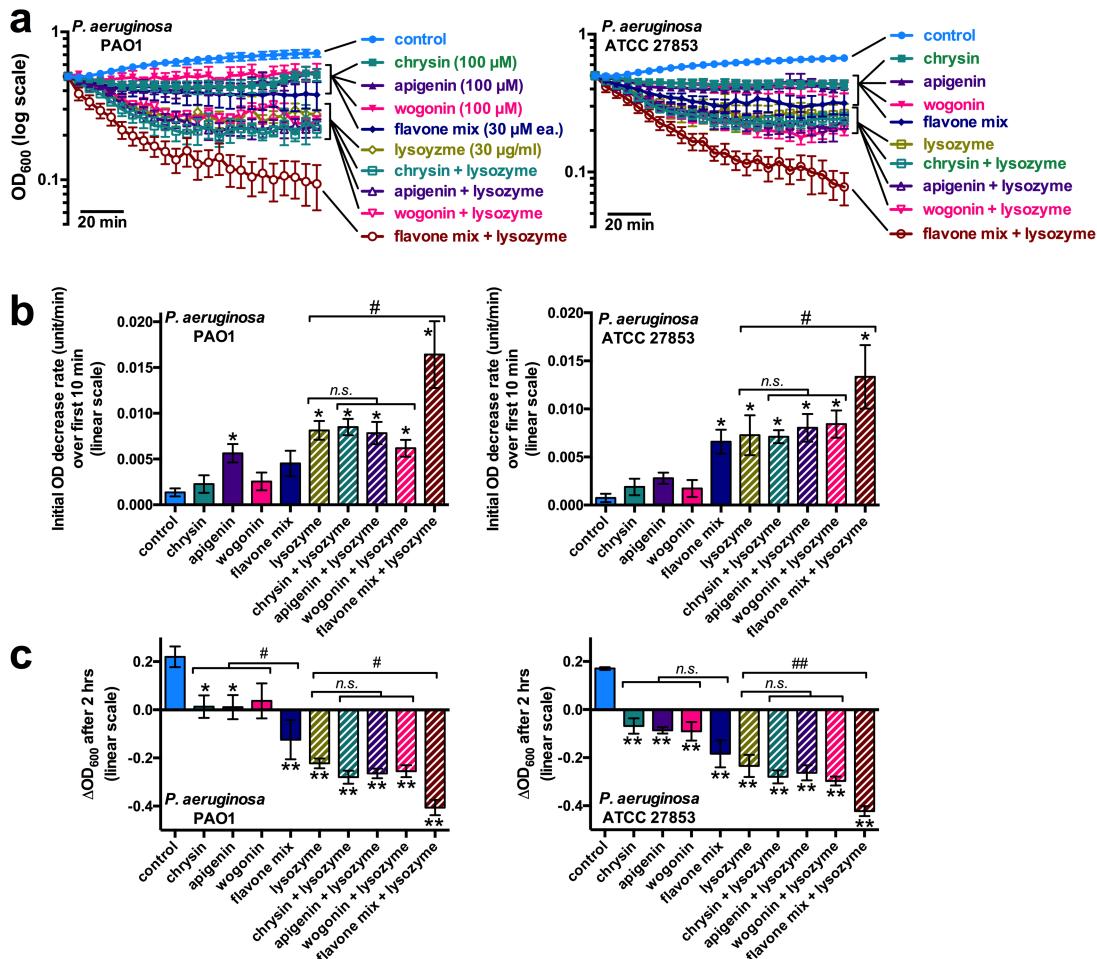


Fig 3. Synergistic anti-bacterial effects of flavones in combination with the airway antimicrobial protein lysozyme. (A) Traces of planktonic growth of *P. aeruginosa* in the presence of flavones ± lysozyme. Note the greatest OD₆₀₀ decrease (bacterial lysis) occurred with lysozyme and flavone mix combined. (B) Bar graphs of the initial OD decrease rate (OD₆₀₀ units/min) from A ($n = 3\text{--}6$ experiments for each condition). (C) Bar graphs showing ΔOD_{600} after 2 hours from A. For B and C, asterisks denote significance vs. control (LB only) by one-way ANOVA, Dunnett's post-test ($* = p < 0.05$, $** = p < 0.01$); # and ## indicates $p < 0.05$ and 0.01 , respectively, between bracketed bars (one-way ANOVA, Bonferroni post-test).

<https://doi.org/10.1371/journal.pone.0185203.g003>

the presence of EDTA to help disrupt the outer lipopolysaccharide-containing gram-negative layer, rapid lysis is observed with lysozyme treatment at low ionic strength [76, 77]. Rapid lysis of *P. aeruginosa* (decrease in OD₆₀₀) was observed in the presence of recombinant human lysozyme ± flavones (Fig 3A–3C). While individual flavones (apigenin, chrysin, and wogonin) caused minimal lysis under these conditions and did not enhance lysozyme-mediated lysis, a mixture of these flavones (100 μM each) promoted lysis approximately 50% as well as lysozyme after 2 hrs. (Fig 3C). Moreover, the flavone mixture significantly enhanced lysozyme-mediated *P. aeruginosa* lysis (Fig 3B and 3C). Data are presented as both initial rate of lysis (Fig 3B) as well as the change in OD₆₀₀ after 2 hrs from the starting OD of 0.5 (Fig 3C).

To confirm that the decrease in OD₆₀₀ observed reflected bacterial lysis, we quantified GFP release from a strain of *P. aeruginosa* PAO1 that expresses soluble GFP, PAO-GFP. After exposure to lysozyme ± flavones for 10 min, cells were centrifuged and supernatant was collected for measurement of fluorescence. Lysozyme significantly increased supernatant fluorescence

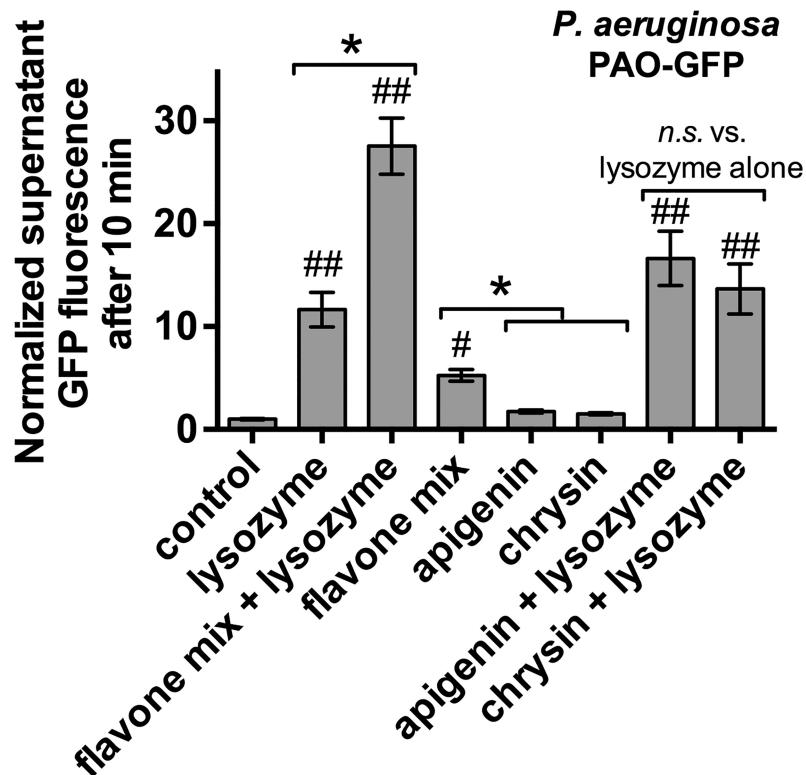


Fig 4. Confirmation of bacterial lysis by measurement of GFP release from PAO-GFP. Bar graph showing normalized fluorescence (control = 1) from PAO-GFP cultures incubated in lysozyme lysis buffer with addition of lysozyme and/or flavones as indicated and described in the text. Flavone mix contained 100 μ M each apigenin, chrysin, and wogonin. Apigenin and chrysin were used alone at 300 μ M to compare an equal number of moles of flavone molecules. Synergistic effects of the flavone mixture were observed both alone and combined with lysozyme. Significance determined by one-way ANOVA with Bonferroni post-test; # and ## indicate $p < 0.05$ and $p < 0.01$, respectively, compared with control; * indicates $p < 0.05$ between bracketed groups.

<https://doi.org/10.1371/journal.pone.0185203.g004>

over control conditions (lysis buffer alone), which was enhanced by addition of the flavone mix (100 μ M each apigenin, chrysin, and wogonin) (Fig 4). The flavone mix by itself caused a significant increase in GFP release that was not observed by an equal amount of a single flavone (300 μ M apigenin or 300 μ M chrysin) (Fig 4). Neither 300 μ M apigenin nor 300 μ M chrysin significantly increased lysozyme-mediated lysis (Fig 4), supporting that there is a synergistic effect of mixing flavone compounds.

An enhancement of cell membrane damage by flavones was also confirmed by staining with 1-N-phenylnaphthylamine (NPN). NPN is a hydrophobic fluorophore that undergoes a quantum yield increase upon transition from a hydrophilic aqueous environment to a hydrophobic phospholipid environment [78], and NPN uptake is frequently used to measure gram-negative outer membrane permeabilization [78–82]. A flavone mix enhanced NPN uptake alone or in combination with lysozyme greater than an equal amount of individual flavones, as found above (Fig 5).

To determine if the effects observed above with recombinant lysozyme translated to enhancement of physiological antimicrobials secreted by airway epithelial cells, we tested the ability of flavones to enhance the antimicrobial activity of Calu-3 cell secretions. Calu-3 cells are a model of airway secretory cells [72], resembling some aspects of both submucosal gland

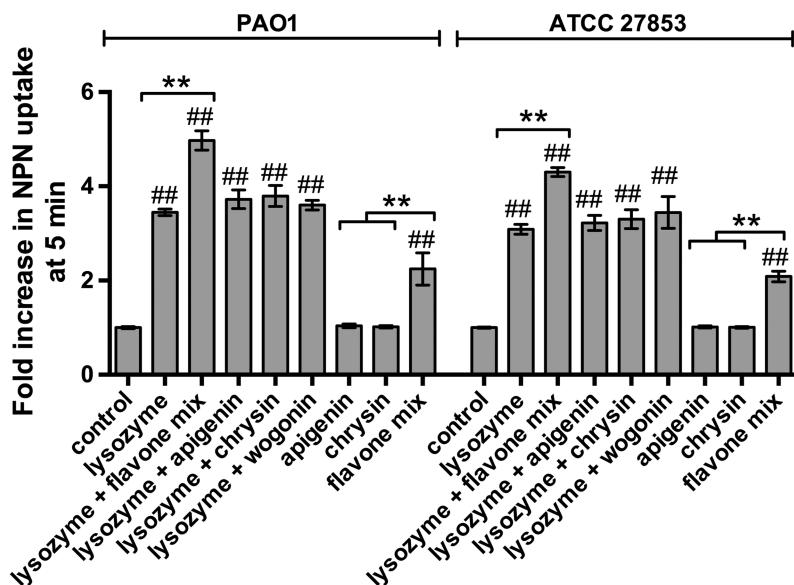


Fig 5. Confirmation of bacterial cell wall damage by NPN uptake. Bacteria were incubated for 5 min with lysozyme ± flavones as indicated and described in the text, followed by measurement of NPN fluorescence, reflecting uptake of NPN into the bacterial phospholipid membrane. Data are expressed as fold increase in NPN fluorescence. Raw fluorescence values are in S1 File. Significance determined by one-way ANOVA with Bonferroni post-test; # and ## indicate $p < 0.05$ and $p < 0.01$, respectively, compared with control; ** indicates $p < 0.01$ between bracketed groups.

<https://doi.org/10.1371/journal.pone.0185203.g005>

serous cells (e.g. secretion of lysozyme [83] and β -defensins [84]) as well as surface epithelial goblet cells (e.g. secretion of Muc5AC [85, 86]). We carried out an antimicrobial assay using apical airway surface liquid (ASL) washings from Calu-3 air-liquid interface (ALI) cultures (ALIs; Fig 6A). We found that the collected Calu-3 washings were potently antibacterial, as they caused a >3 -log reduction in the number of *P. aeruginosa* CFUs recovered even at a 25% dilution (Fig 6B). The antimicrobial effect was partially lost at further dilution (12.5%) and fully lost at 6.25% dilution, but the activity of these low dilutions was significantly enhanced by the presence of a flavone mixture (Fig 6B).

We also tested the ability of flavones to enhance the activity of ASL washings from primary sinonasal ALIs (Fig 6C), which differentiate into ciliated and goblet cells, mimicking the *in vivo* epithelium and secreting a similar array of antimicrobial peptides [50, 53, 87, 88]. ALI cultures also contain solitary chemosensory cells, which express T2R bitter taste receptors and regulate secretion of antimicrobial peptides from surrounding cells [50]. Primary sinonasal ALIs were stimulated apically with denatonium benzoate, a bitter compound that activates the T2Rs in solitary chemosensory cells (T2Rs 10, 30, and/or 46) and stimulates rapid secretion of stored antimicrobial peptides, including β -defensins 1 and 2 [50]. β -defensins are small cationic proteins that permeabilize bacterial cell membranes. Flavones had no significant effect on CFUs recovered when mixed with control (un-stimulated, PBS-only-treated) ASL (Fig 6D), but potentiated the antibacterial effects of denatonium-stimulated ASL when diluted to 25% and 12.5% (Fig 6D).

Discussion

Here, we found specifically that a mixture of flavones can enhance the bacteriolytic activity of recombinant human lysozyme against a common gram-negative opportunistic pathogen, *P.*

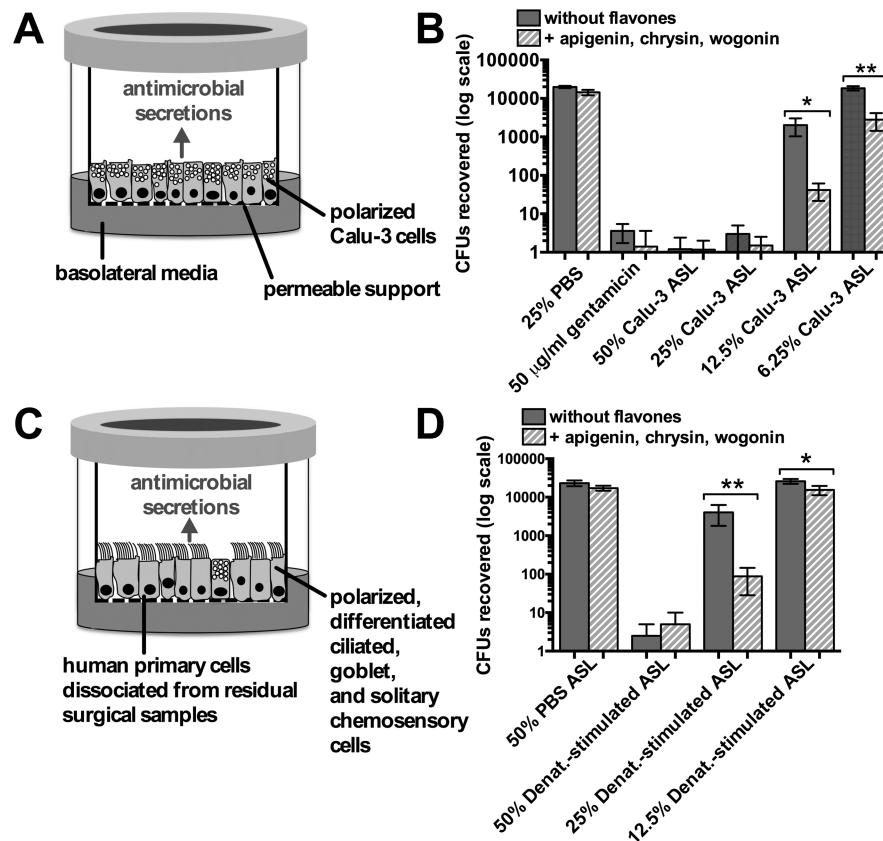


Fig 6. Antimicrobial effects of Calu-3 and primary cell airway surface liquid (ASL) are enhanced by flavones. (A) Calu-3 air-liquid interface cultures (ALIs) recapitulate a polarized secretory epithelium with polarized secretion of antimicrobial peptides and mucus, similar in composition to that of airway submucosal exocrine gland serous acinar cells [65, 72, 89]. (B) Bar graph shows number of colony forming units (CFUs) recovered from bacteria mixed with dilutions of Calu-3 ASL washings. As negative control, bacteria were incubated with 25% PBS (first column) not in contact with Calu-3 cells; 50 μg/mL gentamicin in 25% PBS was used as positive control. Antimicrobial activity was enhanced at lower dilutions of Calu-3 ASL (12.5% and 6.25%) in the presence of the flavone mix (50 μM each apigenin, chrysanthemum, and wogonin). (C) Primary sinonasal epithelial cultures recapitulate the surface airway epithelium, with differentiated ciliated, goblet, and solitary chemosensory cells, likewise with polarized secretion of antimicrobial peptides and mucus. (D) Bar graph shows CFUs when *P. aeruginosa* were mixed with ASL washings from primary sinonasal ALI cultures stimulated with denatonium benzoate (10 mM). Asterisks denote significance determined by one-way ANOVA, Bonferroni post-test of paired columns (each condition ± flavone; * p < 0.05 and ** p < 0.01).

<https://doi.org/10.1371/journal.pone.0185203.g006>

aeurginosa, and flavones can enhance the bactericidal activity of endogenous human respiratory cell secretions, of which lysozyme is a major component [65, 83, 89, 90]. Together with our previous study [32], our data suggest flavone compounds might modulate respiratory innate immunity through multiple mechanisms (Fig 7). These compounds may have some clinical utility to enhance antibiotic efficacy or enhance endogenous innate immunity.

To our knowledge, such synergistic activities of a mixture of different but highly similar-structured flavonoids have not been previously reported. While synergistic activities with antibiotics have been reported, synergy with endogenous airway antimicrobials is also a novel finding. While our data suggest that flavones have only very low-level antibacterial activity on their own, they may have a substantial ability to enhance the efficacy of innate antimicrobials secreted by human airway cells. Coupled with their previously demonstrated anti-inflammatory activities [18, 21, 91, 92] and activation of T2R-mediated innate immune responses [32],

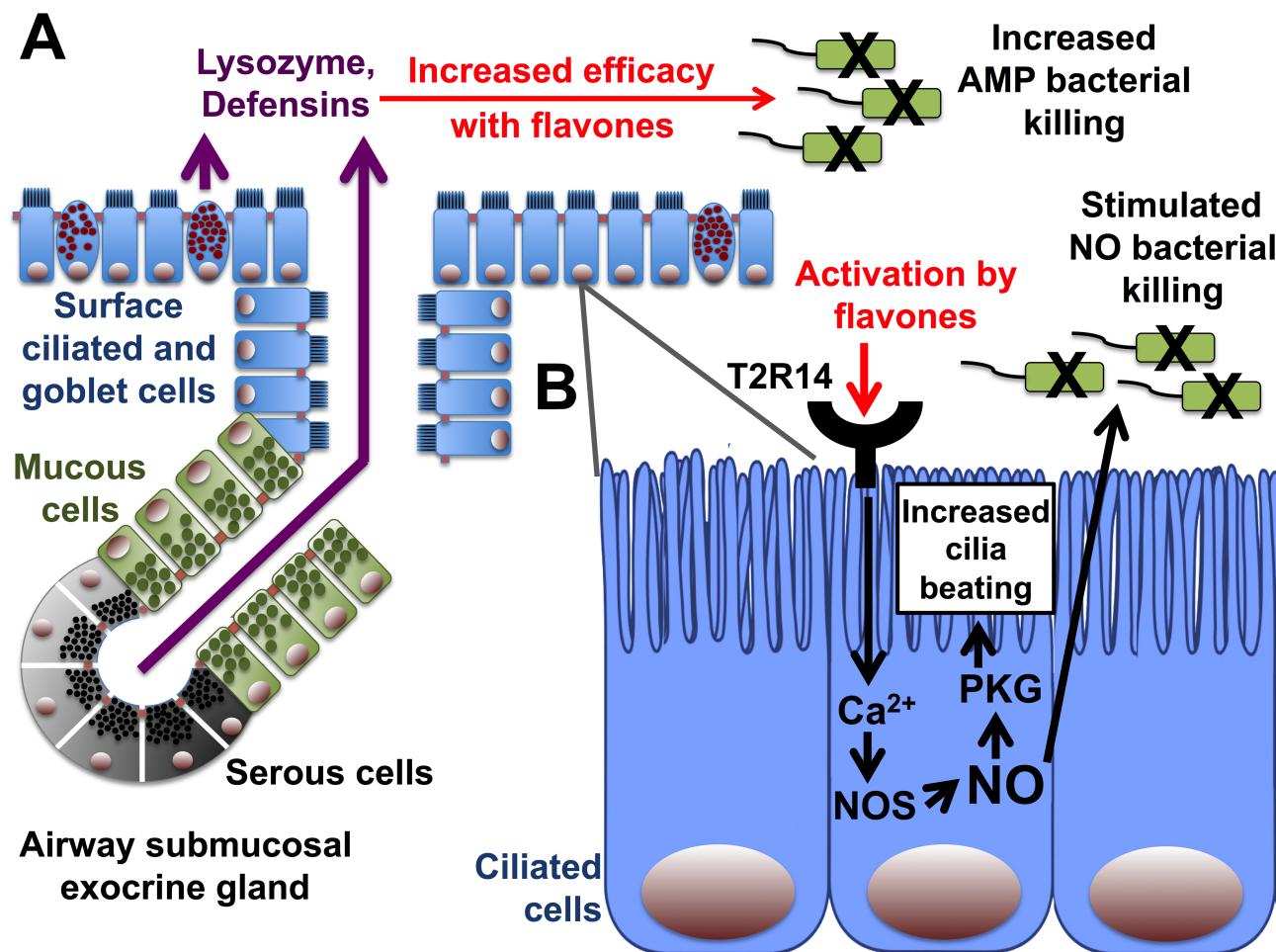


Fig 7. Multiple mechanisms of modulation of respiratory epithelial innate immunity by flavones. (A) Lysozyme is primarily secreted by serous cells of airway submucosal exocrine glands [72]. Defensins are secreted by surface epithelial cells as well as glands. Here, we show that flavones increase the efficacy of these and possibly other secreted antimicrobial peptides (AMPs). (B) We showed previously that flavones also activate the bitter taste receptor T2R14, expressed in both sinonasal [32] and bronchial cilia [99]. T2R14 activation in sinonasal cilia increases nitric oxide synthase (NOS)-mediated production of NO, which increases ciliary beating through protein kinase G (PKG) to promote bacterial clearance and directly diffuses into the airway surface liquid to kill bacteria [53].

<https://doi.org/10.1371/journal.pone.0185203.g007>

flavones are an attractive novel class of compounds to investigate as potential topical therapeutics for respiratory infections. The potential clinical utility of these compounds is most strongly supported by their ability to enhance bactericidal efficacy of secretions from human airway cells independent of their NO-generating function on T2R receptors [32], as the flavones were mixed with airway cell secretions after removal from the epithelial cells. Future high-throughput screening of flavone compounds using *in vitro* assays as described here may identify the most efficacious single compounds or mixtures to activate these effects, suggesting which compounds to prioritize for future *in vivo* testing.

Materials and methods

Reagents

Unless indicated, all reagents, solutions, and protocols used were as previously described [49, 50, 53, 87, 88]. Stock solutions of flavones (Cayman Chemical, Ann Arbor, MI) were

made at 100 or 120 mM in DMSO ($\geq 1000x$). Final DMSO concentrations were always $\leq 0.3\%$, which had no effect alone on bacterial growth kinetics in any assay tested. Reagents not specifically listed below were obtained from Sigma-Aldrich (St. Louis, MO).

Bacterial and fungal culture and antibacterial assays

Pseudomonas aeruginosa strains PAO1 (HER-1018; ATCC BAA-47) and ATCC 27853 (Boston 41501) were from American Type Culture Collection (Manassas, VA) cultured in LB medium (Gibco/Thermo Scientific). Methicillin-resistant *Staphylococcus aureus* (MRSA) strain M2 [93] and clinical isolates of MRSA and coagulase-negative *Staphylococcus* (isolated by the Philadelphia VA Medical Center Microbiology Laboratory) were grown in tryptic soy broth (TSB; Gibco/Thermo Scientific). *Candida albicans* strain HGFP3[56] (kindly provided by Drs. S. Sundstrom, N. Kavanaugh, and K. Ribbeck) was grown in YPD broth (Gibco/Thermo Scientific). Antibiotics (gentamicin, methicillin, and penicillin/streptomycin mixture) were obtained from Sigma. PAO-GFP, a gift from Dr. N. Cohen (University of Pennsylvania), expresses soluble GFP on a gentamicin-resistant plasmid and was grown in media containing 50 $\mu\text{g}/\text{mL}$ gentamicin to maintain selection.

For planktonic growth assays, an overnight log-phase culture was diluted to a density of 0.1, with 10 mL total volume per sample. Cultures were grown in 15 mL tubes at 37°C with shaking (180 RPM); 1 mL of solution was removed at each time point (up to 10 time points) and assayed for optical density (OD) at 600 nm in a spectrophotometer. Biofilm assays were carried out in 96-well plates as previously described [28, 94]. After incubation for 48 hrs, microtiter plates were washed with distilled water, followed by staining with 1% crystal violet for approximately 30 min. After a second washing, biofilm mass and crystal violet were solubilized by incubation in 30% acetic acid for 30 min with shaking, and read on a plate reader at 590 nm. Pyocyanin extraction was carried out as previously described [95]. Briefly, 8 mL of supernatant from an overnight culture (grown in cation-adjusted Mueller-Hinton broth, normalized to $\text{OD}_{600} = 1$) was mixed with 3 mL of chloroform. After vortexing and centrifugation, pyocyanin was extracted from the resulting organic chloroform phase with 1 mL of 0.2 N HCl, with absorbance of the acidified pyocyanin read at 520 nm in a plate reader (Spark 10M, Tecan, Männedorf, Switzerland). All values were blanked to LB that had undergone the same extraction procedure. Lysozyme lysis assays were conducted essentially as described [76, 77, 96]. Bacteria were washed and resuspended (OD 0.5) in 20 mM Tris-HCl, 1 mM EDTA, pH 8.0 with recombinant human lysozyme \pm flavones before monitoring OD_{600} between incubations at 37°C. For GFP release assays, lysozyme lysis experiments were carried out as above in 1 mL volume; after 10 min, samples were centrifuged to pellet bacteria (10,000 g for 15 min) and supernatant GFP fluorescence was measured on a Tecan Spark 10M plate reader (485 excitation, 535 emission). Background was estimated by measuring lysis buffer alone and was subtracted from each experimental value before normalization to control conditions (supernatant from cells in lysis buffer alone, no lysozyme).

Bacterial NPN fluorescence assay was modified from previous descriptions [79–82]. *Pseudomonas* were grown to an OD_{600} of 0.5 in LB, centrifuged, and resuspended at half volume of 10 mM HEPES, 5 mM glucose, 0.1 mM EDTA, pH 8. Bacteria were then aliquoted and mixed with an equal volume of diluted airway surface liquid secretions or antibiotics, and then pipetted into a plate reader containing an equal volume of 25% PBS containing 20 μM NPN (final NPN 10 μM , final OD_{600} 0.25). Samples were then incubated for 10 min and read on a Tecan 10M plate reader at 350 nm excitation and 450 nm emission. Emission wavelength was chosen to minimize any endogenous fluorescence of flavones used. Samples were read in triplicate, with averages of at least 3 independent experiments reported. At these wavelengths,

flavones at concentrations used exhibited no significant fluorescence compared with buffer alone.

CFU antimicrobial assays with ASL washings were carried out similarly to a previously published protocol [50, 90] and modified based on our own antimicrobial ASL protocols used in our lab [50]. Cultures were washed copiously with PBS and transferred to antibiotic-free MEME for 48 hrs. before use. Calu-3 cell secretions were collected from 3 week old ALIs stimulated basolaterally with 100 μM isoproterenol for 72 hours, followed by washing of the apical surface with 30 μL 25% PBS. While washing a 1.1 cm² ALIs with 30 μL significantly dilutes the ASL fluid (~1 μL per cm² of surface area [97]), washings retained antibacterial activity and were thus sufficient to be used for this assay. ASL washings (30 μL per culture) were pooled and mixed with bacteria resuspended in 25% PBS, adjusted to 0.1 OD, then diluted 1:1000 in 25% PBS. Bacteria/ASL mixture was divided and flavone mixture (1000x stock in DMSO) was added. Bacteria and ASL mixture was incubated statically in a 96-well plate at 37°C for 2 hrs, followed by 4 serial 10-fold dilutions and spot plating onto LB plates. After overnight incubation at 37°C, CFUs were manually counted.

For primary sinonasal cultures, we carried out an antimicrobial assay as previously described [50]. ASL cultures (0.33 cm² surface area) were transferred to antibiotic free medium for 48 hrs. and washed copiously on the day of the experiment to remove already-secreted antimicrobials. ALIs were then treated on the apical side with 30 μL of 25% PBS ± 10 mM denatonium benzoate and incubated for 30 min at 37°C. ASL was then collected from 3–4 cultures from the same patient and pooled, followed by further dilution with 25% PBS and/or mixing 1:1 with bacteria diluted in 25% PBS (0.1 OD overnight culture diluted 1:1000 in 25% PBS ± flavones). Bacteria and ASL were incubated for 2 hrs at 37°C followed by dilution and spot plating as described above for Calu-3 cells.

Generation of Calu-3 and primary sinonasal air-liquid interface (ALI) cultures

Calu-3 bronchial epithelial cells were obtained from ATCC and cultured in T75 flasks in minimal essential medium (MEM) with Earl's salts and 1 mM L-glutamine, 10% fetal bovine serum, and 1% cell culture penicillin/streptomycin mix. Cells were lifted with 0.25% trypsin and plated on 1.1 cm² cell culture inserts (Greiner BioOne Thincerts, transparent, 0.4 μm pore size). Cells were grown to confluence for 5 days, followed by apical exposure to air and subsequent 3–4 weeks for full differentiation/polarization before use. Only ALIs with transepithelial resistances (TEERs) of 250–300 Ω·cm² were used.

For primary cells, all experimental protocols were carried out in accordance with the University of Pennsylvania School of Medicine guidelines regarding use of residual clinical material in research, U.S. Department of Health and Human Services code of federal regulation Title 45 CFR 46.116, and the Declaration of Helsinki. Patients undergoing sinonasal surgery for either sinonasal disease (e.g. chronic rhinosinusitis) or other procedures (e.g. transnasal approaches to the skull base) were recruited from the Department of Otorhinolaryngology at the University of Pennsylvania with full IRB approval (#800614) and written informed consent was obtained for all participating. Inclusion criteria included patients over 18 years of age who were undergoing medically necessary sinonasal surgery. Exclusion criteria included a history of systemic diseases (e.g. granulomatosis with polyangiitis, cystic fibrosis), immunodeficiencies (e.g., common variable immune deficiency), or use of antibiotics, oral corticosteroids, or anti-biologics (e.g. Xolair) within one month of surgery. Human sinonasal epithelial cells were enzymatically dissociated and grown to confluence in proliferation medium (DMEM/Ham's F-12 plus BEBM; Clonetics, Cambrex, East Rutherford, NJ) for 7 days as previously described

[53, 98]. Confluent cells were dissociated and seeded on porous polyester membranes coated with BSA, type I bovine collagen, and fibronectin in Corning Transwell cell culture inserts in LHC basal medium (Invitrogen). Culture medium was removed from the upper compartment and basolateral media was changed to differentiation medium (1:1 DMEM:BEBM) containing hEGF (0.5 ng/mL), epinephrine (5 ng/mL), BPE (0.13 mg/mL), hydrocortisone (0.5 ng/mL), insulin (5 ng/mL), triiodothyronine (6.5 ng/mL), and transferrin (0.5 ng/mL), supplemented with 100 U/mL penicillin, 100 g/mL streptomycin, 0.1 nM retinoic acid, and NuSerum (BD Biosciences, San Jose, CA) as previously described [53, 98].

Data analysis and statistics

One-way analysis of variance (ANOVA) was performed in GraphPad Prism with appropriate post-tests as indicated; $p < 0.05$ was considered statistically significant. For comparisons of all samples within a data set, Tukey-Kramer post-test was used. For preselected pair-wise comparisons, Bonferroni post-test was performed. For comparisons to a single control group, one-way ANOVA with Dunnett's post-test was used. All other data analysis was performed in Microsoft Excel. For all figures, one asterisk or pound sign (*) or (#) indicates $p < 0.05$ and two asterisks or pound signs (**) or (##) indicates $p < 0.01$ respectively; "n.s." indicates no statistical significance. All data are presented as mean \pm SEM.

Supporting information

S1 Fig. Flavones have minimal effects on *Candida albicans* growth.
(PDF)

S2 Fig. Effects of flavones on planktonic *Staphylococcus* growth.
(PDF)

S3 Fig. Effects of ionic strength on flavone + antibiotic cell lysis.
(PDF)

S4 Fig. Flavones do not affect *P.aeruginosa* biofilm formation but do reduce pyocyanin production.
(PDF)

S1 File. Data values from the main text.
(PDF)

Acknowledgments

The authors thank P. Sundstrom (Dartmouth Medical School), N. Kavanaugh (MIT), and K. Ribbeck (MIT) for HGFP3 *C. albicans*, N. Cohen (University of Pennsylvania) for PAO-GFP, and L. Chandler (Philadelphia VA Medical Center) for clinical nasal isolates of *S. aureus* and *S. epidermidis*. The authors also thank M. Victoria (University of Pennsylvania) for excellent technical assistance.

Author Contributions

Conceptualization: Benjamin M. Hariri, Bei Chen, Nithin D. Adappa, James N. Palmer, David W. Kennedy, Robert J. Lee.

Data curation: Benjamin M. Hariri, Bei Chen, Nithin D. Adappa, James N. Palmer, David W. Kennedy.

Formal analysis: Benjamin M. Hariri.

Funding acquisition: Robert J. Lee.

Investigation: Benjamin M. Hariri, Derek B. McMahon, Bei Chen, Robert J. Lee.

Methodology: Derek B. McMahon, Bei Chen, Nithin D. Adappa, James N. Palmer, David W. Kennedy.

Project administration: Robert J. Lee.

Supervision: Robert J. Lee.

Writing – original draft: Benjamin M. Hariri, Robert J. Lee.

Writing – review & editing: Benjamin M. Hariri, Nithin D. Adappa, James N. Palmer, David W. Kennedy, Robert J. Lee.

References

1. Stevens WW, Lee RJ, Schleimer RP, Cohen NA. Chronic rhinosinusitis pathogenesis. *J Allergy Clin Immunol.* 2015; 136(6):1442–53. <https://doi.org/10.1016/j.jaci.2015.10.009> PMID: 26654193
2. Lee RJ, Cohen NA. Role of the bitter taste receptor T2R38 in upper respiratory infection and chronic rhinosinusitis. *Curr Opin Allergy Clin Immunol.* 2015; 15(1):14–20. <https://doi.org/10.1097/ACI.0000000000000120> PMID: 25304231
3. Hariri BM, Cohen NA. New insights into upper airway innate immunity. *Am J Rhinol Allergy.* 2016; 30(5):319–23. <https://doi.org/10.2500/ajra.2016.30.4360> PMID: 27657896
4. Alanis AJ. Resistance to antibiotics: are we in the post-antibiotic era? *Arch Med Res.* 2005; 36(6): 697–705. Epub 2005/10/12. <https://doi.org/10.1016/j.arcmed.2005.06.009> PMID: 16216651
5. Cherry DK, Woodwell DA. National Ambulatory Medical Care Survey: 2000 summary. *Adv Data.* 2002; (328):1–32. Epub 2003/03/29. PMID: 12661586
6. Ly N, McCaig LF. National Hospital Ambulatory Medical Care Survey: 2000 outpatient department summary. *Adv Data.* 2002;(327):1–27. Epub 2003/03/29. PMID: 12661587
7. Ray NF, Baraniuk JN, Thamer M, Rinehart CS, Gergen PJ, Kaliner M, et al. Healthcare expenditures for sinusitis in 1996: contributions of asthma, rhinitis, and other airway disorders. *J Allergy Clin Immunol.* 1999; 103(3 Pt 1):408–14. Epub 1999/03/09. PMID: 10069873
8. Bhattacharyya N, Grebner J, Martinson NG. Recurrent acute rhinosinusitis: epidemiology and health care cost burden. *Otolaryngol Head Neck Surg.* 2012; 146(2):307–12. <https://doi.org/10.1177/0194599811426089> PMID: 22027867
9. Marcinkiewicz J, Strus M, Pasich E. Antibiotic resistance: a "dark side" of biofilm-associated chronic infections. *Pol Arch Med Wewn.* 2013; 123(6):309–13. Epub 2013/07/06. PMID: 23828150
10. Kennedy JL, Borish L. Chronic rhinosinusitis and antibiotics: the good, the bad, and the ugly. *Am J Rhinol Allergy.* 2013; 27(6):467–72. Epub 2013/11/28. <https://doi.org/10.2500/ajra.2013.27.3960> PMID: 24274221.
11. Settipane RA, Peters AT, Chandra R. Chapter 4: Chronic rhinosinusitis. *Am J Rhinol Allergy.* 2013; 27 Suppl 1:S11–5. Epub 2013/06/05. <https://doi.org/10.2500/ajra.2013.27.3925> PMID: 23711032
12. Manes RP, Batra PS. Bacteriology and antibiotic resistance in chronic rhinosinusitis. *Facial Plast Surg Clin North Am.* 2012; 20(1):87–91. Epub 2011/11/22. <https://doi.org/10.1016/j.fsc.2011.10.010> PMID: 22099621
13. Godoy JM, Godoy AN, Ribalta G, Largo I. Bacterial pattern in chronic sinusitis and cystic fibrosis. *Otolaryngol Head Neck Surg.* 2011; 145(4):673–6. Epub 2011/04/28. <https://doi.org/10.1177/0194599811407279> PMID: 21521896
14. Bhattacharyya N, Kepnes LJ. Assessment of trends in antimicrobial resistance in chronic rhinosinusitis. *Ann Otol Rhinol Laryngol.* 2008; 117(6):448–52. Epub 2008/07/24. <https://doi.org/10.1177/000348940811700608> PMID: 18646442
15. Kingdom TT, Swain RE Jr. The microbiology and antimicrobial resistance patterns in chronic rhinosinusitis. *Am J Otolaryngol.* 2004; 25(5):323–8. Epub 2004/08/31. PMID: 15334396
16. Cushnie TP, Lamb AJ. Recent advances in understanding the antibacterial properties of flavonoids. *Int J Antimicrob Agents.* 2011; 38(2):99–107. <https://doi.org/10.1016/j.ijantimicag.2011.02.014> PMID: 21514796

17. Orhan DD, Ozcelik B, Ozgen S, Ergun F. Antibacterial, antifungal, and antiviral activities of some flavonoids. *Microbiol Res.* 2010; 165(6):496–504. <https://doi.org/10.1016/j.micres.2009.09.002> PMID: 19840899
18. Martens S, Mithofer A. Flavones and flavone synthases. *Phytochemistry.* 2005; 66(20):2399–407. <https://doi.org/10.1016/j.phytochem.2005.07.013> PMID: 16137727
19. Havsteen BH. The biochemistry and medical significance of the flavonoids. *Pharmacol Ther.* 2002; 96(2–3):67–202. PMID: 12453566
20. Seleem D, Pardi V, Murata RM. Review of flavonoids: A diverse group of natural compounds with anti-*Candida albicans* activity in vitro. *Arch Oral Biol.* 2016. <https://doi.org/10.1016/j.archoralbio.2016.08.030> PMID: 27659902
21. Jiang N, Doseff AI, Grotewold E. Flavones: From Biosynthesis to Health Benefits. *Plants (Basel).* 2016; 5(2). <https://doi.org/10.3390/plants5020027> PMID: 27338492.
22. Wu D, Kong Y, Han C, Chen J, Hu L, Jiang H, et al. D-Alanine:D-alanine ligase as a new target for the flavonoids quercetin and apigenin. *Int J Antimicrob Agents.* 2008; 32(5):421–6. <https://doi.org/10.1016/j.ijantimicag.2008.06.010> PMID: 18774266
23. Sato Y, Shibata H, Arai T, Yamamoto A, Okimura Y, Arakaki N, et al. Variation in synergistic activity by flavone and its related compounds on the increased susceptibility of various strains of methicillin-resistant *Staphylococcus aureus* to beta-lactam antibiotics. *Int J Antimicrob Agents.* 2004; 24(3):226–33. <https://doi.org/10.1016/j.ijantimicag.2004.02.028> PMID: 15325425
24. Dao TT, Oh JW, Chi YS, Kim HP, Sin KS, Park H. Synthesis and PGE2 inhibitory activity of vinylated and allylated chrysins analogues. *Arch Pharm Res.* 2003; 26(8):581–4. PMID: 12967189
25. Liu LX, Durham DG, Richards RM. Vancomycin resistance reversal in enterococci by flavonoids. *J Pharm Pharmacol.* 2001; 53(1):129–32. PMID: 11206187
26. Caltagirone S, Rossi C, Poggi A, Ranelletti FO, Natali PG, Brunetti M, et al. Flavonoids apigenin and quercetin inhibit melanoma growth and metastatic potential. *Int J Cancer.* 2000; 87(4):595–600. PMID: 10918203
27. Sato M, Fujiwara S, Tsuchiya H, Fujii T, Iinuma M, Tosa H, et al. Flavones with antibacterial activity against cariogenic bacteria. *J Ethnopharmacol.* 1996; 54(2–3):171–6. PMID: 8953432
28. Hariri BM, Payne SJ, Chen B, Mansfield C, Doghramji LJ, Adappa ND, et al. In vitro effects of anthocyanidins on sinonasal epithelial nitric oxide production and bacterial physiology. *Am J Rhinol Allergy.* 2016; 30(4):261–8. <https://doi.org/10.2500/ajra.2016.30.4331> PMID: 27456596.
29. Suresh Babu K, Hari Babu T, Srinivas PV, Hara Kishore K, Murthy US, Rao JM. Synthesis and biological evaluation of novel C (7) modified chrysins analogues as antibacterial agents. *Bioorg Med Chem Lett.* 2006; 16(1):221–4. <https://doi.org/10.1016/j.bmcl.2005.09.009> PMID: 16213726
30. Ullah Mughal E, Ayaz M, Hussain Z, Hasan A, Sadiq A, Riaz M, et al. Synthesis and antibacterial activity of substituted flavones, 4-thioflavones and 4-iminoflavones. *Bioorg Med Chem.* 2006; 14(14):4704–11. <https://doi.org/10.1016/j.bmc.2006.03.031> PMID: 16603364
31. Patil RH, Babu RL, Naveen Kumar M, Kiran Kumar KM, Hegde SM, Nagesh R, et al. Anti-Inflammatory Effect of Apigenin on LPS-Induced Pro-Inflammatory Mediators and AP-1 Factors in Human Lung Epithelial Cells. *Inflammation.* 2016; 39(1):138–47. <https://doi.org/10.1007/s10753-015-0232-z> PMID: 26276128
32. Hariri BM, McMahon DB, Chen B, Freund JR, Mansfield CJ, Doghramji LJ, et al. Flavones modulate respiratory epithelial innate immunity: anti-inflammatory effects and activation of the T2R14 receptor. *J Biol Chem.* 2017; 292(20):8484–97. *Epub Apr 3.* <https://doi.org/10.1074/jbc.M116.771949> PMID: 28373278
33. Seo HS, Sikder MA, Lee HJ, Ryu J, Lee CJ. Apigenin Inhibits Tumor Necrosis Factor-alpha-Induced Production and Gene Expression of Mucin through Regulating Nuclear Factor-Kappa B Signaling Pathway in Airway Epithelial Cells. *Biomol Ther (Seoul).* 2014; 22(6):525–31. <https://doi.org/10.4062/biomolther.2014.094> PMID: 25489420.
34. Dao TT, Chi YS, Kim J, Kim HP, Kim S, Park H. Synthesis and inhibitory activity against COX-2 catalyzed prostaglandin production of chrysins derivatives. *Bioorg Med Chem Lett.* 2004; 14(5):1165–7. <https://doi.org/10.1016/j.bmcl.2003.12.087> PMID: 14980657
35. Zhang XW, Li WF, Li WW, Ren KH, Fan CM, Chen YY, et al. Protective effects of the aqueous extract of *Scutellaria baicalensis* against acrolein-induced oxidative stress in cultured human umbilical vein endothelial cells. *Pharm Biol.* 2011; 49(3):256–61. <https://doi.org/10.3109/13880209.2010.501803> PMID: 20979538
36. Schrader KK. Plant Natural compounds with antibacterial activity towards common pathogens of pond-cultured channel catfish (*Ictalurus punctatus*). *Toxins (Basel).* 2010; 2(7):1676–89. <https://doi.org/10.3390/toxins2071676> PMID: 22069655.

37. Tan CX, Schrader KK, Khan IA, Rimando AM. Activities of wogonin analogs and other flavones against *Flavobacterium columnare*. *Chem Biodivers.* 2015; 12(2):259–72. <https://doi.org/10.1002/cbdv.201400181> PMID: 25676507
38. Sikder MA, Lee HJ, Mia MZ, Park SH, Ryu J, Kim JH, et al. Inhibition of TNF-alpha-induced MUC5AC mucin gene expression and production by wogonin through the inactivation of NF-kappaB signaling in airway epithelial cells. *Phytother Res.* 2014; 28(1):62–8. <https://doi.org/10.1002/ptr.4954> PMID: 23463646
39. Kim JO, Sikder MA, Lee HJ, Rahman M, Kim JH, Chang GT, et al. Phorbol ester or epidermal growth-factor-induced MUC5AC mucin gene expression and production from airway epithelial cells are inhibited by apigenin and wogonin. *Phytother Res.* 2012; 26(12):1784–8. <https://doi.org/10.1002/ptr.4650> PMID: 22388989
40. Yeh CH, Yang ML, Lee CY, Yang CP, Li YC, Chen CJ, et al. Wogonin attenuates endotoxin-induced prostaglandin E2 and nitric oxide production via Src-ERK1/2-NFKappaB pathway in BV-2 microglial cells. *Environ Toxicol.* 2014; 29(10):1162–70. <https://doi.org/10.1002/tox.21847> PMID: 23362215
41. Yao J, Pan D, Zhao Y, Zhao L, Sun J, Wang Y, et al. Wogonin prevents lipopolysaccharide-induced acute lung injury and inflammation in mice via peroxisome proliferator-activated receptor gamma-mediated attenuation of the nuclear factor-kappaB pathway. *Immunology.* 2014; 143(2):241–57. <https://doi.org/10.1111/imm.12305> PMID: 24766487
42. Yeh CH, Shih HC, Hong HM, Lee SS, Yang ML, Chen CJ, et al. Protective effect of wogonin on proinflammatory cytokine generation via Jak1/3-STAT1/3 pathway in lipopolysaccharide stimulated BV2 microglial cells. *Toxicol Ind Health.* 2015; 31(10):960–6. <https://doi.org/10.1177/0748233713485886> PMID: 23592745
43. Uckoo RM, Jayaprakasha GK, Vikram A, Patil BS. Polymethoxyflavones Isolated from the Peel of Miaray Mandarin (*Citrus miaray*) Have Biofilm Inhibitory Activity in *Vibrio harveyi*. *J Agric Food Chem.* 2015; 63(32):7180–9. <https://doi.org/10.1021/acs.jafc.5b02445> PMID: 26140409
44. Kuroda Y, Ikeda R, Yamazaki T, Ito K, Uda K, Wakabayashi K, et al. Activation of human bitter taste receptors by polymethoxylated flavonoids. *Biosci Biotechnol Biochem.* 2016; 80(10):2014–7. <https://doi.org/10.1080/09168451.2016.1184558> PMID: 27379685
45. Roland WS, van Buren L, Gruppen H, Driesse M, Gouka RJ, Smit G, et al. Bitter taste receptor activation by flavonoids and isoflavonoids: modeled structural requirements for activation of hTAS2R14 and hTAS2R39. *J Agric Food Chem.* 2013; 61(44):10454–66. <https://doi.org/10.1021/jf403387p> PMID: 24117141
46. Roland WS, Vincken JP, Gouka RJ, van Buren L, Gruppen H, Smit G. Soy isoflavones and other isoflavonoids activate the human bitter taste receptors hTAS2R14 and hTAS2R39. *J Agric Food Chem.* 2011; 59(21):11764–71. <https://doi.org/10.1021/jf202816u> PMID: 21942422
47. Lee RJ, Cohen NA. Bitter taste bodyguards. *Sci Am.* 2016; 314(2):38–43. PMID: 26930826
48. Lee RJ, Cohen NA. Taste receptors in innate immunity. *Cell Mol Life Sci.* 2015; 72(2):217–36. <https://doi.org/10.1007/s00018-014-1736-7> PMID: 25323130.
49. Lee RJ, Chen B, Redding KM, Margolskee RF, Cohen NA. Mouse nasal epithelial innate immune responses to *Pseudomonas aeruginosa* quorum-sensing molecules require taste signaling components. *Innate Immun.* 2014; 20(6):606–17. Epub 2013/09/21. <https://doi.org/10.1177/1753425913503386> PMID: 24045336
50. Lee RJ, Kofonow JM, Rosen PL, Siebert AP, Chen B, Doghramji L, et al. Bitter and sweet taste receptors regulate human upper respiratory innate immunity. *J Clin Invest.* 2014; 124(3):1393–405. Epub 2014/02/18. <https://doi.org/10.1172/JCI72094> PMID: 24531552
51. Lee RJ, Cohen NA. Bitter and sweet taste receptors in the respiratory epithelium in health and disease. *J Mol Med (Berl).* 2014; 92(12):1235–44. <https://doi.org/10.1007/s00109-014-1222-6> PMID: 25391251
52. Lee RJ, Cohen NA. The emerging role of the bitter taste receptor T2R38 in upper respiratory infection and chronic rhinosinusitis. *Am J Rhinol Allergy.* 2013; 27(4):283–6. Epub 2013/07/26. <https://doi.org/10.2500/ajra.2013.27.3911> PMID: 23883809
53. Lee RJ, Xiong G, Kofonow JM, Chen B, Lysenko A, Jiang P, et al. T2R38 taste receptor polymorphisms underlie susceptibility to upper respiratory infection. *J Clin Invest.* 2012; 122(11):4145–59. <https://doi.org/10.1172/JCI64240> PMID: 23041624
54. Hamilos DL. Host-microbial interactions in patients with chronic rhinosinusitis. *J Allergy Clin Immunol.* 2014; 133(3):640–53 e4. Epub 2013/12/03. <https://doi.org/10.1016/j.jaci.2013.06.049> PMID: 24290275
55. Brook I. Microbiology of chronic rhinosinusitis. *Eur J Clin Microbiol Infect Dis.* 2016. <https://doi.org/10.1007/s10096-016-2640-x> PMID: 27086363

56. Kavanaugh NL, Zhang AQ, Nobile CJ, Johnson AD, Ribbeck K. Mucins suppress virulence traits of *Candida albicans*. *MBio*. 2014; 5(6):e01911. <https://doi.org/10.1128/mBio.01911-14> PMID: 25389175.
57. O'Malley YQ, Reszka KJ, Spitz DR, Denning GM, Britigan BE. *Pseudomonas aeruginosa* pyocyanin directly oxidizes glutathione and decreases its levels in airway epithelial cells. *Am J Physiol Lung Cell Mol Physiol*. 2004; 287(1):L94–103. Epub 2004/03/17. <https://doi.org/10.1152/ajplung.00025.2004> PMID: 15020296
58. Kanthakumar K, Taylor G, Tsang KW, Cundell DR, Rutman A, Smith S, et al. Mechanisms of action of *Pseudomonas aeruginosa* pyocyanin on human ciliary beat in vitro. *Infect Immun*. 1993; 61(7): 2848–53. Epub 1993/07/01. PMID: 8390405.
59. Hingley ST, Hastie AT, Kueppers F, Higgins ML, Weinbaum G, Shryock T. Effect of ciliostatic factors from *Pseudomonas aeruginosa* on rabbit respiratory cilia. *Infect Immun*. 1986; 51(1):254–62. Epub 1986/01/01. PMID: 3079733.
60. Schwarzer C, Fu Z, Fischer H, Machen TE. Redox-independent activation of NF- κ B by *Pseudomonas aeruginosa* pyocyanin in a cystic fibrosis airway epithelial cell line. *J Biol Chem*. 2008; 283(40): 27144–53. <https://doi.org/10.1074/jbc.M709693200> PMID: 18682396.
61. Schwarzer C, Fischer H, Kim EJ, Barber KJ, Mills AD, Kurth MJ, et al. Oxidative stress caused by pyocyanin impairs CFTR Cl⁻ transport in human bronchial epithelial cells. *Free Radic Biol Med*. 2008; 45(12):1653–62. <https://doi.org/10.1016/j.freeradbiomed.2008.09.011> PMID: 18845244.
62. Parker D, Prince A. Innate immunity in the respiratory epithelium. *Am J Respir Cell Mol Biol*. 2011; 45(2):189–201. Epub 2011/02/19. <https://doi.org/10.1165/rcmb.2011-0011RT> PMID: 21330463
63. Tewfik MA, Latterich M, DiFalco MR, Samaha M. Proteomics of nasal mucus in chronic rhinosinusitis. *Am J Rhinol*. 2007; 21(6):680–5. <https://doi.org/10.2500/ajr.2007.21.3103> PMID: 18201447
64. Dajani R, Zhang Y, Taft PJ, Travis SM, Starner TD, Olsen A, et al. Lysozyme Secretion by Submucosal Glands Protects the Airway From Bacterial Infection. *Am J Respir Cell Mol Biol*. 2005. <https://doi.org/10.1165/rcmb.2005-0059OC> PMID: 15746432
65. Dubin RF, Robinson SK, Widdicombe JH. Secretion of lactoferrin and lysozyme by cultures of human airway epithelium. *Am J Physiol Lung Cell Mol Physiol*. 2004; 286(4):L750–5. <https://doi.org/10.1152/ajplung.00326.2003> PMID: 15003937
66. Duszyk M. CFTR and lysozyme secretion in human airway epithelial cells. *Pflugers Arch*. 2001; 443 Suppl 1:S45–9. <https://doi.org/10.1007/s004240100643> PMID: 11845302
67. Raphael GD, Jeney EV, Baraniuk JN, Kim I, Meredith SD, Kaliner MA. Pathophysiology of rhinitis. Lactoferrin and lysozyme in nasal secretions. *J Clin Invest*. 1989; 84(5):1528–35. <https://doi.org/10.1172/JCI114329> PMID: 2681268
68. Chen PH, Fang SY. Expression of human beta-defensin 2 in human nasal mucosa. *Eur Arch Otorhinolaryngol*. 2004; 261(5):238–41. <https://doi.org/10.1007/s00405-003-0682-z> PMID: 14504864
69. Lee SH, Kim JE, Lim HH, Lee HM, Choi JO. Antimicrobial defensin peptides of the human nasal mucosa. *Ann Otol Rhinol Laryngol*. 2002; 111(2):135–41. <https://doi.org/10.1177/000348940211100205> PMID: 11860065
70. Singh PK, Jia HP, Wiles K, Hesselberth J, Liu L, Conway BA, et al. Production of beta-defensins by human airway epithelia. *Proc Natl Acad Sci U S A*. 1998; 95(25):14961–6. PMID: 9843998
71. Bals R, Wang X, Wu Z, Freeman T, Bafna V, Zasloff M, et al. Human beta-defensin 2 is a salt-sensitive peptide antibiotic expressed in human lung. *J Clin Invest*. 1998; 102(5):874–80. <https://doi.org/10.1172/JCI2410> PMID: 9727055
72. Lee RJ, Foskett JK. Ca²⁺ signaling and fluid secretion by secretory cells of the airway epithelium. *Cell Calcium*. 2014; 55(6):325–36. Epub 2014/04/08. <https://doi.org/10.1016/j.ceca.2014.02.001> PMID: 24703093
73. Lee RJ, Limberis MP, Hennessy MF, Wilson JM, Foskett JK. Optical imaging of Ca²⁺-evoked fluid secretion by murine nasal submucosal gland serous acinar cells. *J Physiol*. 2007; 582(Pt 3):1099–124. <https://doi.org/10.1113/jphysiol.2007.131995> PMID: 17525116
74. Bowes D, Corrin B. Ultrastructural immunocytochemical localisation of lysozyme in human bronchial glands. *Thorax*. 1977; 32(2):163–70. PMID: 325682
75. Klockars M, Reitamo S. Tissue distribution of lysozyme in man. *J Histochem Cytochem*. 1975; 23(12): 932–40. <https://doi.org/10.1177/23.12.1104708> PMID: 1104708
76. Voss JG. Lysozyme Lysis of Gram-Negative Bacteria without Production of Spheroplasts. *J Gen Microbiol*. 1964; 35:313–7. <https://doi.org/10.1099/00221287-35-2-313> PMID: 14179677
77. Levashov PA, Sedov SA, Shipovskov S, Belogurova NG, Levashov AV. Quantitative turbidimetric assay of enzymatic gram-negative bacteria lysis. *Anal Chem*. 2010; 82(5):2161–3. <https://doi.org/10.1021/ac902978u> PMID: 20136093

78. Helander IM, Mattila-Sandholm T. Fluorometric assessment of gram-negative bacterial permeabilization. *J Appl Microbiol.* 2000; 88(2):213–9. PMID: [10735988](#)
79. Alakomi HL, Skytta E, Saarela M, Mattila-Sandholm T, Latva-Kala K, Helander IM. Lactic acid permeabilizes gram-negative bacteria by disrupting the outer membrane. *Appl Environ Microbiol.* 2000; 66(5): 2001–5. PMID: [10788373](#).
80. Johnson L, Mulcahy H, Kanevets U, Shi Y, Lewenza S. Surface-localized spermidine protects the *Pseudomonas aeruginosa* outer membrane from antibiotic treatment and oxidative stress. *J Bacteriol.* 2012; 194(4):813–26. <https://doi.org/10.1128/JB.05230-11> PMID: [22155771](#).
81. Wang J, Chou S, Xu L, Zhu X, Dong N, Shan A, et al. High specific selectivity and Membrane-Active Mechanism of the synthetic centrosymmetric alpha-helical peptides with Gly-Gly pairs. *Sci Rep.* 2015; 5:15963. <https://doi.org/10.1038/srep15963> PMID: [26530005](#).
82. Lv Y, Wang J, Gao H, Wang Z, Dong N, Ma Q, et al. Antimicrobial properties and membrane-active mechanism of a potential alpha-helical antimicrobial derived from cathelicidin PMAP-36. *PLoS One.* 2014; 9(1):e86364. <https://doi.org/10.1371/journal.pone.0086364> PMID: [24466055](#).
83. Babu PB, Chidekel A, Shaffer TH. Protein composition of apical surface fluid from the human airway cell line Calu-3: effect of ion transport mediators. *Clin Chim Acta.* 2004; 347(1–2):81–8. <https://doi.org/10.1016/j.cccn.2004.04.006> PMID: [15313144](#)
84. Platz J, Beisswenger C, Dalpke A, Koczulla R, Pinkenburg O, Vogelmeier C, et al. Microbial DNA induces a host defense reaction of human respiratory epithelial cells. *J Immunol.* 2004; 173(2): 1219–23. PMID: [15240713](#)
85. Berger JT, Voynow JA, Peters KW, Rose MC. Respiratory carcinoma cell lines. MUC genes and glycoconjugates. *Am J Respir Cell Mol Biol.* 1999; 20(3):500–10. <https://doi.org/10.1165/ajrcmb.20.3.3383> PMID: [10030849](#)
86. Shen BQ, Finkbeiner WE, Wine JJ, Mrsny RJ, Widdicombe JH. Calu-3: a human airway epithelial cell line that shows cAMP-dependent Cl⁻ secretion. *Am J Physiol.* 1994; 266(5 Pt 1):L493–501. PMID: [7515578](#)
87. Lee RJ, Workman AD, Carey RM, Chen B, Rosen PL, Doghramji L, et al. Fungal Aflatoxins Reduce Respiratory Mucosal Ciliary Function. *Sci Rep.* 2016; 6:33221. <https://doi.org/10.1038/srep33221> PMID: [27623953](#)
88. Lee RJ, Chen B, Doghramji L, Adappa ND, Palmer JN, Kennedy DW, et al. Vasoactive intestinal peptide regulates sinonasal mucociliary clearance and synergizes with histamine in stimulating sinonasal fluid secretion. *FASEB J.* 2013; 27(12):5094–103. Epub 2013/08/13. <https://doi.org/10.1096/fj.13-234476> PMID: [23934280](#)
89. Joo NS, Lee DJ, Winges KM, Rustagi A, Wine JJ. Regulation of antiprotease and antimicrobial protein secretion by airway submucosal gland serous cells. *J Biol Chem.* 2004; 279(37):38854–60. <https://doi.org/10.1074/jbc.M407077200> PMID: [15234967](#)
90. Zhang Y, Reenstra WW, Chidekel A. Antibacterial activity of apical surface fluid from the human airway cell line Calu-3: pharmacologic alteration by corticosteroids and beta(2)-agonists. *Am J Respir Cell Mol Biol.* 2001; 25(2):196–202. <https://doi.org/10.1165/ajrcmb.25.2.4211> PMID: [11509329](#)
91. Gamet-Payrastre L, Manenti S, Gratacap MP, Tulliez J, Chap H, Payrastre B. Flavonoids and the inhibition of PKC and PI 3-kinase. *Gen Pharmacol.* 1999; 32(3):279–86. PMID: [10211581](#)
92. Ferriola PC, Cody V, Middleton E Jr. Protein kinase C inhibition by plant flavonoids. Kinetic mechanisms and structure-activity relationships. *Biochem Pharmacol.* 1989; 38(10):1617–24. PMID: [2730676](#)
93. Harro JM, Daugherty S, Bruno VM, Jabra-Rizk MA, Rasko DA, Shirtliff ME. Draft Genome Sequence of the Methicillin-Resistant *Staphylococcus aureus* Isolate MRSA-M2. *Genome Announc.* 2013; 1(1). <https://doi.org/10.1128/genomeA.00037-12> PMID: [23405318](#).
94. O'Toole GA. Microtiter dish biofilm formation assay. *J Vis Exp.* 2011;(47). <https://doi.org/10.3791/2437> PMID: [21307833](#).
95. Essar DW, Eberly L, Hader A, Crawford IP. Identification and characterization of genes for a second anthranilate synthase in *Pseudomonas aeruginosa*: interchangeability of the two anthranilate synthases and evolutionary implications. *J Bacteriol.* 1990; 172(2):884–900. PMID: [2153661](#).
96. Prokhorenko IR, Zubova SV, Ivanov AY, Grachev SV. Interaction of Gram-negative bacteria with cationic proteins: Dependence on the surface characteristics of the bacterial cell. *Int J Gen Med.* 2009; 2:33–8. PMID: [20360884](#).
97. Widdicombe JH. Regulation of the depth and composition of airway surface liquid. *J Anat.* 2002; 201(4): 313–8. <https://doi.org/10.1046/j.1469-7580.2002.00098.x> PMID: [12430955](#).
98. Lai Y, Chen B, Shi J, Palmer JN, Kennedy DW, Cohen NA. Inflammation-mediated upregulation of centrosomal protein 110, a negative modulator of ciliogenesis, in patients with chronic rhinosinusitis. *J*

- Allergy Clin Immunol. 2011; 128(6):1207–15 e1. Epub 2011/10/11. <https://doi.org/10.1016/j.jaci.2011.09.001> PMID: 21982113
99. Shah AS, Ben-Shahar Y, Moninger TO, Kline JN, Welsh MJ. Motile cilia of human airway epithelia are chemosensory. Science. 2009; 325(5944):1131–4. Epub 2009/07/25. <https://doi.org/10.1126/science.1173869> PMID: 19628819