

Interactions between nutrition and reproduction in the management of the mature male ruminant

G. B. Martin^{1,2†}, D. Blache^{1,2}, D. W. Miller³ and P. E. Vercoe^{1,2}

¹UWA Institute of Agriculture M082, The University of Western Australia, Crawley, Western Australia 6009, Australia; ²School of Animal Biology, The University of Western Australia, Crawley, Western Australia 6009, Australia; ³School of Veterinary and Biomedical Sciences, Murdoch University, Murdoch, WA 6150, Australia

(Received 30 September 2009; Accepted 5 November 2009; First published online 7 January 2010)

In mature male sheep and goats, changes in feed intake seem to have little effect on gonadal endocrine function but induce profound changes on sperm production. These outcomes are due to changes in size of the seminiferous tubules and in spermatogenic efficiency. Except with severe underfeeding, there are only minor changes in the endocrine function of the testis (testosterone production) unless season-long treatments are imposed. For cattle, nutrition clearly affects testicular development and the production of spermatozoa in young bulls, as it does in other species but, after the period of rapid growth has ended, there appears to be little or no response to nutrition. We are developing a clear picture of the metabolic signals, neuroendocrine processes and hormonal control systems that are involved, particularly for the mature male sheep. The energetic components of the diet, rather than protein, seem to be responsible, so we have envisaged a model of the relationship between energy balance and reproduction that has 4 'dimensions': genotype, structure (organs), communication (chemical and neural signals, nutrient sensing) and time (dynamics, metabolic memory, programming). We have linked these perspectives to 'resource allocation theory' and incorporated them into strategies for 'clean, green and ethical animal production'. In contrast to the clear outcomes with respect to spermatogenesis, the effects of nutrition on sexual behaviour are more difficult to define, perhaps because the behaviour is affected by a complex mix of physiological factors and because of flawed methods for quantifying male behaviour. For example, sexual behaviour is compromised by severe feed restriction, but male sexual behaviour requires intensive motor activity so a decline in libido could be caused by general weakness rather than specific nutritional limitations. The interaction between sexual activity and feeding behaviour also complicates the issue under field conditions. At the other end of the scale, overweight males can show reduced sexual success because they have difficulty courting and mounting. For this reason, exercise can enhance the fertilising capacity of rams. This will be important in extensive mating systems where males need to assemble and guard a harem and then mate many times for several weeks. For artificial insemination centres, there seems to be very few data on the nutritional management of males, but problems with overfed animals appear to be a risk. Future research should concentrate on the intra-testicular systems mediating the effects of nutrition on the production of spermatozoa.

Keywords: testis, spermatogenesis, nutrition, hormones (reproductive, metabolic), behaviour

Implications

In mature rams and bucks, but not bulls, nutrition exerts major effects on testicular mass. The change in mass reflects changes in the amount of seminiferous tissue and thus spermatogenic capacity. Spermatogenic efficiency is also altered, so the overall outcome is a major change (up to 250%) in the rate of production of spermatozoa. Except for extreme undernutrition, these effects are not linked to major changes in testosterone production or in sexual behaviour. Male small ruminants

should thus be fed correctly for 2 months before mating. This is a critical component of 'clean, green and ethical' systems of animal production.

Introduction

The ruminants developed seasonal reproductive strategies in response to the pressures of natural selection imposed by their environment for millions of years, long before humans began to domesticate them in the pre-dawn of the agricultural age. Despite 11 000 years of environmental management and directed breeding, many of the characteristics

† E-mail: Graeme.Martin@uwa.edu.au

of these original strategies are still with us. One of the most persistent is sensitivity to nutritional inputs, primarily because an adequate supply of nutrients, from body reserves or from diet, is absolutely essential for lactation. Avoiding nutritional scarcity during this critical stage of the mammalian reproductive process becomes a challenge because the supply of forage varies greatly throughout the year. In many regions, the periods of scarcity are predictable from year to year, but the actual season of scarcity might be winter (Temperate regions), autumn (Mediterranean regions) or spring (some Sub-tropical regions). Semi-arid and arid regions offer the greatest challenge because the timing is unpredictable for the peaks and troughs in forage supply. Considering this variety of environments and the fact that wild and domesticated ruminant species are found in all of them, it is no surprise that the problem of adequate nutrient supply at lactation has been solved by a variety of reproductive strategies, particularly when the duration of pregnancy varies as much as it does between the large and small ruminants.

In temperate regions, a spring birth has a very clear and consistent advantage, so photoperiodic time measurement, because it is the most reliable way to determine whether winter or summer is approaching, exerts a very powerful influence over reproductive activity (Lincoln and Short, 1980). However, the majority of the world's sheep and goats live between 35 degree North and 35 degree South (Lindsay, 1991), where the photoperiod-driven breeding patterns that characterise these species are often poorly co-ordinated with changes in forage supply (Sutama and Edey, 1985; Martin *et al.*, 2002). In these environments, many mammals have developed 'opportunistic' reproductive strategies that can cope with the problem (Bronson, 1985). The control systems of their reproductive axis respond to photoperiod, but they are more holistic and flexible than those of animals from temperate regions because they are heavily influenced by other inputs, particularly nutrition and socio-sexual stimuli.

Our studies extended the concept developed by Bronson (1985) to two ruminants that have been introduced to Australia, both of which exhibit a considerable degree of opportunism – the Merino sheep (of Mediterranean origins) and the Australian cashmere goat (recently evolved from the feral goat population). The role of socio-sexual stimuli in the opportunistic breeding strategies of these animals has been reviewed elsewhere (Walkden-Brown *et al.*, 1999). In this review, we will deal with the nutritional inputs into the processes that control reproductive function. We will focus primarily on the mature males and the endpoint will be the gametogenic and endocrine activity of the testes. Although our own studies are largely limited to the Australian goats and sheep, we will integrate them with work on other genotypes and environments, and we will also address related issues for cattle. In all cases, we will consider how this information affects animal industry.

Foetal life and puberty

In extensive management systems, pregnant females are often forgotten between mating and the final few weeks of

pregnancy because it has traditionally been thought that there are no serious demands on their reserves until the foetus reaches a significant size. However, this ignores two major factors: (i) the role of metabolic inputs into embryo and foetal development ('foetal programming'); (ii) the nutritional requirements for the development of the placenta (review: Bell, 1984). In both cases, the consequences of nutritional mis-management may not become evident until much later, after birth or even after sexual maturity, but are associated with poor life-time performance (review: Bell, 1984). The life-time productivity of the tissues that provide commodities (skin follicles; precursor muscle cells; mammary tissue) can be compromised, as can the development of reproductive tissues (review: Martin *et al.*, 2004). Thus, male lambs born to ewes that were underfed during the second half of pregnancy have fewer Sertoli cells in their testes at birth (Bielli *et al.*, 2002), and retardation of intrauterine growth can delay the onset of puberty in male lambs (Da Silva *et al.*, 2001). Further investigation is needed but it seems likely that the outcome in both cases would be a reduction in spermatogenic capacity in adult life.

The roles of postnatal nutrition in progress towards puberty in male farm animals has been reviewed by Brown (1994). We will focus on mature animals for the remainder of this review, but we would like to acknowledge the major contributions in recent years from the laboratory of John Kastelic who have studied metabolic input into pubertal development of young bulls, and have clearly demonstrated that superior calfhood nutrition, probably through the action of metabolic hormones, promotes gonadotrophin secretion and thus testicular growth, and an earlier onset of spermatogenesis (see Barth *et al.*, 2008). There is very broad agreement between these observations and our own on sexually mature male sheep, as we shall see below.

Mature males – testicular responses to nutrition

Nutritional influences on reproduction in farm animals were probably recognised soon after domestication. In the historical overview preceding his study, Clark (1934) was prepared to cite Aristotle and Darwin has having considered the concept, but gave Walter Heape the credit for providing the first experimental evidence of 'flushing' in English sheep in 1899. However, it was Clark (1934) himself who showed that flushing increased lambing rate by increasing ovulation rate. This phenomenon has long intrigued reproductive physiologists and continues to dominate research on the nutritional influences on reproduction to this day. By contrast, relatively little attention was paid to the male. It was recognised that the production of gametes by mature males is affected by nutrition but, after reviewing the ruminant literature, Moule (1963) was led to conclude: '...so far no workers seem to have attempted a systematic study of the overall effects of nutrition on semen production in male domestic animals'. There seemed to be no literature at all on male goats. For mature bulls, Moule missed the work of Flipse and Almquist (1961), leaving the study of a

single 5-year-old bull by Mann and Walton (1953) who found that undernutrition reduced the concentrations of fructose and citrate in seminal plasma but did not affect the sperm output.

The most interesting of the earlier work on mature animals was perhaps that of Akira Mori, who carried out a controlled experiment on large groups of severely underfed mature rams in Japan during the Second World War. When his work was published many years later, it showed that semen quality and sperm production were improved within a month or 2 months by supplements that led to a weight gain (Mori, 1959). Mori was convinced that protein shortage had caused the infertility in his flocks, but he did not test the concept formally with nutritionally rigorous diets. This sort of problem, and the worry expressed by Moule (1963) that many experiments on 'nutrition' concerned only pathological effects, requires us to define the term as we shall use it in this review. Deficiencies in trace elements (e.g. zinc, Martin *et al.*, 1994b) or vitamins (Robinson *et al.*, 2006), heavy metal toxicities, or imbalances in essential amino or fatty acids, have long been known to affect male reproductive function adversely (Leatham, 1975), but for the present review, we need to constrain the discussion as much as possible to shifts in energy or protein balance of similar magnitude and duration to those that would be experienced by animals grazing natural forage. Generally, we will avoid the extremes of this range too, some of which lead to total gonadal dysfunction (the animals studied by Mori (1959) must have been close to this). Similarly, we will not consider responses to fasting because of the confounding effects of stress.

Nutrition and the production of spermatozoa

The study by Mori (1959) included Merino rams and his data were soon supported by two further studies with this breed. Salamon (1964) used high-frequency semen collections and Setchell *et al.* (1965) used a histological assessment of the seminiferous tubules and epididymis, but they both came to the same conclusion – the daily rate of sperm production is reduced by underfeeding and improved by refeeding. Importantly, the quality of the semen produced, measured as sperm count and sperm motility, is also decreased by undernutrition for a period of time greater than the 7-week duration of spermatogenesis (Parker and Thwaites, 1972; Robinson *et al.*, 2006). Goats were studied by Hiroe and Tomizuka (1965), who also found that loss of body mass was associated with reductions in the output of spermatozoa. There were some negative findings too (Tilton *et al.*, 1964), but subsequent studies (Parker and Thwaites, 1972; Braden *et al.*, 1974; Oldham *et al.*, 1978; Alkass *et al.*, 1982; Martin *et al.*, 1987; Cameron *et al.*, 1988) led to the wide acceptance of strong, direct relationships between plane of nutrition, testicular mass and the number of spermatozoa available for ejaculation, for the small ruminants at least (Figure 1).

For cattle, the picture is more complex. Most of the older literature describes experiments with young *Bos taurus*

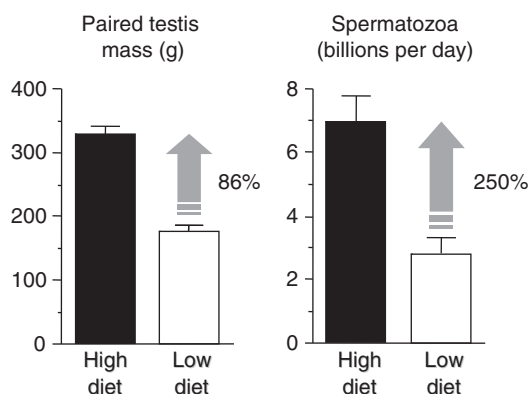


Figure 1 Effect of nutrition on testicular growth and the production of spermatozoa in 1-year-old Merino rams ($P < 0.01$ for both). The diets led to liveweight gains of 17.2 kg (High) and -5.8 kg (Low) after 9 weeks ($P < 0.001$). Semen was collected daily with an artificial vagina and the number of spermatozoa per ejaculate was averaged for the ninth week. The rate of production of spermatozoa was calculated from numbers of stages VI, VII and VIII spermatids in testicular homogenates following castration at the end of the experiment. Testicular mass was determined after slaughter. Redrawn after Cameron *et al.* (1988).

bulls that are up to 2 years old and post-pubertal but still growing rapidly (review Brown, 1994). In such animals, nutrition clearly affects testicular development and the production of spermatozoa in much the same way as it does in mature sheep and goats (Davies *et al.*, 1957; Flipse and Almquist, 1961; Van Demark and Mauger, 1964; Van Demark *et al.*, 1964; Gauthier and Berbigier, 1982). The same applies to *Bos indicus* bulls (Rekwot *et al.*, 1987; Tegegne *et al.*, 1992). As mentioned above, the Kastelic laboratory has added an extra level of sophistication to our understanding of the effects of nutrition over the first 18 months of life (review: Barth *et al.*, 2008). However, after the period of rapid growth has ended, there appears to be little or no response to nutrition (Flipse and Almquist, 1961; Van Demark and Mauger, 1964; Van Demark *et al.*, 1964; Coulter *et al.*, 1987). Caution is needed with these older studies on mature bulls. For example, in the experiment by Van Demark and colleagues, the 'low' plane of nutrition had little effect on body weight, as well as testicular size, even over 12 months, although production of spermatozoa was reduced (Van Demark and Mauger, 1964; Van Demark *et al.*, 1964). In the other cases, the 'high' planes seemed to constitute overfeeding, as the sperm production was markedly decreased compared to 'medium' or 'low' planes (Coulter *et al.*, 1987; Mwansa and Makarechian, 1991). In addition, the energy and protein balance of the experimental diets requires clarification, as both have been manipulated in these studies, but not compared in the one experiment.

Nutritional effects on spermatogenic tissues

In rams, changing nutrition alters not only the total mass of testicular tissue, but also the efficiency with which the gametes are produced by that tissue. This is evident from

the fact that proportional changes in the sperm production are consistently greater than changes in testicular size. For example, Oldham *et al.* (1978) found that a 25% increase in testicular size led to an 81% increase in the production of spermatozoa and Cameron *et al.* (1988) found that an 86% increase in testicular size led to a 250% increase in the production of spermatozoa (Figure 1). The number of ejaculated spermatozoa is not affected until 7 weeks after a change in diet, suggesting that the effects on spermatogenic efficiency are exerted after the last spermatogonial division. This is similar to the effects of stimulatory photoperiod in more seasonal breeds of sheep, where efficiency is increased by reducing the rate of degeneration (presumably apoptosis) of germ cells following the mitotic and meiotic divisions of the spermatogenic cycle (Hochereau-de Reviers *et al.*, 1985). From histological studies it was clear that nutrition affects the diameter of the seminiferous tubules, the relative proportion of the testis occupied by the seminiferous tubules and the proportion of the seminiferous tubule occupied by the seminiferous epithelium (Setchell *et al.*, 1965; Oldham *et al.*, 1978).

In a detailed study, Hötzel *et al.* (1998) also showed that seminiferous tubule length and diameter were higher in well-fed rams than in underfed rams. Their most challenging finding was that the number of Sertoli cell nuclei per testis was also affected (well-fed $120 \pm 6 \times 10^8$; underfed $77 \pm 7 \times 10^8$; $P < 0.001$), whereas the proportion of testis occupied by Sertoli cell nuclei was not. This is consistent with changes in the spermatogenic function of the testis, particularly the increase in spermatogenic efficiency, but it is in conflict with the notion that Sertoli cell number is established before puberty in all species, including the ram (Monet-Kuntz *et al.*, 1984) and goat (Hochereau-de Reviers *et al.*, 1986). However, there had been several reports of changes in Sertoli cell number after puberty (see Hötzel *et al.*, 1998) and, recently, Sarah Meachem and her colleagues have addressed this question directly. For hamsters, they provided solid evidence for seasonal changes and have proposed that adult Sertoli cells are not terminally differentiated. Their view is that Sertoli cells can enter a transitional state exhibiting features common to both undifferentiated and differentiated Sertoli cells (Tarulli *et al.*, 2006).

Nutritional effects on testicular interstitial tissues

Hötzel *et al.* (1998) found that the number of Leydig cells per testis in the ram was not affected by diet, but the total volume of Leydig cells per testis was 30% higher in well-fed rams than in underfed rams, indicating a change in the volume of individual cells. This is in broad agreement with the study by Setchell *et al.* (1965) and suggests that the rate of production of testosterone should also be affected. This should be reflected in circulating testosterone concentrations, especially when combined with changes in the testicular blood flow. Blood flow through the testis is closely related to testis mass and thus changes in response to nutrition (Setchell *et al.*, 1965). The relative volume of

blood and lymph vessels was not affected by diet in the study by Hötzel *et al.* (1998), but their absolute volumes increased or decreased as a function of the change in testicular size. These changes in the vasculature and the associated changes in blood flow, accommodate the transport of testosterone to the peripheral circulation (Setchell, 1986 and 1990). Thus, the present data are in agreement with a previous nutritional study in mature rams (Setchell *et al.*, 1965) and suggest that total blood flow was affected while there was probably little or no change in blood flow per unit mass of testis. As a consequence, larger amounts of testosterone produced by the testis would have reached the peripheral circulation in rams fed with the high diet than in those fed with the low diet. In assessing the literature on this topic, an important consideration is the severity and duration of the dietary treatments. For example, Setchell *et al.* (1965) compared rams that were severely underfed for 3 months (body fat was reduced to less than 12% of live mass) with rams that were arguably obese (25% to 49% fat). In the study by Hötzel *et al.* (1998), similar conditions were imposed. Mature cashmere bucks require about 4 months of dietary treatment before testosterone production is affected (Walkden-Brown *et al.*, 1994). Differences in severity and duration explain the disagreements between these studies and other study showing no effect of nutritional treatments on peripheral testosterone concentrations (Ritar *et al.*, 1984; Martin *et al.*, 1994a).

In bulls, the effects of nutrition on testosterone production were clear from the earliest studies based on measurement of fructose and citrate concentrations in seminal plasma (indicators of stimulation of the seminal vesicles by testosterone). From their study of a single mature bull, Mann and Walton (1953) concluded that undernutrition reduced testosterone production and this was supported by Davies *et al.* (1957), although their data was derived from younger animals. Subsequently, these effects in growing bulls were confirmed by direct measurement of testicular testosterone content (Mann *et al.*, 1967) and plasma testosterone concentrations (Gauthier and Berbigier, 1982).

Nutrition and the expression of sexual behaviour in mature males

Despite the clear, mechanistic evidence of links between nutrition and reproductive endocrinology, there is some controversy on the effect of undernutrition on the expression of sexual behaviour of male sheep, mainly due to the methods used to assess libido and the frequency of observations (Parker and Thwaites, 1972). If feed restriction is sufficiently long and severe, such that over 30% of body mass is lost, the expression of normal sexual behaviour is compromised (Parker and Thwaites, 1972). However, sexual behaviour requires fairly intensive motor activity (Banks, 1964) so a decline in libido could be caused by a general weakness in underfed rams rather than energy restriction (Tilbrook and Cameron, 1990). In fact, during mating, motor activity is greater in rams than in ewes, because of the very

active part taken by the rams during courtship (Banks, 1964). Interestingly, it is also greater in rams with smaller testes than in rams with larger testes (Raadsmas and Edey, 1985), suggesting that the cost of reproduction is greater in animals that produce less spermatozoa. Overall, it appears that libido is more sensitive to undernutrition than sperm production but increases in the plane of nutrition stimulate sperm production before affecting libido.

Energy balance is, of course, a reflection of energy expenditure as well as energy intake. Thus, exercise affects testicular size (Thwaites, 1995) and rams mating under field conditions show a loss of testicular weight, a reduction in the sperm production, and increase in proportion of inter-tubular tissue, a decrease in proportion of spermatocytes, round spermatids and decrease in number of spermatozoa in the epididymus (Knight *et al.*, 1987). This might explain why rams have better reproductive performance when kept in intensive systems than when kept in extensive systems (Fourie *et al.*, 2004). However, overfed males show reduced sexual activity possibly because they are overweight and have difficulty expressing courtship or mounting (Okolski, 1975). Thus, regular exercise can increase the fertilising ability of Merino rams mating under field conditions (Combrink and Schoeman, 1993).

Physiological mechanisms

Nutrition affects male reproductive function in two types of physiological process: (a) through the metabolic and reproductive centres of the brain, leading to a change in the output of gonadotrophin-releasing hormone (GnRH); (b) through pathways that seem to be independent of changes in GnRH secretion (Hötzel *et al.*, 1995). The outcome of the combined pathways is the change in testicular mass, primarily seminiferous tissue and in the efficiency of spermatogenesis as described above. We now have a strong understanding of the metabolic inputs to the brain-gonadal axis (nutrients, metabolites and substrates) as well as the endocrine signals from metabolic and storage tissues.

Setchell *et al.* began unravelling the physiological processes by describing the dependence of testicular metabolism on glucose and found that underfeeding reduced the metabolic activity and blood flow of the testis (Setchell *et al.*, 1965; Setchell and Hinks, 1967). They also found that testosterone output was reduced and supported the suggestion from work on rats that the pituitary gonadotrophins had a role in nutritional responses (the 'pseudohyphysectomy' mechanism of the 1940s). However, few other data were available when we began work in this area, arguably no surprise considering how few studies had been done and the limitations of the endocrine techniques of the time.

Progress depended on the availability of an efficient experimental model. We were greatly aided by the availability of an acute nutritional stimulus that would elicit physiological responses – lupin grain, a highly digestible source of energy and protein that had low concentrations of soluble carbohydrates so could be acutely fed to ruminants

in large quantities (e.g. 1 kg/day) without causing acidosis. We combined this treatment with the mature Merino ram, an animal that showed rapid, repeatable testicular responses to changes in food supply at all times of the year. Finally, we are able to use the strong linear relationship between testicular size and the rate of production of spermatozoa to estimate the gametogenic potential of the testis (Amann, 1970; Knight, 1977).

A multi-dimensional control system based on energy homeostasis

Here, we deal with the relationship between metabolic status and the activity of the system that controls the central driver of reproductive function, pulsatile GnRH secretion. We reflect on these relationships within the framework of a model that comprises four interdependent 'dimensions': (i) genetic, (ii) structural, (iii) communicational and (iv) temporal. To illustrate these concepts, we will mainly use the male sheep that has been acutely placed on a high plane of nutrition. Most of the processes involved are autonomic by nature but, to aid clarity of expression, we will occasionally risk the use of anthropomorphic language (e.g. 'decision to reproduce'). These concepts have been presented in detail previously in a cross-model comparison in which we used information from mature male sheep and *post-partum* dairy cows (Blache *et al.*, 2007). Therefore, here, we will cover the topics briefly.

Bioenergetics of reproduction – the concept of 'metabolic status'

Energy costs are attached to all components of the reproductive process, from the expression of specific behaviours, such as sexual or maternal behaviour, to the production of morphological elements, such as gametes, fetuses and milk (for a full review, see Martin *et al.*, 2008). Considering the energetic requirements, the relationship between the metabolic and the reproductive regulatory systems needs to be highly tuned if the probability of success is to be reasonable. Thus, the regulatory processes that link nutrition and reproduction are largely the same regulatory processes that control energy homeostasis.

At any given time, for any given animal, the amount of energy available for reproduction depends on the difference between the amount of energy expended, including the demands for maintenance, and the pool of disposable energy. The pool of disposable energy includes the energy derived from feed intake plus the energy stored in body tissues, especially adipose tissue, liver and muscle. Most authors refer to 'energy balance' but we prefer 'metabolic status' because it includes a strong integrative dimension.

Dimension 1 – the genetic dimension

In male sheep, the effects of metabolic status and dietary manipulation on the reproductive axis differ between genotypes. This genetic dimension accounts for variations in the responses to all environmental inputs. We have previously

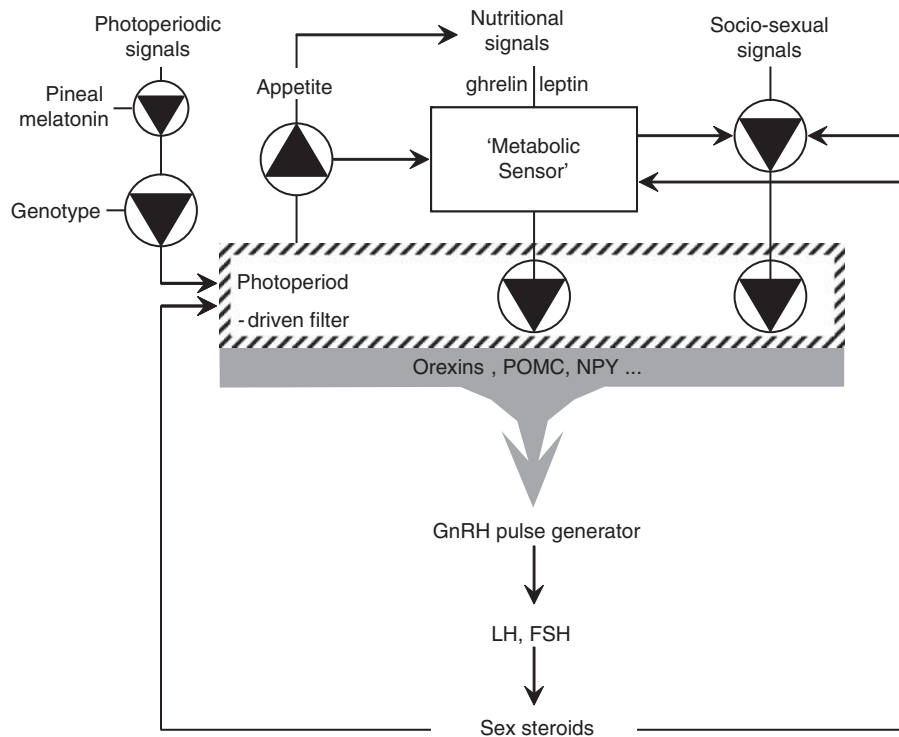


Figure 2 A schema describing the proposed relationships between photoperiodic, nutritional and social cues and the ways that they interact with genotype and steroid feedback in the control of hypothalamic-pituitary-testicular axis in the male sheep. Nutritional input is via ‘metabolic status’, a reflection of the difference between energy expenditure and the sum of energy available from food intake and from energy reserves, as measured by a hypothetical ‘metabolic sensor’. Redrafted after Blache *et al.* (2003).

suggested that photoperiod acts as a ‘filter’ of the effect of nutrition on the reproductive system and that this filtering effect depends on the genotype of the animal (Blache *et al.*, 2003; Figure 2). This is because, in the ram, the effect of nutrition on the reproductive endocrine axis is smaller in breeds that are very responsive to photoperiod (e.g. Suffolk) than in breeds (e.g. Merino) that are less responsive (Martin *et al.*, 1999 and 2002; Hötzel *et al.*, 2003). Interestingly, two lines of evidence suggest that this genetic influence is exerted at the very top of the hierarchy of the reproductive control system – first, pineal function does not differ greatly between Suffolk and Merino rams (Martin *et al.*, 1999 and 2002); second, studies with monozygotic twin sheep suggest that most variation in the activity of the hypothalamic-pituitary axis has environmental causes (Celi *et al.*, 2007).

Dimension 2 – the structural dimension

The brain and the gonads have always been seen as the primary targets for nutritional input into reproductive function but we now accept major roles for the pancreas, liver and adipose tissue (Figure 3). For adipose tissue, this transformation has been spectacular, as it has been elevated from a passive storage site to a vital endocrine organ that produces a number of signals (detailed below). Similarly, the digestive system is also now implicated in the regulatory processes through which nutrition affects reproduction as its endocrine output has been detected and identified. The digestive system also produces very direct metabolic information in the form

of energy metabolites (glucose, fatty acids) and amino acids (detailed below). Effectively, every tissue involved in the regulation of metabolic status is also involved in the communication dimension of the system that regulates reproductive activity.

Dimension 3 – the communication dimension

The systems that regulate the reproductive axis need to be able to respond accurately to changes in metabolic status. This is accomplished through communication processes that are endocrine, neural and nutrient-based.

The GnRH neuroendocrine system – the fundamental controller of reproduction: the secretion of GnRH by the hypothalamus, and consequently luteinizing hormone (LH) and follicle-stimulating hormone (FSH) from the pituitary gland, are central to the reproductive process (review: Martin, 1984). There is a threshold frequency of GnRH pulses above which males will produce sperm and females will ovulate so, effectively, the ‘decision’ of an animal to reproduce or not is implemented through pathways, as yet not described, that control the production of GnRH. The GnRH neuroendocrine system is also the final common pathway via which gonadal activity is usually influenced by external factors, including socio-sexual signals and photoperiodic cues, as well as metabolic status (Figure 2; reviews: Blache *et al.*, 2000, 2002, 2003 and 2006). In mature male sheep, an acute increase in the intake of energy and protein induces, first, an increase in the frequency of pulses of

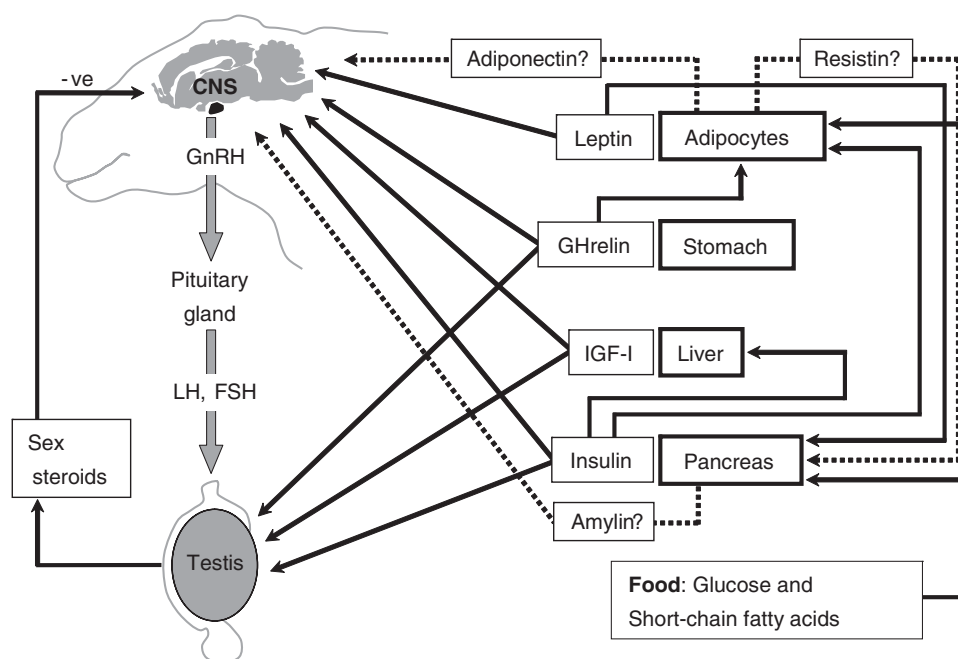


Figure 3 Schematic summary of the potential relationships among the endocrine and neural inputs into the systems that control the reproductive system and mediate the responses to change in metabolic status. For clarity, we have omitted some hormonal systems (growth hormone (GH), prolactin and oxytocin) that are also involved in the control of the metabolic status but do not appear to exert any direct action on gonadotrophin-releasing hormone (GnRH) secretion.

GnRH and LH, and then an increase in the tonic secretion of FSH; an acute reduction of feed intake has the opposite effects (Martin *et al.*, 1994a). That said, it is important to note that tissues central to the control of reproduction, including the pituitary gland and gonads, can also respond independently to metabolic inputs, if not to the extent of switching the reproductive process on or off.

Insulin. Insulin is affected by energy balance and food intake and seems to be involved in the control of reproduction in male sheep because: (i) following an acute increase in dietary allowance, the increase in insulin secretion coincides with the increase in LH pulse frequency (Zhang *et al.*, 2004); (ii) a high plane of nutrition leads to high concentrations of insulin in both plasma and cerebrospinal fluid (CSF) (Miller *et al.*, 1998, Zhang *et al.*, 2004 and 2005); (iii) with a restricted diet or with diabetes, a low dose of insulin infused into the third ventricle increases LH pulse frequency to values similar to those seen in well-fed animals (Miller *et al.*, 1995 and 2002; Tanaka *et al.*, 2000); (iv) insulin receptors are present in the ram hypothalamus (Blache *et al.*, 2002); (v) When GnRH/LH secretion is stimulated by an increase in nutrition, the response is associated with increased amounts of circulating insulin entering the brain, a decrease in hypothalamic expression of the orexigenic neuropeptides, neuropeptide-Y and agouti-related peptide, and an increase in the expression of the anorexigenic peptides pro-opiomelanocortin and cocaine- and amphetamine regulated transcript (Miller *et al.*, 2007). A role for this group of regulatory pathways is supported by studies in other species showing that insulin

receptors in the hypothalamus co-localize with all four peptides (Schwartz, 2006).

Growth hormone (GH) and insulin-like growth factor-1 (IGF-1). Energy balance certainly affects plasma concentrations of GH under a wide variety of conditions but, in male sheep at least, an increase in nutrition induces a decrease in plasma GH concentrations (Miller *et al.*, 1998). Circulating concentrations of IGF-1 are also affected by diet, but concentrations of IGF-1 in CSF are not (Miller *et al.*, 1998) and we have not been able to demonstrate that IGF-1 infusion into the third ventricle affects LH pulse frequency (Blache *et al.*, 2000). It therefore seems likely that, in male sheep at least, IGF-1 is not heavily involved in responses of the critical reproductive centres to changes in nutritional status.

Thyroid hormones. In the mature male sheep, the concentrations of thyroid hormones in plasma and CSF are not affected by the acute elevations of nutrition that increase LH pulse frequency, suggesting that they play no role in the stimulation of GnRH activity (Miller *et al.*, 1998; Zhang *et al.*, 2004 and 2005).

Adipocyte hormones. Adipose tissue was once considered as a passive energy reserve but is now seen as playing an active role in the regulation of food intake, metabolism, immunity, thermoregulation and cardiovascular function, as well as reproduction. Of the 20 or more endocrine products of adipose tissue, leptin appears to exert the most influence over the reproductive axis in ruminants (review: Chilliard

et al., 2005), although future research might reveal key roles for the others. Both short- and long-term changes in metabolic status alter the expression and release of leptin and the sensitivity of gonadal and brain tissues to it, and numerous experiments in female and male sheep have shown that leptin can affect the reproductive neuroendocrine control system (review: Adam *et al.*, 2003; Chilliard *et al.*, 2005). However, the consensus seems to be that leptin plays permissive rather than triggering roles.

Two other adipose hormones, adiponectin and resistin, have yet to receive the same interest as leptin but perhaps should do so because they are exclusively produced by adipose tissue. In contrast to leptin, adiponectin stimulates energy expenditure without any effect on feed intake in rats when it is infused into the cerebral ventricle (Ahima, 2005). In male rat pituitary cells in culture, adiponectin reduces the expression of GnRH receptor and decreases the secretion of LH (Malagon *et al.*, 2006). Resistin has been named as such it seems to mediate insulin resistance (Steppan *et al.*, 2001) and the gene encoding for it has been localised to the rat testis, where its expression is controlled by nutrition, suggesting a potential direct action on the gonads (Nogueiras *et al.*, 2003 and 2004). With respect to ruminants, the resistin gene has been localised to the mammary gland and adipocytes of the cow (Komatsu *et al.*, 2003). It has been proposed that neither resistin nor adiponectin act directly on the GnRH cells or the neuropeptide systems that control GnRH secretion. Instead, resistin inhibits insulin secretion while adiponectin stimulates insulin secretion (Figure 3). Therefore, the balance of secretion of these two adipocyte hormones controls inputs by insulin, and consequently leptin, into the neuropeptidergic regulation of both food intake and reproduction (review: see Budak *et al.*, 2006).

Gastro-intestinal hormones. A most interesting potential candidate in the regulation of reproduction by metabolic status is ghrelin, a gut hormone that is the endogenous ligand of the growth hormone secretagogue (GHS) receptor. In rodents, the distribution of GHS receptors overlaps that of GnRH in the arcuate nucleus (St-Pierre *et al.*, 2003) and ghrelin decreases the secretion of LH (Fernandez-Fernandez *et al.*, 2004). However, in sheep, ghrelin secretion is inhibited by an increase in feed intake and stimulated by fasting (Sugino *et al.*, 2004). Interestingly, ghrelin and GHS receptors are expressed in human and rat testis (Tena-Sempere, 2005) and ghrelin is found in most reproductive tissues in both male and female sheep (Miller *et al.*, 2005). Clearly, further work is required.

Direct nutrient inputs

Many products of digestion enter the circulation and can act as signals in their own right, regulating the secretion of GnRH pulses or perhaps acting directly on the gonad to promote gamete and hormone production. In rams, for example, the volatile fatty acids, a major currency in energy transactions in ruminants, stimulate GnRH secretion (Boukhliq and Martin, 1997; Boukhliq *et al.*, 1997). The role of glucose is

less clear – we detected no GnRH response to intra-abomasal or intravenous infusions of glucose in rams (Miller *et al.*, 1995; Boukhliq *et al.*, 1996 and 1997). This suggests that there are nutritional inputs, perhaps some form of 'nutrient sensing or signalling' that are not associated with whole body energy status but can nevertheless stimulate the hypothalamic-pituitary-gonadal axis. At gonadal level, the ovarian follicles of sheep respond strongly to direct inputs from glucose, fatty acids and a number of metabolic hormones, and there are local intra-follicular nutrient sensing and integrative pathways and mechanisms that modulate gonadotrophin-stimulated folliculogenesis and regulate ovulation rate (review: Scaramuzzi *et al.*, 2006). This concept has not been tested for the testis of the sheep.

Interactions among the elements of the communication dimension

At any given time, metabolic status depends on the status of all three compartments of energy balance – intake, storage and expenditure – and the various hormonal systems that they control or influence (Figure 3). These interactions are perhaps best exemplified by leptin secretion because it is affected by intake and expenditure as well as storage (mass of adipose tissue). In turn, leptin can stimulate the activity of three other endocrine systems that are involved in controlling the reproductive axis: (i) pancreatic insulin (Zieba *et al.*, 2003); (ii) pituitary GH (Henry *et al.*, 2001) and (iii) thyroid hormones (Flier *et al.*, 2000). In addition, leptin secretion is affected by other inputs, such as the products of digestion and absorption, and autonomic neural activity (Pénicaud *et al.*, 2000).

It is important to note that the ultimate level of integration lies within the central nervous system. The brain mechanisms involved in the sensing of metabolic status and in the connection of metabolic status to GnRH neuronal activity are poorly understood and the list of candidate neurochemical links can be bewildering. Receptors for leptin and insulin will always be a focus of attention. The orexins, despite their relevance being questioned (Blache *et al.*, 2003) continue to be of interest because the expression of orexin receptor 2 is dramatically inhibited by high intakes of energy and protein (Blache *et al.*, 2006). We need to test whether this change in sensitivity to orexins is linked to a change in reproductive activity or a change in food intake. At this stage, it seems unlikely that μ -receptor opioidergic systems are involved (Celi *et al.*, 2009) but, as outlined above, the regulatory system does include the neuropeptides that are involved in the control of food intake (Miller *et al.*, 2007).

More recently, the neuropeptide, kisspeptin, has drawn a lot of attention. In female sheep, it has been shown to stimulate GnRH and FSH secretion, to respond to sex steroids, and to play a role in the seasonality of ovulation (Messenger *et al.*, 2005; Smith *et al.*, 2007 and 2008). Importantly, in male rats, the kisspeptin receptor is found in over 75% of GnRH neurons (Irwig *et al.*, 2004). Interestingly, the ligand acts through a membrane receptor so it

could mediate the rapid responses of the GnRH system to acute changes in metabolic status. Kisspeptin could play an important integrative role because research in rodents has suggested that Kiss1 neurons could be the primary transducers of internal and environmental cues that regulate the neuroendocrine reproductive axis (review: Popa *et al.*, 2008). The interactions among peripheral signals (insulin, leptin, ghrelin, adiponectin and resistin) and the neuroendocrine systems (kisspeptin, neuropeptides involved in the control of food intake) need to be revealed if we are to understand the integrative processes that lead the brain to the 'decision to reproduce'.

Dimension 4 – the temporal dimension

There is a range of dynamic aspects in the reproductive responses to nutritional inputs – the effects of time *per se*, as seen in the effects of photoperiod and foetal programming, and the concept of 'metabolic memory' that allows for nutritional information to be carried forward beyond the life of the original metabolic stimulus.

In the ram, the response of the GnRH neurons to an abrupt change in nutrition is initially rapid and robust, but then fades over the next few weeks (Martin *et al.*, 1994a; Zhang *et al.*, 2004). In contrast, the long-term effect of nutrition on the ram testis, measured on a scale of several weeks, seems to be independent of changes in the primary, GnRH-based, control system (Hötzel *et al.*, 1995). On an annual timescale, the role of nutritional inputs, as well as the types of response to those inputs, can vary substantially, especially in genotypes that experience seasonal changes in appetite (Rhind *et al.*, 2002). Suggested inputs into the reproductive control systems that might implement these strategies include actions of leptin or ghrelin in testis, brain expression of neuropeptides affected by photoperiod, innervation of adipocytes by the sympathetic and parasympathetic systems, and autonomic nervous projections from the adipocytes to the brain.

Previous metabolic status influences the reproductive response to an increase in energy availability. With respect to adipose stores, mature rams in low body condition, but not rams in high body condition, show a robust and repeatable increase in LH pulse frequency in response to an increase in intake. In low body condition rams, the leptin response is also blunted but the response to insulin is not (Zhang *et al.*, 2005). These observations suggest that (i) neither insulin nor leptin are necessary for inducing an increase in GnRH pulse frequency in response to an increase in food intake and (ii) leptin secretion does not always respond to an influx of nutrients. Together with studies in dairy cattle (review: Blache *et al.*, 2007), these observations are consistent with the notion of a 'metabolic memory' that modulates the stimulatory effect of nutrient intake according to the level of either energy reserves or energy expenditure. Recently, the processes that might underpin 'metabolic memory' have been investigated in sheep and cattle and the data suggest that leptin and insulin are central to the concept (Chilliard *et al.*, 2005). In addition,

growth factors, such as insulin and growth hormones, also affect leptin secretion in whole animal studies and in isolated adipocytes (Chilliard *et al.*, 2005), and could therefore also be part of 'metabolic memory'. Other endocrine factors secreted by adipose tissue or by the digestive system might also be involved: for example, several metabolic effects have been described for adiponectin in rodents, such as increased insulin resistance and fatty acid oxidation, and reduced glucose output by the liver (review: Diez and Iglesias, 2003). These interactions between endocrine systems and nutrient supply may effectively act as a sort of peripheral integratory mechanism that complements the brain's integratory mechanisms, with the combination being responsible for 'metabolic memory'.

Practical application

Around the world, there is an increasing demand for animal products that are 'clean, green and ethical' (CGE). 'Clean' involves minimising the use of drugs, chemicals and hormones; 'green' involves minimising the impact of the industry on the environment, including the production of greenhouse gases by ruminants; 'ethical' has an obvious focus on animal welfare, but ethical judgement needs to be applied to all practices in the rest of the supply chain, not just the farmers. CGE principles are relevant to all forms of animal production, from low-input extensive grazing systems to high-input intensive systems involving confinement of the animals.

The nutrition-reproduction relationship is encapsulated in the CGE principle through 'focus feeding', a cost-effective way to feed energy at critical points in the reproductive process (Martin *et al.*, 2004). For example, producers can use focus feeding to boost the sperm production before mating, maximise potential litter size by increasing the ovulation rate and maximise postnatal survival and development, and minimise non-productive periods caused by delays in puberty or long *post-partum* anoestrus. Reproductive inefficiencies have always been seen as limiting profitability but there is now an extra imperative in the 'green' aspect of ruminant production systems – females that are not breeding are still producing methane, increasing the amount of greenhouse gases per unit of production of carcass or milk. This issue magnifies the consequences of, in particular, *post-partum* anoestrus in dairy cattle and offspring mortality in the sheep industry (Martin *et al.*, 2009).

For each period of focus feeding, we need to consider both the composition and duration of the diet, and we could use conserved or stored feed, or we could shift the entire reproductive process so that the critical periods are better aligned with peaks and troughs in the availability of forage. As we have discussed above, in balanced diets, this generally concerns the supply of energy. Often, supplements can achieve their aims by supplying a high-intensity stimulus for a very short period (e.g. 'flushing' for a few days to increase ovulation rate in sheep) effectively achieving the same effect as a mild dose of exogenous hormone. We use

the term 'nutritional pharmacology' to describe such phenomena (Martin *et al.*, 2008) and we have now extended the concept to include forage plants that provide beneficial secondary compounds for directly managing methane production and animal health (e.g. gastro-intestinal parasites, acidosis), thus further reducing the use of drugs.

The male in extensive management systems

It should now be a common practice to feed supplements to males for 8 weeks before mating to ensure that their testicular size and sperm production are maximal (review: Martin *et al.*, 2004). Producers should have their males healthy and 'fit not fat' (Combrink and Schoeman, 1993) and should take steps to prevent them from being stressed or losing any condition during the 8-week period leading up to mating. This can be done by grazing management or by supplementation. Quality nutrition for a few males is a reliable, cheap way to guarantee maximum sperm supplies. Some issues need research: (i) Is constant good condition better than an annual 8-week supplement with respect to sperm quantity and quality? (ii) How are sperm supplies affected if animals are losing weight but still in relatively good body condition at mating? (iii) What is the ideal condition score? (iv) We need to extend the focus feeding concept to males of other breeds besides the Merino sheep and the cashmere goat.

The male in intensive management systems, including artificial insemination (AI) centres

In seasonal systems, it should still be common practice to feed supplements to males for 8 weeks before the breeding period to ensure that their testicular size and sperm production are maximal. Unfortunately, it also seems to be common practice to overfeed males in the interests of maximising fertility and body size, especially in AI centres and studs, yet there is little doubt that this, as well as a lack of physical exercise, is detrimental to their performance (Okolski, 1975; Combrink and Schoeman, 1993). Again, more research is needed to extend the 'focus feeding' concept to males of other ruminant genotypes and to ascertain if there is any advantage in maintaining males at a constant body mass throughout the year rather than using an 8-week period of pre-mating supplementation.

Conclusions

Recent years have seen many advances in our understanding of the effects of nutrition on the reproductive system of mature male ruminants. Responses in testicular size due to changes in the size of the seminiferous tubules are also associated with alterations in the efficiency of spermatogenesis, all of which lead to changes in the rate of production of spermatozoa. These effects are rarely accompanied by similar changes in the endocrine function of the testis, as measured by the production of testosterone

or inhibin. In rams, changes in nutrition affect gonadotrophin secretion for only a few weeks, whereas testicular growth is affected for several months. In mature male goats during the non-breeding season, nutrition-induced testicular growth does not seem to be associated with any gonadotrophin response. It therefore seems likely that nutrition-driven testicular growth is at least partly independent of changes in gonadotrophin secretion. The energetic components of the diet, rather than the protein content, seem to be responsible for affecting gonadotrophin secretion in the ram. Variations in the expression of the nutritional responses between sexes, breeds and species probably reflect variations in the role of this environmental factor as a modulator of reproductive function.

Future research should concentrate on the intra-testicular systems mediating the effects of nutrition on the production of spermatozoa, as well as the neural and neuroendocrine processes leading to changes in gonadotrophin secretion. These pursuits will be greatly aided if we can determine how metabolic factors affect the GnRH-dependent and -independent effects on testicular growth. Finally, we need a serious research effort in the area of novel forages that can be used to improve health and also cut down greenhouse gas emissions. All this research will help us develop 'natural systems' for controlling and improving the productivity of farmed ruminants, at the same time greatly improving the image of ruminant industries in society and thus the marketplace.

Acknowledgements

We would like to thank Ms Margaret Blackberry for her endless toil in the laboratory. In addition, these studies, done over many years, could not have been contemplated without the generous assistance of the students and staff in Animal Science at The University of WA. Funding was supplied by the National Health and Medical Research Council, the Australian Research Council, Meat and Livestock Australia, CSIRO, the Australian Wool Corporation and the University of WA.

References

- Adam CL, Archer ZA and Miller DW 2003. Leptin actions on the reproductive neuroendocrine axis in sheep. *Reproduction Supplement* 61, 283–297.
- Ahima RS 2005. Central actions of adipocyte hormones. *Trends in Endocrinology and Metabolism* 16, 307–313.
- Alkass JE, Bryant MJ and Walton JS 1982. Some effects of level of feeding and body condition upon sperm production and gonadotropin concentrations in the ram. *Animal Production* 34, 265–277.
- Amann RP 1970. Sperm production rates. In *The testis, vol 1, development, anatomy and physiology* (ed. WR Gromes, NL Van Demark and AD Johnson), pp. 433–481. Academic Press, London, UK.
- Banks EM 1964. Some aspects of sexual behavior in domestic sheep, *Ovis aries*. *Behaviour* 23, 249–279.
- Barth AD, Brito LF and Kastelic JP 2008. The effect of nutrition on sexual development of bulls. *Theriogenology* 70, 485–494.
- Bell AW 1984. Factors controlling placental and fetal growth and their effects on future production. In *Reproduction in sheep* (ed. DR Lindsay and DT Pearce), pp. 144–152. Cambridge University Press, Cambridge, UK.

- Bielli A, Pérez R, Pedrana G, Milton JTB, Lopez A, Blackberry MA, Duncombe G, Rodriguez-Martinez H and Martin GB 2002. Low maternal nutrition during pregnancy reduces the numbers of Sertoli cells in the newborn lamb. *Reproduction, Fertility and Development* 14, 333–337.
- Blache D, Adam CL and Martin GB 2002. The mature male sheep: a model to study the effects of nutrition on the reproductive axis. In *Large mammals as neuroendocrine models* (ed. DC Skinner, NP Evans and C Doberska) *Reproduction Supplement* 59, pp. 219–233. Society for Reproduction and Fertility, Cambridge, UK.
- Blache D, Chagas LM, Blackberry MA, Vercoe PE and Martin GB 2000. Metabolic factors affecting the reproductive axis in male sheep. *Journal of Reproduction and Fertility* 120, 1–11.
- Blache D, Chagas LM and Martin GB 2007. Nutritional inputs into the reproductive neuroendocrine control system – a multidimensional perspective. In *Reproduction in domestic ruminants VI* (ed. JI Juengel, JF Murray and MF Smith), pp. 123–139. Nottingham University Press, Nottingham, UK.
- Blache D, Zhang S and Martin GB 2003. Fertility in males: modulators of the acute effects of nutrition on the reproductive axis of male sheep. In *Reproduction in domestic ruminants V* (ed. BK Campbell, R Webb, H Dobson and C Doberska), pp. 387–402. Society for Reproduction and Fertility, Cambridge, UK.
- Blache D, Zhang S and Martin GB 2006. Dynamic and integrative aspects of the regulation of reproduction by metabolic status in male sheep. *Reproduction, Nutrition, Development* 46, 379–390.
- Boukhliq R and Martin GB 1997. Administration of fatty acids and gonadotrophin secretion in the mature ram. *Animal Reproduction Science* 49, 143–159.
- Boukhliq R, Martin GB, White CL, Blackberry MA and Murray PJ 1997. Role of glucose, fatty acids and protein in the regulation of testicular growth and the secretion of gonadotrophin, prolactin, somatotrophin and insulin in the mature ram. *Reproduction, Fertility and Development* 9, 515–524.
- Boukhliq R, Miller DW and Martin GB 1996. Relationships between the nutritional stimulation of gonadotrophin secretion and peripheral cerebrospinal fluid (CSF) concentrations of glucose and insulin in rams. *Animal Reproduction Science* 41, 201–204.
- Braden AWH, Turnbull KE, Mattner PE and Moule GR 1974. Effect of protein and energy content of the diet on the rate of sperm production in rams. *Australian Journal of Biological Sciences* 27, 67–73.
- Bronson FH 1985. Mammalian reproduction: an ecological perspective. *Biology of Reproduction* 32, 1–26.
- Brown BW 1994. A review of the nutritional influences on reproduction in boars, bulls and rams. *Reproduction Nutrition Development* 34, 89–114.
- Budak E, Fernández Sánchez M, Bellver J, Cerveró A, Simón C and Pellicer A 2006. Interactions of the hormones leptin, ghrelin, adiponectin, resistin, and PYY3-36 with the reproductive system. *Fertility and Sterility* 85, 1563–1581.
- Cameron AWN, Murphy PM and Oldham CM 1988. Nutrition of rams and output of spermatozoa. *Proceedings of the Australian Society of Animal Production* 17, 162–165.
- Celi P, Miller DW, Blache D and Martin GB 2009. Interactions between nutritional and opioidergic pathways in the control of LH secretion in male sheep. *Animal Reproduction Science* (on line doi:10.1016/j.anireprosci.2009.03.011).
- Celi P, Walkden-Brown SW, Széll AZ, Blache D, Wilkinson HM and Martin GB 2007. Twin efficiency for reproductive variables in monozygotic twin sheep. *Theriogenology* 68, 663–672.
- Chilliard Y, Delavaud C and Bonnet M 2005. Leptin expression in ruminants: nutritional and physiological regulations in relation with energy metabolism. *Domestic Animal Endocrinology* 29, 3–22.
- Clark RT 1934. Studies of reproduction in sheep I. The ovulation rate of the ewe as affected by the plane of nutrition. *Anatomical Record* 60, 125–134.
- Combrink GC and Schoeman SJ 1993. The influence of exercising rams on the lambing performance of a Merino ewe flock. *South African Journal of Animal Science* 23, 24–25.
- Coulter GH, Carruthers TD, Amann RP and Kozub GC 1987. Testicular development, daily sperm production and epididymal sperm reserves in 15-month-old Angus and Hereford bulls: effects of bull strain plus dietary energy. *Journal of Animal Science* 64, 254–260.
- Da Silva P, Aitken RP, Rhind SM, Racey PA and Wallace JM 2001. Influence of placentally mediated fetal growth restriction on the onset of puberty in male and female lambs. *Reproduction* 122, 375–383.
- Davies DV, Mann T and Rowson LEA 1957. Effect of nutrition on the onset of male sex hormone activity and sperm formation in monozygous bull-calves. *Proceedings of the Royal Society of London, Series B, Biological Sciences* 147, 332–351.
- Diez JJ and Iglesias P 2003. The role of the novel adipocyte-derived hormone adiponectin in human disease. *European Journal of Endocrinology* 148, 293–300.
- Fernandez-Fernandez R, Tena-Sempere M, Aguilar E and Pinilla L 2004. Ghrelin effects on gonadotropin secretion in male and female rats. *Neuroscience Letters* 362, 103–107.
- Flier JS, Harris M and Hollenberg AN 2000. Leptin, nutrition, and the thyroid: the why, the wherefore, and the wiring. *Journal of Clinical Investigation* 105, 859–861.
- Flipse RJ and Almquist JO 1961. Effect of total digestible nutrient intake from birth to four years of age on growth and reproductive development and performance of dairy bulls. *Journal of Dairy Science* 44, 905–914.
- Fourie PJ, Schwalbach LM, Neser FWC and Van der Westhuizen C 2004. Scrotal, testicular and semen characteristics of young Dorper rams managed under intensive and extensive conditions. *Small Ruminant Research* 54, 53–59.
- Gauthier D and Berbigier P 1982. The influence of nutritional levels and shade structure on testicular growth and hourly variations of plasma LH and testosterone levels in young Creole bulls in a tropical environment. *Reproduction, Nutrition, Développement* 22, 793–801.
- Henry BA, Goding J, Tilbrook A, Dunshea F and Clarke I 2001. Intracerebroventricular infusion of leptin elevates the secretion of luteinising hormone without affecting food intake in long-term food restricted sheep, but increases growth hormone irrespective of bodyweight. *Journal of Endocrinology* 168, 67–77.
- Hiroe K and Tomizuka T 1965. Effects of nutrition on the characteristics of goat semen. *Bulletin of the National Institute of Animal Industry* 8, 17–24.
- Hochereau-de Reviers MT, Perreau C, Delouis C, Chemineau and Courot M 1986. Effects of photoperiod during foetal life and age on total number of Sertoli cells per testis between birth and adulthood in the goat. *Biology of Reproduction* (suppl. 234), 369 (abstract).
- Hochereau-de Reviers MT, Perreau C and Lincoln GA 1985. Photoperiodic variations of somatic and germ cell populations in the Soay ram testis. *Journal of Reproduction and Fertility* 74, 329–334.
- Hötzel MJ, Markey CM, Walkden-Brown SW, Blackberry MA and Martin GB 1998. Morphometric and endocrine analyses of the effects of nutrition on the testis of mature Merino rams. *Reproduction, Fertility and Development* 113, 217–230.
- Hötzel MJ, Walkden-Brown SW, Blackberry MA and Martin GB 1995. The effect of nutrition on testicular growth in mature Merino rams involves mechanisms that are independent of changes in GnRH pulse frequency. *Journal of Endocrinology* 147, 75–85.
- Hötzel MJ, Walkden-Brown SW, Fisher JA and Martin GB 2003. Determinants of the annual pattern of reproduction in mature male Merino and Suffolk sheep: response to a nutritional stimulus in the breeding and non-breeding season. *Journal of Reproduction and Fertility* 15, 1–9.
- Irwig MS, Fraley GS, Smith JT, Acohido BV, Popa SM, Cunningham MJ, Gottsch ML, Clifton DK and Steiner RA 2004. Kisspeptin activation of gonadotropin releasing hormone neurons and regulation of KiSS-1 mRNA in the male rat. *Neuroendocrinology* 80, 264–272.
- Knight TW 1977. Methods for the indirect estimation of testes weight and sperm numbers in Merino and Romney rams. *New Zealand Journal of Agricultural Research* 20, 291–296.
- Knight TW, Gherardi S and Lindsay DR 1987. Effects of sexual stimulation on testicular size in the ram. *Animal Reproduction Science* 13, 105–115.
- Komatsu T, Itoh F, Mikawa S and Hodate K 2003. Gene expression of resistin in adipose tissue and mammary gland of lactating and non-lactating cows. *Journal of Endocrinology* 178, R1–R5.
- Leatham JH 1975. Nutritional influences on testicular composition and function in mammals. In *Handbook of physiology* (Exec ed. SR Geiger) Section 7, Male reproductive system (Section ed. RO Greep and EB Astwood) vol 5, *Endocrinology* (Vol ed. DW Hamilton and RO Greep), pp. 225–232. American Physiological Society, Washington, USA.
- Lincoln GA and Short RV 1980. Seasonal breeding: nature's contraceptive. *Recent Progress in Hormone Research* 36, 1–52.
- Lindsay DR 1991. Reproduction in the sheep and goat. In *Reproduction in domestic animals*, 4th edition. (ed. PT Cupps), chapter 15, pp. 491–515. Academic Press, San Diego, USA.

- Malagon MM, Rodriguez-Pacheco F, Martinez-Fuentes AJ, Tovar S, Pinilla L, Tena-Sempere M, Dieguez C and Castano JP 2006. Regulation of pituitary cell function by the adipokine adiponectin. *Frontiers in Neuroendocrinology* 27, 35.
- Mann T, Rowson LEA, Short RV and Skinner JD 1967. The relationship between nutrition and androgenic activity in pubescent twin calves, and the effect of orchitis. *Journal of Endocrinology* 38, 455–468.
- Mann T and Walton A 1953. The effect of underfeeding on the genital functions of a bull. *Journal of Agricultural Science (Cambridge)* 43, 343–347.
- Martin GB 1984. Factors affecting the secretion of luteinizing hormone in the ewe. *Biological Reviews* 59, 1–87.
- Martin GB, Blache D and Williams IH 2008. Allocation of resources to reproduction. In *Resource allocation theory applied to farm animal production* (ed. W Rauw), pp. 169–191. Oxford University Press, Oxford, UK.
- Martin GB, Durmic Z, Kenyon PR and Vercoe PE 2009. Landcorp farming limited lecture: 'clean, green and ethical' animal reproduction: extension to sheep and dairy systems in New Zealand. *Proceedings of the New Zealand Society of Animal Production* 69, 140–147.
- Martin GB, Hötzel MJ, Blache D, Walkden-Brown SW, Blackberry MA, Bouklich R, Fisher JA and Miller DW 2002. Determinants of the annual pattern of reproduction in mature male Merino and Suffolk sheep: modification of response to photoperiod by annual cycle of food supply. *Reproduction, Fertility and Development* 14, 165–175.
- Martin GB, Milton JTB, Davidson RH, Banchemo Hunzicker GE, Lindsay DR and Blache D 2004. Natural methods of increasing reproductive efficiency in sheep and goats. *Animal Reproduction Science* 82–83, 231–246.
- Martin GB, Sutherland SRD and Lindsay DR 1987. Effects of nutritional supplements on testicular size and the secretion of LH and testosterone in Merino and Booroola rams. *Animal Reproduction Science* 12, 267–281.
- Martin GB, Tjondronegoro S and Blackberry MA 1994a. Effects of nutrition on testicular size and the concentrations of gonadotrophins, testosterone and inhibin in plasma of mature male sheep. *Journal of Reproduction and Fertility* 101, 121–128.
- Martin GB, Tjondronegoro S, Bouklich R, Blackberry MA, Briegel JR, Blache D, Fisher JA and Adams NR 1999. Determinants of the annual pattern of reproduction in mature male Merino and Suffolk sheep: modification of endogenous rhythms by photoperiod. *Reproduction, Fertility and Development* 11, 355–366.
- Martin GB, White CL, Markey CM and Blackberry MA 1994b. Effects of dietary zinc deficiency on the reproductive system of young male sheep: testicular growth and the secretion of inhibin and testosterone. *Journal of Reproduction and Fertility* 101, 87–96.
- Message S, Chatzidakis EE, Ma D, Hendrick AG, Zahn D, Dixon J, Thresher R, Malinge I, Lomet D, Carlton MBL, Colledge WH, Caraty A and Aparicio SAJR 2005. Kisspeptin directly stimulates gonadotropin-releasing hormone release via G protein-coupled receptor 54. *Proceedings of the National Academy of Sciences (USA)* 102, 1761–1766.
- Miller DW, Blache D, Bouklich R, Curlewis JD and Martin GB 1998. Central metabolic messengers and the effects of diet on gonadotrophin secretion in sheep. *Journal of Reproduction and Fertility* 112, 347–356.
- Miller DW, Blache D and Martin GB 1995. Insulin stimulates LH secretion in rams when infused into the third cerebral ventricle: comparison with the effect of improved nutrition. *Journal of Endocrinology* 147, 321–329.
- Miller DW, Findlay PA, Morrison MA, Raver N and Adam CL 2002. Seasonal and dose-dependent effects of intracerebroventricular leptin on LH secretion and appetite in sheep. *Journal of Endocrinology* 175, 395–404.
- Miller DW, Harrison JL, Bennett EJ, Findlay PA and Adam CL 2007. Nutritional influences on reproductive neuroendocrine output: insulin, leptin and orexigenic neuropeptide signaling in the ovine hypothalamus. *Endocrinology* 148, 5313–5322.
- Miller DW, Harrison JL, Brown YA, Doyle U, Lindsay A, Adam CL and Lea RG 2005. Immunohistochemical evidence for an endocrine/paracrine role for ghrelin in the reproductive tissues of sheep. *Reproductive Biology and Endocrinology* 3, 60.
- Monet-Kuntz C, Hochereau de Reviers MT and Terqui M 1984. Variations in testicular androgen receptors and histology of the lamb testis from birth to puberty. *Journal of Reproduction and Fertility* 70, 203–210.
- Mori A 1959. Studies on the reproductive failure of ram caused by underfeeding. I. On the effects of underfeeding upon the mating potency of ram, and the effects of normal feeding upon its recovery from impotence. *Tohoku Journal of Agricultural Research* 10, 263–281.
- Moule GR 1963. Postpubertal nutrition and reproduction by the male. *Australian Veterinary Journal* 39, 299–304.
- Mwansa PB and Makarechian M 1991. The effect of postweaning level of dietary energy on sex drive and semen quality of young beef bulls. *Theriogenology* 35, 1169–1178.
- Nogueiras R, Barreiro ML, Caminos JE, Gaytan F, Suominen JS, Navarro VM, Casanueva FF, Aguilar E, Toppari J, Dieguez C and Tena-Sempere M 2004. Novel expression of resistin in rat testis: functional role and regulation by nutritional status and hormonal factors. *Journal of Cell Science* 117, 3247–3257.
- Nogueiras R, Gallego R, Gualillo O, Caminos JE, Garcia-Caballero T, Casanueva FF and Dieguez C 2003. Resistin is expressed in different rat tissues and is regulated in a tissue- and gender-specific manner. *FEBS Letters* 548, 21–27.
- Okolski A 1975. Effect of different amounts of protein in the diet on sexual behaviour and properties of semen in rams. *Acta Agraria et Silvestria, Zootechnica* 15, 101–121.
- Oldham CM, Adams NR, Gherardi PB, Lindsay DR and Mackintosh JB 1978. The influence of level of feed intake on sperm producing capacity of testicular tissue in the ram. *Australian Journal of Agricultural Research* 29, 173–179.
- Parker GV and Thwaites CJ 1972. The effects of undernutrition on libido and semen quality in adult Merino rams. *Australian Journal of Agricultural Research* 23, 109–115.
- Pénaud L, Cousin B, Leloup C, Lorsignol A and Casteilla L 2000. The autonomic nervous system, adipose tissue plasticity, and energy balance. *Nutrition* 16, 903–908.
- Popa SM, Clifton DK and Steiner RA 2008. The role of Kisspeptins and GPR54 in the neuroendocrine regulation of reproduction. *Annual Review of Physiology* 70, 213–238.
- Raadsma HW and Edey TN 1985. Mating performance of paddock-mated rams. II. Changes in sexual and general activity during the joining period. *Animal Reproduction Science* 8, 101–107.
- Rekwot PI, Oyedipe EO, Akerejola OO and Kumi-Diaka J 1987. The effect of protein intake on body weight, scrotal circumference and semen production of Bunaji bulls and their Friesian crosses in Nigeria. *Animal Reproduction Science* 16, 1–9.
- Rhind SM, Archer ZA and Adam CL 2002. Seasonality of food intake in ruminants: recent developments in understanding. *Nutrition Research Reviews* 15, 43–65.
- Ritar AJ, Adams NR and Sanders MR 1984. Effect of lupin feeding on LH, testosterone and testes. In *Reproduction in sheep* (ed. DR Lindsay and DT Pearce), pp. 76–78. Cambridge University Press, UK.
- Robinson JJ, Ashworth CJ, Rooke JA, Mitchell LM and McEvoy TG 2006. Nutrition and fertility in ruminant livestock. *Animal Feed Science and Technology* 126, 259–276.
- Salamon S 1964. The effect of nutritional regimen on the potential semen production of rams. *Australian Journal of Agricultural Research* 15, 645–656.
- Scaramuzzi RJ, Campbell BK, Downing JA, Kendall NR, Khalid M, Munoz-Gutiérrez M and Somchit A 2006. A review of the effects of supplementary nutrition in the ewe on the concentrations of reproductive and metabolic hormones and the mechanisms that regulate folliculogenesis and ovulation rate. *Reproduction, Nutrition and Development* 46, 339–354.
- Schwartz MW 2006. Distribution of insulin receptor substrate-2 in brain areas involved in energy homeostasis. *Brain Research* 1112, 169–178.
- Setchell BP 1986. The movement of the fluids and substances in the testis. *Australian Journal of Biological Sciences* 39, 193–207.
- Setchell BP 1990. Local control of testicular fluids. *Reproduction, Fertility and Development* 2, 291–309.
- Setchell BP and Hinks NT 1967. The importance of glucose in the oxidative metabolism of the testis of the conscious ram and the role of the pentose cycle. *Biochemical Journal* 102, 623–630.
- Setchell B, Waites GMH and Lindner HR 1965. Effect of undernutrition on testicular blood flow and metabolism and the output of testosterone in the ram. *Journal of Reproduction and Fertility* 9, 149–162.
- Smith JT, Clay CM, Caraty A and Clarke IJ 2007. KiSS-1 messenger ribonucleic acid expression in the hypothalamus of the ewe is regulated by sex steroids and season. *Endocrinology* 148, 1150–1157.
- Smith JT, Coolen LM, Kriegsfeld LJ, Sari IP, Jaafarzadehshirazi MR, Maltby M, Bateman K, Goodman RL, Tilbrook AJ, Ubuka T, Bentley GE, Clarke IJ and

- Lehman MN 2008. Variation in kisspeptin and gonadotropin-inhibitory hormone expression and terminal connections to GnRH neurons in the brain: a novel medium for seasonal breeding in the sheep. *Endocrinology* 149, 5770–5782.
- St-Pierre DH, Wang L and Tache Y 2003. Ghrelin: a novel player in the gut-brain regulation of growth hormone and energy balance. *News in Physiological Sciences* 18, 242–246.
- Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, Patel HR, Ahima RS and Lazar MA 2001. The hormone resistin links obesity to diabetes. *Nature* 409, 307–312.
- Sugino T, Hasegawa Y, Kurose Y, Kojima M, Kangawa K and Terashima Y 2004. Effects of ghrelin on food intake and neuroendocrine function in sheep. *Animal Reproduction Science* 82–83, 183–194.
- Sutama IK and Edey TN 1985. Reproductive development during winter and spring of Merino ram lambs grown at three different rates. *Australian Journal of Agricultural Research* 36, 461–467.
- Tanaka T, Nagatani S, Bucholtz DC, Ohkura S, Tsukamura H, Maeda KI and Foster DL 2000. Central action of insulin regulates pulsatile luteinizing hormone secretion in the diabetic sheep model. *Biology of Reproduction* 62, 1256–1261.
- Tarulli GA, Stanton PG, Lerchl A and Meachem SJ 2006. Adult Sertoli cells are not terminally differentiated in the Djungarian hamster: effect of FSH on proliferation and junction protein organization. *Biology of Reproduction* 74, 798–806.
- Tegegne A, Entwistle KW and Mukasa-Mugerwa E 1992. Gonadal and extragonadal sperm reserves and testicular histometric characteristics in Zebu and crossbred bulls: effect of dry season nutritional supplementation. *Animal Reproduction Science* 29, 25–33.
- Tena-Sempere M 2005. Exploring the role of ghrelin as novel regulator of gonadal function. *Growth Hormone and IGF Research* 15, 83–88.
- Thwaites CJ 1995. The comparative effects of undernutrition, exercise and frequency of ejaculation on the size and tone of the testes and on semen quality in the ram. *Animal Reproduction Science* 37, 299–309.
- Tilbrook AJ and Cameron AWN 1990. The contribution of the sexual behaviour of rams to successful mating of ewes under field conditions. In *Reproductive physiology of Merino sheep – concepts and consequences* (ed. CM Oldham, GB Martin and IW Purvis), pp. 143–160. School of Agriculture, The University of Western Australia, Perth, Australia.
- Tilton WA, Wamick AC, Cunha TJ, Loggins PE and Shirley RL 1964. Effect of low energy and protein intake on growth and reproductive performance of young rams. *Journal of Animal Science* 23, 645–650.
- Van Demark NL, Fritz GR and Mauger RE 1964. Effect of energy intake on reproductive performance of dairy bulls. 2. Semen production and replenishment. *Journal of Dairy Science* 47, 898–904.
- Van Demark NL and Mauger RE 1964. Effect of energy intake on reproductive performance of dairy bulls. 1. Growth, reproductive organs, and puberty. *Journal of Dairy Science* 47, 798–802.
- Walkden-Brown SW, Martin GB and Restall BJ 1999. Role of male-female interaction in regulating reproduction in sheep and goats. *Journal of Reproduction and Fertility Supplement* 52, 243–257.
- Walkden-Brown SW, Restall BJ, Norton BW, Scaramuzzi RJ and Martin GB 1994. Effect of nutrition on seasonal patterns of LH, FSH and testosterone concentration, testicular mass, sebaceous gland volume and odour in Australian cashmere goats. *Journal of Reproduction and Fertility* 102, 351–360.
- Zhang S, Blache D, Blackberry MA and Martin GB 2004. Dynamics of the responses in secretion of LH, leptin and insulin following an acute increase in nutrition in mature male sheep. *Reproduction, Fertility and Development* 16, 823–829.
- Zhang S, Blache D, Blackberry MA and Martin GB 2005. Body reserves affect the reproductive endocrine responses to an acute change in nutrition in mature male sheep. *Animal Reproduction Science* 88, 257–269.
- Zieba DA, Amstalden M, Macie MN, Keisler DH, Raver N, Gertler A and Williams GL 2003. Divergent effects of leptin on luteinizing hormone and insulin secretion are dose dependent. *Experimental Biology and Medicine* 228, 325–330.