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Correlation of Oxidative Stress Biomarkers and Hematological Parameters in Blood Cancer Patients from Sardinia, Italy

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ABSTRACT

Background: Over the last few decades, there has been a dramatic increase in hematological malignancies (HMs) in the population of Sardinia. It is accepted that oxidative stress biomarkers have been demonstrated to be prognostically important in various neoplastic diseases. The aim of this study is to evaluate serum vitamin E, total antioxidant capacity (TAC), Malondialdehyde (MDA) and reactive oxygen species (ROS) levels in 80 Sardinian patients with different HMs [acute myeloid leukemia (AML)(n=20), myelodysplastic syndromes (MDS) (n=20), Hodgkin lymphoma (HL) (n=20) and non-Hodgkin lymphoma (NHL) (n=20)] on the day of their diagnosis.

Materials and Methods: Samples from all participants were obtained after an overnight fast (at least 10 hours). This study was approved and conducted in accordance with Good Clinical Practice guidelines and the Declaration of Helsinki. Patients and controls provided written, informed consent before entering the study. All study participants' medical history and their medication were documented upon enrolling.

Results: Lower levels of TAC and Vitamin E were observed in most of the studied groups compared to healthy controls (0.41-0.49 mmol/L vs. 0.56 mmol/L) (19.55-28.55 μ mol/L vs. 34.51 μ mol/L). Moreover, higher average MDA levels were observed in HL and NHL patients compared to healthy controls (16.6 ng/ml-17.8 ng/ml vs. 7.4 ng/ml). Additionally, the ROS values of all studied groups were found elevated. Serum TAC showed significant negative correlations with MDA values (R= -0.51; P<0.001). Statistical significance was observed in all hematological parameters, producing either positive or negative correlation with at least one OS biomarker.

Conclusion: The present data suggest that Sardinian patients with HL and NHL on the day of their diagnosis presented the highest OS in comparison to AML and healthy subjects. Moreover, MDS patients presented high OS status. Likewise, our results also indicated that changes in their hematological indices are eminent of their oxidative and antioxidative status.

Keywords: Oxidative stress biomarkers; Total antioxidant capacity; Malondialdehyde; Reactive oxygen species; Hematological malignancies, Sardinia

INTRODUCTION

The genetic peculiarities per se of the Sardinian population have significantly influenced the occurrence of hematological malignancies over the last decades¹. Meanwhile, recent studies have suggested an increased rate of hematological malignancies in the island². Moreover, several types of genetic mutations in this Mediterranean population result in either deficient enzymes or other mutation types in response to oxidative stress^{3,4}. The term 'oxidative stress biomarker' has been defined by the National Institute of Health as "a characteristic that is objectively measured and evaluated as an indicator of pathogenic processes, or pharmacological responses to a therapeutic intervention⁵. Several studies have evaluated oxidative stress (OS) biomarkers⁶ for various disorders such as diabetes⁷, malaria ⁸⁻¹⁰, heart¹¹ and diseases^{12,13}, neurodegenerative along with hemolytic^{14,15} and neoplastic disorders. The last of which includes hematological malignancies, a diverse group of blood cancers with various etiology, incidence, prognosis, and survival. Among them, myelodysplastic syndromes (MDS) represent a unique scenario as oxidative stress can increase due to both transfusion-dependent iron overload and dyserythropoiesis itself.¹⁶ Publications from different groups have presented measurements of different oxidative stress biomarkers for different hematological malignancies such as acute lymphoblastic leukemia (ALL), Hodgkin lymphoma (HL) and Myelodysplastic syndromes¹⁷⁻²³. Battisti et al. measured plasmatic thiobarbituric acid-reactive substances (TBARS), serum protein carbonylation, whole blood catalase (CAT) and superoxide dismutase (SOD) activities in ALL patients²². They found higher TBARS levels and serum protein carbonylation and reduced levels of antioxidants in ALL patients compared to controls. Fracchiolla et al. measured reactive oxygen metabolites (ROMs) levels and total antioxidant capacity (TAC) in oncohematological patients, and they also confirmed the oxidative imbalance in these group of patients²⁴. Despite this, there is not a comprehensive study comparing oxidative stress biomarkers in different hematological malignancies. It is already known that in most of the hematological malignancies, hematopoietic cells can be exposed to a wide spectrum of alterations²⁵. At first, excessive oxidative stress damages biomolecules such as DNA, proteins, and lipids, leading to cellular dysfunction and cell death. Accumulation of such damaging effects on individuals can result in diseases such as hematological malignancies. In addition, cells have multiple mechanisms to protect themselves from stress. These mechanisms include apoptosis, DNA repair, cell cycle regulation, and induction of antioxidant and detoxifying enzymes²⁵. On the other hand, reactive oxygen species (ROS) activate a cascade of events such as activation of tyrosine kinases and the mitogen-activated protein kinase system, followed by the activation of transcription factor subset²⁶⁻²⁸. Antioxidants are induced by oxidative stress to act not simply as scavengers of ROS but also as important regulators oxidative stress responses²⁹. Meanwhile, of oxidative stress often causes apoptosis, in which mitochondrial control has been known to play an essential role³⁰. The dysregulation of antioxidants and apoptosis is deeply involved in the disorders³¹. pathogenesis of hematopoietic Considering the current literature, this study aims to provide more details on this topic, evaluate and compare the antioxidant and oxidative status of sub-grouped Sardinian patients into four prognostically different groups of hematological malignancies. The groups are divided into four categories according to their diagnosis: i) acute myeloid leukemia (AML), ii) Hodgkin (HL), iii) non-Hodgkin (NHL) lymphomas and iv) myelodysplastic syndromes (MDS). As regards the study groups' oxidative status, direct measurements of ROS and malondialdehyde (MDA), an indicator of lipid peroxidation, were measured. Vitamin E and total antioxidant capacity (TAC) were used to evaluate the antioxidant status.

MATERIALS AND METHODS Data collection

Samples from all participants were obtained after an overnight fast (at least 10 hours). This study was approved and conducted in accordance with Good Clinical Practice guidelines and the Declaration of Helsinki. Patients and controls provided written, informed consent before entering the study. All study participants' medical history and their medication were documented upon enrolling.

Table 1 shows the demographic and clinical characteristics of the study groups. A group of 80 Sardinian patients (35 females and 45 males) were examined after blood samples were taken on the day of diagnosis. The results were compared with a sample of forty healthy age-and sex-matched controls, 17 females and 23 males, with an average age of 60.6 ± 10.2 years. Most of our patients were characterized by relevant comorbidities (in details, >90% of them for NHL, MDS and AML and >60% of them for HL), so it was impossible to make a formal comparison between the oxidative status of patients with and without relevant concomitant medical conditions.

The study population consisted of 5 groups (Table 1): (All groups are matched for age and sex):

- 40 healthy controls
- 20 patients diagnosed with Hodgkin lymphoma (HL)
- 20 patients diagnosed with non-Hodgkin lymphoma (NHL), 11 with histologically aggressive (DLBCL (diffuse large B cell lymphoma) and mantle) and 9 with indolent (follicular, MALT (mucosa associated lymphoid tissue) or small lymphocyte lymphoma).
- 20 patients diagnosed with acute myeloid leukemia (AML): 11 with an intermediate cytogenetic risk (normal karyotype), 3 unfavorable (monosomy and deletions in chromosomes 5 and 7), 5 favorable with t (8; 21) or t (15; 17), and one unclassified.
- 20 patients diagnosed with myelodysplastic syndrome (MDS): 10 RCMD (refractory cytopenia with multilineage dysplasia, 4 RARS (refractory cytopenia with ringed sideroblasts), 3 RAEB (refractory cytopenia with excess of blasts), 2 RAUD (refractory cytopenia with unilineage dysplasia), 15q- syndrome.

Table 1. Patient characteristics

Number of patients	80
Median age (years)	65.5 (±13.9)
Sex	
Male	45
Female	35
AML	
ELN stratification risk ¹	
Favorable	3
Intermediate	8
Adverse	5
Not available	3 4
MDS	4
WHO subtype	10
RA	10
RARS	4
RAUD	2
RAEB	3
5 q- syndrome	1
R-IPSS	
Very low	6
Low	9
Intermediate	3
High	1
Very high	1
NHL	
Histology	
MALT	1
FL	3
MCL	2
SLL	4
DLBCL	8
unclassified	2
Stage	-
	1
II	6
	3
III IV	
	7 3
not available	3
B symptoms	
Yes	4
No	13
not available	3
HL	
Stage	
I	2
II	8
III	2
IV	5
not available	3
B symptoms	
Yes	5
No	12
not available	3

Abbreviations: DLBCL: diffuse large B-cell NHL, FL: follicular NHL, IPI: International Prognostic Index, MALT: mucosa associated lymphoid tissue NHL, MCL: mantle cell NHL, MZL: marginal zone NHL, NHL: non-Hodgkin lymphoma, SLL: small lymphocyte NHL.

RA: refractory anemia; RARS: refractory anemia with ringed sideroblasts, RAUD: refractory anemia with unilineage dysplasia; RAEB: refractory anemia with excess blasts; R-IPSS: Revised International Prognostic Scoring System.

¹Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert

Oxidative stress biomarkers

A number of assays have been used to measure the levels of oxidative damage in Sardinian patients with various hematological malignancies. These laboratory tests variously measure excretion rates of damaged biomolecules in the blood. Several markers have been measured because of their accurate and clinically relevant approach to evaluate oxidative damage in many different types of damage. All hematological and biochemical parameters were estimated by spectrophotometric methods.

Total antioxidant capacity (TAC)

Antioxidant activity was assayed spectrophotometrically, as previously described¹⁴ using a TAC Kit (Total Antioxidant Capacity Colorimetric assay kit, produced by Cayman Chemical Co., Ann Arbor, USA) with the TEAC method in the clinical chemistry laboratory of the Alexander Technological Educational Institute, Thessaloniki, Greece.

Vitamin E

To determine vitamin E levels, an already described³² technique was used within HPLC (High-performance liquid chromatography) system. The alpha-tocopherol was measured by the method of high-performance liquid chromatography reverse phase (RP-HPLC).

Malondialdehyde (MDA)

Lipid peroxidation status in serum was determined by estimating malondialdehyde (MDA) as thiobarbituric acid reactive substances at 532nm, prior to dialysis with high-performance liquid chromatography(HPLC)³² in Laboratory of Biological Chemistry, Medical School, Aristotle University of Thessaloniki, Greece.

Reactive oxygen species (ROS)

Intracellular ROS levels in serum were detected using the cell-permeable ROS-sensitive probe 2',7'dichlorodihydrofluorescein diacetate (CM-H₂DCFDA) which causes fluorescence upon oxidation. As already described³⁴, the fluorescence probe was added to serum and the volume was made up with PBS. The oxidation of CM-H₂DCFDA was followed by 52 measuring the fluorescence in 96-well black-walled microplates (Corning^D, Sigma Aldrich) using a SAFAS Xenius (Monaco). The fluorescence is expressed in "% emission" as determined with the software "Xenius". ROS has been expressed as % of the maximal fluorescence obtained with H₂O₂ (3 mM).

Statistical analysis

Data were analyzed using the SPSS version 22 .0 Statistical package. Descriptive statistics presented as mean \pm standard deviation and frequencies as percentages. Kolmogorov–Smirnov analysis verified the normality of the data set. Pearson's chi-square test or chi-square test of association was used to discover if there is a relationship between the categorized data, while Fisher's exact test was used when expected variables were 2% of the total number of variables. Additionally, independent sample t-test was used to compare between means. In all statistical analysis, level of significance (p-value) was set at α =0.05.

RESULTS

TAC, Vitamin E, MDA and ROS levels (Table 2) in the four patients groups (HL, NHL, AML and MDS) and the healthy group were evaluated among the Sardinian population.

Hodgkin (HL) and non-Hodgkin lymphoma (NHL)

The most significant difference between lymphoma samples and healthy controls was seen in patients with HL, where we found lower Vitamin E and TAC levels (average Vitamin E value 19.55 μ mol/L vs 34.51 μ mol/L of the controls, *P* <0.001) (average TAC value 0.41 mmol/L vs 0.56 mmol/L of the controls, *P* <0.001: a 1.76- and 1.37-fold decrease, respectively in comparison to the controls) and markedly higher levels of MDA (average value 17.76 ng/ml vs 7.52 ng/ml of controls, *P* <0.001: a 2.36-fold increase).

NHL samples also showed a similar trend for Vitamin E, TAC levels (average Vitamin E value 20.78 μ mol/L vs 34.51 μ mol/L of the controls, *P* <0.001) (average TAC value 0.45 mmol/L vs. 0.56 mmol/L of controls, *P* <0.001: a 1.66- and 1.25-fold decrease, respectively) and MDA (average value 16.7 ng/ml vs. 7.52ng/ml of the controls, *P* <0.001: a 2.22-fold increase). In particular, among NHL patients greater

differences were demonstrated with normal controls in the indolent rather than aggressive subtype. Respectively, an average TAC value of 0.42 mmol/L vs 0.47 mmol/L (P = 0.005), and an average MDA value of 17.2 ng/ml vs 16.4 ng/ml (P = 0.003), were observed. When comparing all the study

groups, ROS levels in both HL (average value 20% vs 3.7% of controls, *P* <0.0001: a 5.40 fold increase) and NHL (average value 13.4% vs 3.7% of controls, *P* <0.001: a 3.62 fold increase) appeared to be in the higher range (Table 2).

Table 2. Serum levels of oxidative stress biomarkers in healthy controls and AML, NHL, HL and MDS patients. Mean TAC (mmol/L), Vitamin E (µmol/L), MDA (ng/ml) and ROS (%) values

Studied groups (n=120)	TAC (mmol/L)	Vitamin E (μmol/L)	MDA (ng/ml)	ROS (%)
Healthy Controls (n=40)	0.56	34.51	7.52	3.7
	(±0.05)	(±1.45)	(±0.9)	(±1.4)
Acute Myeloid Leukemia (AML) (n=20)	0.49** (±0.06)	25.10*	14.9*	10.6*
		(±2.45)	(±1.1)	(±1.7)
Non-Hodgkin lymphoma (NHL) (n=20)	0.45* (±0.04)	20.78*	16.7*	13.4*
		(±1.75)	(±0.7)	(±0.9)
Hodgkin lymphoma (HL) (n=20)	0.41* (±0.07)	19.55*	17.8*	20*
		(±1.65)	(±0.8)	(±2.5)
Myelodysplastic syndrome (MDS) (n=20)	0.55* (±0.03)	28.55*	13.5*	7.5*
		(±1.45)	(±1.2)	(±1.5)

Abbreviations: TAC: Total antioxidant capacity, MDA: malondialdehyde, ROS: reactive oxygen species; Significant differences between all studied groups (AML, HL, NHL and MDS groups) and healthy controls are marked with *,**. *P < 0.001, **P = 0.003, Data were analyzed using a two-tailed, two-sample Student's t-test.

Acute Myeloid Leukemia (AML)

Lower values of Vitamin E and TAC were found in the AML group, compared to the healthy group (average Vitamin E value 25.10 µmol/L vs 34.51 μ mol/L of the controls, P <0.001) (average TAC value 0.49 mmol/L vs 0.56 mmol/l of the controls, P =0.003: a 1.14-fold decrease). Indeed, MDA levels were higher (average value 14.9 ng/ml vs 7.52 ng/ml, P <0.001: a 1.98 fold increase), without any significant difference between cytogenetic risk groups. In addition, in AML patients there was a moderate negative correlation between MDA and TAC, with r = -0.45 vs. +0.18 compared to the controls. ROS levels of the AML group were significantly higher compared to healthy controls (average value 10.6% vs 3.7%; P < 0.001: a 2.86-fold increase) (Table 2).

Myelodysplastic syndromes (MDS)

Taking into consideration that MDS represent a unique scenario due to the different factors leading to a possible increase in oxidative stress, we compared them specifically with the healthy subjects. MDS patients showed lower levels of Vitamin E (average Vitamin E value 28.55 $\mu mol/L\,vs$

34.51 μ mol/L of the controls, P <0.001) compared to the healthy group (Table 2). Furthermore, there was a moderate negative correlation between MDA and TAC in these patients (R = -0.35; P <0.001). ROS (average value 7.5%) and MDA levels (average value 13.5 ng/ml) were higher compared to the healthy group, (ROS 2.02-fold increase vs controls; MDA 1.79-fold increase vs controls) (Table 2).

Correlation between OS biomarkers and hematological parameters in Hodgkin and non-Hodgkin lymphoma group

In Table 3, we present associations between all the tested OS biomarkers (TAC, Vitamin E, MDA and ROS levels) with the hematological parameters of Hodgkin and non-Hodgin lymphoma patients; the two groups with the higher evaluated oxidative stress values on the day of their diagnosis. A statistically significant positive correlation was observed between their bilirubin levels and i) TAC values and ii) Vitamin E levels (R = 0.65; p <

0.001, R=0.57; p < 0.001); (R = 0.47; p < 0.001, R=0.69; p < 0.001). On the other hand, neutrophil activity seemed to be negatively correlated with TAC activity (R = -0.44; p < 0.001); (R = -0.34; p < 0.001) and Vitamin E levels (R = - 0.38; p < 0.001); (R = -0.37; p < 0.001), respectively. Furthermore, in the HL group, bilirubin levels presented a statistically significant negative correlation with both MDA and ROS values (R = - 0.75; p < 0.001, R= -

0.49; p < 0.001, respectively). Finally, hemoglobin levels showed a statistical significant positive correlation only with the MDA values (R = 0.33; p < 0.001) in the HL patient group. None of the other measured hematological indices correlated with the OS biomarkers presented statistically significant correlations.

Hodgkin and non-Hodgkin lymphoma patients								
Pearson's Correlation		TAC	Vitamin E	MDA	ROS			
		R; p-value	R; p-value	R; p-value	R; p-value			
HGB	HL	0.14;<0.001	0.24; 0.005	0.33; <0.001	-0.09< 0.001			
	NHL	-0.13; <0.001	0.18; 0.007	0.07;<0.001	-0.09; 0.07			
WBC	HL	-0.36; 0.008	-0.19; 0.007	-0.02; < 0.001	-0.12; < 0.001			
	NHL	0.25; <0.001	-0.09; 0.014	0.01; < 0.001	0.37; < 0.001			
Bilirubin	HL	0.65; < 0.001	0.57; < 0.001	-0.75 < 0.001	-0.27; 0.04			
	NHL	0.47; < 0.001	0.69; < 0.001	-0.39< 0.001	-0.11; < 0.001			
Ferritin	HL	-0.11; 0.03	0.12; 0.15	-0.32< 0.001	-0.26 ; 0.05			
	NHL	-0.47; < 0.001	0.08; 0.17	-0.08< 0.001	-0.001 ; 0.14			
NEU	HL	-0.44; < 0.001	-0.38; < 0.001	0.14< 0.001	-0.20 ; 0.05			
	NHL	-0.34; < 0.001	-0.37; < 0.001	-0.25< 0.001	0.27; < 0.001			

*Correlation is significant at p value < 0.05 (2-tailed)

Abbreviations: WBC-White blood cells; HGB-Haemoglobin; NEU-Neutrophils; PLT-platelet count; NEU-neutrophil count; LYM- lymphocyte count;

DISCUSSION

It has already been demonstrated how in some neoplasms oxidative stress is characterized by an imbalance between the production of ROS and a biological system's ability to repair itself^{35,36}. This study has the potential to evaluate the oxidant and the antioxidant status of Sardinian patients with different blood and bone marrow disorders. The HL patient group demonstrated the greatest oxidant status, whereas patients with MDS showed the least. In agreement with our study, Bur et al.²⁰ also suggest that significant OS exists in HL and especially in patients with more aggressive forms of the disorder. Moreover, high ROS and MDA production in HL patients has been associated with a significant decrease in antioxidant defence mechanisms³⁷. Also, our study showed that the bilirubin system of HL patients seems to be affected by their oxidative status. Regarding MDS patients, Pimkova et al.23 did not observe increased MDA levels which contrasts with our findings in this group of patients. But, Goncalves et al.¹⁶ in agreement with our study observed increased ROS levels and consequently the involvement of oxidative stress to the development of MDS. Among all the analyzed hematological malignancies, we found an increase of oxidative damage, although of varying degrees. Our results indicate that the studied parameters of the OS status in human blood potentially modulated by concomitant are neoplastic disorders, suggesting an overall increase in OS. Importantly, the level of the studied markers, as well as the possible correlation between these parameters and their hematological indices in different patients group might have prognostic relevance for healthy subjects. Moreover, it will be worthy, to evaluate possible correlations between oxidative stress biomarkers and iron levels of MDS patients, which has shown to influence survival in patients with low riskmyelodysplastic syndromes³⁸. Nowadays, it is well known that in these disorders oxidative stress is dependent on labile iron plasma. In fact, iron overload in these patients derives from both blood transfusions and dyserythropoiesis whereby both factors can contribute to establishing an oxidative status in their cells. For this reason, more data are needed to better understand the mechanisms of hematological improvement and evaluate the oxidative stress markers in MDS patients under different iron chelation therapies³⁹.

Our findings may also suggest a possible use of these parameters for screening and increased monitoring of these neoplastic conditions. Further studies are required to determine the source and species of ROS generated by tumor cells and whether the ROS with therapeutic effects originates from the metabolism of normal cells or rather neoplastic cells. The knowledge that OS induces protein carbonylation by inactivating antioxidant enzymes⁴⁰ could be used to investigate an efficient way to boost the endogenous production of antioxidants and incorporate them into diets, thus helping to neutralize the free radicals produced by cellular metabolisms.

CONCLUSION

We suggest the involvement of oxidative stress in HL, NHL, AML and MDS development and prognosis. Patients with HL and NHL presented the worst oxidative status. Moreover, oxidative stress in MDS patients seems to be subtype-dependent. Regarding the tested biomarkers, TAC and MDA may constitute novel markers with value in diagnosis and/or prognosis of these diseases. More studies need to be done in order to clarify the associations between OS biomarkers and hematological parameters in Hodgkin and non-Hodgkin lymphoma group. In conclusion, in all studied groups a negative correlation (R = -0.51; P < 0.001) between TAC and MDA levels was observed, whereby MDA values increased as the TAC levels decreased. In addition, lower Vitamin E levels in all studied groups in comparison with the healthy group confirmed the low values of total antioxidants. Furthermore, ROS levels appeared to be significantly higher in all studied groups compared to the healthy group (P <0.001). The present study contributes to a better understanding of the oxidative basis of neoplastic diseases, as multifactorial and heterogeneous diseases.

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Statement of Ethics

Subjects have given their written informed consent. The study protocol has been approved by the Research Institute's Committee of University of Sassari on human research.

Conflict of interest

The authors declare that they have no conflict of interest

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REFERENCES

1. Broccia G, Deplano W, Dessalvi P, et al. Hematological malignancies in the island of Sardinia, 1974-1993: age and sex distributions and temporal changes in incidence. Hematol Oncol. 2004; 22(3):91-109.

2. Broccia G, Gabbas A, Longinotti M. Newly diagnosed cases of hematologic malignancies in Sardinia in the early 2000: an estimation of their number, age and geographic distribution on the basis of a previous epidemiologic survey. Haematologica. 2005;90(3):429-30.

3. Cao A, Galanello R, Furbetta M, et al. Thalassaemia types and their incidence in Sardinia. J Med Genet. 1978;15(6):443-7.

4. Lenzerini L, Meera Khan P, Filippi G, et al. Characterization of glucose-6-phosphate dehydrogenase variants. I. Occurrence of a G6PD Seattle-like variant in Sardinia and its interaction with the G6PD Mediterranean variant. Am J Hum Genet. 1969;21(2):142-53.

5. Strimbu K, Tavel JA. What are biomarkers? Curr Opin HIV AIDS. 2010; 5(6):463-6.

6. Marrocco I, Altieri F, Peluso I. Measurement and Clinical Significance of Biomarkers of Oxidative Stress in Humans. Oxid Med Cell Longev. 2017;2017:6501046.

7. Pitocco D, Tesauro M, Alessandro R, et al. Oxidative stress in diabetes: implications for vascular and other complications. Int J Mol Sci. 2013; 14(11):21525-21550.

8. Aguilar R, Marrocco T, Skorokhod OA, et al. Blood oxidative stress markers and Plasmodium falciparum malaria in non-immune African children. Br J Haematol. 2014;164(3):438-50.

9. Percário S, Moreira DR, Gomes BA, et al. Oxidative stress in malaria. Int J Mol Sci. 2012; 13(12):16346-72.

10. Pantaleo A, Ferru E, Pau MC, et al. Band 3 Erythrocyte Membrane Protein Acts as Redox Stress Sensor Leading to Its Phosphorylation by p (72) Syk. Oxid Med Cell Longev. 2016;2016:6051093.

11. Costa VM, Carvalho F, Bastos ML, et al. Contribution of catecholamine reactive intermediates and oxidative stress to the pathologic features of heart diseases. Curr Med Chem. 2011; 18(15):2272-314.

12. Liu Z, Zhou T, Ziegler AC, et al. Oxidative Stress in Neurodegenerative Diseases: From Molecular Mechanisms to Clinical Applications. Oxid Med Cell Longev. 2017;2017:2525967.

13. Uttara B, Singh AV, Zamboni P, et al. Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. Curr Neuropharmacol. 2009;7(1):65-74.

14. Tsamesidis I, Fozza C, Vagdatli E, et al. Total antioxidant capacity in Mediterranean beta-thalassemic patients. Adv Clin Exp Med. 2017; 26(5):789-793.

15. Vanella A, Campisi A, Castorina C, et al. Antioxidant enzymatic systems and oxidative stress in erythrocytes with G6PD deficiency: effect of deferoxamine. Pharmacol Res. 1991;24(1):25-31.

16. Goncalves A, Cortesao E, Oliveiros B, et al. Oxidative stress and mitochondrial dysfunction play a role in myelodysplastic syndrome development, diagnosis, and prognosis: a pilot study. Free Radic Res. 2015; 49(9):1081-94.

17. Camargo Cde Q, Borges Dda S, de Oliveira PF, et al. Individuals with hematological malignancies before undergoing chemotherapy present oxidative stress parameters and acute phase proteins correlated with nutritional status. Nutr Cancer. 2015;67(3):463-71.

18. Masutani H. Oxidative stress response and signaling in hematological malignancies and HIV infection. Int J Hematol. 2000;71(1):25-32.

19. Bowen D, Wang L, Frew M, et al. Antioxidant enzyme expression in myelodysplastic and acute myeloid leukemia bone marrow: further evidence of a pathogenetic role for oxidative stress? Haematologica. 2003;88(9):1070-2.

20. Bur H, Haapasaari KM, Turpeenniemi-Hujanen T, et al. Oxidative stress markers and mitochondrial antioxidant enzyme expression are increased in aggressive Hodgkin lymphomas. Histopathology. 2014;65(3):319-27.

21. Wang SS, Davis S, Cerhan JR, et al. Polymorphisms in oxidative stress genes and risk for non-Hodgkin lymphoma. Carcinogenesis. 2006;27(9):1828-34.

22. Battisti V, Maders LD, Bagatini MD, et al. Measurement of oxidative stress and antioxidant status in acute lymphoblastic leukemia patients. Clin Biochem. 2008;41(7-8):511-8.

23.Pimková K, Chrastinová L, Suttnar J, et al. Plasma levels of aminothiols, nitrite, nitrate, and malondialdehyde in myelodysplastic syndromes in the context of clinical outcomes and as a consequence of iron overload. Oxid Med Cell Longev. 2014; 2014: 416028.

24. Fracchiolla NS CA, Orofino N, Novembrino C, et al. Oxidative Stress as Marker of Sepsis in Onco Hematological Patients: A Pilot Study. J Blood Disord Transfus. 2015; 6(5):311.

25. Kuhn V, Diederich L, Keller TCS 4th, et al. Red Blood Cell Function and Dysfunction: Redox Regulation, Nitric Oxide Metabolism, Anemia. Antioxid Redox Signal. 2017; 26(13):718-742.

26. Cosentino-Gomes D, Rocco-Machado N, Meyer-Fernandes JR. Cell signaling through protein kinase C oxidation and activation. Int J Mol Sci. 2012; 13(9): 10697–10721.

27. Reuter S, Gupta SC, Chaturvedi MM, et al. Oxidative stress, inflammation, and cancer: how are they linked? Free Radic Biol Med. 2010; 49(11):1603-16.

28. Gupta RK, Patel AK, Shah N, et al. Oxidative stress and antioxidants in disease and cancer: a review. Asian Pac J Cancer Prev. 2014; 15(11):4405-9.

29. Pisoschi AM, Pop A. The role of antioxidants in the chemistry of oxidative stress: A review. Eur J Med Chem. 2015; 97: 55-74.

30. Dröge W. Free radicals in the physiological control of cell function. Physiol Rev. 2002; 82(1):47-95.

31. Blokhina O, Virolainen E, Fagerstedt KV. Antioxidants, oxidative damage and oxygen deprivation stress: a review. Ann Bot. 2003; 91 Spec No: 179-94.

32. Apostolidou C, Adamopoulos K, Lymperaki E, et al. Cardiovascular risk and benefits from antioxidant dietary intervention with red wine in asymptomatic hypercholesterolemics. Clin Nutr ESPEN. 2015; 10(6):e224-e233.

33. Fucile C, Marini V, Zuccoli ML, et al. HPLC determination of malondialdehyde as biomarker for oxidative stress: application in patients with alcohol dependence. Clin Lab. 2013; 59(7-8):837-41.

34. Oparka M, Walczak J, Malinska D, et al. Quantifying ROS levels using CM-H2DCFDA and HyPer. Methods. 2016;109:3-11.

35. Frijhoff J, Winyard PG, Zarkovic N, et al. Clinical Relevance of Biomarkers of Oxidative Stress. Antioxid Redox Signal. 2015; 23(14):1144-70.

36. Lee JD, Cai Q, Shu XO, et al. The Role of Biomarkers of Oxidative Stress in Breast Cancer Risk and Prognosis: A

Systematic Review of the Epidemiologic Literature. J Womens Health (Larchmt). 2017; 26(5):467-482.

37. Omoti CE, Benedict N, ES I, et al. Total Antioxidant Capacity (TAC) in Patients with Haematological Malignancies in Niger Delta-region of Nigeria. Am J Cancer Sci. 2013; 2: 101-107.

38. Swart Ld, Reiniers C, Bagguley T, et al. Plasma Iron Levels Predict Survival in Patients with Lower-Risk Myelodysplastic Syndromes. Haematologica. 2018; 103(1):69-79.

39. Pilo F, Angelucchi E. A storm in the niche: Iron, oxidative stress and haemopoiesis. Blood Rev. 2018; 32(1):29-35.

40. Udensi UK, Tchounwou PB. Dual effect of oxidative stress on leukemia cancer induction and treatment. J Exp Clin Cancer Res. 2014; 33: 106.

41. Fiaschi T, Chiarugi P. Oxidative stress, tumor microenvironment, and metabolic reprogramming: a diabolic liaison. Int J Cell Biol. 2012; 2012:762825.