

# De Novo Short Read Assembly Algorithm with Low Memory Usage

Yuki Endo<sup>1</sup>, Fubito Toyama<sup>1</sup>, Chikafumi Chiba<sup>2</sup>, Hiroshi Mori<sup>1</sup> and Kenji Shoji<sup>1</sup>

<sup>1</sup>Graduate School of Engineering, Utsunomiya University, Yoto 7-1-2, Utsunomiya, Tochigi, 321-8585, Japan

<sup>2</sup>Faculty of Life and Environmental Sciences, University of Tsukuba, Tennoudai 1-1-1, Tsukuba, Ibaraki 305-8572, Japan

**Keywords:** Bioinformatics, Next Generation Sequencing, De Novo Assembly

**Abstract:** Determining whole genome sequences of various species has many applications not only in biological system, but also in medicine, pharmacy and agriculture. In recent years, the emergence of high-throughput next-generation sequencing technologies has dramatically reduced time and costs for whole genome sequencing. These new technologies provide ultrahigh throughput with lower unit data cost. However, the data are very short length fragments of DNA. Thus, developing algorithms for merging these fragments is very important. Merging these fragments without reference data is called de novo assembly. Many algorithms for de novo assembly have been proposed in recent years. Velvet, one of the algorithms, is famous because it has good performance in terms of memory and time consumption. But memory consumption increases dramatically when the size of input fragments is huge. Therefore, it is necessary to develop algorithm with low memory usage. In this paper, we propose an algorithm for de novo assembly with lower memory. In our experiments using *E.coli* K-12 strain MG 1655, memory consumption of the proposed algorithm was one-third of that of Velvet.

## 1 INTRODUCTION

Determining whole genome sequences of various species has many applications not only in biological system, but also in medicine, pharmacy and agriculture. In recent years, the emergence of high-throughput next-generation sequencer (NGS) technologies has dramatically reduced time and costs for whole genome sequencing. These new technologies provide ultrahigh throughput with lower unit data cost. However, many very small fragments (fewer than 100 base pair) of DNA are sequenced in these technologies. The fragments are typically called reads. The precision of NGS is not perfect. Therefore, the reads obtained by NGS might include sequencing errors. Thus, developing algorithms for merging these reads is very important. Merging these reads without reference data is called de novo assembly.

The de novo assembly algorithms can be classified into two approaches by its features : overlap-layout-consensus (OLC) and de Bruijn graph. OLC approaches relies on an overlap graph. Edena (Hernandez et al., 2008), MIRA (Chevreux et al., 2004), Celera (Miller et al., 2008), SSAKE (Warren et al., 2007), and VCAKE (Jeck et al., 2007) assemblers are based on OLC approach. In OLC strategy, pairwise overlaps are found by all-against-all pair-wise

comparison. Overlap graphs are constructed from pair-wise overlaps. In overlap graphs, a vertex represents a read and an edge denotes an overlap between two connected vertices (reads). Consensus sequences are determined by tracing paths in the graph. On the other hand, Velvet (Zerbino and Birney, 2008), ABySS (Simpson et al., 2009), ALLPATHS (Butler et al., 2008), and SOAPdenovo (Li et al., 2010) assemblers are based on de Bruijn graph approach. In the de Bruijn graph, a vertex represents a sequence of  $k$  bases ( $k$ -mer), where  $k$  is any positive integer. An edge represents an overlap of  $k-1$  bases. Thus, two connected vertices denote a  $k-1$  overlap between their vertices ( $k$ -mers). After the de Bruijn graph was constructed from reads obtained by NGS, contigs are determined by tracing paths in the graph. The de Bruijn graph approach is most widely applied to the short reads from Solexa and SOLiD platforms. In this approach, fixed-length ( $k-1$ ) overlaps are found and redundant  $k$ -mers (subsequence) are compressed. Thus, it is suitable for assembling vast quantities of short reads. Velvet is one of the most popular de novo assembler based on the de Bruijn graph. However memory consumption increases dramatically when the size of input reads is huge (more than several gigabytes). Therefore it is hard to use Velvet for huge scale assembly.

In this paper, we propose an algorithm for huge scale de novo assembly with low memory usage. Although our algorithm is based on de Bruijn graph approach in the same way as Velvet, edge information is not kept in main memory. Thus, the amount of memory can be reduced greatly in our method. In our experiments using *E.coli* K-12 strain MG 1655, the results showed that maximum memory consumption of the proposed algorithm was one-third that of Velvet. Furthermore, the running time of proposed method was also faster than that of Velvet.

## 2 AN ASSEMBLY ALGORITHM WITH LOW MEMORY USAGE

In this paper, we propose an algorithm for huge scale de novo assembly with low memory usage. Figure 1 shows the outline of our algorithm. First, all  $k$ -mers obtained by dividing reads are recorded. At the same time, the number of occurrences of each  $k$ -mer is also counted. Second, the de Bruijn graph is constructed using  $k$ -mers. Then, the graph is partitioned into subgraphs such that the subgraph has a simple path or a simple cycle. The simple path is a path in the graph which does not have repeating vertices or edges. The subgraphs are connected to make a larger simple path. The number of occurrences of a  $k$ -mer is used to select a connection path. Finally, contigs are generated by tracing vertices in the each connected graphs.

### 2.1 Extraction of $k$ -mers

$k$ -mers are extracted from all reads. They are kept in database in memory as “ $k$ -mer integer”. As shown in Table 1,  $k$ -mer integer is a representation of one-to-one correspondence between  $k$ -mer and integer. Specifically, bases “A”, “C”, “G”, and “T” correspond to quaternary digits 0, 1, 2, and 3 respectively. Thus, a  $k$ -mer sequence is expressed as a quaternary numeral. For example, a 5-mer base sequence “ACGTA” is converted to the quaternary number 01230. Then, it is converted to the decimal number 108. Thus, 108 is  $k$ -mer integer corresponding to “ACGTA”. If a  $k$ -mer sequence is represented by a string, memory would be needed  $k$  bytes. In the  $k$ -mer integer representation, the amount of memory for  $k$ -mer sequences can be reduced to one fourth of that of the string representation. Using  $k$ -mer integer is not only superior to memory usage, but also to processing time. Forward and reverse complement  $k$ -mer sequences are regarded as the same sequence in our method. Either of the two complementary sequences is registered in the database on main memory.

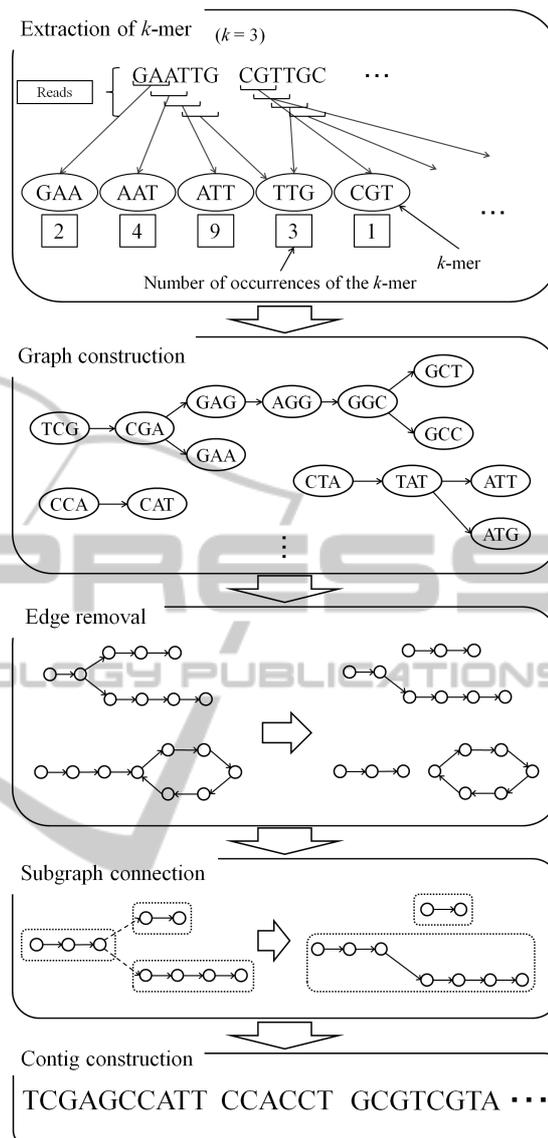


Figure 1: Outline of the proposed method.

In this work,  $k$ -mer integer and the number of occurrences of the  $k$ -mer corresponding to the  $k$ -mer integer are kept in main memory. In order to achieve low memory usage, other data (such as edges in de Bruijn graph) is not kept in the memory.  $K$ -mer sequence in which the number of occurrences of the sequence is small (less than a threshold) is not used in the graph construction because it is suspected that such  $k$ -mer sequences contain sequencing errors. In our experiments, the threshold was set to 5. Figure 2 shows the extraction of  $k$ -mer and the contents of database in the proposed method.

Table 1:  $k$ -mer sequence corresponding to  $k$ -mer integer (in case of 5-mer).

$k$ -mer	$k$ -mer integer		
	Quaternary	Binary	(decimal)
AAAAA	0	0	0
AAAAC	1	1	1
AAAAG	2	10	2
AAAAT	3	11	3
AAACA	10	100	4
AAACC	11	101	5
AAACG	12	110	6
AAACT	13	111	7
AAAGA	21	1000	8
AAAGC	22	1001	9

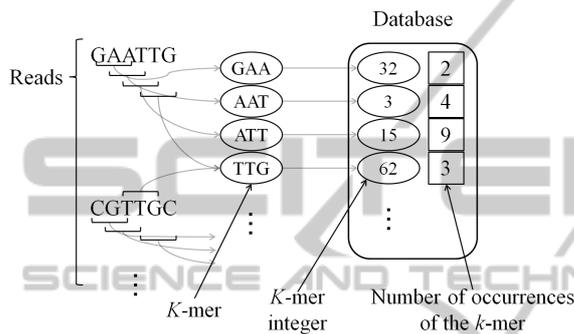


Figure 2: Extraction of  $k$ -mer and the contents of database in the proposed method (in case of 3-mer).

## 2.2 Graph Construction

The de Bruijn graph is constructed using  $k$ -mers. In the de Bruijn graph, each vertex represents a  $k$ -mer. An edge represents an overlap of  $k-1$  bases. Thus, two connected vertices denote a  $k-1$  overlap between their vertices ( $k$ -mers). For example, there is an edge between the two vertices corresponding to “ACGTA” and “CGTAC”. The direction of the edge is from “ACGTA” to “CGTAC”.

The contigs are generated by tracing vertices in the de Bruijn. However, de Bruijn graph constructed from short reads has numerous branches and cycles. Therefore, it is difficult to find the paths from which contigs are constructed. The details of the method to find out such paths are described in Section 2.3.

In conventional algorithms using de Bruijn graph, when the graph is constructed, edge information about which vertices are connected to each other is also kept in main memory. Since there are many edges in the graph, keeping the edge information consumes a huge amount of memory. In this paper, the edge information is not kept in main memory. Thus, although computational time for assembly is increased, memory usage can be reduced. The existence of the edge is calculated only when it is required. Specifically, the vertices which are connected

by a directed edge from a vertex have only 4 types of  $k$ -mers because the  $k$ -mers which are represented by the connected vertices overlap by  $k-1$  bases as shown in Fig. 3. Thus, the connected vertices ( $k$ -mer sequences) can be obtained by checking four values of  $k$ -mer integer in the database. By using this method, although processing time to check the values of  $k$ -mer integer is increased, memory for keeping connection edge data is not required. Only the data of vertices are kept in the database. Thus, the amount of memory can be reduced greatly in our method. The graph construction is finished by registering the values of  $k$ -mer integer from  $k$ -mer sequences and the number of occurrences of each  $k$ -mer on our database.

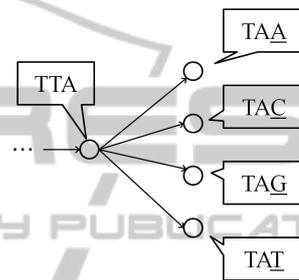


Figure 3: Example of 4 types of  $k$ -mers which are connected to the current vertex (in case of 3-mer).

## 2.3 Edge Removal

As mentioned in Section 2.2, the constructed graph has numerous branches and cycles. Consequently, it is important to select the edges on the path from which a contig is constructed. Figure 4 shows examples of branches. A vertex has multiple edges in Fig. 4 (a). A vertex has multiple edges from other vertices in Fig. 4 (b). Although the outdegree of the vertices with branches is two in Fig. 4, the maximum number of the out degree is four. Figure 4 (c) shows an example of a cycle. Actually, these branches and cycles are intricately intertwined. A path from the directed graph in which branches and cycles are included is needed to generate a contig. It is necessary to determine the unique simple path based on some criteria. The edge removal process is as follows.

1. A start vertex ( $k$ -mer) which has the largest number of occurrences is selected.
2. The start vertex is set to the current vertex.
3. The existence of the vertices which are connected to the current vertex is checked.
  - (a) If one connected vertex is found, the vertex is set to the current vertex. Go to 3.
  - (b) If multiple connected vertices are found, one of them is set to the current vertex. The details of

- the vertex selection are described in later in this section. Go to 3.
- (c) If the connected vertex is not found, the current vertex is regarded as the end vertex. Go to 4.
4. The existence of the vertices which are not selected yet is checked.
- (a) If the vertices which are not selected are found, the new start vertex which has the largest number of occurrences is selected from the vertices which are not selected yet. Go to 2.
  - (b) If the vertices which are not selected are not found, the process is finished.

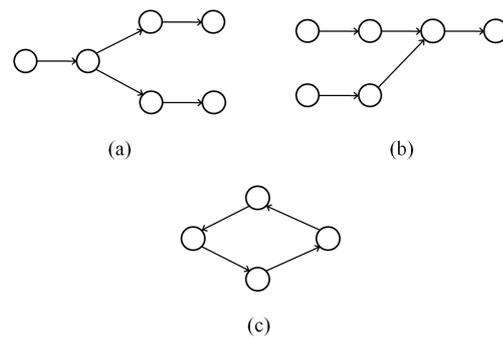


Figure 4: Examples of branches and cycle.

In this process, the vertices which are included in a path are assigned to same label. A path from the start vertex to the end vertex represents a subgraph. Multiple subgraphs are created in this process;

The details of the vertex selection in which branches and cycles are included are described as below. When there are multiple outgoing edges from the current vertex as shown Fig. 4 (a), the edge connected to the vertex in which the number of occurrences of  $k$ -mer is the largest is selected and the other outgoing edges are removed. When there are multiple incoming edges as shown in Fig. 4 (b), the edge is selected by the same process and other incoming edges are removed. In this edge selection process, when the difference of occurrences of  $k$ -mer between the selected vertex and unselected vertices is less than a threshold, the current vertex is regarded as the end vertex and new start vertex is selected again. When the label of the selected vertex is the same as that of the current vertex as shown in Fig. 4 (c) (in the case of cycle), the current vertex is regarded as the end vertex. In many cases, a cycle is a part of a path as shown in Fig. 5. The vertices which are in the cycle are assigned to the new label in this case. Thus, a cycle is separated from the current scanning path.

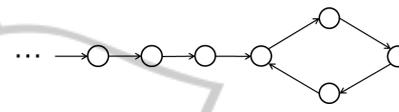


Figure 5: Example of a cycle which is a part of a path.

Furthermore, there is the case in which the selected vertex has been assigned to other label as shown in Fig. 6. In this case, all the vertices in the path which has the selected vertex are reassigned to the label of the current vertex.

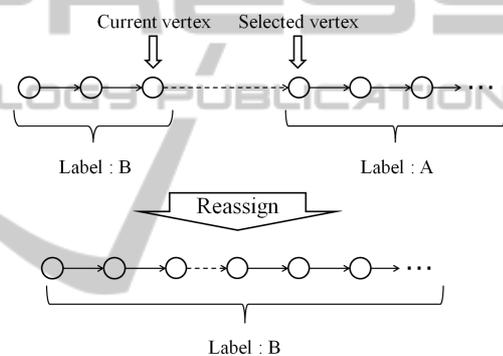


Figure 6: Example of the label reassignment.

## 2.4 Subgraph Connection and Contig Construction

To construct a longer path, subgraphs obtained by the process described in previous section are connected. The outline of the subgraph connection process is as follows. First, a subgraph with simple path is selected. The subgraphs with the longest path is selected from subgraphs which have not been selected.

However the subgraph with a simple cycle can be selected more than once. This subgraph is set to the start subgraph. Next, the subgraph in which the start vertex or the end vertex are connected to the end vertex or the start vertex of the selected subgraph are searched. The details of connection of the subgraph with a simple cycle are described in later in this section. This process is realized by checking  $k$ -mer integer described in the previous section. If such subgraph is found, the start (end) vertex is connected to the end (start) vertex, and the two subgraph are merged into a single subgraph. This graph expanding process is repeated while merging graph exists. If there are multiple subgraphs which can be connected, the subgraph with longer simple path is connected. Figure 7 shows an example of connecting subgraphs. The example in this figure is the case of left side connection. The same process is repeated on the right side.

In addition, the subgraph with a simple cycle is connected as shown in Fig. 8. In this figure, The vertex  $v_a$  is start vertex of subgraph  $G_{path}$  with a simple path. The vertex  $v_b$  is contained in subgraph  $G_{cycle}$

with a simple cycle. If the  $k$ -mers of the vertex  $v_a$  and  $v_b$  overlap each other, the vertex  $v_c$  which connected to  $v_b$  is checked. ( $v_c$  is also contained  $G_{cycle}$ .)  $v_b$  and  $v_c$  are regarded as the start( or end) vertex. Then, the  $G_{path}$  and  $G_{cycle}$  are merged. If the vertex  $v_c$  is connected to other vertex which is included in other subgraph, These subgraphs are merged again as shown in Fig. 8.

Edge between end vertex and start vertex was found.

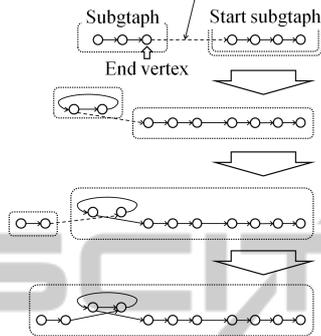


Figure 7: Example of subgraphs connection.

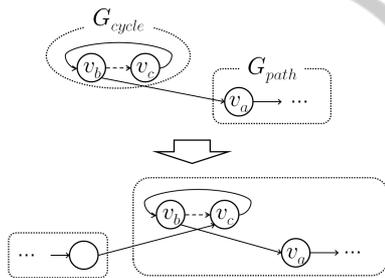


Figure 8: Example of the connection of subgraph with a simple cycle.

After the subgraph connection, the list of the vertices is obtained by tracing the path which is included in a subgraph. The contig is generated by merging  $k$ -mers which are referenced from the vertices as shown in Fig. 9. The final contigs are obtained by repeating this process for all subgraphs. In our experiments, the contigs which are longer than a threshold are output. In our experiments, the threshold was set to length of the reads.

### 3 EXPERIMENTAL RESULTS

To evaluate the performance of the proposed method, we compare the performance of our method with that of Velvet which is one of the most popular de novo assembler based on the de bruijn graph. *E.coli* K-12 strain MG 1655 for which the complete DNA sequence is known was used in our experiments. The

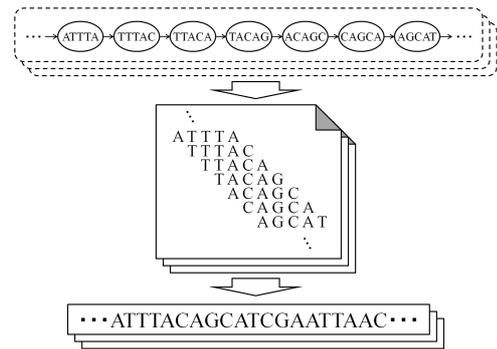


Figure 9: Generation of contigs.

sequence length is approximately 4.6Mbp. The real dataset with 35bp reads was made by a computer from the complete DNA sequence of *E.coli* K-12 strain MG 1655. Experiments were conducted using reads (35bp) that generated by the NGS from the genome. Experiments were run with the  $k$ -mer size of 19, 21, 23, 25, 27, 29, and 31. We assessed the maximum memory consumption, the running time, the contig length, and the accuracy of contigs.

Figure 10 and Fig. 11 show the maximum memory usage and the running time for each  $k$ -mer, respectively. As shown in Fig. 10, the memory usage of the proposed algorithm is one-third of that of Velvet. Therefore, the purpose of this research was achieved because the amount of memory was reduced greatly. Moreover the average running time of the proposed method was approximately 41% of that of Velvet. In the proposed method, the connection between vertices is checked each time when a path is traced. Thus, the running time to find the connection between vertices is increased. However, since the path tracing algorithms for resolving branches and cycles is very simple, the average running time was faster in the proposed method. In addition, the running time was shortened according to the increase of the  $k$ -mer size. On the other hand, the change of the  $k$ -mer size had hardly influence on the memory.

Table 2 shows the results of the contig quantity of the proposed method and Velvet. Maximum length, N50 length, the number of contigs, and total bases are shown in Table 2. The N50 length is defined as the length of the shortest contig such that the sum of contigs of equal length or longer is at least 50% of the total length of all contigs. Table 3 shows the results of the accuracy of the contigs. Percentage of the genome covered and average error rate are shown in Table 3. The maximum length of the proposed method was shorter than that of Velvet as shown in Table 2. Moreover, N50 length of the proposed method was shorter than that of Velvet. The main reason for these results is that the path tracing algorithm for re-

solving branches and cycles is very simple in the proposed method. Thus, the maximum length and N50 length would be better by modifying the path tracing algorithm. On the other hand, the error rate of the proposed method was lower than that of Velvet. The genome coverage was slightly lower than that of Velvet. These results indicate that there are not large differences in the contig quantity from path tracing algorithms.

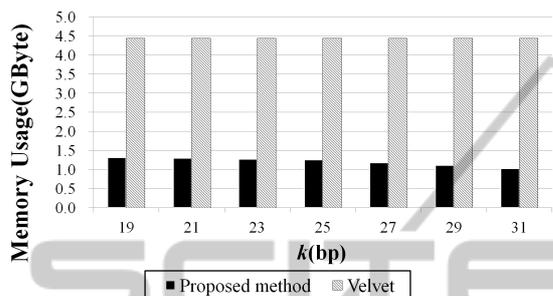


Figure 10: Comparison of maximum memory consumption.

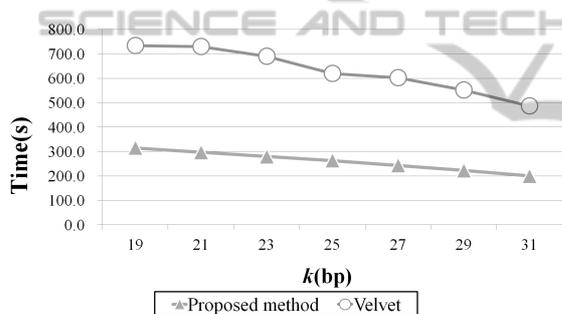


Figure 11: Comparison of running time.

Table 2: Comparison of quantity of contigs.

	Maximum length (bp)	N50 (bp)	# of contigs	Total (bp)
Proposed method	74,708	17,038	631	4,560,202
Velvet	81,421	22,870	754	4,544,229

Table 3: Comparison of precision of contigs.

	Genome covered (%)	Average error rate (%)
Proposed method	99.59	5.71
Velvet	99.89	7.29

## 4 CONCLUSIONS

In this paper, we propose an algorithm for huge scale de novo assembly with low memory usage. In our experiments using *E.coli* K-12 strain MG 1655, the results showed that maximum memory consumption of the proposed algorithm was one-third that of Velvet.

Furthermore, the running time of proposed method was also faster than that of Velvet. These results showed that the proposed method outperformed Velvet for the memory and the running time. On the other hand, contig quality obtained by the proposed method was slightly worse than that of Velvet. To improve the accuracy of the contigs, we need to modifying the path tracing algorithm in the future works.

## REFERENCES

Butler, J., MacCallum, I., Kleber, M., Shlyakhter, I. A., Belmonte, M. K., Lander, E. S., Nusbaum, C., and Jaffe, D. B. (2008). ALLPATHS: de novo assembly of whole-genome shotgun microreads. *Genome Res.*, 18(5):810–820.

Chevreaux, B., Pfisterer, T., Drescher, B., Driesel, A. J., Muller, W. E., Wetter, T., and Suhai, S. (2004). Using the miraEST assembler for reliable and automated mRNA transcript assembly and SNP detection in sequenced ESTs. *Genome Res.*, 14(6):1147–1159.

Hernandez, D., Francois, P., Farinelli, L., Osteras, M., and Schrenzel, J. (2008). De novo bacterial genome sequencing: millions of very short reads assembled on a desktop computer. *Genome Res.*, 18(5):802–809.

Jeck, W. R., Reinhardt, J. A., Baltrus, D. A., Hickenbotham, M. T., Magrini, V., Mardis, E. R., Dangl, J. L., and Jones, C. D. (2007). Extending assembly of short DNA sequences to handle error. *Bioinformatics*, 23(21):2942–2944.

Li, R., Zhu, H., Ruan, J., Qian, W., Fang, X., Shi, Z., Li, Y., Li, S., Shan, G., Kristiansen, K., Li, S., Yang, H., Wang, J., and Wang, J. (2010). De novo assembly of human genomes with massively parallel short read sequencing. *Genome Res.*, 20(2):265–272.

Miller, J. R., Delcher, A. L., Koren, S., Venter, E., Walenz, B. P., Brownley, A., Johnson, J., Li, K., Mobarry, C., and Sutton, G. (2008). Aggressive assembly of pyrosequencing reads with mates. *Bioinformatics*, 24(24):2818–2824.

Simpson, J. T., Wong, K., Jackman, S. D., Schein, J. E., Jones, S. J., and Birol, I. (2009). ABySS: a parallel assembler for short read sequence data. *Genome Res.*, 19(6):1117–1123.

Warren, R. L., Sutton, G. G., Jones, S. J., and Holt, R. A. (2007). Assembling millions of short DNA sequences using SSAKE. *Bioinformatics*, 23(4):500–501.

Zerbino, D. R. and Birney, E. (2008). Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res.*, 18(5):821–829.