

Effects of γ -aminobutyric acid on feed intake, growth performance and expression of related genes in growing lambs

D. M. Wang, B. Chacher, H. Y. Liu, J. K. Wang, J. Lin and J. X. Liu[†]

MoE Key Laboratory of Molecular Animal Nutrition, Institute of Dairy Sciences, College of Animal Sciences, Zhejiang University, Hangzhou 310058, P. R. China

(Received 17 January 2014; Accepted 31 July 2014; First published online 7 November 2014)

This study was conducted to investigate the effects of rumen-protected γ -aminobutyric acid (GABA) on feed intake, growth performance and expression of related genes in growing lambs. A total of 24 lambs weaned at age of 50 days were divided into four block of six based on their BW, six lambs within a block were allocated to three pairs, which were then assigned randomly to three treatments with addition of rumen-protected GABA at levels of 0, 70 or 140 mg/day for 6 weeks. Dry matter intake was recorded weekly in three consecutive days, and BW was recorded every two weeks. At the end of the trial, four lambs from each group were slaughtered, and duodenum and ileum mucosa were obtained for measurement of mRNA abundance of GABA receptor and cholecystokinin receptor. Dry matter intake was higher (P < 0.01) in the lambs fed 140 mg/day GABA than that in the control or 70 mg GABA-fed lambs. Average daily gain and nutrients digestibility were not different (P > 0.05) among treatments. Lambs fed 140 mg/day GABA had higher mRNA abundance of GABA-B receptor (P < 0.01) in duodenum mucosa. Serum CCK content was lower (P < 0.01) in lambs fed 140 mg/day GABA than that GABA may enhance feed intake by regulating GABA- and cholecystokinin-related genes.

Keywords: γ -aminobutyric acid, γ -aminobutyric acid-B receptor, cholecystokinin-2 receptor, feed intake, growing lambs

Implications

 γ -aminobutyric acid has shown to increase dry matter intake in both monogastric and ruminant animals. Addition of 140 mg/day rumen-protected γ -aminobutyric acid enhanced dry matter intake, probably by regulating mRNA abundance of cholecystokinin-2 and GABA-B receptors and then serum cholecystokinin content.

Introduction

Central injection of γ -aminobutyric acid (GABA) receptor agonists stimulates feed intake in sheep (Girard *et al.*, 1985). It is reported that baclofen (GABA-B agonist) may alleviate the inhibitory effect of exogenous peripheral cholecystokinin (CCK), a peptide hormone that plays important roles as gastrointestinal hormone and neurotransmitter, on dry matter intake (DMI) in rats (Ebenezer, 1995), indicating that GABA and CCK may have associative effects on DMI. Moreover, motility in duodenum can be inhibited by activating GABA-A receptor, or be activated by stimulating GABA-B receptor (Fargeas *et al.*, 1988). In contrast, motility in ileum can be activated by stimulating GABA-A receptor, or be restrained by activation of GABA-B receptor (Ong and Kerr, 1982). The reduced motility of gastrointestinal tracts may cause inhibited digestion and lower DMI (Xing and Chen, 2004).

Dietary GABA, instead of injection of GABA or its receptors, may increase DMI and milk protein yield in dairy cows, and the increased DMI seemed to be related to CCK metabolism (Wang *et al.*, 2013). However, it is questionable if dietary GABA may inhibit CCK synthesis of type I cells, a kind of specialized endocrine cells that produce and secrete hormones, in the duodenum and ileum. Thus, the objective of the current study was to investigate the effects of dietary rumenprotected GABA on DMI, average daily weight gain (ADG), serum variables (CCK and GABA) and mRNA abundance of the related genes in duodenum and ileum of growing lambs.

Material and methods

Animals, feeds and experimental design

A total of 24 male Huzhou lambs, weaned at the age of 50 days, with an initial BW of 13.5 kg (s.d. 2.09), were used. All the lambs were divided into four block of six based on their BW, six lambs within a block were allocated to

[†] E-mail: liujx@zju.edu.cn

three pairs, which were then assigned randomly to three treatments with addition of rumen-protected GABA at levels of 0, 70 and 140 mg/day, respectively. The rumenprotected GABA was used because the unprotected GABA was guickly degraded within the rumen (Wang et al., 2010) and non-protected GABA had little effect on feed intake and lactation performance in transition cows (Wang et al., 2013). The procedure for preparation of the rumen-protected GABA was described elsewhere (Wang et al., 2010). Each pen of two lambs was kept in an individual pen $(2.5 \times 2 \text{ m})$ and bedded on steel frame during the feeding trial lasting for 6 weeks. All the lambs were fed a total mixed ration (TMR) twice a day at 0700 and 1530 h, and had free access to drinking water. In brief, the TMR was prepared by cutting and blending the individual feed ingredients listed in Table 1, and then fed to the sheep in each pen. The daily amount of GABA was mixed with 1 g of corn and fed in two equal portions by top-dress feeding to individual sheep, to ensure that both lambs in the pen consumed the correct dose of GABA.

Sampling procedures and measurements

DMI and ADG. The amount of feed given was calculated to allow 10% orts. Feed offered and refused were recorded for three consecutive days (2^{nd} , 3^{rd} and 4^{th} days) weekly, to determine DMI. Nutrients compositions are listed in Table 1. The lambs were weighed on days 0, 14, 28 and 42 of the trial. Data were used to calculate ADG (mean of ADG for the period of days 0–14, 14–28 and 28–42). All measurements were conducted at 0630 h before feeding.

Composition of the TMR and feces. The TMR samples were collected from each feeding on 2^{nd} , 3^{rd} and 4^{th} days weekly. The samples were dried in an air-forced oven at 60°C for 48 h

 Table 1 Ingredients and composition of the experimental basal diet

Ingredients	DM basis (g/kg)
Bean curd residue	319
Distillers grains	223
Corn silage	110
Soybean straw	99.8
Corn	82.4
Wheat bran	73.4
Cottonseed meal	34.1
Barley	27.6
Premix ¹	21.0
Calcium carbonate	3.4
Salt	3.4
Calcium bicarbonate	2.2
Nutrient composition (g/kg DM)	
NDF	351
ADF	193
СР	162
AIA	16

DM = dry matter; AIA = acid insoluble ash.

¹Formulated to provide (per kilogram of DM): Co, 200 mg; Cu, 15 000 mg; I, 600 mg; Fe, 8000 mg; Mg, 100 000 mg; Mn, 30 000 mg; Se, 200 mg; Zn, 40 000 mg; vitamin A, 4 000 000 IU; vitamin D, 400 000 IU; vitamin E, 8000 IU. and then ground for later analysis of dry matter (DM, Association of Official Analytical Chemists, 1997, method 930.15) and CP (Association of Official Analytical Chemists, 1997, method 928.08). Contents of NDF and ADF were expressed inclusive of residual ash (Van Soest *et al.*, 1991; Association of Official Analytical Chemists, 1997, method 973.18). Apparent digestibilities of nutrients were estimated using acid insoluble ash (AIA) as an internal marker based on the concentration of AIA in the diet and feces, where AIA was determined. Fecal samples of each pen were collected on the 4th day biweekly, and then composited to be analyzed for chemical composition.

Serum variables. Blood samples were collected from the neck vein at 1100 h on days 14, 28 and 42 of the trial period, respectively, and then centrifuged at $3000 \times g$ for 10 min to collect serum. Serum samples were stored at -20° C for later analysis of contents of GABA and CCK as the methods described elsewhere (Wang *et al.*, 2013).

Slaughter trial and sampling

One animal of each pen was electrically stunned and slaughtered. Intestinal sections (3 cm for each section) from the duodenum and ileum were washed with double distilled water. The epithelium was separated from the underlying muscle layers and stored in liquid nitrogen until analyzed.

RNA extraction and determination of mRNA abundance

Total RNA was isolated from duodenum and ileum mucosa according to TRIzol reagent (Invitrogen, Life Technologies, Carlsbad, CA, USA). Reverse transcription reactions (20 μ l) consisted of 16 μ l total RNA, 1 μ l of RNAse inhibitor (Promega, Madison, WI, USA), 1 μ l dNTPs (Sigma, St. Louis, MO, USA) and 2 μ l of 5 \times M-MLV RT reaction buffer (Promega) were used to get cDNA. The optimal reverse transcription procedure was 20°C for 5 min, followed by 42°C for 60 min and then 70°C for 5 min. Reactions were immediately stopped by putting reaction system on ice. The cDNA was stored at -20°C for real-time PCR.

Real-time PCR was performed with ABI PRISM 7700 sequence detection system (ABI Biosystems, Foster City, CA, USA) according to optimized PCR protocols. A total volume of 20 µl PCR reaction system was composited by 10 µl SYBR Green PCR Master Mix, 2.0 µl primer (1.0 µl forward and 1.0 µl reverse), 2.0 µl cDNA template and 6 µl sterile superstilled water. The following protocol was employed for the PCR reaction: denature at 95°C for 3 min, 42 cycles of amplification (94°C for 30 s, 51°C for 30 s, 72°C for 60 s) and finally extension at 72°C for 7 min. The C_t value was recorded and the relative expression level ($2^{-\Delta\Delta C_t}$) was calculated for determination of related mRNA abundance. Primers used in the current study are given in Supplementary Table S1.

Statistical analysis

All data were analyzed using repeated measures by PROC MIXED model (SAS Release 6.12, 1988). Treatment, pen

Item	Rumen-protected GABA (mg/day)				<i>P</i> value	
	0	70	140	s.e.m.	Linear	Quadratic
DMI (g/day)	1146	1172	1248	14.2	<0.01	0.15
ADG (g/day)	284	282	310	15.6	0.22	0.45
Feed efficiency ¹	0.25	0.24	0.25	0.011	0.95	0.65
Digestibility (%)						
Dry matter	67.7	67.5	66.4	1.47	0.54	0.80
CP	63.3	63.5	64.5	1.53	0.65	0.85
NDF	55.1	54.6	55.0	1.47	0.99	0.81
ADF	38.7	37.6	37.4	0.91	0.41	0.76

Table 2 Effects of dietary rumen-protected γ -aminobutyric acid (GABA) on dry matter intake (DMI), average daily weight gain (ADG), feed efficiency and nutrients digestibility in growing lambs

¹Feed efficiency = ADG/DMI.

Table 3 Effects of dietary rumen-protected γ -aminobutyric acid (GABA) on the mRNA abundances of cholecystokinin (CCK), its receptors and GABA receptors in epithelium of duodenum and ileum in growing lambs

Item	Rumen-protected GABA (mg/day)				P value	
	0	70	140	s.e.m.	Linear	Quadratic
Serum content						
CCK (pg/ml)	684 ^A	650 ^A	412 ^B	25.1	< 0.01	< 0.01
GABA (mmol/l)	27.7	28.8	28.6	0.74	0.33	0.46
Gene expression						
ССК						
In duodenum	0.27	0.24	0.30	0.076	0.80	0.64
In ileum	0.28	0.24	0.25	0.062	0.39	0.93
CCK-1 receptor						
In duodenum	0.18	0.16	0.21	0.045	0.38	0.40
In ileum	0.30	0.27	0.29	0.052	0.91	0.69
CCK-2 receptor						
In duodenum	2.03 ^A	1.07 ^{AB}	0.77 ^B	0.161	< 0.01	0.15
In ileum	1.15	0.99	0.93	0.208	0.72	0.72
GABA-A receptor						
In duodenum	1.40	1.55	1.18	0.143	0.33	0.19
In ileum	1.10	0.97	0.95	0.198	0.96	0.82
GABA-B receptor						
In duodenum	0.46 ^B	0.43 ^B	0.94 ^A	0.051	< 0.01	0.01
In ileum	0.54	0.45	0.56	0.138	0.56	0.55

^{A,B}Values with different superscripts are significantly different (P < 0.01).

(sheep for variable at slaughter) within treatment, block and sampling time were included in the model as main effects and treatment × time was included as the interaction. Linear and quadratic effects of treatment were tested for DMI, ADG, serum contents of GABA and CCK, and gene expression related with GABA and CCK using orthogonal polynomial contrasts, respectively. Data are presented as the least squares mean, and differences were considered significant at P < 0.05.

Results and discussion

Addition of GABA increased DMI of the lambs linearly (P < 0.01, Table 2). The ADG and feed conversion efficiency did not differ among treatments (P > 0.05), though the lambs

fed 140 mg/day rumen-protected GABA grew faster (numerically, by 9.1%) than others. We acknowledge that low replication may account for insufficient power to detect differences in growth rate.

Dietary GABA had no effect on digestibility of DM, CP, NDF and ADF (P > 0.05, Table 2). The rumen degradation rate of rumen-protected GABA used in the current study was about 15% (Wang *et al.*, 2010). Thus, the released GABA in rumen may have been about 10.5 or 21 mg/day in lambs fed 70 or 140 mg/day rumen-protected GABA, respectively. Unprotected GABA had limited effects on rumen pH, volatile fatty acids (acetate, propionate and butyrate) and ammonia-nitrogen concentrations (unpublished). Thus, dietary rumen-protected GABA did not exert significant effect on nutrients digestibility. Serum CCK content decreased (linear, P < 0.01 and quadratic, P < 0.01), while serum GABA was not affected by GABA (P > 0.05, Table 3). The mRNA abundance of CCK in duodenum and ileum was not decreased (P > 0.05, Table 3), indicating that the decreased serum CCK contents may be attributed to the less CCK synthesis and release by type I cells in the mucosal epithelium of the systems other than the small intestines. The quadratic effect (P < 0.01) of dietary GABA on serum CCK content indicated that higher dietary GABA may be needed for inhibiting effect on CKK synthesis.

As shown in Table 3, addition of GABA did not affect mRNA abundances of all the related genes in ileum (P > 0.05), but decreased the mRNA abundance of CCK-2 receptor (linearly, P < 0.01) and increased the GABA-B receptor in duodenum (linear, P < 0.01 and quadratic, P = 0.01). Ong and Kerr (1982) observed that GABA-A and GABA-B receptors had different responses to type of GABA or their receptor agonists. In the current study, the GABA-B receptor in the duodenum seemed to be more sensitive to dietary GABA than GABA-A receptor did. Activation of GABA-B receptor by GABA-B agonist could increase food intake in rats (Ebenezer, 1995), while intake-inhibitory effect induced by CCK was highly related with activation of type A CCK-1 receptor in rats (Moran *et al.*, 1992). Thus, the depressed expression of CCK-2 receptor gene and up-regulated expression of GABA-B receptor gene in duodenum may contribute to the increased DMI in sheep induced by GABA in the current study. However, how dietary GABA regulated the CCK secretion by type I cells is unclear from the current work and warrants for further study.

Conclusion

Dietary addition of rumen-protected GABA linearly increased DMI but did not affect ADG in growing sheep. The increased DMI induced by higher level of GABA may be attributed to the enhanced expression of GABA-B receptor gene, and the decreased expression of CCK-2 receptor gene in duodenal mucosa.

Acknowledgments

This study was supported by the grant from the China Agriculture Research System (CARS-37). The authors thank Pangda Sheep Farm for providing and caring the sheep. All the members of Institute of Dairy Science, Zhejiang University (Zhejiang, China) are acknowledged for their assistance in sampling.

Supplementary material

To view supplementary material for this article, please visit http://dx.doi.org/10.1017/S1751731114002651

References

Association of Official Analytical Chemists 1997. Official methods of analysis, vol. 5, 16th edition. AOAC, Gaithersburg, MD, USA.

Ebenezer IS 1995. Intraperitoneal administration of baclofen increases consumption of both solid and liquid diets in rats. European Journal of Pharmacology 273, 183–185.

Fargeas MJ, Fioramonti J and Bueno L 1988. Central and peripheral action of GABAA and GABAB agonists on small intestine motility in rats. European Journal of Pharmacology 150, 163–169.

Girard CL, Seoane JR and Matte JJ 1985. Studies of the role of gammaaminobutyric acid in the hypothalamic control of feed intake in sheep. Canadian Journal of Physiology and Pharmacology 63, 1297–1301.

Moran TH, Ameglio PJ, Schwartz GJ and McHugh PR 1992. Blockade of type A, not type B, CCK receptors attenuates satiety actions of exogenous and endogenous CCK. The American Journal of Physiology 262, R46–R50.

Ong J and Kerr DI 1982. GABAA- and GABAB-receptor-mediated modification of intestinal motility. European Journal of Pharmacology 86, 9–17.

SAS 1988. SAS/STAT user's guide (Release 6.12). SAS Institute Inc., Cary, NC.

Van Soest PJ, Bobertson JB and Lewis BA 1991. Methods of dietary fiber, NDF and non-starch polysaccharides in relation to animal nutrition. Journal of Dairy Science 74, 3583–3597.

Wang DM, Liu HY, Wang C and Liu JX 2010. Preparation and stability of rumen-protected γ -aminobutyric acid. Chinese Journal of Animal Nutrition 22, 1451–1456.

Wang DM, Liu HY, Wang C, Liu JX and Ferguson JD 2013. Effects of rumenprotected gamma-aminobutyric acid on feed intake, performance and antioxidative status in transition cows. Livestock Science 153, 66–72.

Xing J and Chen JD 2004. Alterations of gastrointestinal motility in obesity. Obesity Research 12, 1723–1732.