LETTER 587

## Potential of (2E,7E)-Nonadienedioates in Asymmetric Synthesis: Construction of Homopipecolic Acid and an Aminoester Building Block for Peptide Nucleic Acids

Narciso M. Garrido,\* Alfonso G. Rubia, Carlos Nieto, David Díez

Departamento de Química Orgánica, Universidad de Salamanca, Plaza de los Caídos 1-5, 37008 Salamanca, Spain Fax +34(923)294574; E-mail: nmg@usal.es

Received 19 December 2009

Dedicated with respect and affection to Gerry Pattenden, an inspiring scientist, on the occasion of his 70th birthday

**Abstract:** A convenient, asymmetric synthesis of (R)-homopipe-colic acid methyl ester and an homochiral peptide nucleic acid (PNA) monomer building block are described, starting from the orthogonally disubstituted (2E,7E)-nonadienedioate. The approach involves stereoselective Michael monoaddition of (R)-N-benzyl-N- $\alpha$ -methylbenzylamide to the unsaturated ester as the key step, and subsequent transformation of the remaining double bond of the unsaturated acid.

Key words:  $\beta$ -amino acids, asymmetric synthesis, Michael additions, PNA, homopipecolic acid, orthogonally substituted dienedioate

Every synthetic route starts from a particular substrate that lends itself to the retrosynthetic scheme planned. The capacity of a substrate to participate in a wide range of synthetic pathways depends on the potential of their structure: cycles, chains and functional groups. Molecules with functional groups that react selectively are attractive from this point of view: the better the chemical orthogonality of the functional groups, the larger the spectrum of synthetic transformations possible and, consequently, the range of accessible targets.

Cyclic  $\beta$ -amino acids such as (R)-homopipecolic acid (Scheme 1) have a number of interesting features that have been used to develop synthons of natural products<sup>1</sup>

and key intermediates in β-lactam structures.<sup>2</sup> Synthetic oligonucleotides (Scheme 1; DNA/RNA) have been considered as potential gene-targeted therapeutic agents (antisense and antigene).<sup>3</sup> Peptide nucleic acids (PNAs) were first reported in 1991 as DNA mimics<sup>3c</sup> and, since this time, a vast number of studies have been reported covering their synthesis, properties and potential applications. Among the known oligonucleotide analogues, acyclic N-(2-aminoethyl)glycyl peptide nucleic acids (Scheme 1; PNA I) or those derived from base-containing  $\delta$ -amino acid derivatives4 (Scheme 1; PNA II), are found to be very good mimics of DNA/RNA. Within this area, a steadily growing group of analogues in which the sugarphosphate backbone is replaced by a polyamide backbone, is emerging, mainly as a consequence of the intriguing base-pairing properties of their prototype PNA. In this context, we envisaged the synthesis of the amino acid building block mononer 2 in PNA III.

We have demonstrated<sup>5</sup> the use of chiral lithium ( $\alpha$ -methylbenzyl)benzylamide [(R)-**3** or (S)-**3**] to initiate asymmetric conjugate addition cyclisation of octa-2,6-dienedioate and nona-2,7-dienedioate to generate chiral cyclopentane and cyclohexane derivatives **4** and **5**, respectively. The We have also developed strategies to stereoselectively obtain double- (**7**) and mono-addition (**6**, **8**, **9** and **10**) products (Scheme 2), where the Z-double bond

$$(R)\text{-homopipecolic acid} \begin{tabular}{ll} HO-P=O & H(OH) \\ HO-P=O &$$

## Scheme 1

SYNLETT 2010, No. 4, pp 0587–0590 Advanced online publication: 08.02.2010 DOI: 10.1055/s-0029-1219375; Art ID: D37209ST © Georg Thieme Verlag Stuttgart · New York 588 N. M. Garrido et al. LETTER

plays a crucial role on the (Z,E)-dienedioate as a vehicle for  $\gamma$ -deprotonation. We have proposed **9** to be an intermediate in an approach to **1**.5a

Here, as shown in the retrosynthetic analysis (Scheme 3), we focused on the potential of (2E,7E)-nonadienedioate 11 as an orthogonally functionalised starting material in which the groups show differing reactivity towards lithium (R)-N-benzyl-N- $\alpha$ -methylbenzylamide [(R)-3]. This selectivity is exploited synthetically by modifying the residual functionality to give either homopipecolic acid methyl ester (formerly synthesised using other protocols<sup>6</sup>) or to construct a thymine long-chain  $\beta$ -amino acid PNA monomer 2 for use in oligomerisation to form PNA. Both goals were developed in an enantiocontrolled way.

**Scheme 2** Reagents and conditions: (a) (*R*)-**3** (1.2 equiv), THF, -78 °C; (b) (*R*)-**3** (3 equiv), THF, -78 °C; (c) *t*-BuOK, *t*-BuOH.

The synthesis of homopipecolic methyl ester (Scheme 4) started with addition of lithium (R)-N-benzyl-N- $\alpha$ -methylbenzylamide [(R)-3] to the orthogonally functionalised substrate 11, to provide adduct 12 (de >95%)<sup>7</sup> stereoselectively (*vide infra*) in good yield, in accordance with the literature. Acid salt generation enriches electron-density on the conjugated olefin, averting nucleophilic attack at this centre. The next step required ozonolysis of 12, however, since reports in the literature recommended prior esterification, 12 was treated with TMSCHN<sub>2</sub> to provide the corresponding diester 13. Attempts at ozonolysis of 13 were unsuccessful, leading instead to decomposition of the starting material as a consequence of N-oxide

CO<sub>2</sub>Me
$$CO_2$$
H

Ph
 $CO_2$ Me
 $CO_2$ H

11

 $CO_2$ H

NHBoc

 $CO_2$ Me

 $CO_2$ H

 $CO_2$ H

 $CO_2$ H

 $CO_2$ H

 $CO_2$ H

 $CO_2$ H

 $CO_2$ Me

 $CO_2$ H

 $CO_2$ H

 $CO_2$ Me

 $CO_2$ Me

**Scheme 3** Proposed strategy for the synthesis of (*R*)-homopipecolic acid and monomers **1** and **2** for PNA synthesis

formation<sup>11</sup> provoking a Cope elimination. However, treatment of **13** with anhydrous HCl followed by ozonolysis and reduction with Me<sub>2</sub>S gave aldehyde **15**. Finally, hydrogenolytic debenzylation over Pearlmans catalyst induced cyclisation to the imine, which underwent reduction to (R)-homopipecolic methyl ester in situ {[ $\alpha$ ]<sub>D</sub><sup>26</sup> -3.6 (c 0.32, CHCl<sub>3</sub>); Lit.<sup>6c</sup> for the enantiomer [ $\alpha$ ]<sub>D</sub><sup>26</sup> +3.9 (c 0.64, CHCl<sub>3</sub>)} in 50% overall yield.

**Scheme 4** Reagents and conditions: (a) Lithium (*R*)-*N*-benzyl-*N*-α-methylbenzylamine [(*R*)-**3**; 3.6 equiv], THF, –78 °C; (b) TMSCHN<sub>2</sub>, benzene–MeOH (1:1), 30 min; (c) HCl (g); (d) O<sub>3</sub>, then Me<sub>2</sub>S; (e) Pd(OH)<sub>2</sub>/C, H<sub>2</sub> (4 atm), EtOAc.

Scheme 5 Reagents and conditions: (a) Pd(OH)<sub>2</sub>/C, H<sub>2</sub> (4 atm), Boc<sub>2</sub>O, EtOAc, 3 d; (b) BH<sub>3</sub>·THF, THF, 20 °C, 60 min; (c) CBr<sub>4</sub>, Ph<sub>3</sub>P, CH<sub>2</sub>Cl<sub>2</sub>, 45 min; (d) thymine, TBAI, K<sub>2</sub>CO<sub>3</sub>, DMF, 70 °C, 6 h.

The route towards the PNA monomer started from 12 (Scheme 5). Reacting a mixture of 12, Boc<sub>2</sub>O and Pearlmans catalyst in ethyl acetate for three days under hydrogen (4 atm), accomplished a one-pot amine-debenzylation, Boc-reprotection and hydrogenation of the olefin in 62% yield. Subsequent selective reduction of the carboxylic acid with borane, followed by treatment with CBr<sub>4</sub>/PPh<sub>3</sub>, and finally, treatment with thymine, K<sub>2</sub>CO<sub>3</sub>, and TBAI in refluxing DMF<sup>12</sup> provided the target compound 2.<sup>13</sup> However, the poor nucleophilicity of thymine resulted in a relatively low yield in the final displacement (38%).

In summary, we have achieved the synthesis of two valuable products as important building blocks: (R)-homopipecolic methyl ester and a PNA-monomer containing a long-chain  $\beta$ -amino acid backbone. Both products were elaborated in a divergent fashion starting from (2E,7E)-nonadienedioate monoester 11, which is a readily accessible bifunctional substrate that exhibits orthogonal behaviour towards aza-Michael stereocontrolled addition of chiral lithium ( $\alpha$ -methylbenzyl)benzylamide. The residual functionality can then undergo a range of possible synthetic transformations, demonstrating the power of this protocol.

## Acknowledgment

The authors are grateful for financial support from the Spanish MICINN (EUI2008-00173), MEC (CTQ2009-11172/BQU), the FSE and Junta de Castilla y León (Spain): (SA001A09) and excellence GR-178. The authors also thank Dr. A. M. Lithgow for work on the NMR spectra and Dr. César Raposo for the mass spectra. C.N. thanks Junta de Castilla y León for a FPI doctoral fellowship.

## **References and Notes**

- (a) Back, T. G.; Hamilton, M. D. Org. Lett. 2002, 4, 1779.
   (b) Morley, C.; Knight, D. W.; Share, A. C. J. Chem. Soc., Perkin Trans. 1 1994, 2903.
- (2) Avenoza, A.; Busto, J. H.; Cativiela, C.; Corzana, F.; Peregrina, J. M.; Zurbano, M. M. J. Org. Chem. 2002, 67, 598; and references cited therein.
- (3) (a) Nielsen, P. E. Peptide Nucleic Acids: Protocols and Applications; Horizon Biosience: Norfolk, 2004, 318.
  (b) Nielsen, P. E.; Hyrup, B. Bioorg. Med. Chem. 1996, 1, 5.
  (c) Nielsen, P. E.; Egholm, M.; Berg, R. H.; Buchardt, O. Science 1991, 1497.
- (4) Saviethri, D.; Leumann, Ch.; Scheffold, R. Helv. Chim. Acta 1996, 79, 288.
- (5) (a) Garrido, N. M.; Díez, D.; Domínguez, S. H.; Sanchez, M. R.; García, M.; Urones, J. G. Molecules 2006, 11, 435.
  (b) Urones, J. G.; Garrido, N. M.; Díez, D.; El Hammoumi, M. M.; Domínguez, S. H.; Casaseca, J. A.; Davies, S. G.; Smith, A. D. Org. Biomol. Chem. 2004, 2, 364. (c) Garrido, N. M.; El Hammoumi, M. M.; Díez, D.; García, M.; Urones, J. G. Molecules 2004, 9, 373. (d) Urones, J. G.; Garrido, N. M.; Díez, D.; Domínguez, S. H.; Davies, S. G. Tetrahedron: Asymmetry 1999, 10, 1173. (e) Urones, J. G.; Garrido, N. M.; Díez, D.; Domínguez, S. H.; Davies, S. G. Tetrahedron: Asymmetry 1997, 8, 2683.
- (6) (a) Davies, S. G.; Fletcher, A. M.; Roberts, P. M.; Smith, A. D. Tetrahedron 2009, 65, 10192. (b) Davies, S. G.; Díez, D.; Domínguez, S. H.; Garrido, N. M.; Kruchinin, D.; Price, P. D.; Smith, D. Org. Biomol. Chem. 2005, 3, 1284.
  (c) Chippindale, A. M.; Davies, S. G.; Iwamoto, K.; Parkin, R. M.; Smethurst, C. A. P.; Smith, A. D.; Rodriguez-Solla, H. Tetrahedron 2003, 59, 3253. (d) O'Brien, P.; Porter, D. W.; Smith, N. M. Synlett 2000, 1336. (e) Kato, Y.; Wakabayashi, T.; Watanabe, K. Synth. Commun. 1977, 7, 239
- (7) Analysis of the crude product by <sup>1</sup>H NMR (400 MHz) confirmed it to be diastereomerically pure as no trace was found of any other stereoisomer. An ee >95% is consistent with the high optical purity of the lithium amide used.
- (8) Davies, S. D.; Smith, A. D.; Price, P. D. *Tetrahedron: Asymmetry* **2005**, *16*, 2833; and references cited therein.

590 N. M. Garrido et al. LETTER

(9) (a) Prior, W. A.; Giamalva, D.; Church, D. F. J. Am. Chem. Soc. 1983, 105, 6858. (b) Prior, W. A.; Giamalva, D.; Church, D. F. J. Am. Chem. Soc. 1985, 107, 2793.

- (10) Analysis of the crude product by  $^{1}H$  NMR (400 MHz) confirmed it to be diastereomerically pure as no trace was found of any other stereoisomer. The (2S, $\alpha R$ ) diastereisomer has been prepared by non-stereoselective monoaddition of  $\alpha$ -methylbenzylamine to (2E,8E)-decadienedioate followed by treatment with benzyl chloride. See ref. 5d
- (11) Hanessian, S.; Snacéau, J. Y.; Chemla, P. Tetrahedron 1995, 51, 6669.
- (12) Lenzi, A.; Reginato, G.; Taddei, M. *Tetrahedron Lett.* **1995**, 36, 1713.
- (13) Typical procedure: A suspension of thymine (108.6 mg, 0.861 mmol), TBAI (34.4 mg, 0.086 mmol) and K<sub>2</sub>CO<sub>3</sub> (59.6 mg; 0.431 mmol) in DMF (5 mL) was stirred for 30 min, then heated to 70 °C for 30 min. Bromide 18 (17 mg,

0.04 mmol) was added and the resulting mixture was stirred for 6 h at 70 °C. Then the mixture was cooled to 0 °C, filtered through Celite® and the filter pad was washed with EtOAc. The filtrate was washed with H<sub>2</sub>O, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent was removed. Purification of the crude product by flash chromatography (hexane–Et<sub>2</sub>O, 1:4) provided **2** (8 mg, 38%) as an oil. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 1.20-1.57$  (m, 10 H, H-4, H-5, H-6, H-7, H-8), 1.42 [s, 9-H, C(CH<sub>3</sub>)<sub>3</sub>], 1.92 (s, 3 H, Me- $C5'),\, 2.50\ (m,\, 2\ H,\, H\text{-}2),\, 3.65\text{-}3.70\ (m,\, 2\ H,\, H\text{-}9),\, 3.67\ (s,\, H,\, H\text{-}1),\, 3.67\ (s,\, H,\, H\text{-}2),\, 3.67\ (s,\, H,\, H\text{-}2),$ 3 H, OCH<sub>3</sub>), 3.89 (m, 1 H, H-3), 4.93 (d, J = 8.7 Hz, 1 H,NH), 6.97 (s, 1 H, H6'), 8.22 (s, 1 H, H-3'). IR (neat): 3365,  $2931, 2857, 1736, 1712, 1483, 1366, 1166, 1094 \text{ cm}^{-1}.$ <sup>13</sup>C NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 12.59, 26.12, 26.48, 28.60, 29.10, 32.14, 34.71, 39.44, 47.62, 48.70, 51.87, 77–79, 110.78, 140.67, 150.93, 164.33, 172.38. HRMS: m/z calcd for C<sub>20</sub>H<sub>33</sub>N<sub>3</sub>O<sub>6</sub>: 434.2261; found: 434.2260.