

Effects of excess metabolizable protein on ovarian function and circulating amino acids of beef cows: 2. Excessive supply in varying concentrations from corn gluten meal

T. C. Geppert^{1a}, A. M. Meyer², G. A. Perry³ and P. J. Gunn^{1†}

¹Department of Animal Science, Iowa State University, Ames, IA 50011, USA; ²Division of Animal Sciences, University of Missouri, Columbia, MO 65211, USA; ³Department of Animal Sciences, South Dakota State University, Brookings, SD 57007, USA

(Received 10 February 2016; Accepted 8 August 2016; First published online 9 September 2016)

In the dairy industry, excess dietary CP is consistently correlated with decreased conception rates. However, amount of excess CP effects on reproductive function in beef cattle is largely undefined. The objective of this experiment was to determine the effects of excess metabolizable protein (MP) supplementation from a moderately abundant rumen undegradable protein (RUP) source (corn gluten meal: 62% RUP) on ovarian function and circulating amino acid (AA) concentrations in beef cows consuming low quality forage. Non-pregnant, non-lactating beef cows (n = 16) were allocated by age, BW and body condition score (BCS) to 1 of 2 isocaloric supplements designed to maintain BW for 60 days. Cows had ad libitum access to corn stalks and were individually offered a corn gluten meal-based supplement daily at 125% (MP125) or 150% (MP150) of National Research Council (NRC) MP requirements. After a 20-day supplement adaptation period, cows were synchronized for ovulation. After 10 days of synchronization, follicular growth was reset with gonadotropin releasing hormone. Daily thereafter, transrectal ultrasonography was performed to diagram ovarian follicular waves, and blood samples were collected for hormone, metabolite and AA analyses. After 7 days of observation of estrus, corpus luteum (CL) size was determined via ultrasound. Data were analyzed using the MIXED procedures of SAS. No differences ($P \ge 0.21$) in BW and BCS existed throughout the study; however, plasma urea N at ovulation was greater (P = 0.04) in MP150. Preovulatory ovarian follicle size at dominance, duration of dominance, size at spontaneous luteolysis, length of proestrus and wavelength were not different ($P \ge 0.11$) between treatments. However, ovulatory follicles were larger (P = 0.04) and average antral follicle count was greater (P = 0.01) in MP150 than MP125. Estradiol concentration and ratio of estradiol to ovulatory follicle volume were not different due to treatment ($P \ge 0.25$). While CL volume 7 days post-estrus was greater (P < 0.01) in MP150 than MP125, circulating progesterone 7 days post-estrus and ratio of progesterone to CL volume were not different ($P \ge 0.21$). Total AA were not different ($P \ge 0.76$) at study initiation or completion; however, as a percent of total AA, branched-chain AA at ovulation were greater (P = 0.02) in MP150. In conclusion, supplementation of CP at 150% of NRC MP requirements from a moderately undegradable protein source may enhance growth of the ovulatory follicle and subsequent CL compared with MP supplementation at 125% of NRC MP requirements.

Keywords: branched-chain amino acids, beef cow, follicle, plasma urea nitrogen, protein

Implications

Beef cows offered metabolizable protein (MP) at 150% of requirements from a moderately abundant, rumenundegradable protein source had increased ovulatory follicle diameter, antral follicle counts (AFC) and corpus luteum (CL) development compared with 125% MP. These data indicate that increasing concentrations of excess CP around the time of ovulation do not negatively impact reproductive function in beef cows as previously hypothesized. To the contrary, some aspects of reproduction may be enhanced by excess CP, which may be linked to altered amino acid (AA) profiles in the bloodstream; however, further research is necessary to determine which specific AAs should be fed to improve reproductive performance.

Introduction

When high protein feedstuffs are supplemented as an energy source to beef cows, total dietary protein may likely exceed

^a Present address: South Dakota State University Extension, 1800 E. Spruce St., Mitchell, SD 57301, USA.

[†] Email: pgunn@iastate.edu

150% of National Research Council (NRC) requirements (Gunn *et al.*, 2014c). This is of particular concern as excess dietary protein has been associated with reduced fertility in dairy cattle (Butler *et al.*, 1996) as abundance of ammonia and urea in the reproductive tract alters uterine pH (Elrod and Butler, 1993) making a less favorable environment for embryo development. However, more recent studies have demonstrated that supplementation of distillers grains as an energy source, resulting in excess dietary protein, may be beneficial to reproductive performance of primiparous beef heifers (Gunn *et al.*, 2014c) and pregnancy rates of beef cows (Gunn *et al.*, 2014a).

More specifically, when moderately rumen undegradable protein (RUP) source (62% RUP) was offered to equal ~150% of NRC MP requirements ovulatory follicle diameter of non-lactating beef cows were enhanced (Gunn et al., 2014b; Geppert et al., 2016). However, research is warranted to determine if amount of excess MP from a RUP feedstuff differentially affects ovarian function of beef cows. Therefore, the objective of this experiment was to determine if the amount of excess dietary protein (125% MP v. 150% MP) from a moderately rumen undegradable feedstuff affected ovarian function of beef cows consuming processed corn stalks. We hypothesized that increasing the amount of excess MP would enhance ovulatory follicle diameter, CL development, hormone profile and alter AA profile around the time of ovulation, compared with excess MP at a decreased supplementation rate.

Material and methods

Animals and diets

All protocols and procedures used were approved by the Iowa State University Institutional Animal Care and Use Committee. The project was conducted at Zumwalt Station Research Unit in Ames, Iowa from May 2014 to July 2014. In order to evaluate the effects of excess amounts of MP from a moderately undegradable protein source on ovarian function around ovulation, 16 non-pregnant, non-lactating, cycling, Angus and Angus-Simmental multiparous beef cows were used. Cows were stratified by age $(6.36 \pm 2 \text{ years})$, body condition score (BCS; 4.93 ± 0.34 (1 = emaciated, 9 = obese; Wagner *et al.*, 1988)) and BW (552 ± 36 kg) and allotted to 1 of 2 isocaloric dietary supplements differing in amount of excess MP inclusion and fed at a rate of 1 kg/ animal per day (Table 1). All cows were offered ad libitum access to low quality forage (processed corn stalks; 6% CP and 80% NDF) and individually supplemented once daily with a moderately undegradable protein supplement (corn aluten meal: 62% RUP) for 60 days (Figure 1). Diets were formulated so that when supplements were combined with estimated ad libitum corn stalk consumption, total dietary MP would be: (1) 125% (MP125) or (2) 150% (MP150) of requirements (NRC, 2000). Diet formulation was based on near-infrared spectroscopy feedstuff analysis before study initiation (Dairyland Laboratories Inc., Arcadia, WI, USA) and designed to maintain similar cow BW and BCS throughout **Table 1** Supplement provided to beef cows consuming ad libitum corn stalks¹

	Treatment ²		
Item	MP125	MP150	
DM intake (kg/day)			
Corn silage	0.25	0.25	
Corn gluten meal ³	0.31	0.67	
Cracked corn	0.32	-	
Mineral	0.11	0.11	
Calculated nutrient intake of supplement			
CP (kg/day)	0.26	0.43	
Rumen undegradable protein (kg/day)	0.14	0.25	
Net energy for maintenance (Mcal/kg)	1.62	1.74	
Net energy for gain (Mcal/kg)	1.12	1.13	

DM = dry matter.

¹Corn stalk nutrient analysis (% DM basis): 51% TDN; 6% CP; 50% ADF; 80% NDF. When combined with estimated corn stalk intake, total dietary metabolizable protein (MP) intake for MP125 and MP150 = 570 and 680 g/day, respectively. Requirement for 590 kg cow (average starting weight of cows enrolled in study) = 455 g/day for maintenance (NRC, 2000).

²Treatment included *ad libitum* access to corn stalks, followed by daily supplementation of MP at 125% (MP125) or 150% (MP150) of MP requirements for non-pregnant, non-lactating multiparous beef cows.

³Nutrient analysis (% DM basis): 67% CP, 62% rumen undegradable protein, 2.4% fat.

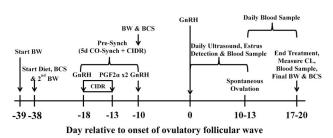


Figure 1 Experimental design for dietary treatments (MP125 = 125% metabolizable protein (MP) com gluten meal; MP150 = 150% com gluten meal outlining daily tasks and data collected relative to onset of ovulatory follicular wave in beef cows. Protein supplementation and *ad libitum* corn stalks began on day–38 and were offered until day 37, for a total of 60 days. BCS = body condition score; CIDR = controlled intravaginal drug releasing device; CL = corpus luteum; GnRH = gonadotropin releasing hormone; $PGF_{2\alpha} = prostaglandin F_{2\alpha}$.

the study. Although diets were not isonitrogenous, balancing diets using MP also ensured that the rumen degradable protein (RDP) requirements were met or exceeded for cows in both treatments.

Cows were allowed *ad libitum* access to processed corn stalks (51% total digestible nutrients (TDN); 50% ADF; 80% NDF and 6% CP) in a pen setting (three pens). Corn stalk intake was estimated using the following formula (NRC, 2000) for non-pregnant beef cows: dry matter intake = ((Shrunk $BW^{0.75} \times (0.04997 \times net energy for maintenance^2 + 0.03840)/$ net energy for maintenance) × (temperature adjustment) × (mud adjustment)), where adjustments for temperature and mud were 1 and 1, respectively. Although corn stalk refusal was unable to be accurately quantified due to feeding system, corn stalk delivery was similar across all pens and exceeded estimated intake.

Geppert, Meyer, Perry and Gunn

Daily individual supplementation took place at 0800 h inside a facility where 10 side-by-side stanchions were located. All cows from one treatment (n = 8) were restrained in stanchions and allowed to consume supplement individually. Cows were allowed adequate time to consume supplement while daily samples and measurements were being recorded. After supplement consumption and data collection (<1 h), cows were released from stanchions and commingled in for corn stalk consumption.

Performance characterization

Initial BCS was assessed on the day before trial initiation and was recorded as the average of scores from two trained investigators throughout all time points. In addition, initial BW was recorded as the average BW measured on the day before and the first day of the trial, before supplement delivery. Additional BCS and BW were taken once monthly, with BW being averaged on 2 consecutive days before treatment delivery when cows had restricted access to corn stalks for 12 h before BW collection to minimize variation in gut fill. Final BCS was taken on the last day of supplementation, and final BW was taken before supplementation on the day before and the last day of the study.

Pre-synchronization

Treatment experimental design is outlined in Figure 1. After 20 days of initiating dietary treatments, cows were synchronized with the 5-day CO-synch + controlled intravaginal drug release (CIDR) protocol. At synchronization initiation cows were administered 100 µg of gonadotropin releasing hormone (GnRH, Cystorelin; Merial Limited, Duluth, GA, USA) and received a CIDR (Zoetis Animal Health, Florham Park, NJ, USA). After 5 days, the CIDR was removed, 25 mg prostaglandin $F_{2\alpha}$ (PGF_{2 α}, Lutalyse; Zoetis Animal Health, New York, NY, USA) was administered and an Estrotect heat detection aid (Rockway Inc., Spring Valley, WI, USA) was placed on the tailhead of each cow. A second 25-mg dose of $PGF_{2\alpha}$ was administered 8 h after CIDR removal to ensure complete luteolysis of the young, accessory CL induced by the first administration of GnRH (Bridges et al., 2012). Estrus detection was performed for 72 h following CIDR removal by trained personnel. Cows were monitored for visual estrus twice daily for 30 min each at 0630 and 1700 h. After 72 h of CIDR removal and initial $PGF_{2\alpha}$ injection, all cows received 100 µg GnRH, and heat detection aid activity was recorded and removed.

Ovarian follicular wave characterization

After 10 days of synchronization protocol completion, $100 \mu g$ of GnRH was administered to initiate growth of a new follicular wave. Beginning at GnRH administration and daily thereafter at 0800 h, cows were subject to transrectal ultrasonography (Ibex Portable Ultrasound, variable MHz linear array transducer; E.I. Medical Imaging, Loveland, CO, USA) for complete characterization of a single follicular wave. Location and size of all antral follicles $\geq 3 \text{ mm}$ in diameter were recorded each day by drawing sketches of

each ovary. Transrectal ultrasound examinations were performed by the same investigator for the duration of the ultrasound period, and estrus was detection was performed twice daily by trained personnel at 0630 and 1700 h with the aid of heat detector patches (Rockway Inc.). All follicle measurements were made using the internal caliper function of the ultrasound, with final reported follicle diameters being the average of the greatest two cross-sectional perpendicular measurements of the follicle.

Day of dominance and size at dominance were categorized on the day which the largest growing follicle was at least 8 mm in diameter and at least 1 mm larger in diameter than any other growing follicle of the same wave (Ginther *et al.*, 1997). The dominant follicle was documented as the largest growing follicle in the wave of interest, and the secondary follicle was the second largest growing follicle in the same wave. Duration of dominance was retrospectively determined as the number of days from attainment of dominance until day of ovulation. Daily ovarian ultrasound ended upon successful ovulation as confirmed by disappearance of the ovulatory follicle from the wave of interest, preceded by visual display of estrus.

When estrus and subsequent ovulation were both confirmed, ultrasound measurements were halted until 7 days post-estrus, when ultrasound was performed once again to measure the total CL volume. Volume of the CL was determined using the internal caliper function of the ultrasound and formula for a rotary ellipsoid (V = 4/3 Π *ab*², where *a* = longitudinal axis and *b* = transverse axis). If lacuna(e) was present, the volume was calculated as previously stated and then subtracted from the total CL volume. After final CL measurement, dietary treatments ended.

Day of spontaneous luteolysis during the ovulatory follicle wave was retrospectively determined via radioimmunoassay (RIA) and defined as the day on which circulating progesterone concentrations were <50% of the concentration on the previous day (Ginther *et al.*, 2007). This also allowed ovulatory follicle size at spontaneous luteolysis and growth from luteolysis to ovulation to be determined. Length of proestrus was also retrospectively determined as the hours between spontaneous luteolysis and expression of estrus. Wave emergence was tracked by tracing the dominant follicle back to a cohort of follicles ≤ 4 mm in diameter, and wave length was calculated as total days from wave emergence to ovulation. AFC were totaled daily and all days were averaged to calculate average AFC of the entire wave.

Plasma analyses

During the 20-day dietary adaptation period, coccygeal blood samples were collected at time of feeding twice weekly (Monday and Thursday), and plasma was stored for later analysis for plasma urea nitrogen (PUN) concentration. During the ultrasound period until final CL measurement 7 days post-estrus, coccygeal blood samples were taken daily for plasma analysis of PUN, estradiol-17 β and progesterone concentrations. Approximately 4 ml of blood was collected via coccygeal venipuncture in a 6 ml EDTA vacutainer (10.8 mg

of EDTA; BD Vacutainer; Becton, Dickinson and Co., Franklin Lakes, NJ, USA) and immediately placed on ice. Samples were centrifuged at $1750 \times g$ for 25 min at 4°C, and plasma was recovered and transferred into 5 ml polystyrene tubes and frozen at -20° C until respective analyses were conducted.

Plasma samples collected during the ultrasound period were analyzed for circulating progesterone concentrations to determine day of spontaneous luteolysis and subsequent CL progesterone secretion. Concentrations were determined using a commercially available RIA kit (Coat-A-Count; Siemens Healthcare Diagnostics Inc., Los Angeles, CA, USA). Across three assays, the average intra-assay CV was 1.2%, with the inter-assay CV from pooled samples containing 0.59 and 5.82 ng/ml, was 9.8% and 6.7%, respectively. Average sensitivity across the assays were 0.14 ng/ml (95% confidence interval). Plasma samples collected from the day of spontaneous luteolysis until ovulation were analyzed for circulating concentrations of estradiol-17 β via RIA at South Dakota State University following methods described by Perry and Perry (2008). Across three assays, the average intra-assay CV was 4.5% and the average inter-assay CV from a pooled serum sample containing 7.9 pg/ml was 8%. The average sensitivity across three assays was 0.49 pg/ml (95% confidence interval).

Plasma samples were analyzed for PUN using a commercially available assay (Urea Nitrogen Procedure No. 0580; Stanbio Laboratory, Boerne, TX, USA). Samples and standards were prepared, loaded into a 96-well plate and read on an Eon Microplate Spectrophotometer at 520 nm (BioTek Instruments Inc., Winooski, VT, USA). Across eight assays, the average intra-assay CV was 5.3%, with an inter-assay CV from pooled serum containing 7.49 mg/dl of urea N was 10.9%. Preprandial blood samples taken at treatment initiation and around time of estrus (day 15) were analyzed for circulating AA concentrations at North Dakota State University (Fargo, ND, USA) via methods of ultra performance liquid chromatography (Lemley *et al.*, 2013).

Statistical analysis

Data were analyzed using the MIXED procedure of SAS 9.3 (SAS Institute Inc., Cary, NC, USA), with the main effect of treatment and experimental unit of animal. For ovarian follicular wave characteristics, hormone and metabolite analyses, the covariates of age and individual change in BW were included in the model as covariates. The MIXED procedures of SAS for REPEATED measures were utilized to analyze ovulatory follicle growth, progesterone and PUN concentrations. The covariance structures compound symmetry, heterogeneous compound symmetry, autoregressive order one, unstructured and ante-dependence were compared. The covariance structure resulting in the smallest Bayesian information criterion was used for the final analyses. The effects of treatment and day, as well as the interaction of treatment and day were analyzed in the model. Length of time from visual display of estrus to final CL measurement was also run as a covariate in the repeated analyses; however, it was removed from the model due to

insignificance (P > 0.10). The simple effects within day were determined using the slice function of SAS. The correlation procedure of SAS was used to determine relationships among variables of interest. For all analyses, significance was noted at $P \le 0.05$, with tendencies identified at P > 0.05 and ≤ 0.10 . It should be noted two cows from the MP125 treatment and three cows from the MP150 treatment failed to reach progesterone concentrations >1 ng/ml after ovulation; therefore, all data from those five cows were removed from the study.

Results

Performance and ovulatory follicle wave characteristics

Performance characteristics of BW and BCS are presented in Table 2. As designed, BW and BCS were similar ($P \ge 0.74$) between treatments at initiation of the study. Furthermore, with use of isocaloric dietary treatments, final BW and BCS were not different ($P \ge 0.73$) at the conclusion of the trial. Ovulatory follicle wave and CL characteristics are presented in Table 3. The preovulatory follicle parameters of wave length, day of luteolysis; size at dominance, duration of dominance and growth post-dominance as well as size at spontaneous luteolysis, growth post-luteolysis and duration of proestrus were not different ($P \ge 0.11$) between treatments. However, cows from the MP150 treatment had areater maximum ovulatory follicle diameter (P = 0.04) and average AFC (P = 0.01) compared with the MP125 treatment. In addition, there was a significant (P < 0.02) treatment × day interaction for ovulatory follicle size postemergence, as cows from MP150 treatment had larger $(P \leq 0.05)$ ovulatory follicles on days 10 and 11 postemergence than MP125 cows (Figure. 2). Lastly, CL volume 7 days post-estrus was greater (P < 0.01) in MP150 than MP125 treatment cows. Ovulatory follicle size was positively correlated to CL volume (P = 0.03, r = 0.64).

Plasma analyses

Circulating estradiol concentrations at luteolysis, as well as peak concentration before onset of estrus were not different

 Table 2 Effects of excess amounts of metabolizable protein (MP)

 supplementation on BW and body condition score in beef cows

	Treat	ment ¹		
ltem	MP125	MP150	SEM	<i>P</i> -value
BW (kg)				
Initial	549	557	16.96	0.74
Final	561	551	19.57	0.73
Body condition score ²				
Initial	4.95	4.90	0.16	0.84
Final	4.84	4.83	0.11	0.99

¹Treatment included *ad libitum* access to corn stalks, followed by daily supplementation of MP at 125% (MP125) or 150% (MP150) of MP requirements for non-pregnant, non-lactating multiparous beef cows.

²Scale of 1 = emaciated to 9 = obese, Wagner *et al.* (1988).

ltem	Treatment ¹			
	MP125	MP150	SEM ²	<i>P</i> -value
Ovulatory follicle size at dominance ³ (mm)	8.31	8.53	0.32	0.65
Dominance duration ⁴ (days)	6.65	6.62	1.27	0.99
Ovulatory follicle growth post-dominance ⁵ (mm)	4.37	6.81	0.94	0.11
Dominant follicle size at luteolysis ⁶ (mm)	10.30	12.10	0.92	0.25
Dominant follicle growth post-luteolysis (mm)	2.08	3.31	0.68	0.29
Proestrus duration ⁷ (h)	56.68	63.31	9.38	0.63
Ovulatory follicle diameter (mm)	12.60	15.28	0.73	0.04
Maximum secondary follicle diameter (mm)	7.90	7.92	0.69	0.99
Follicular wave length (days)	10.73	10.73	0.87	0.99
Total ovarian antral follicle count (AFC)	13.72	16.68	0.62	0.01
Corpus luteum volume 7 days post-estrus (cm ³)	1.17	6.08	0.56	<0.01

 Table 3 Effects of excess amounts of metabolizable protein (MP) supplementation on ovulatory follicle wave and corpus luteum characteristics in beef cows

¹Treatment included *ad libitum* access to corn stalks, followed by daily supplementation of metabolizable protein (MP) at 125% (MP125) or 150% (MP150) of requirements for non-pregnant, non-lactating multiparous beef cows.

²Greater SEM presented (MP125: n = 6; MP150: n = 5).

³Dominance obtained when largest growing follicle was at least 1 mm larger than any other growing follicle and at least 8 mm in diameter. ⁴Period between attainment of dominance until ovulation.

⁵Growth of ovulatory follicle between dominance and ovulation.

⁶Luteolysis defined as first day on which circulating progesterone concentrations were <50% of previous day concentration.

⁷Period between luteolysis and expression of estrus.

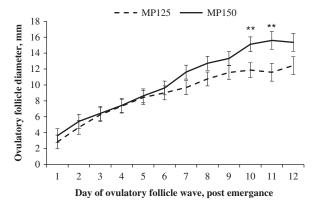


Figure 2 Effect of excess amounts of metabolizable protein (MP) supplementation (MP125 = 125% and MP150 = 150% of requirements) on ovulatory follicle diameter in beef cows. A treatment × day interaction (P < 0.02) was observed. *P*-values for treatment and day were 0.13 and <0.001, respectively. Days on which ovulatory follicle diameter was significantly different between treatments ($P \leq 0.05$) denoted with **.

between dietary treatments ($P \ge 0.51$; Table 4). Furthermore, when the ratio of estradiol to ovulatory follicle volume was assessed, no difference between treatments existed (P = 0.98). In addition, circulating progesterone and the ratio of progesterone to CL volume were not different ($P \ge 0.21$) due to dietary treatment. Peak concentrations of estradiol were not related to dominant follicle size (P = 0.21); however, progesterone concentrations 7 days post estrus tended to be positively associated with CL volume (P = 0.07; r = 0.56). Initial PUN concentrations were not different (P = 0.17; Table 4) between treatments. However, there was a treatment × day interaction detected (P < 0.01; Figure. 3), as PUN concentrations were greater (P < 0.05) in MP150 compared with MP125 cows after onset of the ovulatory follicular wave.

AA concentrations at the time of ovulation are located in Table 5. At trial initiation before protein supplementation, total AA, as well as total essential AA and non-essential AA were similar ($P \ge 0.81$; data not shown) between dietary treatments. Also at trial initiation, when samples were analyzed as a percent of total AA, data was not different (P > 0.14) between treatments. At ovulation, plasma samples showed no difference ($P \ge 0.77$) in total circulating AA or total essential and non-essential AA concentrations between treatments. However, circulating total phenylalanine tended to be greater (P = 0.09) in the MP150 than MP125 treatment. In addition, when expressed as a percent of total AA, branched-chain amino acids (BCAA) were greater (P = 0.02) in MP150 compared with MP125.

Discussion

The objective of this study was to determine if MP fed at 125% of MP requirements from a moderately undegradable source would have similar effects on beef cow ovarian function as supplementation at 150%. Diets containing increased CP have historically resulted in elevated circulation of urea N. This may be of concern as PUN concentrations above 19 mg/dl have been associated with suppressed conception and pregnancy rates in dairy cows (Butler *et al.*, 1996). However, feeding CP at 150% of NRC requirements has not been shown to result in PUN exceeding 19 mg/dl in beef cattle (Rusche *et al.*, 1993; Gunn *et al.*, 2014c). Although PUN varies with feeding and collection time, these

	Treatment ¹			
Item	MP125	MP150	SEM ²	<i>P</i> -value
Peak estradiol-17 β (pg/ml)	5.86	6.97	1.10	0.51
Estradiol-17 β : ovulatory follicle volume (pg/ml per mm)	0.46	0.46	0.08	0.98
Progesterone 7 days post-estrus (ng/ml)	2.23	3.39	0.63	0.25
Progesterone: corpus luteum volume (ng/ml per•cm ³)	1.70	0.36	0.65	0.21
Pre-treatment PUN (mg/dl)	2.20	2.86	0.32	0.17
PUN at ovulation (mg/dl)	5.94	8.59	0.82	0.04

Table 4 Effects of excess amounts of metabolizable protein (MP) supplementation on estradiol, progesterone and plasma urea nitrogen concentrations in beef cows

PUN = plasma urea nitrogen.

¹Treatment included ad libitum access to corn stalks, followed by daily supplementation of MP at 125% (MP125) or 150% (MP150) of requirements for non-pregnant, non-lactating multiparous beef cows. ²Greater SEM presented (MP125: n = 6; MP150: n = 5).

MP125 MP150 12 10 Plasma Urea Nitrogen, mg/dI 8 6 4 2 0 -38 -35 -31 -28 -24 -20 -16 -13 -9 -5 -2 1 5 8 12 15 19 22 Day relative to onset of ovulatory folliclular wave

Figure 3 Effect of excess amounts of metabolizable supplementation (MP; MP125 = 125% and MP150 = 150% of requirements) on circulating plasma urea N (PUN) concentrations in beef cows. There was a treatment × day interaction (P < 0.001) observed. P-values for treatment and day were 0.02 and <0.001, respectively. Days on which PUN was different between treatments ($P \le 0.05$ and P < 0.10) are denoted by ** and *, respectively.

studies showed no negative associations between PUN and reproductive parameters.

Few studies have explored the relationship between excess RUP, PUN and reproductive parameters in beef cattle. However, it was recently elucidated that excess CP from a more concentrated rumen undegradable feedstuff (150% RUP via corn gluten meal) enhanced ovulatory follicle size of non-lactating females compared with 150% MP from a more degradable source (soybean meal; Geppert et al., 2016). Gunn et al. (2014b) also reported enhanced ovulatory follicle diameter, AFC and estradiol concentrations from cows fed 150% MP when compared with 100% MP.

In the present study, supplementation of RUP at 150% of requirements resulted in greater PUN circulation than 125% after 40 days of supplementation, which was expected. Nonetheless, while PUN concentrations were elevated compared with baseline in both treatments after supplementation, circulating PUN was still relatively low compared with concentrations known to be associated with decreased reproductive performance which is likely due to preprandial collection of blood samples for PUN analysis.

Table 5 Effects of excess amounts of metabolizable protein (MP) supplementation on circulating amino acid concentrations at ovulation in beef cows

	Treat	ment ¹		
ltem	MP125	MP150	SEM ²	<i>P</i> -value
Amino acid (µmol/l)				
Total	1490	1526	84.59	0.76
Non-essential	869.18	898.57	50.45	0.77
Glycogenic	946.13	968.27	54.26	0.77
Ketogenic	189.72	198.54	17.23	0.71
Branched-chain	610.59	668.83	25.81	0.16
Essential	620.71	636.51	46.09	0.81
Histidine	49.90	48.79	2.69	0.77
Arginine	48.16	53.58	7.24	0.59
Threonine	36.47	33.37	4.59	0.63
Lysine	58.71	51.05	9.81	0.58
Methionine	17.44	17.07	1.59	0.87
Valine	143.90	147.88	7.72	0.71
Isoleucine	69.75	66.31	6.36	0.70
Leucine	131.01	147.49	10.18	0.26
Phenylalanine	45.80	51.43	2.18	0.09
Tryptophan	19.57	19.53	1.95	0.99
Amino acid (% of total)				
Non-essential	58.52	58.27	1.57	0.91
Glycogenic	63.69	36.40	1.46	0.89
Ketogenic	12.64	12.99	0.69	0.71
Branched-chain	41.04	44.05	0.80	0.02
Essential	41.48	41.73	1.57	0.91
Histidine	3.36	3.21	0.12	0.40
Arginine	3.21	3.48	0.38	0.61
Threonine	2.42	2.15	0.22	0.40
Lysine	3.84	3.31	0.51	0.47
Methionine	1.16	1.11	0.06	0.53
Valine	9.67	9.76	0.38	0.86
Isoleucine	4.63	4.35	0.26	0.44
Leucine	8.80	9.68	0.52	0.25
Phenylalanine	3.10	3.38	0.13	0.15
Tryptophan	1.29	1.30	0.10	0.98

¹Treatment included *ad libitum* access to corn stalks, followed by daily supplementation of MP at 125% (MP125) or 150% (MP150) of requirements for nonpregnant, non-lactating multiparous beef cows. ²Greater SEM presented (MP125: n = 6; MP150: n = 5).

Ovulatory follicle wave characteristics

Previous studies reported that excess CP supplementation yielded larger ovulatory follicles in lactating cows (Lents et al., 2008) and first-calf heifers (Gunn et al., 2014c). In agreement with the current study, recent research in our lab (Gunn et al., 2014b; Geppert et al., 2016) reported that excess CP sourced from corn gluten meal, a moderately abundant RUP source, supplemented at 150% of NRC MP requirements enhanced ovulatory follicle size compared with diets containing 100% of MP requirements from gluten meal and 150% of MP requirements from soybean meal (64% RDP) in non-lactating, non-pregnant beef cows, respectively. Ovulatory follicle size has been associated with enhanced pregnancy success, as spontaneous ovulation of large follicles in beef heifers were more likely to result in a pregnancy than females ovulating smaller follicles (Perry et al., 2007). However, pregnancy success may not be correlated with follicle size when spontaneous ovulation occurs (Perry et al., 2005). The disconnect between spontaneous ovulation beef heifers v. cows could be due to differences in age and stage of development of the females. Thus, non-lactating beef cows having reduced nutrient demands when compared with developing heifers may be able to utilize excess MP to increase ovarian follicular development.

Greater ovulatory follicle diameter could potentially be due to the increased proportion of bypass protein in MP150 *v*. MP125, increasing circulating insulin concentrations which have been previously associated with enhanced dominant follicle size (Ciccioli *et al.*, 2003), and working through growth hormone (GH) and IGF-1 pathways to control follicular growth (Silva *et al.*, 2009). However, this is speculative as insulin, GH and IGF-1 was not measured in the present study.

It has been previously demonstrated, as is here, that excess dietary RUP supplementation yields greater average AFC (Gunn et al., 2014b). Greater AFC may result in improved fertility due to improved oocyte competence compared with lower AFC cows (Ireland et al., 2011). However, the exact mechanism of how AFC is mediated is unknown. Cushman et al. (2014) suggested that AFC may be predetermined before birth, and may also be influenced through developmental programming. Yet, to our knowledge this study and research by Gunn et al. (2014b) are the first reports suggesting that postnatal diet may alter AFC. While initial AFC measurements were not taken in either of the previously mentioned studies, similar AFC results in two separate experiments is intriguing. Because the primordial germ cell population is established before birth, enhancing follicle recruitment or AFC may shorten the reproductive lifespan of a female by depleting the follicular reserve more quickly. Yet, as AFC begins decreasing in bovine females after 5 years of age (Cushman et al., 2009), perhaps excess protein fed at 150% MP requirements may be beneficial to follicular recruitment in females past peak AFC production by increasing IGF-1 and FSH receptors, aiding the transition from secondary to antral follicles (Silva et al., 2009).

Cows induced to ovulate large follicles have been reported to develop larger CL compared with cows induced to ovulate small follicles (Vasconcelos *et al.*, 2001). As the CL develops from the granulosa and theca cells of the ovulatory follicle, larger follicles tend to have more follicular cells and greater estradiol exposure which influence the developing luteal tissue after ovulation (Atkins *et al.*, 2013). Smaller CL development in MP125 cows was expected after ovulating smaller follicles. However, normal development of the CL is also dependent on blood flow which is also associated with AA (arginine) through nitric oxide production (Tamanini, *et al.*, 2003). Although we hypothesized that differences in CL volume may be due to circulating AA, the lack of differences in arginine and its metabolites (citrulline and ornithine, data not shown) do not support this hypothesis.

Hormones and amino acids

Hormones. Given differing ovulatory follicle sizes, it was surprising that we did not observe a difference in estradiol concentrations due to treatment. One would expect greater circulating estradiol concentrations at ovulation of the larger follicles (Vasconcelos et al., 2001; Perry et al., 2005; Busch et al., 2008); however, similar proestrus intervals between treatments may have mitigated differences in estradiol production (Bridges et al., 2010). In addition, as size of the ovulatory follicle has been associated with subsequent CL volume (Vasconcelos et al., 2001), a correlation between CL volume and circulating progesterone has also been observed in beef cows (Echternkamp et al., 2009). However, regardless of follicle and CL size in the present study, no difference in subsequent circulating progesterone was observed, potentially due to altered vascularization of the subsequent CL (Tamanini and De Ambrogi, 2004) and hormone priming (McNatty et al., 1981) altering luteal formation in both treatments (Busch et al., 2008).

Despite the lack of differences in estradiol and progesterone concentrations, the decreased production of essential hormones in both treatments is slightly concerning as certain concentrations must be present in order to successfully maintain normal estrous cycles and attain pregnancy. As a positive correlation between estradiol concentration at estrus and subsequent progesterone concentration at 7 days postestrus exists (Jinks *et al.*, 2012), it was not surprising that hormone concentrations followed similar patterns in the current study. Other authors have observed no alterations to hormone production after excess RUP supplementation (Gunn *et al.*, 2014b; Geppert *et al.*, 2016); therefore, suppressed estradiol and progesterone concentrations regardless of treatment in this study may have been due to seasonal effects of heat stress as reviewed by Rensis and Scaramuzzi (2003).

Amino acids. Recent research in our lab has demonstrated that excess CP is capable of shifting circulating AA concentrations in beef cows consuming low quality forage (Geppert *et al.*, 2016). In particular, increased leucine in the aforementioned study and BCAA in the current study may influence ovarian function. Specifically, greater BCAA in circulation has been shown to enhance ovarian tissue synthesis through the mechanistic target of rapamycin activation pathway (Greiwe *et al.*, 2001), as observed in bovine luteal tissue (Zhang *et al.*, 2011). However, as this is one of this first reports on the relationship between AA and ovarian function in beef cows, further investigation into these and other mechanisms is warranted.

Conclusion

In summary, feeding excess RUP at 125% and 150% MP resulted in differential effects on ovarian reproductive parameters. While circulating hormone concentrations were similar between treatments, ovulatory follicle and CL parameters were enhanced by excess RUP at 150% of MP requirements compared with 125%. Furthermore, cows fed 150% MP had greater BCAA circulation, which may have a potential role in the increased growth of ovulatory follicles and CL development. Therefore, supplementation of CP at 150% of NRC MP requirements from a moderately undegradable protein source appears to enhance growth of the ovulatory follicle and the subsequent CL compared with MP supplementation at 125% of requirements. While effects of excess RUP at the ovarian level were assessed here, a continuation of this study should be conducted to further investigate the impacts of excess MP from a moderately RUP on overall fertility.

Acknowledgments

The authors wish to thank David Bruene, Sam Lei, Marsha Kapphahn, Allie Lundberg, Cassie Elliott and Shannon Ure for their assistance with animal care, data collection and lab analyses.

References

Atkins JA, Smith MF, MacNeil MD, Jinks EM, Abreu FM, Alexander LJ and Geary TW 2013. Pregnancy establishment and maintenance in cattle. Journal of Animal Science 91, 722–733.

Bridges GA, Ahola JK, Brauner C, Cruppe LH, Currin JC, Day ML, Gunn PJ, Jaeger JR, Lake SL, Lamb GC, Marquezini GHL, Peel RK, Radunz RE, Stevenson JS and Whittier WD 2012. Determination of the appropriate delivery of prostaglandin $F_{2\alpha}$ in the five-day CO-Synch + controlled intravaginal drug release protocol in suckled beef cows. Journal of Animal Science 90, 4814–4822.

Bridges GA, Mussard ML, Burke CR and Day ML 2010. Influence of length of proestrus on fertility and endocrine function in female cattle. Animal Reproduction Science 117, 208–215.

Busch DC, Atkins JA, Bader JF, Schafer DJ, Patterson DJ, Geary TW and Smith MF 2008. Effect of ovulatory follicle size and expression of estrus on progesterone secretion in beef cows. Journal of Animal Science 86, 553–563.

Butler WR, Calaman JJ and Beam SW 1996. Plasma and milk urea nitrogen in relation to pregnancy rate in lactating dairy cattle. Journal of Animal Science 74, 858–865.

Ciccioli NH, Wettemann RP, Spicer LJ, Lents CA, White RF and Keisler DH 2003. Influence of body condition at calving and postpartum nutrition on endocrine function and reproductive performance of primiparous beef cows. Journal of Animal Science 81, 3107–3120.

Cushman RA, Allan MF, Kuehn LA, Snelling WM, Cupp AS and Freetly HC 2009. Evaluation of antral follicle count and ovarian morphology in crossbred beef cows: Investigation of influence of stage of the estrous cycle, age and birth weight. Journal of Animal Sciene 871, 971–1980. Cushman RA, McNeel AK and Freetly HC 2014. The impact of cow nutrient status during the second and third trimesters on age at puberty, antral follicle count, and fertility of daughters. Livestock Science 162, 252–258.

Echternkamp SE, Cushman RA and Allan MR 2009. Size of ovulatory follicles in cattle expressing multiple ovulations naturally and its influence on corpus luteum development and fertility. Journal of Animal Science 87, 3556–35568.

Elrod CC and Butler WR 1993. Reduction of fertility and alteration of uterine pH in heifers fed excess ruminally degradable protein. Journal of Animal Science 71, 694–701.

Geppert TC, Meyer AM, Perry GA and Gunn PJ 2016. Effects of excess metabolizable protein on ovarian function and circulating amino acids of beef cows: 1. Excessive supply from corn gluten meal or soybean meal. Animal, published online doi:10.1017/S1751731116001889.

Ginther OJ, Kot K, Kulick LJ and Wiltbank MC 1997. Emergence and deviation of follicles during the development of follicular waves in cattle. Theriogenology 48, 75–87.

Ginther OJ, Silva LA, Araujo RR and Beg MA 2007. Temporal associations among pulses of 13,14-Dihydro-15-keto-PGF2alpha, luteal blood flow, and luteolysis in beef cattle. Biology of Reproduction 76, 506–513.

Greiwe JS, Kwon G, McDaniel ML and Semenkovich CF 2001. Leucine and insulin activate p70 S6 kinase through different pathways in human skeletal muscle. American Journal of Physiology, Endocrinoloy, and Metabolism 281, E466–E471.

Gunn PJ, Lemenager RP and Bridges GA 2014a. Using corn stover and dried distillers grains with soluble to conserve stockpiled forages and improve reproductive performance and progeny growth in fall-calving beef cows. Professional Animal Scientist 30, 215–224.

Gunn PJ, Lemenager RP and Bridges GA 2014b. Excess rumen undegradable protein alters parameters of reproductive function in beef cows. Animal Industry Report AS 660, ASL R2852. Retrieved on 5 May 2015 from http://lib.dr.iastate. edu/ans_air/vol660/iss1/17.

Gunn PJ, Schoonmaker JP, Lemenager RP and Bridges GA 2014c. Feeding excess crude protein to gestating and lactating beef heifers: Impact on parturition, milk composition, ovarian function, reproductive efficiency and pre-weaning progeny growth. Livestock Science 167, 435–448.

Ireland JJ, Smith GW, Scheetz D, Jimenez-Krassel F, Folger JK, Ireland JLH, Mossa F, Lonergan P and Evans ACO 2011. Does size matter in females? An overview of the impact of the high variation in the ovarian reserve on ovarian function and fertility, utility of anti-Mllerian hormone as a diagnostic marker for fertility and causes of variation in the ovarian reserve. Reproduction, Fertility and Development 23, 1–14.

Jinks EM, Smith MF, Atkins JA, Pohler KG, Perry GA, MacNeil MD, Roberts AJ, Waterman RC, Alexander LJ and Gear TW 2012. Preovulatory estradiol and the establishment and maintenance of pregnancy in suckled beef cows. Journal of Animal Science 91, 1176–1185.

Lemley CO, Camacho LE, Meyer AM, Kapphahn M, Caton JS and Vonnahme KA 2013. Dietary melatonin supplementation alters uteroplacental amino acid flux during intrauterine growth restriction in ewes. Animal 7, 1500–1507.

Lents CA, White FJ, Ciccioli NH, Wettemann RP, Spicer LJ and Lalman DL 2008. Effects of body condition score at parturition and postpartum protein supplementation on estrous behavior and size of the dominant follicle in beef cows. Journal of Animal Science 86, 2549–2556.

McNatty KP, Gibb M, Dobson C, Thurley DC and Findlay JK 1981. Changes in the concentration of gonadotrophic and steroidal hormones in the antral fluid of ovarian follicles throughout the oestrous cycle of the sheep. Australian Journal of Biological Science 34, 67–80.

National Research Council (NRC) 2000. Nutrient requirements of beef cattle, 7th revised edition. National Academy Press, Washington, DC, USA.

Perry GA and Perry BL 2008. Effect of preovulatory concentrations of estradiol and initiation of standing estrus on uterine pH in beef cows. Domestic Animal Endocrinology 34, 333–338.

Perry GA, Smith MF, Lucy MC, Green JA, Parks TE, MacNeil MD, Roberts AJ and Geary TW 2005. Relationship between follicle size at insemination and pregnancy success. Proceedings of the National Academy of Science of the USA 102, 5268–5273.

Perry GA, Smith MF, Roberts AJ, MacNeil MD and Geary TW 2007. Relationship between size of the ovulatory follicle and pregnancy success in beef heifers. Journal of Animal Science 85, 684–689.

Rensis FD and Scaramuzzi RJ 2003. Heat stress and seasonal effects on reproduction in the dairy cow – a review. Theriogenology 60, 1139–1151.

Geppert, Meyer, Perry and Gunn

Rusche WC, Cochran RC, Corah LR, Stevenson JS, Harmon DL, Brandt RT Jr. and Minton JE 1993. Influence of source and amount of dietary protein on performance, blood metabolites, and reproductive function of primiparous beef cows. Journal of Animal Science 71, 557–563.

Silva JR, Figueiredo VJR and Van Den Hurk R 2009. Involvement of growth hormone (GH) and insulin-like growth factor (IGF) system in ovarian folliculogenesis. Theriogenology 71, 1193–1208.

Tamanini C, Basini G, Grasselli F and Tirelli M 2003. Nitric oxide and the ovary. Journal of Animal Science 81 (suppl. 2), E1–E7.

Tamanini C and De Ambrogi M 2004. Angiogenesis in developing follicle and corpus luteum. Domestic Animal Reproduction 39, 206–216.

Vasconcelos JLM, Sartori R, Oliveira HN, Guenther JG and Wiltbank MC 2001. Reduction in size of the ovulatory follicle reduces subsequent luteal size and pregnancy rate. Theriogenology 56, 307–314.

Wagner JJ, Lusby KS, Oltjen JW, Kakestraw J, Wettermann RP and Walters LE 1988. Carcass composition in mature Hereford cows: estimation and effect on daily metabolizable energy requirement during winter. Journal of Animal Science 66, 603–612.

Zhang Z, Debing Y, Dingzhong Y and Zhengchao W 2011. Activation of PI3K/mTOR signaling pathway contributes to induction of vascular endothelial growth factor by hCG in bovine developing luteal cells. Animal Reproduction Science 125, 42–48.