

RESEARCH ARTICLE

# Linking systemic angiogenic markers to synovial vascularization in rheumatoid arthritis

Agathe Leblond<sup>1</sup>, Sonia Pezet<sup>1</sup>, Anne Priscille Trouvin<sup>2</sup>, Muriel Elhai<sup>1,2</sup>, Virginie Gonzalez<sup>1</sup>, Yannick Allanore<sup>1,2</sup>, Jérôme Avouac<sup>1,2\*</sup>

**1** Université Paris Descartes, Sorbonne Paris Cité, INSERM U1016 and CNRS UMR8104, Institut Cochin, Paris, France, **2** Université Paris Descartes, Sorbonne Paris Cité, Service de Rhumatologie A, Hôpital Cochin, Paris, France

\* [jerome.avouac@cch.aphp.fr](mailto:jerome.avouac@cch.aphp.fr)



## Abstract

### Background

Neoangiogenesis is a crucial event to promote the development of the hyperplastic proliferative pathologic synovium in Rheumatoid arthritis (RA). Ultrasound (US) is sensitive for detection of power Doppler (PD) vascularization.

### Objective

To explore the associations between a set of complementary circulating angiogenic markers and a comprehensive US assessment in patients with RA.

### Patients and methods

Serum levels of eight angiogenic markers were measured by quantitative ELISAs in a total of 125 patients with RA, who were all systematically assessed in parallel by PDUS, performed on 32 joints.

### Results

Serum levels of soluble Vascular Cell Adhesion Molecule-1 (sVCAM-1) and Tie-2 were more likely to be increased in patients with synovial hyperemia detected on at least one joint (Power Doppler grade  $\geq 1$ ). sVCAM-1, Tie-2 and Angiostatin concentrations gradually increased together with the grade of the semiquantitative PDUS scale and concentrations of these three markers were markedly increased in patients with moderate to marked hyperemia (Power Doppler grade 2 and 3). Levels of sVCAM-1, Tie-2, and Angiostatin correlated with a global arthritis sum score, defined by the sum of the semiquantitative PDUS scores for all joints examined. Levels of Tie-2 and Placenta Growth Factor (PIGF) were associated with PDUS features indicating residual disease activity.

## OPEN ACCESS

**Citation:** Leblond A, Pezet S, Trouvin AP, Elhai M, Gonzalez V, Allanore Y, et al. (2018) Linking systemic angiogenic markers to synovial vascularization in rheumatoid arthritis. PLoS ONE 13(9): e0203607. <https://doi.org/10.1371/journal.pone.0203607>

**Editor:** Chi Zhang, University of Texas Southwestern Medical Center, UNITED STATES

**Received:** March 28, 2018

**Accepted:** August 3, 2018

**Published:** September 6, 2018

**Copyright:** © 2018 Leblond et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information files.

**Funding:** Grant supports were supplied by: “Appel d’Offre Interne de l’Hôpital Cochin – MERRI Performances des Etablissements de Santé dans la Promotion de la Recherche” and “Arthritis Foundation”.

**Competing interests:** The authors have declared that no competing interests exist.

## Conclusion

Our results support the relevance of measuring serum levels of vascular markers to evaluate the intensity and extent of synovial vascularization. Angiogenic markers, and particularly Tie-2, could be a valuable surrogate of active synovitis and their place in relation to PDUS in clinical practice deserve further investigation.

## Introduction

Rheumatoid arthritis (RA) is the most common cause of chronic inflammatory arthritis with a prevalence ranging from 0.5% to 1% of the adult population worldwide [1, 2]. It is an autoimmune disease with a complex pathogenesis implicating innate and adaptive immunity together with angiogenesis [3].

The synovium is the primary site of RA-related inflammatory process, with infiltration of blood-derived inflammatory cells at the interface between cartilage and bone. One of the most noticeable signs of synovitis is the amount of synovial vascularization related to angiogenesis and vasculogenesis, which are critical for synovial proliferation and invasiveness. This invasive and destructive front promotes the development of bone and cartilage destruction.

Formation of new vessels consists of several complementary processes including activation, proliferation and migration of endothelial cells. This phenomenon is mediated by the differential regulation of angiogenic mediators and inhibitors [4–6].

Neoangiogenesis leads, together with inflammation-induced vasodilation of preexistent blood vessels, to increased blood flow in affected joints. Previous studies showed the considerable ability of highly sensitive power Doppler Ultrasound (PDUS) to improve the scoring of synovitis by detecting extended synovial vasculature [7, 8]. In addition, persistent synovial vascularity, assessed by power Doppler ultrasound (PDUS), has been linked to increased risk of disease flares and structural joint damages [9, 10].

Only scarce data are currently available regarding correlations between systemic angiogenic activity, measured by angiogenic factors in the serum, and the amount of local synovial vascularization measured by Doppler ultrasound [11]. Previous studies were characterized by limited sample size and assessed a restricted set of angiogenic markers, focusing mainly on VEGF. Moreover, the relationship between residual synovial vascularization and circulating angiogenic marker levels in patients with low disease activity (LDA) has not been explored in detail so far.

Acute phase reactants (ESR and CRP) are largely used in clinical practice but may not continually reflect the persistence of synovitis. Thus, novel biomarkers that exhibit higher associations with inflammation and angiogenesis in RA patients are warranted. The objective of the present study was to seek for associations between synovial vascularity assessed by PDUS and a panel of 8 serum vascular markers, reflecting different angiogenic processes, such as endothelial cell activation, proliferation, survival, growth and migration, as well as vessel maturation and stabilization.

## Patients and methods

### Study design

Cross-sectional study.

## Inclusion criteria

Consecutive patients with RA, >18 years of age, fulfilling the 1987 American College of Rheumatology (ACR) or the 2010 ACR/European League Against Rheumatism (EULAR) classification for RA, who have attended partial hospitalization program at the department of Rheumatology A, Cochin Hospital, over a 11-month period (May 2016 to April 2017) [12]. All included patients agreed to participate in the study after informed consent, which was recorded in the medical source file. The protocol and the informed consent document have received Institutional Review Board/Independent Ethics Committee (IRB/IEC) approval before initiation of the study (“Comité de Protection des Personnes” Paris Ile de France I).

## Data collection from RA patients

History-taking, physical examination, laboratory tests, and review of medical files were systematically performed to collect data from RA patients. Current and past medication use was obtained from information provided by patients, and based on the review of medical records. RA disease activity was assessed using the Disease Activity Score based on evaluation of 28 joints (DAS28) [13], using Erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) [14]. Surrogate measures of cumulative disease activity were health status and joint destruction. Health status was measured by the self-administered Stanford Health Assessment Questionnaire (HAQ). Systematic hand and foot x-rays were performed to measure joint destruction, defined by the presence of erosions.

## Laboratory tests

Laboratory studies were obtained in RA patients on the morning of hospital visit. They included complete blood cell count, Westergren erythrocyte sedimentation rate (ESR, considered elevated above 28mm hour<sup>-1</sup>), CRP concentration (considered elevated if above 10mg/l), and serum creatinine concentration. Rheumatoid factor (RF) and second-generation anti-cyclic citrullinated peptide (anti-CCP2) antibodies were detected by enzyme-linked immunosorbent assay (ELISA).

## Ultrasonography (US) assessment

All of the power Doppler and greyscale ultrasound (PDUS) examinations were performed using a multiplanar technique in accordance with the EULAR guidelines for musculoskeletal ultrasound in rheumatology. PDUS was performed, the day blood samples were collected, by two rheumatologists trained in musculoskeletal US who were blinded to clinical evaluations (APT and ME). Consensus between rheumatologists was obtained before the beginning of the study on both the technique and the US findings, reported in a standardized form. The equipment was a 7–15 MHz linear array transducer (Toshiba Aplio). Power Doppler settings were standardized with a pulse repetition frequency of 750Hz, a gain of 50–53 dB and a low wall filter. US examination was performed on 32 (16 paired) joints of both hands (MCPs 1–5 and PIPs 1–5), both wrists (radio-ulnar, medio-carpal and radio-carpal) and both forefeet (MTPs 1–5).

The presence of hypoechoic synovial hyperplasia (SH) and joint effusion (JE), both assessed using greyscale, and of synovial vascularization, assessed using power Doppler (PD), was scored using semiquantitative scales. The presence of synovitis (SH and PD, without JE) was scored for each joint according to the semiquantitative OMERACT-EULAR-US composite PDUS scale, giving a score of 0–3 for each joint (0 = absence, no synovial hyperemia, 1 = mild, hyperemia in less than 1/3 of the synovial surface area; 2 = moderate, hyperemia in less than 2/

3 of the synovial surface area; 3 = marked, in more than 2/3 of the synovial surface area). A global synovitis score, derived from the Global OMERACT-EULAR Synovitis Score (GOESS), was calculated for the 16 paired joints, using the sum of the composite PDUS scores for all joints examined, giving a potential score of 0–96 for the 16 paired joints [15].

### Angiogenic marker measurement

Peripheral blood was collected in a vacutainer tube in the morning, at the same time as samples collected from hospitalized patients for routine analysis. Peripheral blood was allowed to clot by leaving it undisturbed at room temperature. Serum was then prepared by centrifuging whole blood at 1,000–2,000 x g for 10 minutes in a refrigerated centrifuge. Serum concentrations of the following eight angiogenic markers—Vascular Endothelial Growth Factor (VEGF), Placenta Growth Factor (PlGF), Tie-2, Angiopoietin-1, soluble Vascular Cell Adhesion Molecule-1 (sVCAM-1), Interleukin-8 (IL-8, CXCL8), CYR61 (CCN1) and Angiostatin—were measured by quantitative ELISAs (R&D Systems, Minneapolis, MN and RayBiotech, Norcross, GA), according to manufacturer recommendations. Biological function, intra-assay / inter-assay coefficients of variation, recovery and linearity are provided for each marker in [S1 Table](#).

### Statistical analyses

All data are expressed as mean values  $\pm$  standard deviation (SD), unless stated otherwise. Statistical analysis was performed using GraphPad Prism 7.0a software (San Diego, CA). For a two-group comparison, unpaired or paired t-test was used. One-way analysis of variance followed by Tuckey's multiple comparison tests was performed to compare data among three or more independent groups. Correlations were assessed using Spearman's rank correlation test. Differences in frequency were examined using the chi-square test. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ , and \*\*\*\* $P < 0.0001$ .

## Results

### Study population

From the 140 consecutive RA patients initially selected, 15 patients were excluded because of incomplete assessment (absence or incomplete PDUS assessment and/or absence of angiogenic marker measurements). A total of 125 patients (106 females, 84.8%) were finally included, with a mean age of  $58.7 \pm 15.9$  years and a mean disease duration of  $15.2 \pm 11.5$  years. Although the mean DAS-28 was  $3.31 \pm 1.59$ , in favor of low disease activity, half of RA patients had a DAS28  $> 3.2$ . The majority of patients received conventional synthetic DMARDs (116 patients, 92.8%), corticosteroids (87 patients, 69.6%), and targeted biologic therapies (81 patients, 64.8%), reflecting tertiary center recruitment. Detailed characteristics are provided in [Table 1](#).

### Levels of angiogenic markers

Levels of the 8 markers are provided in [S2 Table](#). As expected, serum levels of PlGF, a member of the VEGF sub-family correlated with VEGF. Levels of the matricellular protein CYR61 positively correlated with molecules involved in cell adhesion (sVCAM-1, IL-8) or cell death (Angiostatin), and negatively correlated with Angiopoietin-1, implicated in vessel maturation, adhesion, migration, and survival. Levels of soluble Tie-2, a crucial mediator of angiogenic process, correlated with IL-8 and Angiostatin. All correlations between the different markers are presented in [S3 Table](#).

Table 1. Study population.

		Patients with rheumatoid arthritis (n = 125)
<b>Demographics</b>	Age (years), mean ± SD	58.7±15.9
	Females, n (%)	106 (84.8)
<b>Disease characteristics</b>	Disease duration (years), mean ± SD	15.2±11.5
	Positive rheumatoid factor, n (%)	102 (81.6)
	Positive anti-CCP2 antibodies, n (%)	107 (85.6)
	Erosions on hand/foot x-rays, n (%)	75 (60.0)
<b>Disease activity:</b>	Tender Joint Count, mean±SD	2.7±4.1
	Swollen Joint Count, mean ±SD	3.1±4.5
	DAS28, mean ±SD	3.31±1.59
	DAS28<2.6, n (%)	37 (29.6)
	DAS28 >3.2, n (%)	62 (49.6)
	DAS28 >5.1, n (%)	12 (9.6)
	DAS28 CRP, mean ±SD	3.02±1.39
	DAS28 CRP<2.6, n (%)	51 (40.8)
	DAS28 CRP>3.2, n (%)	44 (35.2)
	DAS28 CRP>5.1, n (%)	8 (6.4)
	ESR (mmH1), mean ±SD	18.4±14.7
	ESR>28 mmH1, n (%)	28 (22.4)
	CRP (mg/L), median (range)	8.7±21.9
	CRP >10 mg/L, n (%)	25 (20.0)
<b>Function</b>	HAQ, median (range)	0.97±0.82
	HAQ >1.5, n (%)	37 (29.6)
<b>Ultrasound assessment</b>	Hand synovitis, n (%)	80 (64.0)
	Wrist, n (%)	45 (36.0)
	MCP joints, n (%)	63 (50.4)
	PIP joints, n (%)	39 (31.2)
	Hand tenosynovitis, n (%)	29 (23)
	MTP joint synovitis, n (%)	39 (31.2)
	Positive doppler signal, n (%)	53 (42.4)
	+ n (%)	22 (17.6)
	++ n (%)	16 (12.8)
+++ n (%)	15 (12.0)	
<b>Treatment received</b>	Current corticosteroid use, n (%)	87 (69.6)
	Current conventional DMARD use, n (%)	116 (92.8)
	Current anti-TNF-α use, n (%)	28 (22.4)
	Current rituximab use, n (%)	32 (25.6)
	Current tocilizumab use, n (%)	12 (9.6)
	Current abatacept use, n (%)	9 (7.2)

SD: Standard Deviation, DAS: Disease Activity Score, ESR: Erythrocyte Sedimentation Rate, CRP: C-reactive protein, HAQ: Health Assessment Questionnaire, MCP Metacarpophalangeal, PIP: Proximal Interphalangeal, MTP Metatarsophalangeal, DMARD: Disease Modifying Anti-Rheumatic Drug, TNF-α: Tumor Necrosis Factor-α

<https://doi.org/10.1371/journal.pone.0203607.t001>

### PDUS assessment

Reliability was tested on static images of hand, wrist and forefoot joints obtained from 20 consecutive RA patients. Inter-observer reliability for SH and PD evaluation was defined by κ coefficients of 0.72 and 0.75, respectively. Synovitis was detected in 84 patients with RA (67.2%). Among these patients, 53 patients (42.4%) had positive Doppler signal, including 31 with moderate to marked hyperemia. The global synovitis score ranged from 0 to 52 with a mean value of 5.4±9.9; 29 patients had a global synovitis score >7, corresponding to the 75<sup>th</sup> percentile value. This cut-off provided the best sensitivity and specificity for active disease, defined by a DAS28 >5.1 (sensitivity: 87.5%, specificity 88%, area under the ROC curve 0.89).

Detailed PDUS evaluation is presented in [Table 1](#).

## Levels of circulating angiogenic markers are associated with synovial vascularization assessed by PDUS

**Angiogenic biomarker levels according to the presence of synovial hyperemia.** We first compared levels of angiogenic biomarkers between patients without or with increased synovial vascularization detected by PDUS (Fig 1A–1H). We observed that serum levels of sVCAM-1 ( $808 \pm 293$  ng/mL vs.  $697 \pm 240$  ng/mL,  $P = 0.022$ ) (Fig 1C) and Tie-2 ( $16.2 \pm 7.5$  ng/mL vs.  $13.8 \pm 4.9$  ng/mL,  $P = 0.038$ ) (Fig 1D) were more likely to be increased in patients with synovial hyperemia detected on at least one joint (Power Doppler grade  $\geq 1$ ).

We next considered relevant synovitis, defined as grade 3 EULAR-OMERACT combined scoring system (Grade 3 Synovial Hypertrophy and  $\leq$  Grade 3 power Doppler signal or Grade 1 or 2 Synovial Hypertrophy and a Grade 3 power Doppler signal) [16]. Forty-nine patients fulfilled this definition. As previously observed, serum levels of sVCAM-1 ( $816 \pm 297$  ng/mL vs.  $715 \pm 260$  ng/mL,  $P = 0.045$ ) and Tie-2 ( $16.3 \pm 7.3$  ng/mL vs.  $13.9 \pm 5.3$  ng/mL,  $P = 0.041$ ) were more likely to be increased in patients with relevant synovitis.

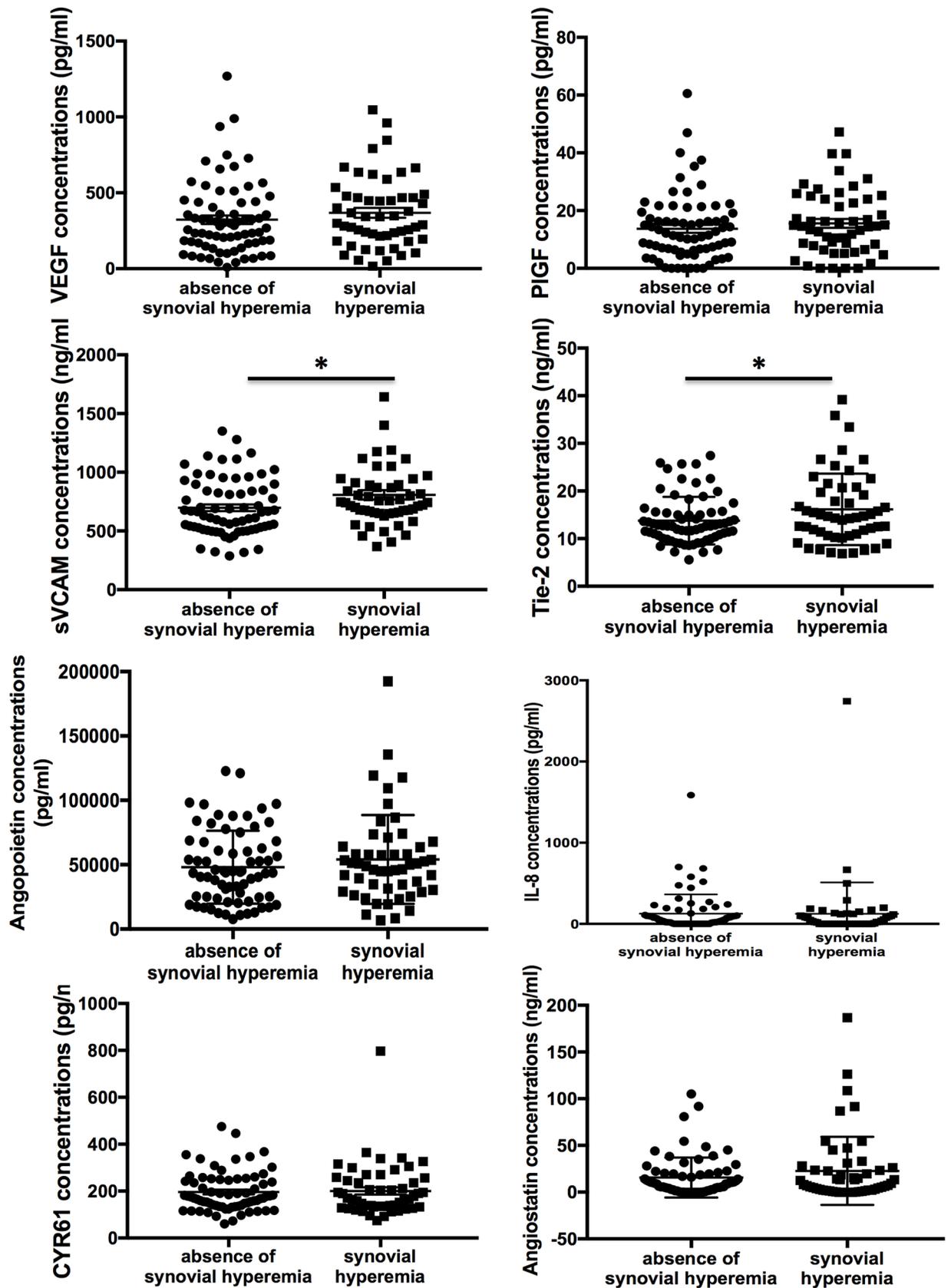
**Angiogenic biomarker levels according to the extent of synovial vascularization.** We next stratified these results according to the extent and intensity of synovial vascularization assessed by PDUS (Table 2). Interestingly, serum levels of sVCAM-1, Tie-2 and Angiostatin gradually increased together with the grade of the semiquantitative PDUS scale (Table 2 and Fig 2A–2C). Indeed, concentrations of these markers were markedly increased in patients with moderate to marked hyperemia (Power Doppler grade 2 and 3) compared to RA patients with absence or mild hyperemia on PDUS (Fig 2D–2F).

Since our previous analysis focused on the highest PDUS semiquantitative scale detected for each patient, we next studied the association between angiogenic marker levels and the global arthritis sum score, to take into account the number of synovitis detected by PDUS and the extent of their vascularization. This global arthritis score correlated with serum levels of sVCAM-1 ( $r = 0.20$ ,  $P = 0.028$ ), Tie-2 ( $r = 0.28$ ,  $P = 0.001$ ), and Angiostatin ( $r = 0.25$ ,  $P = 0.006$ ) (S1A–S1C Fig). In addition, patients with a global arthritis score  $>7$  were more likely to have increased sVCAM-1 ( $832 \pm 272$  ng/mL vs.  $704 \pm 227$  ng/mL,  $P = 0.013$ ), Tie 2 ( $17.9 \pm 8.3$  ng/mL vs.  $13.7 \pm 5.2$  ng/mL,  $P = 0.002$ ), Angiopoietin-1 levels ( $61049 \pm 40829$  pg/mL vs.  $47530 \pm 27002$  pg/mL,  $P = 0.043$ ) and Angiostatin ( $29.3 \pm 45.8$  ng/mL vs.  $15.5 \pm 20.5$  ng/mL,  $P = 0.025$ ) than patients with a score  $\leq 7$  (Fig 3A–3H).

**Levels of Tie-2 and PlGF are associated with PDUS features indicating residual activity in patients with low disease activity.** We next aimed to determine whether angiogenic markers might be used to detect residual activity in patients with low disease activity and remission. Among the 81 patients with a DAS28  $\leq 3.2$ , 22 patients had synovial hyperemia detected on at least one joint (Power Doppler grade 1 in 13 patients, grade 2 in 6 patients and grade 3 in 3 patients). Patients with synovial hyperemia on at least one joint were more likely to have significantly increased levels of PlGF ( $18.9 \pm 11.2$  pg/mL vs.  $13.1 \pm 9.5$  pg/mL,  $P = 0.022$ ) (Fig 4B) and Tie-2 ( $15.7 \pm 5.8$  ng/mL vs.  $12.6 \pm 3.4$  ng/mL,  $P = 0.004$ ) (Fig 4D) than patients with absence of synovial hyperemia. No significant difference was observed regarding other angiogenic markers (Fig 4A, Fig 4C, Fig 4E–4H).

Among the 51 patients in remission with a DAS28  $< 2.6$ , only 10 patients had synovial hyperemia detected on at least one joint (Power Doppler grade 1 in 7 patients, grade 2 in 1 patients and grade 3 in 2 patients). Concentrations of the different angiogenic markers did not significantly differ in patients with or without synovial hyperemia.

**Angiogenic biomarker levels according to the treatment received.** Tie-2 serum levels were significantly decreased in the 32 patients treated with rituximab ( $12.2 \pm 4.6$  vs.  $17.4 \pm 8.2$ ,  $P = 0.003$ ). In line with this observation, a significant reduction of the global arthritis score



**Fig 1. Levels of angiogenic biomarkers between patients without or with synovial hyperemia detected by Power Doppler.** Statistical test: two-sided unpaired t-test. \*  $P < 0.05$ .

<https://doi.org/10.1371/journal.pone.0203607.g001>

( $1.5 \pm 3.2$  vs.  $7.0 \pm 9.9$   $P = 0.004$ ) and of the proportion of patients with a global arthritis score  $> 7$  (3/32, 9.4% vs. 14/44, 31.8%,  $P = 0.021$ ) was observed in patients treated with rituximab, compared to the 44 patients treated with conventional synthetic DMARDs only. A trend toward significance was also observed for a lower proportion of patients with moderate to marked hyperemia (Power Doppler grade 2 and 3) upon rituximab (5/32, 15.6% vs. 15/44, 34.1%,  $P = 0.072$ ).

Angiostatin serum levels were significantly decreased in the 28 patients treated with TNF- $\alpha$  inhibitors ( $8.8 \pm 10.3$  vs.  $26.3 \pm 37.6$ ,  $P = 0.045$ ). A marked reduction of the global arthritis score ( $1.8 \pm 3.3$  vs.  $7.0 \pm 9.9$   $P = 0.010$ ) and of the proportion of patients with a global arthritis score  $> 7$  (3/28, 10.7% vs. 14/44, 31.8%,  $P = 0.041$ ) was observed in patients treated with TNF $\alpha$  inhibitors, compared to the 44 patients treated with conventional synthetic DMARDs only. A trend toward significance was also observed for a lower proportion of patients with moderate to marked hyperemia (Power Doppler grade 2 and 3) upon TNF $\alpha$  inhibitors (4/28, 14.3% vs. 15/44, 34.1%,  $P = 0.065$ ).

No significant variation was observed regarding the other markers according to treatment received.

**Angiogenic marker levels according to RA disease characteristics.** Patients with DAS28-CRP  $> 5.1$  were more likely to have significantly higher Tie-2 ( $21.8 \pm 10.1$  vs.  $13.9 \pm 5.1$  ng/mL,  $P = 0.002$ ), CYR61 ( $288 \pm 219$  pg/ml vs.  $180 \pm 177$ ,  $P = 0.026$ ) and Angiostatin ( $45.8 \pm 63.4$  vs.  $17.5 \pm 25.7$ ,  $P = 0.047$ ) serum concentrations, as compared to patients with a DAS28-CRP  $\leq 3.2$ . No association was observed between other angiogenic markers and disease activity.

VEGF correlated with ESR ( $r = 0.18$ ,  $P = 0.045$ ) and CRP ( $r = 0.26$ ,  $P = 0.004$ ) levels. CYR61 correlated with age ( $r = 0.31$ ,  $P < 0.001$ ) and disease duration ( $r = 0.23$ ,  $P = 0.017$ ). Increased

**Table 2. Levels of angiogenic markers according to the intensity and extent of synovial hyperemia detected by Power Doppler.**

	Absence of synovial hyperemia (n = 72)	Power Doppler Grade 1 (n = 22)	Power Doppler Grade 2 (n = 16)	Power Doppler Grade 3 (n = 15)	P-value
VEGF (pg/ml), mean (SD)	309 ± 213	365 ± 222	359 ± 282	382 ± 212	NS
PIGF (pg/ml), mean (SD)	13.0 ± 10.2	17.5 ± 11.4	15.0 ± 12.5	13.4 ± 9.0	NS
sVCAM-1 (ng/ml), mean (SD)	697 ± 240	666 ± 149	793 ± 189	943 ± 307	* 0.002 ** 0.003
Tie2 (ng/ml), mean (SD)	13.8 ± 4.9	12.9 ± 4.3	15.3 ± 8.7	21.3 ± 7.7	* <0.001 ** <0.001 *** 0.025
Angiopoietin (pg/ml), mean (SD)	51170 ± 3985	51334 ± 6741	47346 ± 6878	61969 ± 11712	NS
IL8 (pg/ml), mean (SD)	105 ± 163	60 ± 85	74 ± 170	93.99	NS
CYR61 (pg/ml), mean (SD)	196 ± 85	202 ± 87	218 ± 168	181 ± 59	NS
Angiostatin (ng/ml), mean (SD)	15.5 ± 20.4	11.2 ± 14.5	21.1 ± 24.9	47.8 ± 58.1	* <0.001 ** <0.001 *** 0.039

Statistical test: One-way analysis of variance followed by Tuckey's multiple comparison tests

\* Power Doppler grade 3 vs. absence of synovial hyperemia

\*\* Power Doppler grade 3 vs. Power Doppler grade 1

\*\*\* Power Doppler grade 3 vs. Power Doppler grade 2

<https://doi.org/10.1371/journal.pone.0203607.t002>

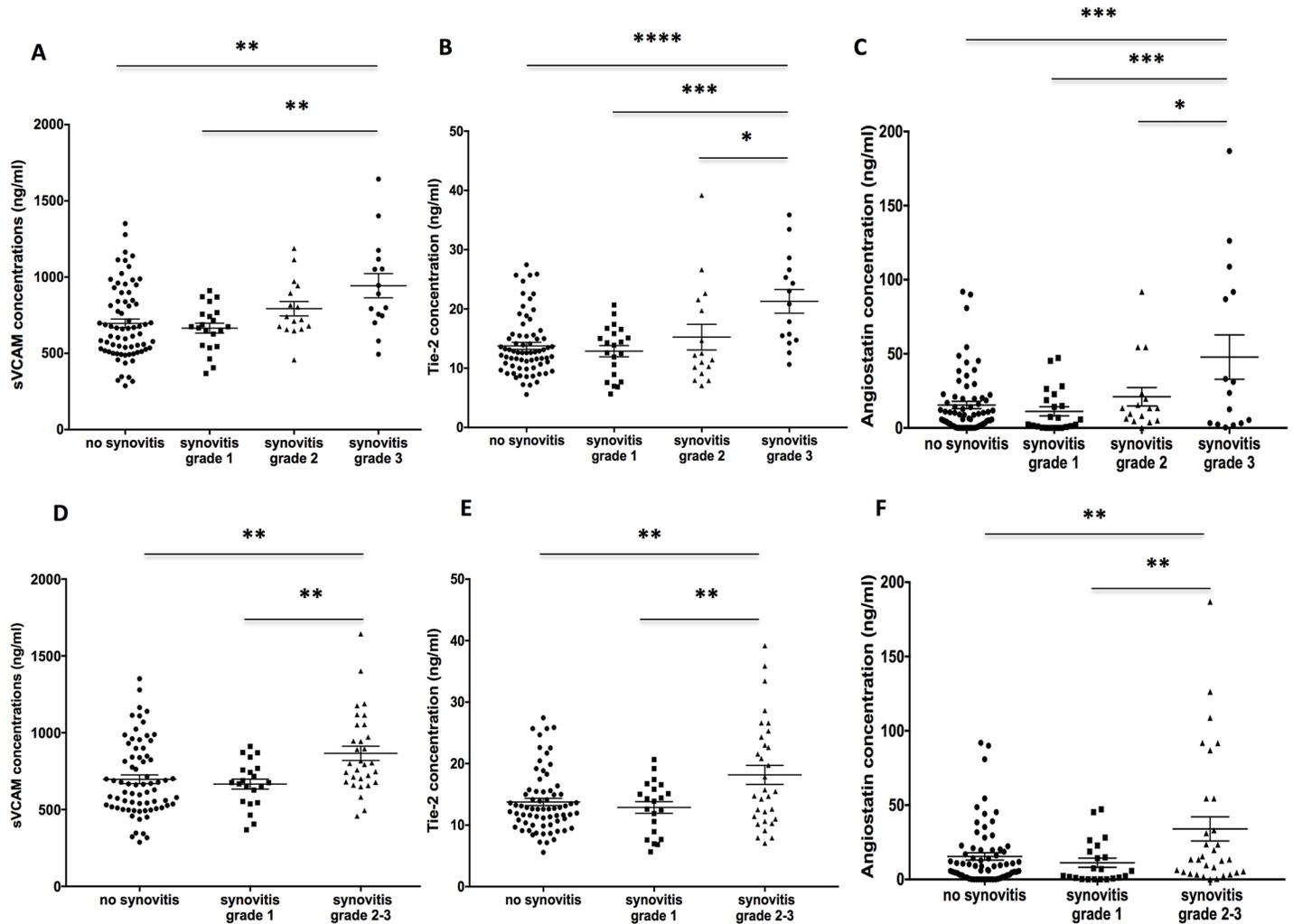


Fig 2. Levels of sVCAM-1 (A and D), Tie-2 (B and E) and Angiostatin (C and F) according to the intensity and extent of synovial vascularization assessed by Power Doppler Ultrasound. Statistical test: ANOVA followed by Tuckey's multiple comparisons test. \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$ , \*\*\*\*  $P < 0.0001$ .

<https://doi.org/10.1371/journal.pone.0203607.g002>

CYR61 levels were observed in patients with bone erosions ( $218 \pm 125$  pg/ml vs.  $179 \pm 69$  pg/ml,  $P = 0.041$ ) and HAQ > 1.5 ( $270 \pm 170$  pg/ml vs.  $181 \pm 69$  pg/ml,  $P < 0.001$ ).

## Discussion

This cross-sectional study of 125 RA patients supports the relevance of measuring serum levels of vascular markers to evaluate the intensity and extent of synovial vascularization and, thus, disease activity [17].

Tie-2, a receptor tyrosine kinase expressed primarily in endothelial cells and fundamental for vascular development, was identified as one of the most promising angiogenic marker in our cohort of RA patients, since it increased parallel to the intensity and extent PDUS scale and correlated with the PDUS global arthritis sum score. These findings are consistent with elevated Tie2 expression in human RA synovium [18]. In addition, serum Tie-2 levels were found increased in systemic sclerosis (SSc), another complex disease characterized by widespread microangiopathy [19, 20]. Moreover, a dysregulation of Ang/Tie2 was observed in the bleomycin mouse model, reflecting early and inflammatory stages of SSc, which did not apply



**Fig 3. Levels of angiogenic markers according to the values of the global synovitis score.** Statistical test: two-sided unpaired t-test. \*  $P < 0.05$ , \*\*  $P < 0.01$ .

<https://doi.org/10.1371/journal.pone.0203607.g003>

for the non-inflammatory tight skin mouse model, highlighting the link between Tie-2 and inflammation, as it was observed in our study [19].

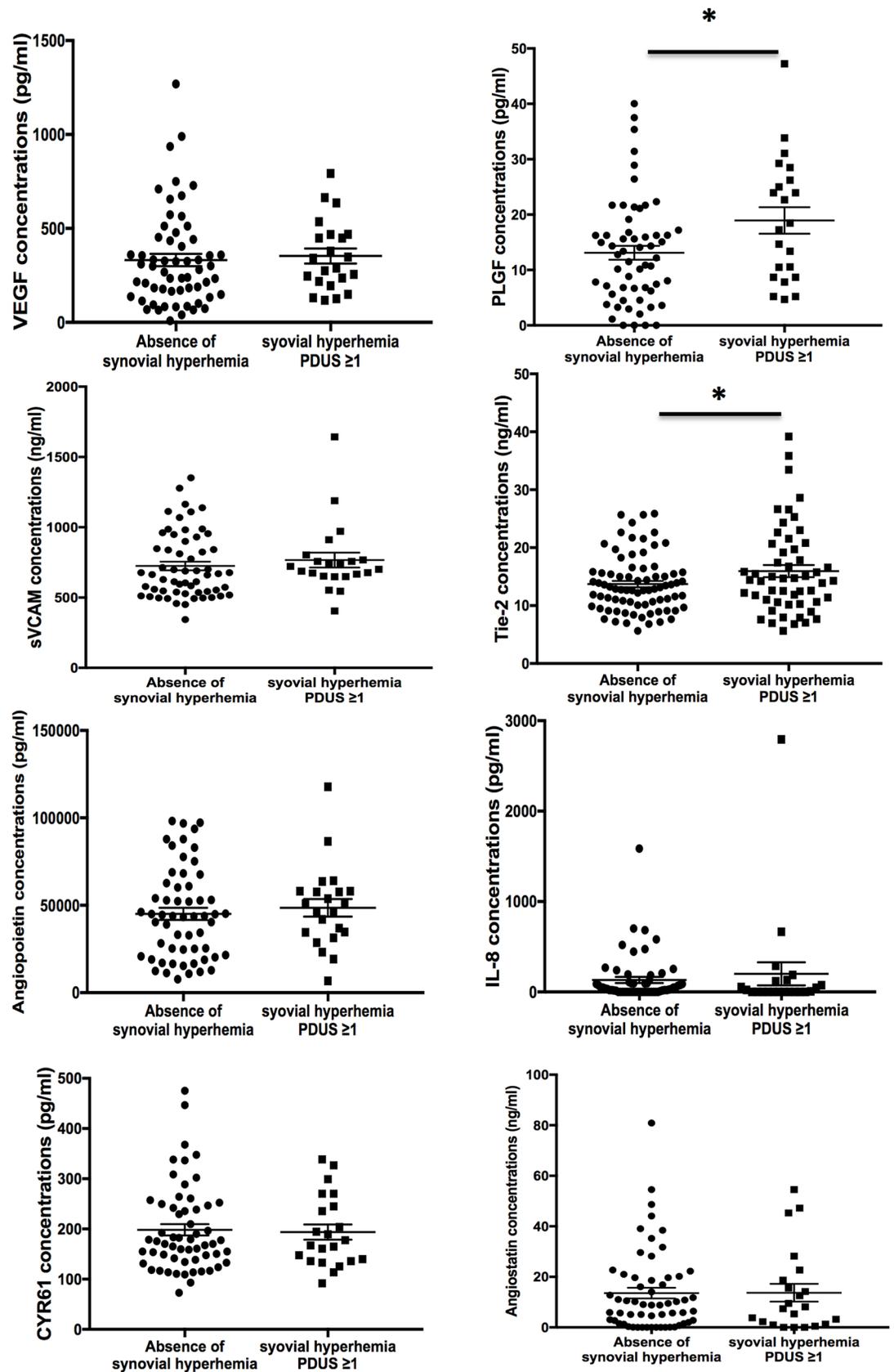
Tie-2 levels markedly decreased together with the PDUS global arthritis score in patients treated with rituximab and were found increased in RA patients in LDA with residual inflammatory activity, together with PIGF levels. In addition, increased PIGF levels were detected in patients with subclinical synovitis. Consistently, higher levels of angiogenic markers, including PIGF, have previously been reported in patients with stringent clinical remission and ultrasound-defined active synovitis [21]. Our findings are consistent with the good correlation previously observed between PDUS and Tie-2 mRNA levels measured in the synovial tissue [22]. These findings also sustain previous data underlining the importance of Tie-2 in the development of synovial neovascularization. Local engagement of synovial Tie-2 signaling is observed during the earliest phases of RA and links the proinflammatory cytokine TNF- $\alpha$  to pathologic angiogenesis [18, 23]. Moreover, gene therapy with soluble Tie-2 receptor inhibits neovascularization and arthritis development and protects against bone destruction in a mouse model of collagen-induced arthritis [24].

VCAM-1 is important in leukocyte trafficking and its increased expression is associated with a number of chronic inflammatory diseases, including RA [25]. Moreover, sVCAM-1 may be important in triggering angiogenesis in the initial stages of RA [26]. sVCAM-1 serum levels gradually increased together with the intensity of synovial inflammation assessed by PDUS. This result is in line with increased sVCAM-1 concentrations in RA [25, 27], especially in patients with early and active disease [28, 29], as well as other complex connective tissue disorders like SSc or systemic lupus erythematosus (SLE) [30–32].

sVCAM-1 is a validated biomarker of cardiovascular disease (CVD) and atherogenesis [33], which is of particular importance since CVD has emerged in these recent years as a cause of morbidity and mortality in patients with RA [14, 34]. Our study links for the first time this endothelial activation marker to synovial vascularity assessed by PDUS, highlighting that sVCAM-1 might also be useful as a marker of synovial inflammation that deserves further evaluation in longitudinal studies. This finding is sustained by the arthritogenic role of sVCAM-1 in the development of experimental inflammatory arthritis in the MRL-Fas(lpr) mouse model [35].

Angiostatin has given promising results in cancer therapy trials, as well as preclinical arthritis studies [36]. Serum levels of this anti-angiogenic fragment of plasminogen was found associated with the extent of synovial vascularity in the present study, which is consistent with increased synovial Angiostatin levels observed in patients with inflammatory arthritis [37] and the positive correlation observed in a preliminary study between Angiostatin and cumulative effusion scores evaluated by ultrasonography [37]. Elevated Angiostatin serum levels were also detected in other complex connective tissue diseases like SSc and SLE [38–40].

Being a critical proangiogenic factor, VEGF was expected to be associated with synovial vascularization. However, VEGF levels were not associated with the PDUS semi-quantitative scale and no correlation was observed between VEGF and the global arthritis sum score in the present study. So far, there is no agreement on the association between VEGF and PDUS. Most of previous studies on VEGF had limited sample size and heterogeneous numbers of joints have been evaluated [11, 17, 41–43]. Two prospective studies failed to correlate serum VEGF concentrations with PDUS findings or scores of disease activity [11, 41]. One possible explanation could be variable time response of both factors. Thus, the examination of a high



**Fig 4. Levels of angiogenic markers according to the intensity and extent of synovial hyperemia detected by Power Doppler in the 81 patients with low disease activity.** Statistical test: two-sided unpaired t-test. \*  $P < 0.05$ .

<https://doi.org/10.1371/journal.pone.0203607.g004>

number of joints performed in the herein study is consistent with previous data, suggesting that VEGF is not a powerful marker of active synovitis in RA patients [44, 45].

One novelty of our study was to measure for the first-time serum levels of CYR61, a matrix-cellular protein that is essential for the proper development of the cardiovascular system and the control of angiogenesis. Although we failed to link CYR61 to synovial vascularization in the present study, CYR61 correlated with disease duration and increased CYR61 levels were observed in patients with high disease activity, bone erosions and HAQ > 1.5. These findings are consistent with the implication of CYR61 signaling pathway in the development of bone erosion recently described, and the evaluation of CYR61 as a biomarker of structural progression in RA should be further considered [46].

There is no single gold standard for quantifying the level of disease activity in RA. Three composite scores are used for monitoring disease evolution: disease activity score (DAS 28), simple disease activity index (SDAI) and clinical disease activity index (CDAI). The disadvantage of these scores is the degree of subjectivity of some of the criteria. Moreover, a significant proportion of the patients with negative inflammatory tests still have active disease [47], highlighting the need to develop additional tools for a more effective monitoring of the disease. PDUS has demonstrated validity in longitudinal assessment and monitoring of disease activity in RA. It has the ability to detect subclinical synovitis not appreciated by clinical examination alone. This examination also correlates significantly with clinical findings and inflammatory markers, along with synovial histopathology in patients with RA [48]. However, there are certain limitations, including the lack of standardization of PDUS scoring and settings, which can limit the use of this technique in clinical practice. This variability for RA synovitis scores with PDUS [49] may partly explain the lower levels of sVCAM-1 and angiostatin in patients with power Doppler grade 1 compared to patients without power Doppler signals.

Based on our results, angiogenic markers may be used as a surrogate of active synovitis, and their precise place in relation to PDUS deserves further investigation. Angiogenic markers might be more relevant in situations where there are concerns regarding PDUS image acquisition and interpretation, like flash artifacts. The remaining controversies in the number of joints to be assessed for monitoring of disease may also support the first-line use of angiogenic markers to evaluate disease activity. Angiogenic markers might also be considered as an alternative to PDUS in centers where accessibility to this examination is still an issue.

The present study has several strengths. A high number of patients were included and carefully assessed and phenotyped in a tertiary center with a long-lasting experience in RA evaluation and care. Two experienced sonographers performed careful PDUS investigation. The study was performed in a routine clinical setting, which reinforces the external validity of the results. An important joint number was assessed by PDUS, but the addition of even more joints might potentially have modified the findings. Our study is limited by its observational design and any pathogenic link emerged from this type of study should be taken very cautiously. The low number of patients in remission with synovial hyperemia may have prevented the identification of angiogenic markers of subclinical synovitis in patients in remission. Prospective studies will be needed to assess the temporal relationship between disease activity and angiogenic markers.

In conclusion, serum levels of the angiogenic markers Tie-2, sVCAM-1 and Angiostatin were strongly associated with synovial vascularization and inflammation assessed by PDUS among patients with established RA. Moreover, Tie-2 and PIGF were associated with persistent

disease activity. These data highlight the possibility to identify surrogate serum angiogenic markers of active synovitis, and their pertinence needs to be confirmed in longitudinal studies.

## Supporting information

**S1 Table. Biological function, intra-assay and inter-assay coefficients of variation, recovery and linearity of each angiogenic marker.**

(DOCX)

**S2 Table. Levels of angiogenic markers in RA patients.**

(DOCX)

**S3 Table. Correlation between angiogenic markers in RA patients.**

(DOCX)

**S1 Fig. Correlations between serum levels of sVAM-1 (A), Tie2 (B) and Angiostatin (C) with the global synovitis score.** Statistical test: Spearman's rank correlation test.

(TIFF)

## Author Contributions

**Conceptualization:** Yannick Allanore, Jérôme Avouac.

**Formal analysis:** Agathe Leblond, Sonia Pezet, Anne Priscille Trouvin, Muriel Elhai, Virginie Gonzalez, Jérôme Avouac.

**Funding acquisition:** Yannick Allanore, Jérôme Avouac.

**Investigation:** Agathe Leblond, Sonia Pezet, Anne Priscille Trouvin, Muriel Elhai, Virginie Gonzalez, Yannick Allanore, Jérôme Avouac.

**Supervision:** Yannick Allanore, Jérôme Avouac.

**Validation:** Yannick Allanore, Jérôme Avouac.

**Writing – original draft:** Agathe Leblond, Jérôme Avouac.

**Writing – review & editing:** Agathe Leblond, Sonia Pezet, Anne Priscille Trouvin, Muriel Elhai, Virginie Gonzalez, Yannick Allanore, Jérôme Avouac.

## References

1. Criscione LG, E.W SC. Tumor necrosis factor-alpha for the treatment of rheumatic diseases. *Curr Opin Rheumatol.* 2002; 14:204–11. PMID: [11981314](https://pubmed.ncbi.nlm.nih.gov/11981314/)
2. Firestein GS. Starving the synovium: angiogenesis and inflammation in rheumatoid arthritis. *The Journal of Clinical Investigation.* 1999; 103:3–4. <https://doi.org/10.1172/JCI5929> PMID: [9884327](https://pubmed.ncbi.nlm.nih.gov/9884327/)
3. McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. *N Engl J Med.* 2011; 365(23):2205–19. <https://doi.org/10.1056/NEJMra1004965> PMID: [22150039](https://pubmed.ncbi.nlm.nih.gov/22150039/).
4. Leblond A, Allanore Y, Avouac J. Targeting synovial neoangiogenesis in rheumatoid arthritis. *Autoimmun Rev.* 2017; 16(6):594–601. <https://doi.org/10.1016/j.autrev.2017.04.005> PMID: [28414154](https://pubmed.ncbi.nlm.nih.gov/28414154/).
5. Koch AE. Review: angiogenesis: implications for rheumatoid arthritis. *Arthritis Rheum.* 1998; 41(6):951–62. [https://doi.org/10.1002/1529-0131\(199806\)41:6<951::AID-ART2>3.0.CO;2-D](https://doi.org/10.1002/1529-0131(199806)41:6<951::AID-ART2>3.0.CO;2-D) PMID: [9627005](https://pubmed.ncbi.nlm.nih.gov/9627005/).
6. Walsh SJ. Effects of non-mining occupational silica exposure on proportional mortality from silicosis and systemic sclerosis. *J Rheumatol.* 1999; 26(10):2179–85. PMID: [10529136](https://pubmed.ncbi.nlm.nih.gov/10529136/).
7. Naredo E, Bonilla G, Gamero F, Uson J, Carmona L, Laffon A. Assessment of inflammatory activity in rheumatoid arthritis: a comparative study of clinical evaluation with grey scale and power Doppler ultrasonography. *Ann Rheum Dis.* 2005; 64(3):375–81. <https://doi.org/10.1136/ard.2004.023929> PMID: [15708891](https://pubmed.ncbi.nlm.nih.gov/15708891/); PubMed Central PMCID: [PMCPMC1755396](https://pubmed.ncbi.nlm.nih.gov/PMCPMC1755396/).

8. Taylor PC, Steuer A, Gruber J, Cosgrove DO, Blomley MJ, Marsters PA, et al. Comparison of ultrasonographic assessment of synovitis and joint vascularity with radiographic evaluation in a randomized, placebo-controlled study of infliximab therapy in early rheumatoid arthritis. *Arthritis Rheum.* 2004; 50(4):1107–16. <https://doi.org/10.1002/art.20123> PMID: 15077292.
9. Scire CA, Montecucco C, Codullo V, Epis O, Todoerti M, Caporali R. Ultrasonographic evaluation of joint involvement in early rheumatoid arthritis in clinical remission: power Doppler signal predicts short-term relapse. *Rheumatology (Oxford).* 2009; 48(9):1092–7. <https://doi.org/10.1093/rheumatology/kep171> PMID: 19561156.
10. Saleem B, Brown AK, Quinn M, Karim Z, Hensor EM, Conaghan P, et al. Can flare be predicted in DMARD treated RA patients in remission, and is it important? A cohort study. *Ann Rheum Dis.* 2012; 71(8):1316–21. <https://doi.org/10.1136/annrheumdis-2011-200548> PMID: 22294638.
11. Nordal HH, Brokstad KA, Solheim M, Halse AK, Kvien TK, Hammer HB. Calprotectin (S100A8/A9) has the strongest association with ultrasound-detected synovitis and predicts response to biologic treatment: results from a longitudinal study of patients with established rheumatoid arthritis. *Arthritis Res Ther.* 2017; 19(1):3. <https://doi.org/10.1186/s13075-016-1201-0> PMID: 28081709; PubMed Central PMCID: PMC5234113.
12. Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum.* 1988; 31(3):315–24. Epub 1988/03/01. PMID: 3358796.
13. van der Heijde DM, van 't Hof MA, van Riel PL, Theunisse LA, Lubberts EW, van Leeuwen MA, et al. Judging disease activity in clinical practice in rheumatoid arthritis: first step in the development of a disease activity score. *Ann Rheum Dis.* 1990; 49(11):916–20. Epub 1990/11/01. PMID: 2256738; PubMed Central PMCID: PMC1004262.
14. Avouac J, Meune C, Chenevier-Gobeaux C, Dieude P, Borderie D, Lefevre G, et al. Inflammation and Disease Activity are Associated with High Circulating Cardiac Markers in Rheumatoid Arthritis Independently of Traditional Cardiovascular Risk Factors. *J Rheumatol.* 2014; 41(2):248–55. Epub 2013/12/18. <https://doi.org/10.3899/jrheum.130713> PMID: 24334650.
15. D'Agostino MA, Boers M, Wakefield RJ, Berner Hammer H, Vittecoq O, Filippou G, et al. Exploring a new ultrasound score as a clinical predictive tool in patients with rheumatoid arthritis starting abatacept: results from the APPRAISE study. *RMD Open.* 2016; 2(1):e000237. <https://doi.org/10.1136/rmdopen-2015-000237> PMID: 27175297; PubMed Central PMCID: PMC54860864.
16. D'Agostino MA, Terslev L, Aegerter P, Backhaus M, Balint P, Bruyn GA, et al. Scoring ultrasound synovitis in rheumatoid arthritis: a EULAR-OMERACT ultrasound taskforce-Part 1: definition and development of a standardised, consensus-based scoring system. *RMD Open.* 2017; 3(1):e000428. <https://doi.org/10.1136/rmdopen-2016-000428> PMID: 28948983; PubMed Central PMCID: PMC5597799.
17. Strunk J, Rumbaur C, Albrecht K, Neumann E, Muller-Ladner U. Linking systemic angiogenic factors (VEGF, angiogenin, TIMP-2) and Doppler ultrasound to anti-inflammatory treatment in rheumatoid arthritis. *Joint Bone Spine.* 2013; 80(3):270–3. <https://doi.org/10.1016/j.jbspin.2012.09.001> PMID: 23098925.
18. DeBusk LM, Chen Y, Nishishita T, Chen J, Thomas JW, Lin PC. Tie2 receptor tyrosine kinase, a major mediator of tumor necrosis factor alpha-induced angiogenesis in rheumatoid arthritis. *Arthritis Rheum.* 2003; 48(9):2461–71. <https://doi.org/10.1002/art.11213> PMID: 13130465.
19. Moritz F, Schniering J, Distler JHW, Gay RE, Gay S, Distler O, et al. Tie2 as a novel key factor of microangiopathy in systemic sclerosis. *Arthritis Res Ther.* 2017; 19(1):105. <https://doi.org/10.1186/s13075-017-1304-2> PMID: 28545512; PubMed Central PMCID: PMC5445339.
20. Dunne JV, Keen KJ, Van Eeden SF. Circulating angiopoietin and Tie-2 levels in systemic sclerosis. *Rheumatol Int.* 2013; 33(2):475–84. <https://doi.org/10.1007/s00296-012-2378-4> PMID: 22461185.
21. Ramirez J, Ruiz-Esquide V, Pomes I, Celis R, Cuervo A, Hernandez MV, et al. Patients with rheumatoid arthritis in clinical remission and ultrasound-defined active synovitis exhibit higher disease activity and increased serum levels of angiogenic biomarkers. *Arthritis Res Ther.* 2014; 16(1):R5. <https://doi.org/10.1186/ar4431> PMID: 24398122; PubMed Central PMCID: PMC3978423.
22. Kelly S, Bombardieri M, Humby F, Ng N, Marrelli A, Riahi S, et al. Angiogenic gene expression and vascular density are reflected in ultrasonographic features of synovitis in early Rheumatoid Arthritis: an observational study. *Arthritis Res Ther.* 2015; 17:58. <https://doi.org/10.1186/s13075-015-0567-8> PMID: 25889955; PubMed Central PMCID: PMC4476089.
23. van de Sande MG, de Launay D, de Hair MJ, Garcia S, van de Sande GP, Wijbrandts CA, et al. Local synovial engagement of angiogenic TIE-2 is associated with the development of persistent erosive rheumatoid arthritis in patients with early arthritis. *Arthritis Rheum.* 2013; 65(12):3073–83. <https://doi.org/10.1002/art.38128> PMID: 23982963.

24. Chen Y, Donnelly E, Kobayashi H, Debusk LM, Lin PC. Gene therapy targeting the Tie2 function ameliorates collagen-induced arthritis and protects against bone destruction. *Arthritis Rheum.* 2005; 52(5):1585–94. <https://doi.org/10.1002/art.21016> PMID: 15880817.
25. Klimiuk PA, Fiedorczyk M, Sierakowski S, Chwiecko J. Soluble cell adhesion molecules (sICAM-1, sVCAM-1, and sE-selectin) in patients with early rheumatoid arthritis. *Scand J Rheumatol.* 2007; 36(5):345–50. <https://doi.org/10.1080/03009740701406460> PMID: 17963163.
26. Koch AE, Halloran MM, Haskell CJ, Shah MR, Polverini PJ. Angiogenesis mediated by soluble forms of E-selectin and vascular cell adhesion molecule-1. *Nature.* 1995; 376(6540):517–9. <https://doi.org/10.1038/376517a0> PMID: 7543654.
27. Littler AJ, Buckley CD, Wordsworth P, Collins I, Martinson J, Simmons DL. A distinct profile of six soluble adhesion molecules (ICAM-1, ICAM-3, VCAM-1, E-selectin, L-selectin and P-selectin) in rheumatoid arthritis. *Br J Rheumatol.* 1997; 36(2):164–9. PMID: 9133922.
28. Macias I, Garcia-Perez S, Ruiz-Tudela M, Medina F, Chozas N, Giron-Gonzalez JA. Modification of pro- and antiinflammatory cytokines and vascular-related molecules by tumor necrosis factor- $\alpha$  blockade in patients with rheumatoid arthritis. *J Rheumatol.* 2005; 32(11):2102–8. PMID: 16265686.
29. Klimiuk PA, Sierakowski S, Latosiewicz R, Cylwik JP, Cylwik B, Skowronski J, et al. Soluble adhesion molecules (ICAM-1, VCAM-1, and E-selectin) and vascular endothelial growth factor (VEGF) in patients with distinct variants of rheumatoid synovitis. *Ann Rheum Dis.* 2002; 61(9):804–9. <https://doi.org/10.1136/ard.61.9.804> PMID: 12176805; PubMed Central PMCID: PMC1754213.
30. Kuryliszyn-Moskal A, Klimiuk PA, Sierakowski S. Soluble adhesion molecules (sVCAM-1, sE-selectin), vascular endothelial growth factor (VEGF) and endothelin-1 in patients with systemic sclerosis: relationship to organ systemic involvement. *Clin Rheumatol.* 2005; 24(2):111–6. <https://doi.org/10.1007/s10067-004-0987-3> PMID: 15349798.
31. Robak E, Kulczycka L, Sysa-Jedrzejowska A, Wierzbowska A, Robak T. Circulating proangiogenic molecules PIGF, SDF-1 and sVCAM-1 in patients with systemic lupus erythematosus. *Eur Cytokine Netw.* 2007; 18(4):181–7. <https://doi.org/10.1684/ecn.2007.0103> PMID: 17964973.
32. Howe HS, Kong KO, Thong BY, Law WG, Chia FL, Lian TY, et al. Urine sVCAM-1 and sICAM-1 levels are elevated in lupus nephritis. *Int J Rheum Dis.* 2012; 15(1):13–6. <https://doi.org/10.1111/j.1756-185X.2012.01720.x> PMID: 22324942.
33. Shai I, Pischon T, Hu FB, Ascherio A, Rifai N, Rimm EB. Soluble intercellular adhesion molecules, soluble vascular cell adhesion molecules, and risk of coronary heart disease. *Obesity (Silver Spring).* 2006; 14(11):2099–106. <https://doi.org/10.1038/oby.2006.245> PMID: 17135628.
34. Meune C, Touze E, Trinquart L, Allanore Y. High risk of clinical cardiovascular events in rheumatoid arthritis: Levels of associations of myocardial infarction and stroke through a systematic review and meta-analysis. *Arch Cardiovasc Dis.* 103(4):253–61. Epub 2010/07/27. doi: S1875-2136(10)00077-X [pii] <https://doi.org/10.1016/j.acvd.2010.03.007> PMID: 20656636.
35. Oishi H, Mizuki S, Terada M, Kudo M, Araki K, Araki M, et al. Increased expression of soluble form of vascular cell adhesion molecule-1 aggravates autoimmune arthritis in MRL-Fas(lpr) mice. *Pathol Int.* 2007; 57(11):734–40. <https://doi.org/10.1111/j.1440-1827.2007.02165.x> PMID: 17922685.
36. Szekanecz Z, Besenyei T, Paragh G, Koch AE. New insights in synovial angiogenesis. *Joint Bone Spine.* 2010; 77(1):13–9. <https://doi.org/10.1016/j.jbspin.2009.05.011> PMID: 20022538; PubMed Central PMCID: PMC2910514.
37. Gok M, Erdem H, Gogus F, Yilmaz S, Karadag O, Simsek I, et al. Relationship of ultrasonographic findings with synovial angiogenesis modulators in different forms of knee arthritides. *Rheumatol Int.* 2013; 33(4):879–85. <https://doi.org/10.1007/s00296-012-2452-y> PMID: 22811011.
38. Almeida I, Oliveira Gomes A, Lima M, Silva I, Vasconcelos C. Different contributions of angiostatin and endostatin in angiogenesis impairment in systemic sclerosis: a cohort study. *Clin Exp Rheumatol.* 2016;34 Suppl 100(5):37–42. PMID: 26885625.
39. Wu T, Du Y, Han J, Singh S, Xie C, Guo Y, et al. Urinary angiostatin—a novel putative marker of renal pathology chronicity in lupus nephritis. *Mol Cell Proteomics.* 2013; 12(5):1170–9. <https://doi.org/10.1074/mcp.M112.021667> PMID: 23345539; PubMed Central PMCID: PMC3650329.
40. Arriens C, Wren JD, Munroe ME, Mohan C. Systemic lupus erythematosus biomarkers: the challenging quest. *Rheumatology (Oxford).* 2017; 56(suppl\_1):i32–i45. <https://doi.org/10.1093/rheumatology/kew407> PMID: 28013203; PubMed Central PMCID: PMC5850341.
41. Strunk J, Heinemann E, Neeck G, Schmidt KL, Lange U. A new approach to studying angiogenesis in rheumatoid arthritis by means of power Doppler ultrasonography and measurement of serum vascular endothelial growth factor. *Rheumatology (Oxford).* 2004; 43(12):1480–3. <https://doi.org/10.1093/rheumatology/keh380> PMID: 15353607.
42. Kurosaka D, Hirai K, Nishioka M, Miyamoto Y, Yoshida K, Noda K, et al. Clinical significance of serum levels of vascular endothelial growth factor, angiotensin-1, and angiotensin-2 in patients with

- rheumatoid arthritis. *J Rheumatol*. 2010; 37(6):1121–8. <https://doi.org/10.3899/jrheum.090941> PMID: 20436077.
43. Kawashiri SY, Kawakami A, Iwamoto N, Fujikawa K, Satoh K, Tamai M, et al. The power Doppler ultrasonography score from 24 synovial sites or 6 simplified synovial sites, including the metacarpophalangeal joints, reflects the clinical disease activity and level of serum biomarkers in patients with rheumatoid arthritis. *Rheumatology (Oxford)*. 2011; 50(5):962–5. <https://doi.org/10.1093/rheumatology/keq415> PMID: 21186172.
  44. Grothey A, Galanis E. Targeting angiogenesis: progress with anti-VEGF treatment with large molecules. *Nat Rev Clin Oncol*. 2009; 6(9):507–18. <https://doi.org/10.1038/nrclinonc.2009.110> PMID: 19636328.
  45. Semerano L, Clavel G, Assier E, Denys A, Boissier MC. Blood vessels, a potential therapeutic target in rheumatoid arthritis? *Joint Bone Spine*. 2011; 78(2):118–23. Epub 2010/09/21. <https://doi.org/10.1016/j.jbspin.2010.06.004> PMID: 20851025.
  46. Huang TL, Mu N, Gu JT, Shu Z, Zhang K, Zhao JK, et al. DDR2-CYR61-MMP1 Signaling Pathway Promotes Bone Erosion in Rheumatoid Arthritis Through Regulating Migration and Invasion of Fibroblast-Like Synoviocytes. *J Bone Miner Res*. 2017; 32(2):407–18. <https://doi.org/10.1002/jbmr.2993> PMID: 27653023.
  47. Sokka T, Pincus T. Erythrocyte sedimentation rate, C-reactive protein, or rheumatoid factor are normal at presentation in 35%-45% of patients with rheumatoid arthritis seen between 1980 and 2004: analyses from Finland and the United States. *J Rheumatol*. 2009; 36(7):1387–90. <https://doi.org/10.3899/jrheum.080770> PMID: 19411389.
  48. Bhasin S, Cheung PP. The Role of Power Doppler Ultrasonography as Disease Activity Marker in Rheumatoid Arthritis. *Dis Markers*. 2015; 2015:325909. <https://doi.org/10.1155/2015/325909> PMID: 26063952; PubMed Central PMCID: PMC4433665.
  49. Terslev L, Naredo E, Aegerter P, Wakefield RJ, Backhaus M, Balint P, et al. Scoring ultrasound synovitis in rheumatoid arthritis: a EULAR-OMERACT ultrasound taskforce-Part 2: reliability and application to multiple joints of a standardised consensus-based scoring system. *RMD Open*. 2017; 3(1):e000427. <https://doi.org/10.1136/rmdopen-2016-000427> PMID: 28948984; PubMed Central PMCID: PMC5597800.