

SNPs detected in the yak *MC4R* gene and their association with growth traits

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MC4R (melanocortin 4 receptor) is expressed in the appetite-regulating areas of the brain and takes part in leptin signaling pathways. Sequencing of the coding region of the *MC4R* gene for 354 yaks identified the following five single nucleotide polymorphisms (SNPs): SNP1 (273C > T), SNP2 (321 G > T), SNP3 (864 C > A), SNP4 (1069G > C) and SNP5 (1206 G > C). SNP1, SNP2 and SNP3 were synonymous mutations, whereas SNP4 and SNP5 were missense mutations resulting in amino acid substitutions (V286L and R331S). Pairwise linkage disequilibrium (LD) analysis indicated that two pairs of SNPs, SNP2 and SNP5 ($r^2 = 0.81027$) and SNP4 and SNP5 ($r^2 = 0.53816$), exhibited higher degrees of LD. CC genotype of SNP4, CGACG and CTCCC haplotypes for all SNPs were associated with increased BW of animals that were 18 months old and with the average daily gain. The secondary structure and transmembrane region prediction of the yak *MC4R* protein suggested that SNP4 was correlated with influential changes in the seventh transmembrane domain of the *MC4R* protein and with the functional deterioration or even incapacitation of *MC4R*, which may contribute to the increased feed intake, BW and average daily gain of the yaks with CC genotypes. The data from this study suggested that 1069G > C SNP of the *MC4R* gene could be used in marker-assisted selection of growth traits in the Maiwa yak breed.

Keywords: Maiwa yak, *MC4R* gene, SNPs, growth traits

Implication

In this study, we sequenced the coding region of the *MC4R* gene for 354 yaks, and five single nucleotide polymorphisms (SNPs) were identified, among which SNP4 (1069G > C) and SNP5 (1206 G > C) were missense mutations and resulted in amino acid substitutions (286V > L and 331R > S). SNP4 was assumed to correlate with influential changes in the seventh transmembrane domain of the *MC4R* protein and with the functional deterioration or even incapacitation of the *MC4R* gene. Among the three genotypes (GG, GC and CC) of SNP4, CC genotype was associated with increased BW of animals that were 18 months old and with the average daily gain. The data from this study suggested that 1069G > C SNP of the *MC4R* gene can act as a useful marker for the selection of growth traits in the Maiwa yak breed.

Introduction

BW is an important trait for meat production in livestock animals, and genetic factors play important roles in regulating

energy balance. Several genes have been identified to be involved in the neural signaling pathway of energy homeostasis (Barsh *et al.*, 2000; Zhang *et al.*, 2009). Among these genes, the *MC4R* (melanocortin 4 receptor) gene encodes a G-protein-coupled receptor expressed in the appetite-regulating areas of the brain, which takes part in leptin signaling pathways and leads to a decrease in feed intake because of agonist stimulation (Seeley *et al.*, 2004). Mutations of this gene were found to be the most common cause for hereditary human obesity (Pérusse *et al.*, 2005). Moreover, significant associations were found between *MC4R* genotypes and backfat, growth rates and feed intake in some breeds or lines of livestock animals. A functional mutation (Asp298Asn) was associated with daily food intake, growth and fat deposition traits in some commercial pig lines (Hernandez-Sanchez *et al.*, 2003; Houston *et al.*, 2004; Van den Maagdenberg *et al.*, 2007). Variants (–293C > G and –129A > G) in the 5'-untranslated region of the bovine *MC4R* gene were associated with two growth traits in Nanyang cattle breed (Zhang *et al.*, 2009). Therefore, the *MC4R* gene could be an effective candidate gene for growth-related traits selection in livestock breeding.

Bos grunniens (yak) is the most important domesticated species with outstanding adaptability to the alpine climates

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of the Qinghai-Tibetan Plateau, and the number of yaks in China account for over 90% of those distributed all over the world (Cai *et al.*, 2011). As in other animals, the yak *MC4R* gene is a single-exon gene containing the entire 332 amino acid coding sequence (999 bp) and exhibits abundant variations with other mammalian species (Cai *et al.*, 2011; Cai and Mipam, 2013). However, the variation patterns of the *MC4R* gene in the yak population and their association with growth traits have not been reported thus far. In this study, we aimed to investigate the single nucleotide polymorphisms (SNPs) in the *MC4R* gene and analyze their association with growth traits in the Maiwa yak population.

Material and methods

Animals and DNA sample

In total, 354 yaks were sampled and each individual was 12 months old (BW: 72 ± 4 kg). The sampled yaks were genetically unrelated and randomly selected from a breeding population of Maiwa yak in Hongyuan county, Sichuan province of China. Most of the yaks included in this study population were selected from different pastures at least 50 km away from each other. These yaks were of similar body size and grazed on alpine meadow grassland from May to October, during which abundant feed grass is available. The BW and average daily gain (from the 12th to the 18th month) for these 18-month-old yaks were determined, and ear tissue samples were collected and maintained in an ice box before being transported to laboratory. Genomic DNA of each individual was extracted from the ear muscle tissue using the normal method of phenol-chloroform extraction (Sambrook *et al.*, 1989). The sampling and protocol for this research were approved by animal use and ethics committee of the Southwest University of Science and Technology and Southwest University for Nationalities.

MC4R gene sequencing and mutation analysis

PCR was performed using the primers MC4R-F (5'-TGGGA CATTATTCACAGCAG-3') and MC4R-R (5'-CCTACACAG AAGAAAAAGCT-3'), designed by Cai *et al.* (2011) to amplify a fragment of 1238 bp in length covering the ORF region of the yak *MC4R* gene. PCR was performed in a 50 μ l reaction mixture containing 200 ng of genomic DNA, 1.5 mM MgCl₂, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 0.6 units of Taq polymerase (TaKaRa), 10 μ M of each primer and 0.2 mM of each dNTP. Thermal cycling was performed in a PTC-200 thermocycler (MJ Research, Inc., Watertown, MA, USA) under the following conditions: 4 min denaturation at 94°C, followed by 30 cycles of 30 s at 94°C, 30 s at 55°C, 30 s at 72°C and a final 7-min extension at 72°C, before cooling to 12°C for 10 min. PCR products were purified using a Qiagen QIAquick PCR purification kit and were sequenced in an ABI 3730 automated sequencer at BeiJing Zixi Bio Tech Co. Ltd., Changping, Beijing, China. Both strands of the PCR product were completely sequenced.

The *MC4R* gene sequences of 354 yaks were aligned and edited in Clustal X with parameters set to default (Thompson

et al., 1997). Polymorphic sites of nucleotide and amino acid sequences of the coding region of the *MC4R* gene were explored and analyzed using MEGA 4 (Kumar *et al.*, 2008), and Tajima's *D* test (Tajima 1989) for neutrality was performed in DnaSP Version 5.0 (Librado and Rozas, 2009).

SNP genotyping and analysis

The genotypes of each yak from different SNPs of the *MC4R* gene were determined by examining the sequencing chromatographs. Gene frequency, genotype frequency and deviation from the Hardy-Weinberg equilibrium (HWE) for each SNP site were analyzed using the online program SNPStats (<http://bioinfo.iconcologia.net/snpstats/start.htm>). This program was also used to measure the pairwise linkage disequilibrium (LD) pattern between SNPs based on multi-allelic *D'* (coefficient of LD) and *r*² (correlation coefficient) and estimate dominant haplotypes for all SNPs identified in the gene *MC4R*.

Statistical association analysis

The associations between *MC4R* SNP genotypes/major haplotypes and growth traits were analyzed using mixed models in the software SPSS 17.0. The models are as follows:

Model 1:

$$Y_{ijklm} = \mu + Sex_i + Age_j + Family_k + Pasture_l + Genotype_m + e_{ijklm},$$

Model 2:

$$Y_{ijklmn} = \mu + Sex_i + Age_j + Family_k + Pasture_l + Additive_m + Dominance_n + e_{ijklmn},$$

Model 3:

$$Y_{ijklm} = \mu + Sex_i + Age_j + Family_k + Pasture_l + Haplotype_m + e_{ijklm}.$$

In these models, Y_{ijklm} or Y_{ijklmn} represents the phenotypic value of the growth trait; μ is the least square mean; Sex_i indicates the fixed effect for the i^{th} sex; Age_j is the fixed effect for the j^{th} age; $Family_k$ indicates the fixed effect for the k^{th} family; $Pasture_l$ denotes the fixed effect for the l^{th} pasture; $Genotype_m$ denotes the fixed effect for the m^{th} genotype; $Additive_m$ indicates the fixed effect for the m^{th} additive effect; $Dominance_n$ indicates the fixed effect for the n^{th} dominance effect; $Haplotype_m$ denotes the fixed effect for the m^{th} haplotype; and e_{ijklm} or e_{ijklmn} is the random error.

Results

Variations and SNP genotypes in the *MC4R* gene

Yak *MC4R* is a single-exon gene containing the entire 999 bp sequence coding 332 amino acids (Cai *et al.*, 2011). In total, 354 *MC4R* gene sequences of yak were examined, and mutation analysis revealed five SNP sites. Among the five SNPs, SNP1, SNP2 and SNP3 were synonymous C > T, G > T and C > A substitutions at the positions 273, 321 and 864 (AF265221.1; Haegeman *et al.*, 2001), respectively (Table 1).

Table 1 SNPs, alleles and corresponding genotypes detected in the *MC4R* gene of *Bos grunniens*

| SNP/location in AF265221.1* | AA exchange** | Allele | Allele frequency | Genotype | Genotype frequency | P-value*** |
|-----------------------------|---------------|--------|------------------|----------|--------------------|------------|
| SNP1 C > T/273 | A20A | C | 0.54 | C/C | 0.31 | 0.42 |
| | | T | 0.46 | C/T | 0.44 | |
| | | | | T/T | 0.24 | |
| SNP2 G > T/321 | S36S | G | 0.98 | G/G | 0.96 | 1 |
| | | T | 0.02 | G/T | 0.04 | |
| SNP3 C > A/864 | L217L | C | 0.93 | C/C | 0.85 | 1 |
| | | A | 0.07 | C/A | 0.15 | |
| SNP4 G > C/1069 | V286L | G | 0.91 | G/G | 0.86 | 0.09 |
| | | C | 0.09 | G/C | 0.10 | |
| | | | | C/C | 0.04 | |
| SNP5 G > C/1206 | R331S | G | 0.97 | G/G | 0.94 | 1 |
| | | C | 0.03 | G/C | 0.06 | |

SNP = single nucleotide polymorphism.

*The location of each SNP was nominated according to the bovine *MC4R* gene (GenBank accession number: AF265221.1).

**Amino acid exchanges were determined according to yak *MC4R* (GenBank accession number: HM051376).

***Exact test for Hardy–Weinberg equilibrium.

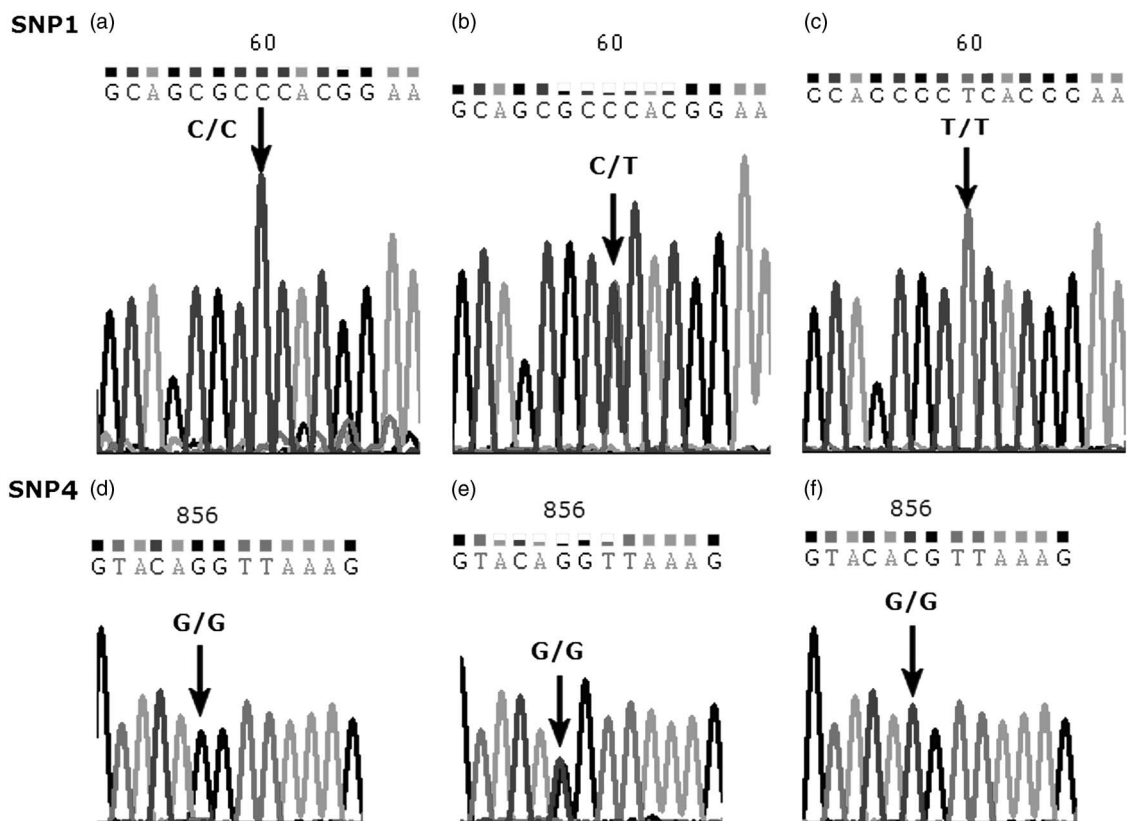


Figure 1 Sequencing chromatographs showing the genotypes determined from SNP1 and SNP4. a = homozygote C/C; b = heterozygote C/T; c = homozygote T/T; d = homozygote G/G; e = heterozygote G/C; and f = homozygote C/C. SNP = single nucleotide polymorphism.

However, SNP4 and SNP5 were both G > C missense mutations at positions 1069 and 1206 (AF265221.1), respectively, and these two sites resulted in amino acid substitutions (V286L and R331S), determined according to yak *MC4R* (HM051376) (Cai *et al.*, 2011; Table 1).

Genotyping the 354 yaks in the five SNP sites revealed that homozygotes were dominant in four sites, which could be displayed by the homozygote frequency as high as 96%

(G/G), 85% (C/C), 86% (G/G) and 94% (G/G) in SNP2, SNP3, SNP4 and SNP5, respectively. In SNP1, however, the frequency of the heterozygote (44%) was higher than that of each homozygote (Table 1). The genotypes determined from the sequencing chromatographs of SNP1 and SNP4 are shown in Figure 1. All these SNPs indicated no significant deviation from HWE ($P > 0.05$); therefore, the yak population sampled in this study were in an equilibrium state.

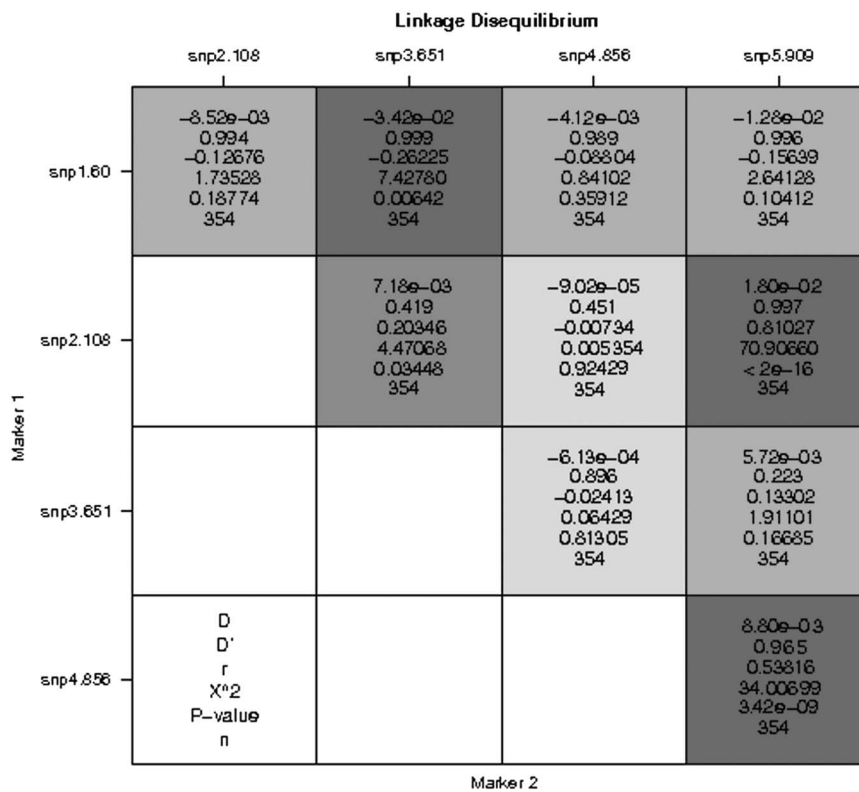


Figure 2 Linkage disequilibrium (LD) plot for the five single nucleotide polymorphisms (SNPs) of the yak *MC4R* gene. Color scheme is in accordance with *D*, *D'* and *r*² scheme from SNPStats program. The numerical data in each block donates pairwise value of *D*, *D'* and *r*². The blank block represents the equal value of *D*, *D'* and *r*² between the same pairs of SNPs. The five SNPs are SNP1.273, SNP2.321, SNP3.864, SNP4.1069 and SNP5.1206, in which the number after each SNP represents the nucleotide site in the bovine *MC4R* gene (GenBank accession number: AF265221.1).

| Haplotype frequencies estimation (n=354) | | | | | | | |
|--|---------|----------|----------|----------|----------|--------|----------------------|
| | snp1.60 | snp2.108 | snp3.651 | snp4.856 | snp5.909 | Total | Cumulative frequency |
| 1 | T | G | C | G | G | 0.463 | 0.463 |
| 2 | C | G | C | G | G | 0.4428 | 0.9058 |
| 3 | C | G | A | C | G | 0.0664 | 0.9722 |
| 4 | C | T | C | C | C | 0.0109 | 0.9831 |
| 5 | C | G | C | C | C | 0.0093 | 0.9924 |
| 6 | C | T | A | C | C | 0.0076 | 1 |
| 7 | T | G | A | G | G | 0 | 1 |

Figure 3 Haplotypes and estimated frequencies for the five single nucleotide polymorphisms (SNPs) of the yak *MC4R* gene.

Linkage disequilibrium of the SNPs

Pairwise LD analysis based on *D'* (coefficient of LD) and *r*² (correlation coefficient) showed that different degrees of LD were present in the five SNP locations (Figure 2). Higher degrees of LD existed between SNP2 and SNP5, as indicated by the higher *r*² values (*r*² = 0.81027). Similarly, the higher *r*² values (*r*² = 0.53816) also illustrated that higher degree of LD existed between SNP4 and SNP5. However, lower extent of LD was discovered between other pairwise SNPs (*r*² < 0.20346),

including SNP1-SNP2, SNP1-SNP3, SNP1-SNP4, SNP1-SNP5, SNP2-SNP3, SNP2-SNP4, SNP3-SNP4 and SNP3-SNP5, which suggested that there might be higher mutation rates or multiple recombination hotspots in the yak *MC4R* gene.

In total, seven haplotypes were identified for all SNPs in the yak *MC4R* gene (Figure 3), among which TGCGG and CGCGG were dominant with a higher frequency of 46.3% and 44.3%, respectively. However, other types of haplotype were rare as exemplified by low frequencies (<6.64%).

Table 2 *MC4R* genotypes corresponding to the least square means, and estimated genetic effects of growth traits in the Maiwa yak population

| Growth traits | SNPs | Genotypes | Genotype means \pm s.e. | Genetic effect | | | | |
|--------------------|----------------|-----------|---------------------------|----------------------------|----------------------------|---------------------|--------------------|-------------------|
| | | | | Additive effect \pm s.e. | Dominant effect \pm s.e. | | | |
| BW (12 months) | SNP1 | C/C | 73.68 \pm 4.32 | 0.943 \pm 0.342* | 0.351 \pm 0.157 | | | |
| | | C/T | 72.39 \pm 2.68 | | | | | |
| | | T/T | 71.80 \pm 5.35 | | | | | |
| | SNP2 | G/G | 71.29 \pm 1.35 | | | | | |
| | | G/T | 73.56 \pm 9.31 | | | | | |
| | SNP3 | C/C | 71.67 \pm 2.03 | | | | | |
| | | C/A | 73.71 \pm 7.46 | | | | | |
| | SNP4 | G/G | 70.67 \pm 2.33 | | | -1.725 \pm 0.504* | -0.185 \pm 0.074 | |
| | | G/C | 72.58 \pm 5.01 | | | | | |
| | | C/C | 74.12 \pm 8.74 | | | | | |
| | SNP5 | G/G | 70.62 \pm 2.13 | | | | | |
| | | G/C | 73.78 \pm 7.02 | | | | | |
| | BW (18 months) | SNP1 | C/C | | | 103.82 \pm 5.78 | 1.455 \pm 0.431* | 0.505 \pm 0.127 |
| | | | C/T | | | 102.87 \pm 4.35 | | |
| | | | T/T | | | 100.91 \pm 6.92 | | |
| SNP2 | | G/G | 101.52 \pm 1.41 | | | | | |
| | | G/T | 103.67 \pm 8.97 | | | | | |
| SNP3 | | C/C | 100.03 \pm 3.08 | | | | | |
| | | C/A | 103.91 \pm 8.00 | | | | | |
| SNP4 | | G/G | 101.95 \pm 3.24 | -5.835 \pm 0.537** | -0.915 \pm 0.371 | | | |
| | | G/C | 106.87 \pm 5.67 | | | | | |
| | | C/C | 113.62 \pm 8.63** | | | | | |
| SNP5 | | G/G | 98.92 \pm 3.05 | | | | | |
| | | G/C | 101.79 \pm 8.42 | | | | | |
| Average daily gain | | SNP1 | C/C | 0.17 \pm 0.03 | 0.005 \pm 0.003 | 0.005 \pm 0.002 | | |
| | | | C/T | 0.17 \pm 0.02 | | | | |
| | | | T/T | 0.16 \pm 0.05 | | | | |
| | SNP2 | G/G | 0.17 \pm 0.01 | | | | | |
| | | G/T | 0.17 \pm 0.07 | | | | | |
| | SNP3 | C/C | 0.16 \pm 0.02 | | | | | |
| | | C/A | 0.17 \pm 0.05 | | | | | |
| | SNP4 | G/G | 0.17 \pm 0.02 | -0.025 \pm 0.008* | | | -0.005 \pm 0.05 | |
| | | G/C | 0.19 \pm 0.05 | | | | | |
| | | C/C | 0.22 \pm 0.07* | | | | | |
| | SNP5 | G/G | 0.16 \pm 0.02 | | | | | |
| | | G/C | 0.16 \pm 0.06 | | | | | |

*Least square means significantly different among the genotypes ($P < 0.05$).

**Least square means significantly different among the genotypes ($P < 0.01$).

Association of SNP genotypes/major haplotypes with growth traits

Statistical association analyses between *MC4R* genotypes and growth traits of Maiwa yak revealed that SNP1, SNP2, SNP3 and SNP5 showed no effects on BW of animals at 18 months of age and on average daily gain as well. However, missense mutations G > C at position 856 of the SNP4 gene may suggest that the change in *MC4R* protein and the CC genotype determined from SNP4 were associated with increased BW of animals aged 18 months (CC-113.62 \pm 8.63 v. GC-106.87 \pm 5.67 and GG-101.95 \pm 3.24; $P < 0.01$) and with average daily gain (CC-0.22 v. GC-0.19 and GG-0.17; $P < 0.05$) (Table 2). Two major haplotypes for all SNPs, CGACG and CTCCC were associated with significantly increased BW of animals aged 18 months ($P < 0.01$) and with the average daily gain ($P < 0.05$) (Table 3).

Table 3 Association between the major haplotypes and growth traits in the Maiwa yak population

| Haplotypes | BW \pm s.e. (12 months) | BW \pm s.e. (18 months) | Average daily gain \pm s.e. |
|------------|------------------------------|------------------------------|----------------------------------|
| TGCCG | 72.65 \pm 2.31 | 100.43 \pm 3.65 | 0.15 \pm 0.02 |
| CGCGG | 74.13 \pm 3.47 | 104.32 \pm 4.02 | 0.17 \pm 0.02 |
| CGACG | 74.67 \pm 6.90 | 113.07 \pm 7.02** | 0.21 \pm 0.06* |
| CTCCC | 76.07 \pm 8.21 | 114.01 \pm 9.31** | 0.21 \pm 0.08* |

*Least square means significantly different among the haplotypes ($P < 0.05$).

**Least square means significantly different among the haplotypes ($P < 0.01$).

Analyses of the genetic effects of different SNP genotypes indicated that additive effects were significant or extremely significant at SNP1 ($P < 0.05$) and SNP4 ($P < 0.01$) (Table 2),

but there were no significant dominance effects at these loci. The negative dominance effects in SNP4 may indicate negative heterosis. All absolute values of the additive effect were higher than those of the dominance effect at the two loci, which suggested that additive effects played more important roles than dominance effects.

Discussion

Current situation and difficulties in yak breeding

Yaks are one of the most important domesticated species in China; however, studies on comprehensive and accurate assessment of yak genetic resources have been greatly restricted. The lack of reliable DNA markers from yak genome was one of the important reasons, which also resulted in stagnation of marker-assisted selection in yak breeding. On the other hand, yaks are semi-grazing and semi-feeding animals in different periods of the year and it is difficult to record and obtain reliable data of quantitative traits from these animals. Considering these conditions, we selected 12-month-old yaks with similar body size (BW: 72 ± 4 kg) and these animals were grazed on grasslands from May to October, during which abundant feed grass was available.

The missense mutation 1069 G > C in the MC4R gene and its association with growth traits

The missense mutation 1069 G > C in the *MC4R* gene and its association with growth traits have been studied in different cattle breeds. Five polymorphisms were detected at position 19 (C > A), 20 (A > T), 83 (T > C), 128 (G > A) and 1069 (G > C) in the *MC4R* gene of eight Chinese cattle breeds, among which 1069 (G > C) was significantly associated with backfat thickness (Huang *et al.*, 2010). The missense

mutation 1069 G > C in the *MC4R* gene of Chinese Qinchuan cattle was significantly associated with live weight, carcass weight, backfat thickness and marbling score (Liu *et al.*, 2010). In the *MC4R* gene of Korean cattle (Hanwoo), 1069 G > C significantly affected backfat thickness (Seong *et al.*, 2012). However, no significant association was found between the 1069 G > C locus of *MC4R* and residual feed intake, feed conversion ratio and average daily gain in a Simmental bull population (Du *et al.*, 2013).

The yak *MC4R* is a single-exon gene containing the entire 332 amino acid coding sequence (999 bp; Cai *et al.*, 2011). Analysis of 354 sequences of the *MC4R* gene showed five SNP sites, representing 0.5% of the total gene sequence analyzed (999 bp). Therefore, the sequences of *MC4R* of yaks showed low levels of polymorphisms in this study, which was due to the important function of *MC4R* in regulating food intake behavior. Low levels of variation in the *MC4R* gene could also be explained by the view that *MC4R* might have been subjected to high levels of continuous purifying selection during yak evolution and variations that would influence the activity of metabolism also had been subjected to purifying selection mechanism during yak evolution (Hughes *et al.*, 2009; Cai *et al.*, 2011).

Pairwise LD analysis of the five SNPs showed that higher degrees of LD only existed between SNP2 and SNP5 ($r^2 = 0.81027$), similarly higher degrees of LD was also examined between SNP4 and SNP5 ($r^2 = 0.56715$). However, lower extent of LD was discovered between other pairwise SNPs, including SNP1-SNP2, SNP1-SNP3, SNP1-SNP4, SNP1-SNP5, SNP2-SNP3, SNP2-SNP4, SNP3-SNP4 and SNP3-SNP5. Domestication and breeding improvement were involved in the selection for specific alleles at candidate genes, which resulted in reduced genetic diversity and increased LD relative to unselected genes (Yamsaki *et al.*, 2005; Zhou *et al.*, 2008). In this

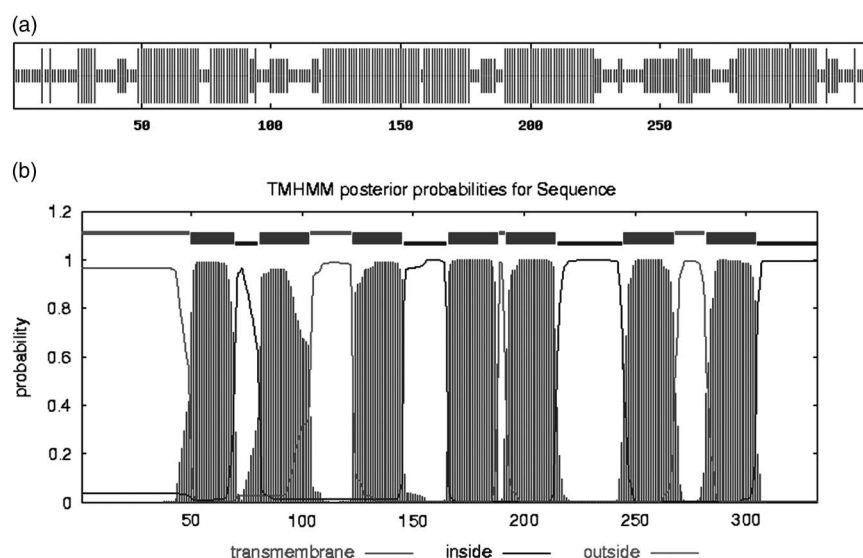


Figure 4 Prediction of the secondary structure and transmembrane region of the yak melanocortin 4 receptor (MC4R) protein. a = the secondary structure prediction of the yak MC4R gene; the blue, red and purple vertical lines stand for α -helix, extended chains and random coils, respectively. b = the transmembrane region prediction for the yak MC4R; the red, blue and purple areas stand for transmembrane regions, inside regions of membrane and outside regions of membrane, respectively.

study, low levels of LD together with a negative Tajima's *D* (-1.08498 , $P > 0.10$) further indicated low levels of polymorphisms in the yak *MC4R* gene and yaks may have been subjected to purifying (negative) selection during their evolution. This was also correlated with the fact that yaks have been grazed livestock for thousands of years and less domestication, artificial selection or breeding improvements were imposed on and they have undergone a long history of natural selection during their pasture grazing and extensive management.

Missense mutations caused amino acid substitution in the protein and secondary or spatial structural changes in the protein, which frequently gave rise to functional changes of the protein. Among the five SNPs identified in this study, two (SNP4 and SNP5) were missense mutations and resulted in amino acid substitutions (V286L and R331S), determined according to yak *MC4R* (HM051376) (Cai *et al.*, 2011). Association analyses between SNP genotypes and the growth traits indicated that SNP4 may have caused the functional change of *MC4R*, because CC genotype determined from SNP4, CGACG and CTCCC haplotypes for all SNPs were associated with increased BW of animals aged 18 months and with average daily gain. The secondary structure (<http://npsa-pbil.ibcp.fr/cgi-bin/npsa-automat.pl>) and transmembrane region prediction (<http://www.cbs.dtu.dk/services/TMHMM/>) of the yak *MC4R* protein revealed that the replacement of valine with leucine at position 286 of the *MC4R* protein was within the seventh transmembrane domain (Figure 4). In this sense, we assumed that SNP4 resulted in influential changes in the seventh transmembrane domain of the *MC4R* protein as well as in the functional deterioration or even incapacitation of the *MC4R* protein, which might contribute to the increased feed intake, BW and average daily gain of the yaks with CC genotypes. Therefore, all these data suggested that 1069 G > C SNP of the *MC4R* gene could be used in marker-assisted selection of growth traits in Maiwa yak breed.

Acknowledgments

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