

# Do genetically modified crops affect animal reproduction? A review of the ongoing debate

W. Zhang and F. Shi<sup>†</sup>

Laboratory of Animal Reproduction, College of Animal Science and Technology, Nanjing Agricultural University, Weigang 1, Nanjing 210095, People's Republic of China

(Received 20 July 2010; Accepted 16 November 2010; First published online 28 January 2011)

In the past few years, genetically modified (GM) crops aimed at producing food/feed that became part of the regular agriculture in many areas of the world. However, we are uncertain whether GM food and feed can exert potential adverse effects on humans or animals. Of importance, the reproductive toxicology of GM crops has been studied using a number of methods, and by feeding GM crops to a number species of animals to ensure the safety assessment of GM food and feed. It appears that there are no adverse effects of GM crops on many species of animals in acute and short-term feeding studies, but serious debates of effects of long-term and multigenerational feeding studies remain. The aims of this review are to focus on the latest (last 3 to 4 years) findings and debates on reproduction of male and female animals after feeding daily diets containing the GM crops, and to present the possible mechanism(s) to explain their influences.

Keywords: genetically modified crop, reproductive toxicity, exogenous protein and DNA, anti-nutrient compound

## **Implications**

Genetically modified (GM) plants are now under development and rapid commercial use in all over the world. However, we are uncertain whether GM food and feed can exert potential reproductive toxicology on humans or animals. In this review, we focus on the latest (last 3 to 4 years) findings and debates on reproduction of male and female animals after feeding daily diets containing the GM crops, and to present the possible mechanism(s) to explain their influences. The controversy about the potential reproductive toxicology of GM foods is complex, which certainly would require further scientific investigation to answer safety concerns.

# Introduction

Genetically modified (GM) crops are identified as crops that use modern techniques of genetic engineering (or biotechnology) to introduce specific genetic material derived from any species of plant, animal, microorganism or even synthetic material into different species of plants by altering genetic material (DNA) coding for herbicide tolerance, insect resistance or a combination of these traits in a way that does not occur naturally. Then, the resulting plants can express

the novel and desirable traits, such as enhanced disease resistance, anti-reversion force or secretion of useful proteins during different stages of the plant growth. In these plants, the genetic insert leads to the production of a gene product, which does not interfere with the overall metabolism of the plant cell, and does not alter the composition of the GM plant except for the introduced trait (World health Organization (WHO), 2002; European Food Safety Authority -Genetically Modified Organism (EFSA GMO) Panel, 2008; Magana-Gomez and delaBarca, 2009; Dhlamini, 2009). With many advantages over conventional crops, GM plants are now under development and rapid commercial use since 1996. On the one hand, the global area of GM crops has increased >80-fold, from 1.7 million hectares in six countries in 1996 to 143 million hectares in 23 countries in 2007. The world's top six producers – the United States, Argentina, Brazil, Canada, India and China – account for >90% of global GM production (James, 2007). On the other hand, more and more types of genetic materials and the systems for GM crops have been created, accompanying the improvement in genetic engineering. These include soybean with improved amino acid composition or potato with enhanced calcium content and other functional foods (Krishnan, 2005; Park et al., 2005; Hirschi, 2009). Owing to the development of GM crops, they are today distributed all over the world, frequently becoming part of human and

<sup>†</sup> E-mail: fxshi@njau.edu.cn

animal diets (Sanvido et al., 2007). As diet is considered one of the most important environmental factors affecting life span, GM crop genomes into which new genes have been inserted by using modern techniques of genetic engineering are very different from conventional crops at the aspect of plant improvement to raise and stabilize yields, to improve resistance to pests and diseases (Dhlamini, 2009). Many discussion forums, studies and publications have been devoted to the safety assessment of GM food and feed. Of importance, reproductive toxicology of GM crops is studied in order to detect whether they will interfere in some way in normal reproduction and induce adverse effects on sexual function and fertility in male and female animals, as well as developmental toxicity in the offspring. Therefore, many studies have been carried out to ensure the safety of GM food via number of methods and species of common animals by feeding a diet containing the novel materials such as a new protein or secondary metabolite. However, it remains debatable whether GM food and feed exert potentially adverse effects on humans or animals. The aims of this review are to focus on the latest (last 3 to 4 years) findings and debates on reproduction of male and female animals after feeding daily diets containing the GM crops, and to present the possible mechanism(s) to explain their influences.

#### The effect of GM crops on female animal reproduction

# Background

In the toxicological investigation of GM crops, various international guidelines have been designed by international organizations like Food and Agriculture Organization (FAO) of The United Nations, World Health Organization (WHO), Organisation for Economic Co-operation and Development (OECD) and the Codex Alimentarius Commission (CAC) to assess the safety of GM foods on the basis of risk analysis concepts and principles. Current approaches are based on internationally approved acute or chronic tests in laboratory animals (rat or mouse) and fast-growing domesticated species such as chicken (FAO/WHO, 2000; OECD, 2007; CAC, 2003a and 2003b; Craig et al., 2008; Sesikeran and Vasanthi, 2008). Especially, the ability of the 90-day rat feeding study to detect the biological/toxicological effects of the new gene product in the GM food (Knudsen and Poulsen, 2007). The effects of GM crops on female animal reproduction are shown in Table 1.

#### Short-term feeding studies

Using animal feeding models, many studies indicated that administration of a large single dose had no acute toxicity on animals as evaluated over a short period of observation. At present, many groups used a short-term (not >1 month) feeding animal model to evaluate safety of GM crops. When feeding different GM crops to mice, no adverse effects were observed in the ovaries fed for 27 days by a GM crop 356043 soybeans contained glyphosate acetyltransferase protein, GAT4601 protein; Delaney *et al.*, 2008a). Moreover, no microscopic pathology was observed in vagina of animals

treated with Cry34Ab1 or Cry35Ab1 proteins for 28 days, even at repeated high dose (1000-fold greater than the highest estimate of human exposure based on the concentrations of these proteins expressed in 59 122 maize grain; Juberg *et al.*, 2009). In livestock, no negative effects were observed for laying hens fed for 21 days or 4 weeks on any parameter measured, including the number of yolks and egg and ovary weight, follicle number, oviductal weight, egg production, egg mass and feed efficiency (Rasmussen *et al.*, 2007; Jacobs *et al.*, 2008).

### Long-term and multigenerational feeding studies

Long-term feeding studies. Compared with the short-term feeding study, the long-term feeding study contained subchronic toxicity (a reduction in the tested animal's life span by  $\sim 10\%$ ) and chronic toxicity, and allowed the investigator to ascertain the variation in responses. At present, many research groups have started to evaluate the safety of GM food and feed in the long-term feeding study.

In the 90-day or 13-week feeding studies in rats, no statistical difference was uncovered in either the relative body and ovary weight of rats fed transgenic corns (Hammond et al., 2006; Healy et al., 2008), maize (MacKenzie et al., 2007; He et al., 2009; Appenzeller et al., 2009a and 2009b), rice (Schroder et al., 2007) or soybeans (Appenzeller et al., 2008; Delaney et al., 2008b); and there were no histopathologic lesions in ovaries from rats fed DAS-59122-7 maize (Cry34Ab1 and Cry35Ab1 proteins; He et al., 2008). Furthermore, Malley et al. (2007) reported higher mean uterus weight during the estrous stage of rats fed maize DAS-59122-7 (Cry34Ab1 and Cry35Ab1 proteins) or 5002B (commercial rodent diets) v. the 33R77 group (non-transgenic reference maize grain), 091 group (non-transgenic nearisogenic maize grain) or 5002A (commercial rodent diets); however, this might be due to the fact that the proportion of rats in proestrus and estrus in the 59122 and 5002B groups was greater than that in the 5002A, 091 and 33R77 maize grain groups, in which a greater proportion of rats were in metestrus and diestrus. The longest long-term feeding study was a 3-year longitudinal study of feeding sheep a diet containing Bt176 (Bacillus thuringiensis) maize. This study indicated that there were no differences in reproductive traits such as fertility and twinning rate, body weight at birth, mortality or daily weight gain up to weaning (90 days of age) in lambs (Trabalza-Marinucci et al., 2008).

As described above for long-term feeding study, their greatest concerns are on ovulation of a normal oocyte, fertilization, uterine status, implantation and prenatal development. However, the female reproductive system is also at risk during fetal development *in utero*, postnatally during puberty and during the female's reproductive lifetime. Given the safety assessment of GM food and feed, Cisterna *et al.* (2008) investigated the ultrastructural and immunocytochemical features of pre-implantation embryos from 10 two-month-old Swiss mice fed a standard diet containing 14% GM soybean or non-GM soybean until weaning (i.e. for 40 to 50 days). Morphological observations revealed that the general aspects

 Table 1 The effect of GM crops on female animal reproduction

| Plant/crop                                  | Inserted protein or trait  | Animal species | Length of the study | Main adverse effects   | Reference                         |
|---|--|----------------|---------------------|--|-----------------------------------|
| Short-term feeding study                    |  |                |                     |  |                                   |
| DP-356ø43-5 soybeans                        | Glyphosate acetyltransferase protein (GAT4601 protein)                             | Mice           | 27 days             | No adverse effects observed in the ovaries.  | Delaney <i>et al.</i> (2008a)     |
| DAS-59122-7 corn                            | Cry34Ab1 and Cry35Ab1 proteins   | Mice           | 28 days             | No adverse effects observed in the ovaries.  | Juberg <i>et al</i> . (2009)      |
| Starlink corn                               | Cry9C protein  | Laying hens    | 21 days             | No negative effects observed on any parameter measured such as the number of yolks and egg weight and, ovary and oviduct weight.   | Rasmussen <i>et al.</i> (2007)    |
| DAS-59122-7 maize                           | Cry34Ab1 and Cry35Ab1 proteins   | Laying hens    | 4-week phases       | No significant difference in egg production and egg mass.  | Jacobs <i>et al.</i> (2008)       |
| Long-term and multigeneration feeding study |  |                |                     |  |                                   |
| MON810 corns                                | Cry1Ab protein   | Rats           | 90 days             | No statistical difference in the relative weight and the relative weight of the ovaries.   | Hammond <i>et al</i> . (2006)     |
| MON88017 corn                               | Cry3Bb1 protein  | Rats           | 13 weeks            | No adverse effects observed in the ovaries.  | Healy <i>et al.</i> (2008)        |
| DAS-ø15ø7-1 maize                           | Cry1F protein  | Rats           | 13 weeks            | No adverse effects observed in the ovaries.  | MacKenzie <i>et al</i> . (2007)   |
| DAS-59122-7 maize                           | Cry34Ab1 and Cry35Ab1 proteins   | Rats           | $\sim$ 90 days      | The proportion of rats in proestrus and estrus was higher in metestrus and diestrus, (there is no adverse effect compared with non-transgenic groups at different stages of estrus stage by two-way analysis). | Malley <i>et al.</i> (2007)       |
| DAS-59122-7 maize                           | Cry34Ab1 and Cry35Ab1 proteins   | Rats           | 90 days             | No statistical difference in the relative weight and the relative weight of the ovaries and no histopathological lesions in the ovaries.   | He <i>et al</i> . (2008)          |
| DP-ø9814ø-6 maize                           | Acetylase GAT4621  | Rats           | 13 weeks            | No adverse effects observed in the ovaries.  | Appenzeller et al. (2009a)        |
| Y642 transgenic maize                       | Lysine-rich protein  | Rats           | 90 days             | No statistical difference in the relative weight and the relative weight of the ovaries and in gross or microscopic pathology.   | He <i>et al</i> . (2009)          |
| DAS-ø15ø7-1xDAS-59122-7<br>maize            | Phosphinothricin-N-<br>acetyltransferase (PAT),<br>Cry1F, Cry34Ab1 and<br>Cry35Ab1 | Rats           | 93 to 94 days       | No statistically significant differences in the relative weight of ovaries and uterus.   | Appenzeller <i>et al.</i> (2009b) |
| KMD1 rice                                   | Cry1Ab protein   | Rats           | 90 days             | No statistical difference in the relative weight and the relative weight of the ovaries, the uterus absolute weight were observed few significant differences compared with non-transgenic rice.               | Schroder <i>et al.</i> (2007)     |
| DP-356ø43-5 soybean                         | Glyphosate<br>acetyltransferase4601<br>(GAT4601)                                   | Rats           | 93 days             | No statistically significant differences in mean relative organ weight in ovaries and uterus and no evidence of altered incidence or severity of pathological changes or lesions was observed.                 | Appenzeller <i>et al.</i> (2008)  |

Effects of GM crops on animal reproduction

Table 1 Continued

| Plant/crop                                       | Inserted protein or trait  | Animal species | Length of the study   | Main adverse effects  | Reference                                  |
|--|--|----------------|---|---|--|
| DP-3ø5423-1 soybean                              | High oleic acid ( <i>gm-fad2-1</i><br>gene)                              | Rats           | 90 days   | No statistical difference in the relative weight and the Relative weight of the ovaries.  | Delaney <i>et al</i> . (2008b)             |
| Bt176 maize                                      | Cry1 protein   | Sheep          | 3 years   | No differences were observed in reproductive traits such as fertility and twin rate, the lambs' BW at birth.  | Trabalza-Marinucci <i>et al.</i><br>(2008) |
| GM soybean                                       | Not described  | Mice           | 40 to 50 days<br>(parent strain)  | Embryo nuclear components is similar, a temporary decrease of pre-mRNA transcription and splicing in two-cell embryos and a resumption in four- and eight-cell embryos.                                     | Cisterna <i>et al.</i> (2008)              |
| GM potato (N-14)                                 | Herbicide resistant bar gene   | Rats           | A five-generation<br>animal study fed<br>for 10 weeks.                                | No GM potato-related changes in reproductive performance, ovaries and uterus weight.  | Rhee <i>et al.</i> (2005)                  |
| GM Bt corn                                       | Herbicide resistant bar gene   | Rats           | Parental generation fed from pregnancy, the other generation fed 3.5 months.          | No signs of adverse effects were seen in clinical appearance of newborns in all three generation. Number of offspring in F1, F2 and F3 generations, birthrate and survival of the offspring did not change. | Kilic and Akay (2008)                      |
| Glyphosate-tolerant or<br>Roundup Ready soybeans | Herbicide resistant bar gene   | Mice           | 28 days   | High level of mortality ( $\sim$ 55, 6%) was observed with pups and 36% of these weighed $<$ 20 g.  | Ermakova (2005)                            |
| NK603 × MON810 corn                              | Cry1Ab and CP4-EPSPS<br>(5-enolpyruvlshimimate-3-<br>phosphate synthase) | Mice           | Multigeneration study (parental generation) and life term study (all fed from birth). | The production parameters average litter size and weight in the 3rd and 4th litters of continuous breeding GM corn were statistically significant compared with non-GM groups.                              | Velimirov <i>et al.</i> (2008)             |

GM = genetically modified.

of embryo nuclear components were similar in the two experimental groups. However, immunocytochemical and *in-situ* hybridization results suggested a temporary decrease in pre-mRNA transcription and splicing in two-cell embryos and a resumption in four- and eight-cell embryos from mice fed the GM soybean. In addition, pre-mRNA maturation seemed to be less efficient in both two-, four- and eight-cell embryos from GM-fed mice than in controls (Cisterna et al., 2008). However, these studies did not provide any information on the source, nutritional composition or any kind of processing of the soybeans used, and the sample size was small for female mice (n = 5), and there was a lack of description of the embryo. The evidence is still far from certain as to whether the long-term consumption of GM foods possesses a possible danger for human or animal health. Therefore, further studies on the effects of GM food components on embryo development are needed.

Multigenerational feeding studies. The limitation of a one-generation test is that the reproductive capacity of chemically exposed rats both prenatally and postnatally is not assessable. However, in a multigeneration test, the postweaning maturation and reproductive capacity of the pups can be evaluated (Francis and Kimmel, 1988). Therefore, more and more reports are being designed to clarify and enlighten the possible effects on animal health of GM crops through multigenerational feeding studies.

One group examined the potential reproductive and developmental toxic effects of rats by using five generation of animals fed a solid pellet containing 5% GM potato and non-GM potato for 10 weeks before mating. They uncovered no GM potato-related changes in reproductive performance, histopathological observations of the reproductive tissues and organ weight was not different. The litter-related indices did not show any GM organism (GMO)-related changes (Rhee et al., 2005). Kilic evaluated the effects of GM Bt corn on the rats that were fed through three generations with either GM corn or its conventional counterpart. No signs of adverse effects were seen in clinical appearance of newborns in all three generations. The dams gave fertile progeny and successfully continued their strips. Number of offspring in F1, F2 and F3 generations, birthrate and survival of the offspring were not changed among groups suggesting their successful reproduction (Kilic and Akay, 2008). Conversely, high levels of mortality (55.6%) and decreases in birth weight of offspring were reported in a GM soybean feeding study in which female rats were fed before mating, during mating and during pregnancy (Ermakova, 2005). However, the study of Ermakova (2007) was in debate and there were certainly no conclusions for their results (Marshall, 2007). In 2008, Velimirov carried out a series of experiments involved in almost every aspect of a multigenerational feeding study. The test diets differed only as to the inclusion of 33% NK603  $\times$  MON810 GM corn  $\nu$ . non-GM corn of a near-isogenic line. They found that the production parameters such as average litter size and weight in the 3rd and 4th litters of continuous breeding GM corn were statistically

significant compared with non-GM groups. In addition, analyses of metabolic pathways by microarrays indicated that the groups differed with regard to some important biochemical pathways, including interleukin (IL) signaling, cholesterol biosynthesis and protein metabolism (Velimirov et al., 2008). Their studies are by far the most meticulous and comprehensive feeding trials to date, and confirm deleterious reproductive and health impacts obtained by scientists independent of the biotech industry and farmer observations in the field. However, the researchers at Monsanto Company think that Dr Velimirov's report lacks sufficient experimental details to fully interpret the results and contains a number of errors that make it unsuitable for risk assessment and/or regulatory purposes (Monsanto Company, 2008).

## The effect of GM crops on male animal reproduction

# Background

The male reproductive system is at risk during fetal development *in utero*, postnatally during puberty and even over the entire life span with targets including testes and accessory organs. In addition, the high rate of cellular proliferation and the unique cellular differentiation within the mammalian testis make it a very sensitive organ that can detect cellular and molecular changes that occur when exposed to a toxicant (Evenson *et al.*, 1980). Therefore, many research groups are concerned about whether the GM food exert negative effects on the male reproductive system in order to ensure the safety of GM food by a number of methods over many species of animals. The effects of GM crops on male animal reproduction are shown in Table 2.

#### Short-term feeding studies

Bt proteins have been shown to be rapidly degraded *in vitro* using simulated gastric fluids (Betz *et al.*, 2000; Momma *et al.*, 2000). Therefore, in a short-term feeding study, there were no statistically significant differences in the testis weight compared with rats fed non-GM soybeans (Delaney *et al.*, 2008a). Moreover, no microscopic pathology was observed in testes of Cry34Ab1 or Cry35Ab1 protein-treatment groups for 28 days, even at a repeated high dose 1000-fold > the highest estimate of human exposure based on the concentrations of these proteins expressed in 59 122 maize grains (Juberg *et al.*, 2009).

# Long-term and multigenerational feeding studies

Long-term feeding studies. In a 90-day or 13-weeks feeding studies on rats, several groups have reported that there were no effects on male reproductive organs such as testes, epididy-mides and prostate in rodents compared with rats receiving non-GM food in long-term study when the diet was treated with different GM foods (Hammond et al., 2006; MacKenzie et al., 2007; Schroder et al., 2007; Appenzeller et al., 2008, 2009a and 2009b; Delaney et al., 2008b; Healy et al., 2008; He et al., 2008 and 2009). However, in MacKenzie et al. (2007 study, there was no statistical difference in the relative weight of the testes, epididymides or prostate. Specifically, the relative

Effects of GM crops on animal reproduction

 Table 2 The effect of GM crops on male reproduction

| Plant/crop                                  | Inserted protein or trait                                 | Animal species | Length of the study  | Main adverse effects   | Reference                         |
|---|---|----------------|--|--|-----------------------------------|
| Short-term feeding study                    |   |                |  |  |                                   |
| DP-356ø43-5 soybeans                        | Glyphosate acetyltransferase<br>Protein (GAT4601 protein) | Mice           | 27 days  | No statistically significant differences in the testes weight.   | Delaney <i>et al</i> . (2008a)    |
| DAS-59122-7 corn                            | Cry34Ab1and Cry35Ab1 protein                              | Mice           | 28 days  | No adverse effects observed in the testes.   | Juberg <i>et al</i> . (2009)      |
| Long-term and multigeneration feeding study |   |                |  |  |                                   |
| MON810 corns                                | Cry1Ab protein  | Rats           | 90 days  | No statistical difference in the relative weight and the relative weight of the testes.  | Hammond <i>et al.</i> (2006)      |
| MON88017 corn                               | Cry3Bb1 protein   | Rats           | 13 weeks   | No adverse effects observed in the testes.   | Healy <i>et al</i> . (2008)       |
| DAS-ø15ø7-1 maize                           | Cry1F protein   | Rats           | 13 weeks   | No adverse effects observed in the testes, epididymides and prostate, the relative kidney weight in the 33% 1507 maize grain group were lower.                                     | MacKenzie <i>et al.</i> (2007)    |
| DAS-59122-7 maize                           | Cry34Ab1 and Cry35Ab1 proteins                            | Rats           | 90 days  | No effect on rodent male organ of reproduction such as testes, epididymides and prostate.  | Malley <i>et al</i> . (2007)      |
| DP-3ø5423-1 soybeans                        | High oleic acid ( <i>gm-fad2-1</i> gene)                  | Rats           | 90 days  | No effect on rodent male organ of reproduction such as testes, epididymides and prostate.  | Delaney <i>et al</i> . (2008b)    |
| DAS-ø15ø7-1xDAS-59122-7<br>maize            | PAT, Cry1F, Cry34Ab1 and<br>Cry35Ab1                      | Rats           | 92 to 93 days  | No statistically significant differences in organ/body weight ratios of testes. No microscopic findings in prostate and urinary bladder.   | Appenzeller et al. (2009b)        |
| DP-ø9814ø-6 maize                           | Acetylase GAT4621   | Rats           | 13 weeks   | No statistically significant differences organ/body weight ratios of testes.   | Appenzeller <i>et al.</i> (2009a) |
| Y642 transgenic maize                       | Lysine-rich protein                                       | Rats           | 90 days  | No statistically significant differences in mean relative organ weight in testes and no differences in gross or microscopic pathology were observed.                               | He <i>et al.</i> (2009)           |
| KMD1 rice                                   | Cry1Ab protein  | Rats           | 90 days  | No effect on rodent male organ of reproduction such as testes, epididymides and prostate.  | Schroder et al. (2007)            |
| DP-3ø5423-1 Soybeans                        | High oleic acid ( <i>gm-fad2-1</i><br>gene)               | Rats           | 90 days  | No effect on rodent male organ of reproduction such as testes, epididymides and prostate.  | He <i>et al</i> . (2008)          |
| DP-356ø43-5 soybean                         | Glyphosate acetyltransferase<br>4601 (GAT4601)            | Rats           | 93 days  | No statistically significant differences in mean relative organ weight in testes and no evidence of altered incidence or severity of pathological changes or lesions was observed. | Appenzeller <i>et al.</i> (2008)  |
| Zea mays L. Bt corn                         | Cry protein   | Mice           | Parent strain fed GM corn<br>and male progeny fed<br>8, 16, 26, 32, 63 and<br>87 days after birth. | No apparent differences in percentages of testicular cell populations (haploid, diploid and tetraploid).   | Brake <i>et al.</i> (2004a)       |

Table 2 Continued

| Plant/crop                                       | Inserted protein or trait  | Animal<br>species | Length of the study  | Main adverse effects  | Reference                      |
|--|--|-------------------|--|---|--------------------------------|
| Glyphosate-tolerant or<br>Roundup Ready soybeans | Bt protein   | Mice              | Parent strain fed GM soybeans, and male progeny fed 8, 16, 26, 32, 63 and 87 days after birth. | No apparent differences in percentages of testicular cell populations (haploid, diploid and tetraploid).  | Brake and Evenson<br>(2004b)   |
| NK603 $	imes$ MON810 corn                        | Cry1Ab and CP4-EPSPS<br>(5-enolpyruvlshimimate-<br>3-phosphate synthase) | Mice              | Multigeneration study (parental generation) and life term study (all fed from birth).          | No pathological findings observed in the testes.  | Velimirov <i>et al.</i> (2008) |
| GM soybean                                       | Not described  | Mice              | Pregnant mice were fed on<br>GM soybean.   | Enlargements in the smooth endoplasmic reticulum in GM-fed mice Sertoli cells. Immunolabelling for Sm antigen, hnRNPs, SC35 and RNA polymerase II is decreased in 2- and 5-month-old GM-fed mice. | Vecchio <i>et al.</i> (2004)   |

kidney weight from male rats receiving the 33% 1507 maize grain (Cry1F protein) were lower than those rats fed diets containing non-GM maize grains. However, the author did not discuss these results (MacKenzie *et al.*, 2007).

Multigenerational feeding studies. Brake designed shortterm mouse study in which pregnant female mice were fed Bt or conventional corn diets: the authors then detected the fetal, postnatal, pubertal or adult testicular development of first generational male mice by dual parameter flow cytometry at time points 8, 16, 26, 32, 63 or 87 days after birth. In addition, they also designed multigenerational mouse study to detect the same endpoints in the 4th generational male mice at the same time points as in the short-term mouse study described above. In this study, no apparent differences in percentages of testicular cell populations (haploid, diploid and tetraploid) were observed between the mice fed the Bt corn diet and those fed the conventional diet (Brake et al., 2004a). The same research group also studied the effect of transgenic soybeans using the same methods. The results showed that the transgenic soybean diet had no negative effect on fetal, postnatal, pubertal or adult testicular development (Brake and Evenson, 2004b).

However, in the Vecchio study (Vecchio et al., 2004), the authors fed pregnant Swiss mice and male litters on a standard laboratory chow containing 14% GM soybean. Then, they evaluated Sertoli cells, spermatogonia and spermatocytes by means of electron microscopy at 2, 5 or 8 months of age Their results indicated that immunolabelling for Sm antigen, hnRNPs, SC35 and RNA polymerase II was decreased in 2- and 5-month-old GM-fed mice, and restored to normal at 8 months. In GM-fed mice of all ages considered, the number of perichromatin granules was higher and the nuclear pore density lower. Moreover, the authors found in GM-fed mice enlargements in the smooth endoplasmic reticulum of Sertoli cells. In an opposing critique, Batista (Batista and Oliveira, 2009) thought that the Vecchio study was flawed. For instance, these authors did not provide any information on the source, nutritional composition or type of soybean processing used, nor did they discuss the appropriateness of the control used, such as whether it was a near-isogenic line that was grown in the same field and under the same environmental conditions. One piece of crucial information would be the isoflavone content of the GM soybean  $\nu$ . the control, because the estrogenic effect of isoflavones per se could be responsible for changes in cell nuclear trafficking (Zhu and Conney, 1998). In addition, we thought Vecchio's study lacks some cell biology experiments such as assessing motility of sperm analyses and sperm count. However, a possible role played by GM crops on the development of male sperm needed more discussions.

## Possible mechanisms for GM effects on animals

GM = genetically modified

Although possessing many advantages compared with conventional crops, there are still doubts as to the safety of GM crops with respect to possible long-term adverse effects on

the environment and human health, as DNA and protein representing the novel constituents in GM crops can be degraded by animals. As it is well known, alterations in dietary agents during the pre-mating period will affect oocyte maturity, blastocyst yield, prenatal survival and the number of offspring born alive (Trosko, 2008; Ashworth et al., 2009). Therefore, given the characteristics of GM crops. DNA and proteins are broken down rapidly into small fragments by digestive enzymes: DNA into fragments and nucleotides within the digestive tract; proteins into polypeptides, peptides and amino acids (Faust and Glenn, 2002). However, now this view faces a challenge, and there are numerous debates about whether exogenous DNA fragments or protein can be absorbed by the gastrointestinal tract and then exist in tissues, resulting in adverse effects. However, there is no direct evidence to support a particular mechanism for an effect of GM crops on reproduction in male and female animals. Therefore, we must focus on some possible factors impacting animal reproduction as follows.

Exogenous protein and effects on gastrointestinal tract
To date, many novel proteins expressed in GM crops were
evaluated by a rigorous safety-assessment process before
the crop is commercialized: assessment of the potential for
the protein to be an allergen or toxin or assessment of
potential toxicity and allergenicity of introduced proteins
(Kier and Petrick, 2008). Importantly, Cry1Ab and 5-enolpyruvlshimimate-3-phosphate synthase (CP4 EPSPS) were
mainly resistance proteins expressed in GM crops. The
amount of transgenic protein ingested by animals (mouse,
rat, livestock, etc.) depends on the concentration of the
protein in the feed, the amount of feed intake and the
duration of daily diet feeding (Alexander et al., 2007).

After feeding GM crops, the digestive fate of new proteins introduced into transgenic crops have been evaluated by examining their in-vivo or in-vitro digestibility in simulated gastric and intestinal fluids by testing for the occurrence of these proteins (or their fragments; Bertrand et al., 2005). Evidence of absorption of Cry1Ab proteins was not obtained from assays of calf tissue extracts including liver, spleen, kidney, mesenteric lymph node and muscle (Chowdhury et al., 2003a). The plasma sample from cows fed non-transgenic maize or transgenic maize (collected before or after 1 or 2 months of feeding) showed no effects of the Cry1Ab protein degraded during digestion in the bovine gastrointestinal tract (Lutz et al., 2005; Paul et al., 2008 and 2010). No toxicity of Cry1Ab at non-physiological high concentrations (100 ng/ml) was observed in short- as well as in long-term experiments as to the viability of rumen epithelial cells (Bondzio et al., 2008). The Powell et al. (2010) study indicated that long-term (3 months) exposure to diets formulated with transgenic papaya did not result in biologically important unintended effects for the gastrointestinal tract.

In contrast, when compared with control maize, MON810 maize induced alterations in the percentage of T and B cells and in sub-populations of CD4+, CD8+,  $\gamma\delta$ T and  $\alpha\beta$ T cells in the gut and peripheral sites at weaning and in adult mice

fed for 30 or 90 days, respectively, An increase in serum IL-6, IL-13, IL-12p70 and MIP-1 $\beta$  after MON810 feeding was also found. These results suggested the importance of the gut and peripheral immune response to GM crop ingestion as well as the age of the consumer in the GMO safety evaluation (Finamore *et al.*, 2008). Another group also found that GM crops had a histological influence in the distal intestine and significant effects in intestinal Na+-dependent D-glucose uptake and SGLT1 protein levels in the region of the pyloric caeca of soy-fed *Atlantic* salmons (Bakke-McKellep *et al.*, 2007 and 2008). Therefore, considering the adverse effects on gastrointestinal tract, it is easy to think that those effects would influence absorption of nutrition and subsequently affect animal reproduction.

On the other hand, the percentage of proteins in foods is relatively small. However, current risk assessment practices recommend evaluating the safety of transgenic proteins, confirming that no potentially toxic proteins are engineered into crops (Parrott et al., 2010). Factors such as pH and pepsin-to-substrate ratio greatly influence the digestion of Cry1Ab proteins suggesting that an in-vitro digestibility test that is new and more physiologically relevant should be involved such that the resistance of a protein to digestion can be studied (Guimaraes et al., 2010). For example, a research group found that significant modifications of some nuclear features in hepatocyte nuclei (Malatesta et al., 2002a) and influence zymogen synthesis and processing in mouse pancreatic acinar cells after mice were fed GM soybean (Malatesta et al., 2002b). Moreover, they reported a significant lowering of nucleoplasmic and nucleolar splicing factors as well as accumulation of perichromatin granules in GM-fed mice (Malatesta et al., 2003). To our knowledge, protein synthesis is dependent upon DNA transcription and mRNA translation in cells. Each specific protein is encoded by an individual gene. Therefore, these findings provide a hypothesis that animals fed a GM crop may exhibit exacerbated effects of some unknown proteins when nuclear modifying or nucleolar splicing factors are lowered.

Albo *et al.* (2007) studied GM maize flour compared with wild type (WT) and some unpredictable differences were detected: (i) glucose and ribitol dehydrogenase spot by 2-D protein gel was unique to Bt maize; (ii) endochitinase A spot was unique to WT maize and (iii) triosephosphate isomerase 1 and one spot of globulin-1 S were overexpressed, whereas cytosolic 3-phosphoglycerate kinase and one spot of aldose reductase were downregulated in Bt maize with respect to WT. In short, some proteins expressed in GM crops are not present in significant quantities in conventional (or non-GM) food and might lack a clear history of safe use. It makes crucial sense that each transgenic food is treated as whole food and not as a single protein, and should be tested directly for toxicity in animals (Dona and Arvanitoyannis, 2009).

Exogenous DNA and horizontal gene transfer (HGT) HGT is the non-sexual or parasexual transfer of genetic material between organisms belonging to the same or different species. Of particular concern are putative recipient

microorganisms in the digestive tracts of the human and animals, which are especially relevant to the above discussion of antibiotic-resistance genes (Craig et al., 2008). Some DNAs in food are degraded during cooking and processing, but others remain intact. Consumed DNA was largely hydrolyzed during digestion (Heritage, 2004). Several studies documented the survival of DNA in food/feed throughout the gastrointestinal tract in pigs (cry1Ab and cry9C gene; Chowdhury et al., 2003b and 2003c), piglets (cry1Ab gene; Chowdhury et al., 2003b) and human intestinal microflora with low levels of the epsps gene (Netherwood et al., 2004; van den Eede et al., 2004). Furthermore, a small fragment of the cry1Ab transgene was detected in liver, spleen, kidney and blood but not in muscle of piglets after feeding GM (MON810) for 35 days (Mazza et al., 2005). Meanwhile, several research groups found that transgenic maize in the presence of ampicillin modified the metabolic profile and microbial population structure of bovine rumen fluid (Koch et al., 2006; Wiedemann et al., 2007). Therefore, it clearly suggests that exogenous DNA fragments or proteins may be absorbed by intestinal tracts or intestinal tract microorganism and made some unknown factors changing and then influenced on the reproductive system.

In contrast, other research groups found that no transgenic DNA was detected in tissues of sheep, pig or fallow deer (Sharma et al., 2006; Guertler et al., 2008). Moreover, no transgenic DNA was detected in blood, ruminal fluid or ruminal bacteria of sheep in a 3-year longitudinal study. However, they found higher expression of Ki-67, a marker of proliferative activation of basal ruminal cells, cell nuclear modifications in the pancreatic acinar cells and hepatocytes and functional modifications in the basal cells (Massimo et al., 2007). Interestingly, recombinant or maize-specific DNA was not detectable in tissue samples of pigs. However, plant DNA fragments were detectable in the investigated pig tissues (Reuter and Aulrich, 2003). The study by Singh et al. (2009) indicated that GM crops may not injure digestive tract and result in affecting other systems by the novel protein and tDNA. Furthermore, Onose et al. (2008) detected the sub-chronic toxicity of chemically-induced gastrointestinal impairment in F344 male rat models with dietarily administered Cry1Ab protein from B. thuringiensis, and there were no significant differences in absolute testis and adrenal weight. These results suggested that Bt protein Cry1Ab was degraded and absorbed in the alimentary canal despite gastrointestinal impairment.

Although the frequencies at which viruses infect a GM plant and recombine with a viral transgene is dependent on a wide range of factors (Keese, 2008), the DNA and protein introduced into biotechnology-derived crops are not different from other sources of DNA in the diet (Animal agriculture's future through biotechnology, 2006). However, there is presently no obvious evidence that mammalian or human cells show altered biological properties due to foreign DNA uptake (Royal Society, 2002), we must nevertheless pay close attention to exogenous DNA and HGT in GM plants as they may pose a risk to human and animal health.

#### Anti-nutrient compounds in GM crops

Nutrition and nutritional value of food and feed are major determinants of human and animal well-being. Thus, ensuring the nutritional quality and equivalence of GM food and feed is of critical importance to man and livestock. In addition, the potential for anti-nutrients (trypsin inhibitor, phytic acid and raffinose) adversely affecting health either directly or indirectly is well known (EFSA GMO Panel, 2008). In the study by Ermakova (2005), high levels of mortality (55.6%) and decreases in weight of offspring were reported in female rats fed GM soybean before mating, during mating and throughout pregnancy. These authors believed that the health of newborns might be affected by toxins, allergens or anti-nutrients in the mother's diet. However, Shepherd et al. (2006) point out that there is no construct specifically induced unintended effects and no consistent differences by targeted compositional analysis. In addition, statistically significant differences between WT controls and eight specific types of fructokinase transgenic potatoes appeared to be random and not associated with any specific construct. However, it is possible that GM varieties could change expression levels of those compounds. A purely speculative possibility is that silent pathways for toxicant and anti-nutrient production could be reactivated by insertion or expression of the new genes (Kaeppler, 2000).

#### Conclusions

Reproductive toxicity testing is an important assessment for safety of GM food and feed. Its primary objective is to detect any effects of GM crops or their metabolites on animal reproductive function, especially on the embryo and fetus, embryonic and fetal implantation and loss, fetal weight and development, and reproductive capacity of offspring (EFSA GMO Panel Working Group, 2008). In recent years, many controversial studies have been published with regard to the effects of GM crops on the reproductive system. As shown in this review, it appears that there is no adverse effect of GM crops observed for many species of animal in acute or short-term feeding studies, but serious debate still surrounds long-term and multigenerational feeding studies are clearly necessary to further investigate on this important issue.

As the definition of GM crops, concerns have been raised with regard to any dietary effects on human or animal health at the aspects of protein coded by the transgene, gene flow, HGT, non-target effects, etc (Craig et al., 2008). When considering the serious debate of long-term and multigenerational feed toxicology studies, one can postulate that an increase in the amount of newly expressed protein or could lead to a toxic effect on reproductive function or stacked after long-term and multigenerational feeding if the protein is potentially toxic. At the same time, transgene expression may change when a transgene is placed in a different genetic background through breeding. Unpredictable alterations and changes in the expression levels of hundreds of genes may occur when specific genes are inserted into different species of plants (Schrijver et al., 2007).

However, these possible factors are not yet thoroughly analyzed and may result in the public's suspicion of GM food and feed safety. It is worth noting that the majority of reports suggest that exogenous DNA fragment or protein may be absorbed by the gastrointestinal tract and exerts some effects on it and the microbial population structure within bovine rumen fluid. In addition, sex differences and phenomena of non-linear dose- or time-related effects of pesticides or drugs may reveal hormone-dependent diseases and the first signs of toxicity (Séralini et al., 2009). In a report of the Soil Association of UK and in several other reviews, it has been shown that GM crops may manifest adverse effects on humans and animals (Azeez and Nunan, 2008; Dona and Arvanitoyannis, 2009; Rickard, 2010). At present, between 20% and 30% of the US public has a negative attitude toward products that contain GM crops, even though farmers in the US have raised crops for more than a decade (International Food Information Council (IFIC) Report, 2008). Kwieciński (2009) point out that application of GM technology in agriculture has caused better political and ideological controversy. Moreover, the Chinese government is expected to begin a \$3.5 billion research and development initiative on GM plants (Stone, 2008). In addition, others contend that not pushing ahead with GM varieties could be more detrimental than any theoretical hazard. The relative safety of the insect- and herbicide-resistant crops that dominate our food today says little about the safety of more complex traits the industry has promised in the future (Mellon, 2010). In short, the controversy about the health safety of GM foods is complex. Good science and its communication are required in order to find solutions (Magana-Gomez and delaBarca, 2009). If combining and analyzing recent findings on reproduction of male and female animals fed daily diets containing GM crops show significance; this may indicate unintended effects, which then certainly would require further scientific investigation to answer safety concerns.

#### Acknowledgement

This study was supported by National Nature Science Foundation of China (No. 30771553) and Basic Research Foundation for Science and Technology of Nanjing Agricultural University, China. We express our gratitude to Dr Reinhold J. Hutz of the Department of Biological Sciences, University of Wisconsin-Milwaukee, Wisconsin 53201, USA, for reading the original manuscript and offering valuable suggestions.

# References

Albo AG, Mila S, Digilio G, Motto G, Aime M and Corpillo S 2007. Proteomic analysis of a genetically modified maize flour carrying Cry1Ab gene and comparison to the corresponding wild-type. Maydica 52, 443–455.

Alexander TW, Reuter T, Aulrich K, Sharma R, Okine EK, Dixon WT and Mcallister T 2007. A review of the detection and fate of novel plant molecules derived from biotechnology in livestock production. Animal Feed Science and Technology 133, 31–62.

Appenzeller LM, Munley SM, Hoban D, Sykes GP, Malley LA and Delaney B 2008. Subchronic feeding study of herbicide-tolerant soybean DP-356043-5 in Sprague-Dawley rats. Food and Chemical Toxicology 46, 2201–2213.

Appenzeller LM, Munley SM, Hoban D, Sykes GP, Malley LA and Delaney B 2009a. Subchronic feeding study of grain from herbicide-tolerant maize DP-098140-6 in Sprague-Dawley rats. Food and Chemical Toxicology 47, 2269–2280.

Appenzeller LM, Malley L, Mackenzie SA, Hoban D and Delaney B 2009b. Subchronic feeding study with genetically modified stacked trait lepidopteran and coleopteran resistant (DAS-01507-1xDAS-59122-7) maize grain in Sprague-Dawley rats. Food and Chemical Toxicology 47, 1512–1520.

Ashworth CJ, Toma LM and Hunter MG 2009. Nutritional effects on oocyte and embryo development in mammals: implications for reproductive efficiency and environmental sustainability. Philosophical Transactions of the Royal Society B: Biological Sciences 364, 3351–3361.

Azeez G and Nunan C 2008. GM crops – the Health Effects, The Soil Association of UK. Bristol, UK. Retrieved April 10, 2010, from http://www.soilassociation.org/LinkClick.aspx?fileticket=SqDvBO1pyEU%3D&tabid=390

Bakke-McKellep AM, Koppang EO, Gunnes G, Sanden M, Hemre GI, Landsverk T and Krogdahl A 2007. Histological, digestive, metabolic, hormonal and some immune factor responses in Atlantic salmon, Salmo salar L., fed genetically modified soybeans. Journal of Fish Diseases 30, 65–79.

Bakke-McKellep AM, Sanden M, Danieli A, Acierno R, Hemre GI, Maffia M and Krogdahl A 2008. Atlantic salmon (Salmo salar L.) parr fed genetically modified soybeans and maize: Histological, digestive, metabolic, and immunological investigations. Research in Veterinary Science 84, 395–408.

Batista R and Oliveira MM 2009. Facts and fiction of genetically engineered food. Trends in Biotechnology 27, 277–286.

Bertrand JA, Sudduth TQ, Condon A, Jenkins TC and Calhoun MC 2005. Nutrient content of whole cottonseed. Journal of Dairy Science 88, 1470–1477.

Betz FS, Hammond BG and Fuchs RL 2000. Safety and advantages of Bacillus thuringiensis-protected plants to control insect pests. Regulatory Toxicology and Pharmacology 32, 156–173.

Bondzio A, Stumpff F, Schon J, Martens H and Einspanier R 2008. Impact of Bacillus thuringiensis toxin Cry1Ab on rumen epithelial cells (REC) – a new in vitro model for safety assessment of recombinant food compounds. Food and Chemical Toxicology 46, 1976–1984.

Brake DG, Thaler R and Evenson DP 2004a. Evaluation of Bt (Bacillus thuringiensis) corn on mouse testicular development by dual parameter flow cytometry. Journal of Agricultural and Food Chemistry 52, 2097–2102.

Brake DG and Evenson DP 2004b. A generational study of glyphosate-tolerant soybeans on mouse fetal, postnatal, pubertal and adult testicular development. Food and Chemical Toxicology 42, 29–36.

Chowdhury EH, Shimada N, Murata H, Mikami O, Sultana P, Miyazaki S, Yoshioka M, Yamanaka N, Hirai N and Nakajima Y 2003a. Detection of Cry1Ab protein in gastrointestinal contents but not visceral organs of genetically modified Bt11-fed calves. Veterinary and Human Toxicology 45, 72–75.

Chowdhury EH, Kuribara H, Hino A, Sultana P, Mikami O, Shimada N, Guruge KS, Saito M and Nakajima Y 2003b. Detection of corn intrinsic and recombinant DNA fragments and Cry1Ab protein in the gastrointestinal contents of pigs fed genetically modified corn Bt11. Journal of Animal Science 81, 2546–2551.

Chowdhury EH, Mikami O, Nakajima Y, Hino A, Kuribara H, Suga K, Hanazumi M and Yomemochi C 2003c. Detection of genetically modified maize DNA fragments in the intestinal contents of pigs fed StarLink CBH351. Veterinary and Human Toxicology 45, 95–96.

Cisterna B, Flach F, Vecchio L, Barabino SM, Battistelli S, Martin TE, Malatesta M and Biggiogera M 2008. Can a genetically-modified organism-containing diet influence embryo development? A preliminary study on pre-implantation mouse embryos. European Journal of Histochemistry 52, 263–267.

Codex Alimentarius Commission (CAC) 2003a. Principles For The Risk Analysis of Foods Derived from Modern Biotechnology CAC/GL 44.

Codex Alimentarius Commission (CAC) 2003b. Guideline for the conduct of food safety assessment of foods derived recombinant-DNA plants CAC/GL 45.

Craig W, Tepfer M, Degrassi G and Ripandelli D 2008. An overview of general features of risk assessments of genetically modified crops. Euphytica 164, 853—880

Delaney B, Zhang J, Carlson G, Schmidt J, Stagg B, Comstock B, Babb A, Finlay C, Cressman RF, Ladics G, Cogburn A, Siehl D, Bardina L, Sampson H and Han Y 2008a. A gene-shuffled glyphosate acetyltransferase protein from Bacillus licheniformis (GAT4601) shows no evidence of allergenicity or toxicity. Toxicological Sciences 102, 425–432.

## Zhang and Shi

Delaney B, Appenzeller LM, Munley SM, Hoban D, Sykes GP, Malley LA and Sanders C 2008b. Subchronic feeding study of high oleic acid soybeans (Event DP-305423-1) in Sprague-Dawley rats. Food and Chemical Toxicology 46, 3808–3817.

Dhlamini Z 2009. Agricultural Biotechnology. In Biosafety of genetically modified organisms: basic concepts, methods and issues (ed. MKA Chowdhury, MI Hoque and A Sonnino), pp. 1–50. FAO, Rome, Italy.

Dona A and Arvanitoyannis IS 2009. Health risks of genetically modified foods. Critical Reviews in Food Science and Nutrition 49, 164–175.

European Food Safety Authority – Genetically Modified Organism (EFSA GMO) Panel 2008. Safety and nutritional assessment of GM plants and derived food and feed: the role of animal feeding trials. Food and Chemical Toxicology 46 (suppl.1), S2–S70.

Ermakova IV 2005. Influence of Genetically Modified Soya on the Birth Weight and Survival of Rat Pups: Preliminary Study. Retrieved May 10, 2010, from http://www.mindfully.org/GE/2005/Modified-Soya-Rats10oct05.htm

Ermakova IV 2007. GM soybeans — revisiting a controversial format. Nature Biotechnology 25, 1351–1354.

Evenson DP, Darzynkiewicz Z and Melamed MR 1980. Comparison of human and mouse sperm chromatin structure by flow cytometry. Chromosoma 78, 225–238.

Food and Agriculture Organization of the United Nations and World Health Organization (FAO/WHO) 2000. Safety aspects of genetically modified foods of plant origin. Report of a Joint FAO/WHO Expert Consultation on Foods Derived from Biotechnology, WHO, Geneva, Switzerland, 37pp.

Faust MA and Glenn BP 2002. Animal feeds from crops derived through biotechnology: farm animal performance and safety. In Biotechnology and Safety Assessment, 3rd edition (ed. JA Taylor and RL Fuchs), pp. 143–189. Academic Press, San Diego, CA, USA.

Finamore A, Roselli M, Britti S, Monastra G, Ambra R, Turrini A and Mengheri E 2008. Intestinal and peripheral immune response to MON810 maize ingestion in weaning and old mice. Journal of Agricultural and Food Chemistry 56, 11533—11539.

Francis EZ and Kimmel GL 1988. Proceeding of the workshop on One-vs -Two generation reproductive effects studies. International Journal of Toxicology 7, 911–925.

Guertler P, Lutz B, Kuehn R, Meyer HHD, Einspanier R, Killerman B and Albrecht C 2008. Fate of recombinant DNA and Cry1Ab protein after ingestion and dispersal of genetically modified maizein comparison to rapeseed by fallow deer (Dama dama). European Journal of Wildlife Research 54, 36–43.

Guimaraes V, Drumare MF, Lereclus D, Gohar M, Lamourette P, Nevers MC, Vaisanen-Tunkelrott ML, Bernard H, Guillon B, Creminon C, Wal JM and Adel-Patient K 2010. In vitro digestion of Cry1Ab proteins and analysis of the impact on their immunoreactivity. Journal of Agricultural and Food Chemistry 58, 3222–3231

Hammond BG, Dudek R, Lemen JK and Nemeth MA 2006. Results of a 90-day safety assurance study with rats fed grain from corn borer-protected corn. Food and Chemical Toxicology 44, 1092–1099.

He XY, Huang KL, Li X, Qin W, Delaney B and Luo YB 2008. Comparison of grain from corn rootworm resistant transgenic DAS-59122-7 maize with non-transgenic maize grain in a 90-day feeding study in Sprague-Dawley rats. Food and Chemical Toxicology 46, 1994–2002.

He XY, Tang MZ, Luo YB, Li X, Cao SS, Yu JJ, Delaney B and Huang KL 2009. A 90-day toxicology study of transgenic lysine-rich maize grain (Y642) in Sprague-Dawley rats. Food and Chemical Toxicology 47, 425–432.

Healy C, Hammond B and Kirkpatrick J 2008. Results of a 13-week safety assurance study with rats fed grain from corn rootworm-protected, glyphosate-tolerant MON 88017 corn. Food and Chemical Toxicology 46, 2517–2524.

Heritage J 2004. The fate of transgenes in the human gut. Nature Biotechnology 22, 170–172.

Hirschi KD 2009. Nutrient biofortification of food crops. Annual Review of Nutrition 29, 401–421.

International Food Information Council (IFIC) 2008. Food Biotechnology: A Study of US consumer Attitudinal Trends Survey. IFIC. Retrieved April 10, 2010, from http://www.ificpubs.org/servlet/Detail?no=46

Jacobs CM, Utterback PL, Parsons CM, Rice D, Smith B, Hinds M, Liebergesell M and Sauber T 2008. Performance of laying hens fed diets containing DAS-59122-7 maize grain compared with diets containing nontransgenic maize grain. Poultry Science 87, 475–479.

James C 2007. Global Status of Commercialized Biotech/GM Crops: 2007. ISAAA Brief No. 37. International Service for the Acquisition of Agri-biotech Applications (ISAA). Ithaca, NY, USA, 16pp.

Juberg DR, Herman RA, Thomas J, Brooks KJ and Delaney B 2009. Acute and repeated dose (28 day) mouse oral toxicology studies with Cry34Ab1 and Cry35Ab1 Bt proteins used in coleopteran resistant DAS-59122-7 corn. Regulatory Toxicology and Pharmacology 54, 154–163.

Kaeppler HF 2000. Food safety assessment of genetically modified crops. Agronomy Journal 92, 793–797.

Keese P 2008. Risks from GMOs due to horizontal gene transfer. Environmental Biosafety Research 7, 123–149.

Kier LD and Petrick JS 2008. Safety assessment considerations for food and feed derived from plants with genetic modifications that modulate endogenous gene expression and pathways. Food and Chemical Toxicology 46, 2591–2605.

Kilic A and Akay MT 2008. A three generation study with genetically modified Bt corn in rats: biochemical and histopathological investigation. Food and Chemical Toxicology 46, 1164–1170.

Knudsen I and Poulsen M 2007. Comparative safety testing of genetically modified foods in a 90-day rat feeding study design allowing the distinction between primary and secondary effects of the new genetic event. Regulatory Toxicology and Pharmacology 49, 53–62.

Koch M, Strobel E, Tebbe CC, Heritage J, Breves G and Huber K 2006. Transgenic maize in the presence of ampicillin modifies the metabolic profile and microbial population structure of bovine rumen fluid in vitro. British Journal of Nutrition 96, 820–829.

Krishnan H 2005. Engineering soybean for enhanced sulfur amino acid content. Crop Science 45, 454–461.

Kwieciński J 2009. Genetically modified abominations? Widespread opposition to GMOs might have deep-seated cultural causes. EMBO Reports 10, 1187–1190.

Lutz B, Wiedemann S, Einspanier R, Mayer J and Albrecht C 2005. Degradation of Cry1Ab protein from genetically modified maize in the bovine gastrointestinal tract. Journal of Agricultural and Food Chemistry 53, 1453–1456.

MacKenzie SA, Lamb I, Schmidt J, Deege L, Morrisey MJ, Harper M, Layton RJ, Prochaska LM, Sanders C, Locke M, Mattsson JL, Fuentes A and Delaney B 2007. Thirteen week feeding study with transgenic maize grain containing event DAS-01507-1 in Sprague-Dawley rats. Food and Chemical Toxicology 45, 551–562.

Magana-Gomez JA and delaBarca AM 2009. Risk assessment of genetically modified crops for nutrition and health. Nutrition Reviews 67, 1–16.

Malatesta M, Biggiogera M, Manuali E, Rocchi MB, Baldelli B and Gazzanelli G 2003. Fine structural analyses of pancreatic acinar cell nuclei from mice fed on genetically modified soybean. European Journal of Histochemistry 47, 385–388.

Malatesta M, Caporaloni C, Gavaudan S, Rocchi MB, Serafini S, Tiberi C and Gazzanelli G 2002a. Ultrastructural morphometrical and immunocytochemical analyses of hepatocyte nuclei from mice fed on genetically modified soybean. Cell Structure and Function 27, 173–180.

Malatesta M, Caporaloni C, Rossi L, Battistelli S, Rocchi MB, Tonucci F and Gazzanelli G 2002b. Ultrastructural analysis of pancreatic acinar cells from mice fed on genetically modified soybean. Journal of Anatomy 201, 409–415.

Malley LA, Everds NE, Reynolds J, Mann PC, Lamb I, Rood T, Schmidt J, Layton RJ, Prochaska LM, Hinds M, Locke M, Chui CF, Claussen F, Mattsson JL and Delaney B 2007. Subchronic feeding study of DAS-59122-7 maize grain in Sprague-Dawley rats. Food and Chemical Toxicology 45, 1277–1292.

Marshall A 2007. GM soybeans and health safety – a controversy reexamined. Nature Biotechnology 25, 981–987.

Mazza R, Soave M, Morlacchini M, Piva G and Marocco A 2005. Assessing the transfer of genetically modified DNA from feed to animal tissues. Transgenic Research 14, 775–784.

Mellon M 2010. Food security: rigorous regulation required. Science 328, 171–172.

Momma K, Hashimoto W, Yoon HJ, Ozawa S, Fukuda Y, Kawai S, Takaiwa F, Utsumi S and Murata K 2000. Safety assessment of rice genetically modified with soybean glycinin by feeding studies on rats. Bioscience, Biotechnology, and Biochemistry 64, 1881–1886.

Monsanto Company 2008. Monsanto Company Response to Austrian Report on Mouse Chronic and Reproduction Studies with NK603 × MON810 Maize. Retrieved April 28, 2010, from http://www.monsanto.com.ar/nuestros\_productos/informacion\_tecnica\_seguridad/documentos/response\_to\_austrian\_gm\_study\_20nov08.pdf

Netherwood T, Martin-Orue SM, O'Donnell AG, Gockling S, Graham J, Mathers JC and Gilbert HJ 2004. Assessing the survival of transgenic plant DNA in the human gastrointestinal tract. Nature Biotechnology 22, 204–209.

Organization for Economic Co-operation and development (OECD) 2007. Consensus document on safety information on transgenic plants expressing Bacillus Thuringiensis – derived insect control proteins. Series on Harmonization of Regulatory Oversight in Biotechnology, No. 42. OECD, Paris, France.

Onose J, Imai T, Hasumura M, Ueda M, Ozeki Y and Hirose M 2008. Evaluation of subchronic toxicity of dietary administered Cry1Ab protein from Bacillus thuringiensis var. Kurustaki HD-1 in F344 male rats with chemically induced gastrointestinal impairment. Food and Chemical Toxicology 46, 2184–2189.

Park S, Kang TS, Kim CK, Han JS, Kim S and Smith RH 2005. Genetic manipulation for enhancing calcium content in potato tuber. Journal of Agricultural and Food Chemistry 53, 5598–5603.

Parrott W, Chassy B, Ligon J, Meyer L, Petrick J, Zhou J, Herman R, Delaney B and Levine M 2010. Application of food and feed safety assessment principles to evaluate transgenic approaches to gene modulation in crops. Food and Chemical Toxicology 48, 1773–1790.

Paul V, Steinke K and Meyer HH 2008. Development and validation of a sensitive enzyme immunoassay for surveillance of Cry1Ab toxin in bovine blood plasma of cows fed Bt-maize (MON810). Analytica Chimica Acta 607, 106–113.

Paul V, Guertler P, Wiedemann S and Meyer HH 2010. Degradation of Cry1Ab protein from genetically modified maize (MON810) in relation to total dietary feed proteins in dairy cow digestion. Transgenic Research 19, 683–689.

Powell M, Wheatley AO, Omoruyi F, Asemota HN, Williams NP and Tennant PF 2010. Comparative effects of dietary administered transgenic and conventional papaya on selected intestinal parameters in rat models. Transgenic Research 19, 511–518.

Rasmussen MA, Sarra AC, Wilhelms K and Scanes CG 2007. Effects of Bt (Bacillus thuringiensis) corn on reproductive performance in adult laying hens. International Journal of Poultry Science 6, 169–171.

Reuter T and Aulrich K 2003. Investigations on genetically modified maize (Bt-maize) in pig nutrition: fate of feed-ingested foreign DNA in pig bodies. European Food Research and Technology 216, 185–192.

Rhee GS, Cho DH, Won YH, Seok JH, Kim SS, Kwack SJ, Lee RD, Chae SY, Kim JW, Lee BM, Park KL and Choi KS 2005. Multigeneration reproductive and developmental toxicity study of bar gene inserted into genetically modified potato on rats. Journal of Toxicology and Environmental Health. Part A 68, 2263–2276.

Rickard C 2010. Response to 'Health risks of genetically modified foods' from Dona and Arvanitoyannis (2009) in Critical Reviews in Food Science and Nutrition (49:164-175). Critical Reviews in Food Science and Nutrition 50, 86–91.

Royal Society 2002. Genetically modified plants for food use and human health-an update. Policy document 4/02. Royal Society, London, UK.

Sanvido O, Romeis J and Bigler F 2007. Ecological impacts of genetically modified crops: ten years of field research and commercial cultivation. Advances in Biochemical Engineering/Biotechnology 107, 235–278.

Schrijver AD, Devos Y, Van den Bulcke M, Cadot P, De Loose M, Reheul D and Sneyers M 2007. Risk assessment of GM stacked events obtained from crosses between GM events. Trends in Food Science & Technology 18, 101–109.

Schroder M, Poulsen M, Wilcks A, Kroghsbo S, Miller A, Frenzel T, Danier J, Rychlik M, Emami K, Gatehouse A, Shu QY, Engel KH, Altosaar I and

Knudsen I 2007. A 90-day safety study of genetically modified rice expressing Cry1Ab protein (Bacillus thuringiensis toxin) in Wistar rats. Food and Chemical Toxicology 45, 339–349.

Séralini GE, de Vendomois JS, Cellier D, Sultan C, Buiatti M, Gallagher L, Antoniou M and Dronamraju KR 2009. How subchronic and chronic health effects can be neglected for GMOs, pesticides or chemicals. International Journal of Biological Sciences 5, 438–443.

Sesikeran B and Vasanthi S 2008. Constantly evolving safety assessment protocols for GM foods. Asia Pacific Journal of Clinical Nutrition 17, 241–244.

Sharma R, Damgaard D, Alexander TW, Dugan ME, Aalhus JL, Stanford K and McAllister TA 2006. Detection of transgenic and endogenous plant DNA in digesta and tissues of sheep and pigs fed Roundup Ready canola meal. Journal of Agricultural and Food Chemistry 54, 1699–1709.

Shepherd LV, McNicol JW, Razzo R, Taylor MA and Davies HV 2006. Assessing the potential for unintended effects in genetically modified potatoes perturbed in metabolic and developmental processes. Targeted analysis of key nutrients and anti-nutrients. Transgenic Research 15, 409–425.

Singh AK, Praveen S, Singh BP, Varma A and Arora N 2009. Safety assessment of leaf curl virus resistant tomato developed using viral derived sequences. Transgenic Research 18, 877–887.

Stone R 2008. China plans \$3.5 billion GM crops initiative. Science 321, 1279. Trabalza-Marinucci M, Brandi G, Rondini C, Avellini L, Giammarini C, Costarelli S, Acuti G, Orlandi C, Filippini G, Chiaradia E, Malatesta M, Crotti S, Antoninia C, Amagliani G, Manuali E, Mastrogiacomo AR, Moscati L, Naceur Haouet M, Gaiti A and Magnani M 2008. A three-year longitudinal study on the effects of a diet containing genetically modified Bt176 maize on the health status and performance of sheep. Livestock Science 113, 178–191.

Trosko JE 2008. Role of diet and nutrition on the alteration of the quality and quantity of stem cells in human aging and the diseases of aging. Current Pharmaceutical Design 14, 2707–2718.

van den Eede G, Aarts H, Buhk HJ, Corthier G, Flint HJ, Hammes W, Jacobsen B, Midtvedt T, van der Vossen J, von Wright A, Wackernagel W and Wilcks A 2004. The relevance of gene transfer to the safety of food and feed derived from genetically modified (GM) plants. Food and Chemical Toxicology 42, 1127–1156.

Vecchio L, Cisterna B, Malatesta M, Martin TE and Biggiogera M 2004. Ultrastructural analysis of testes from mice fed on genetically modified soybean. European Journal of Histochemistry 48, 448–454.

Velimirov A, Binter C and Zentek J 2008. Biological Effects of Transgenic Maize NK603 × MON810 Fed in Long Term Reproduction Studies in Mice. Retrieved April 28, 2010, from http://www.biosicherheit.de/pdf/aktuell/zentek\_studie\_ 2008.pdf

WHO 2002. Foods Derived from Modern Technology: 20 Questions on Genetically Modified Foods. Retrieved October 10, 2010, from http://www.who.int/fsf/GMfood/

Wiedemann S, Gurtler P and Albrecht C 2007. Effect of feeding cows genetically modified maize on the bacterial community in the bovine rumen. Applied and Environmental Microbiology 73, 8012–8017.

Zhu BT and Conney AH 1998. Functional role of estrogen metabolism in target cells: review and perspectives. Carcinogenesis 19, 1–27.